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## New routes towards reutericyclin analogues†

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A range of *N*-acylpyrrolo[3,4-c]isoxazoles and derived *N*-acyltetramides has been prepared *via* a nitrile oxide dipolar cycloaddition approach, as analogues of the acyltetramic acid metabolite reutericyclin, of interest for its antibiotic potential against Gram-positive bacteria including hospital-acquired infections of resistant *Clostridium difficile*.

In order to combat the growing resistance to generally administered antibiotics, such as penicillin and methicillin, the research community is endeavouring to find new compounds that actively inhibit problematic resistant bacteria.<sup>1</sup> This effort has identified a number of potential candidates, one example of which is reutericyclin (1), isolated in 2000 by Jung et al. from Lactobacillius reuteri LTH2584.<sup>2</sup> Reutericyclin belongs to the 3-acyltetramic acid group of natural products (2), characterised by a pyrrolidine-2,4-dione unit carrying an acyl group at C-3.<sup>3</sup> Molecules containing this motif exhibit a range of bio-activities including antibiotic, antitumor, antiviral, antiulcerative, fungicidal and cytotoxic properties.4 Interest in the antibiotic activity of tetramic acids has recently been stimulated by their key relationship to the inducers of bacterial quorum sensing.5 Reutericyclin and derivatives display varying inhibition in Gram-positive bacteria.<sup>2,6</sup> The most interesting of these results is the inhibition of growth of resistant bacterium Clostridium difficile, a leading cause of antibioticassociated diarrhoea in hospitalized patients which can lead to mortalities in persons with a compromised immune system.<sup>7</sup>



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We have, over many years, explored the synthesis of the acyltetramic acid moiety<sup>8</sup> and other cyclic tricarbonyl systems,<sup>9</sup> most recently using pyrroloisoxazoles as masked acyltetramic acids and as core building blocks for peripheral elaboration.<sup>10,11</sup> Our 2nd generation strategy (Scheme 1)<sup>10</sup> uses pyrrolo[3,4-c]isoxazoles 3 (cf. pyrrolo[3,4-d]isoxazoles in our 1st generation approach<sup>11</sup>) formed by cycloaddition of nitrile oxides 4, available in three steps from  $\alpha$ -amino esters, with enamino ester dipolarophiles. We report here significant practical improvements in this strategy (principally in lactam closure) and its application to access novel bicyclic reutericyclin analogues. Reutericyclin has R-configuration at C-5, and is presumably biosynthesised from *R*-leucine,<sup>3,12</sup> however we have conducted our studies in the more readily available, less costly S-series: the chemistry should, of course, be equally applicable to the enantiomeric series.13

The commercially available methyl esters of *S*-valine, *S*-leucine and *S*-phenylglycine were efficiently N-protected (Boc<sub>2</sub>O, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0–20 °C; 99, 97 and 99%, respectively). The protected amino esters (**5a–c**, respectively) were selectively reduced to the corresponding aldehydes using DIBAL-H at -78 °C (91, 93 and 87%), which were converted directly to the oximes **6a–c** (H<sub>2</sub>NOH·HCl, NaOAc, aq. EtOH, 2–8 °C; 86, 79 and 88%) to inhibit potential racemisation (Scheme 2). Treatment with NCS (CHCl<sub>3</sub> reflux) afforded C-chloro-oximes **7**, either used directly (**7a,c**) or isolated (**7b**; 75%); an extended reaction time for chlorination (18 h) led to better results when using the hydroximoyl chlorides **7** (*vide infra*) than in our previous reports.

ChemComm



Scheme 2 Synthesis of pyrroloisoxazoles **10**. (a)  $R^1 = CHMe_2$ ; (b)  $R^1 = CH_2CHMe_2$ ; (c)  $R^1 = Ph$ . Reagents: (i) DIBAL-H, toluene, -78 °C; (ii) H<sub>2</sub>NOH·HCl, NaOAc, aq. EtOH, 2–8 °C; (iii) NCS, CHCl<sub>3</sub> reflux, 18 h; (iv) Et<sub>3</sub>N, CHCl<sub>3</sub> reflux; (v) TFA, 20 °C; 2 M aq. HCl; (vi) T3P, EtOAc, 0–20 °C, 17 h (with **9a,b**) or PS-CDI, Et<sub>3</sub>N, DMF-CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 17 h (with **9c**).

The key dipolar cycloaddition step was performed by addition of Et<sub>3</sub>N to the chloro-oximes in the presence of the pyrrolidine enamine of *tert*-butyl acetoacetate and pyrrolidine (separately prepared; toluene reflux, Dean–Stark conditions; 99%) to form the nitrile oxide *in situ* and complete the cycloaddition (CHCl<sub>3</sub> reflux) to afford isoxazoles **8a,c** (49 and 56% from **6a,c**) and **8b** (60% from **7b**). Simultaneous deprotection of the *N*-Boc amine and *tert*-butyl ester cleavage was achieved by acid treatment (TFA, 20 °C; then 2 M aq. HCl to give hydrochloride salts of better stability for handling and on storage) to leave amino acid salts **9a–c** (99, 70 and 68%).

The final stage in assembly of the pyrroloisoxazoles, closure of the pyrrolo ring, was initially completed by our previously reported method using N-[3-(dimethylamino)propyl]-N'-ethylcarbodiimide hydrochloride (EDCI) (N-hydroxysuccinimide, Et<sub>3</sub>N, DMF, 0-20 °C) which required column chromatography and yielded the pyrroloisoxazoles 10a,b in unreliable yields (ranging 8-60%).<sup>10b</sup> Other peptide coupling reagents were investigated: whilst PyBroP failed, HATU did produce 10a in 40% yield.<sup>14</sup> The variable performance could be improved by using a polystyrene-supported carbodiimide (supplied as PS-CDI; Argonaut Technologies<sup>™</sup>) (Et<sub>3</sub>N, DMF-CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 17 h) that reliably afforded 10a (66%), still however requiring column chromatographic purification and a costly alternative. Finally the simplest and most reliable lactam closure was achieved using the recently commercialised cyclic propylphosphonic anhydride (supplied as T3P; Archemica<sup>™</sup>).<sup>15</sup> Thus a base (Et<sub>3</sub>N) was added to the salts 10a,b in EtOAc followed by T3P (0-20 °C, 17 h). The pyrroloisoxazoles **10a,b** were isolated pure without needing chromatography in good yields (59 and 68%). The phenylglycine-derived 10c was unsuccessful with T3P but could be prepared reliably by the PS-CDI protocol (50%). We have thus revealed two improved protocols for lactam closure to pyrroloisoxazoles 10: using T3P or PS-CDI.<sup>16</sup>

The last stage in the synthesis of the masked reutericyclin analogues was to perform an *N*-acylation. As base we selected to use BuLi (THF, -78 °C). Carboxylic esters were investigated as acylating agents but without success. However, acyl chlorides proved to be effective acylating agents to produce the *N*-acyl



derivatives **11** (Scheme 3).<sup>13</sup> A variety of acyl chlorides were selected including long and short aliphatic chains, an  $\alpha,\beta$ -unsaturated chain, a hindered branched moiety and an aromatic substituent, and all afforded *N*-acyl products **11** in yields of 33–97% (Table 1).‡ Longer chain, aromatic or more hindered acyl chlorides required a slightly longer time for complete reaction than the shorter, unhindered, examples; a standard reaction time of 3 h was eventually employed. The constitution of the *N*-(but-2-enyl)-6-(2-methylpropyl)pyrroloisoxazole **11c** was confirmed by an X-ray crystal structure (Fig. 1).§

This completed the synthesis of the reutericyclin analogues. Next we determined to create some tetramide analogues, by N–O bond cleavage of the pyrroloisoxazole nucleus. This was achieved for bicycles **11a,f** by hydrogenolysis (1 atm H<sub>2</sub>, Pd–C) to afford the enaminoketones (tetramides) **12a,b** (49 and 52%) (Scheme 4). To demonstrate an alternative protocol, and because hydrogenation would be likely to reduce an unsaturated *N*-acyl group,<sup>10b</sup> N–O cleavage of **11l** was accomplished by Mo(CO)<sub>6</sub> (aq. MeCN; then 2 M aq. HCl) to give enaminoketone **12c** (60%).<sup>17</sup>





Fig. 1 X-ray crystal structure of *N*-acyl pyrroloisoxazole **11c**.



Scheme 4 Formation of N-acyltetramides 12 from pyrroloisoxazoles 11.

either returned unchanged enaminoketone (*e.g.*  $H_2O$  at 20 °C or 2 M aq. HCl at reflux; NaNO<sub>2</sub>, 3 M aq.  $H_2SO_4$ ) or led to *N*-deacylation (aq. NaOH, 2 M at reflux or 0.1 M at 20 °C).

In conclusion, we have developed a synthetic route, based on a nitrone 1,3-dipolar cycloaddition, from amino acids to N-acylpyrrolo[3,4-c]isoxazoles 11 as reutericyclin analogues, and presented a diverse selection of 12 novel compounds. Furthermore, we have demonstrated the conversion of these heterobicycles into N-acyltetramides 12. All of these new compounds are currently undergoing biological evaluation.

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## Notes and references

<sup>‡</sup> Typical procedure for *N*-acylpyrrolo[3,4-*c*]isoxazole formation: (S)-5-butyryl-6-isopropyl-3-methyl-5,6-dihydro-4H-pyrrolo[3,4-c]isoxazol-4-one 11a. (S)-6-(1-Methylethyl)-3-methyl-5,6-dihydro-4H-pyrrolo[3,4-c]isoxazol-4-one 10a (50.0 mg, 0.277 mmol) was suspended in dry THF (20 mL) stirred at -78 °C under a nitrogen atmosphere. *n*-Butyl-lithium (0.201 mL, 1.41 M in hexanes, 0.283 mmol) was added and the reaction stirred for 15 min at this temperature, during which time the solution turned yellow. Butanoyl chloride (29.6 mg, 27.0 µL, 0.283 mmol) was then added in two portions over 10 min and the mixture stirred at -78 °C for a further 3 h before quenching by addition of satd. NH<sub>4</sub>Cl solution. The mixture was tested for pH to ensure neutrality had been achieved and then separated between water (20 mL) and EtOAc (25 mL). The organic layer was dried over MgSO4, filtered and concentrated under reduced pressure to produce the title compound 11a as a yellow oil (62 mg, 89%);  $[\alpha]_{D}^{20}$  +36.0 (c 5.00 × 10<sup>-3</sup>, CHCl<sub>3</sub>);  $\nu_{max}$  (CHCl<sub>3</sub>)/cm<sup>-1</sup> 3025, 1725 (C=O), 1689 (C=O), 1650, 1389, 1250, 1131;  $\delta_{H}$  (400 MHz; CDCl<sub>3</sub>) 0.50 (3H, d, J = 6.8, CH(CH<sub>3</sub>)<sub>2</sub>) 0.92 (3H, t, J = 7.2, CH<sub>2</sub>CH<sub>3</sub>), 1.17  $(3H, d, J = 6.8, CH(CH_3)_2), 1.55-1.61 (2H, m, CH_2CH_3), 2.61 (3H, s, 3-CH_3),$ 2.68-2.72 (1H, m, CH(CH<sub>3</sub>)<sub>2</sub>), 2.81, 2.92 (each 1H, dt, J = 7.6, 14.8, CH<sub>2</sub>CO), 5.13 (1H, d, J = 4, CHN);  $\delta_{\rm C}$  (100 MHz; CDCl<sub>3</sub>) 11.8, 12.8, 13.1 (CH<sub>3</sub>), 17.0 (CH<sub>2</sub>CH<sub>3</sub>), 17.9 (CH<sub>3</sub>), 27.0 (CH(CH<sub>3</sub>)<sub>2</sub>), 38.2 (CH<sub>2</sub>CO), 59.9

(CHN), 113.0, 159.5, 165.0 (isoxazole-C), 168.6, 173.2 (CO). HRMS (ESI):  $MNa^+$  273.1211;  $C_{13}H_{18}N_2O_3$  requires  $MNa^+$  273.1210.

§ Crystal data for **11c**:  $C_{13}H_{16}N_2O_3$ , M = 248.28, orthorhombic,  $P2_12_12_1$ , a = 6.9698(12), b = 9.5108(16), c = 19.233(3) Å, V = 1274.9(4) Å<sup>3</sup>, Z = 4,  $\mu$ (Mo-K $\alpha$ ) = 0.71073 Å, 11 350 reflections measured at 150 K on a Bruker APEX 2 CCD diffractometer, 2618 unique data,  $R_{int} = 0.034$ , R[for 2390 data with  $F^2 > 2\sigma(F^2)] = 0.032$ ,  $wR_2$  (all data) = 0.078, 227 parameters. H atoms were freely refined. Absolute structure {x = 0.0(6)} could not be determined reliably. CCDC 959645.

- For example: World Health Organisation, Fact Sheet no. 194, May 2013, http://www.who.int/mediacentre/factsheets/fs194/en/, accessed 09 Aug 2013. U. Theuretzbacher and J. H. Toney, *Curr. Opin. Invest. Drugs*, 2006, 7, 158–166.
- 2 A. Holtzel, M. G. Ganzle, G. J. Nicholson, W. P. Hammes and G. Jung, *Angew. Chem., Int. Ed.*, 2000, **39**, 2766–2768.
- B. J. L. Royles, *Chem. Rev.*, 1995, **95**, 1981–2001; R. Schobert and A. Schlenk, *Bioorg. Med. Chem.*, 2008, **16**, 4203–4221; Y.-C. Jeong and M. G. Moloney, *J. Org. Chem.*, 2011, 1342–1354.
- 4 For leading references: T. Sengoku, Y. Nagae, Y. Ujihara, M. Takahashi and H. Yoda, *J. Org. Chem.*, 2012, 77, 4391–4401; Y.-C. Jeong, M. Anwar, Z. Bikadi, E. Hazai and M. G. Moloney, *Chem. Sci.*, 2013, 4, 1008–1015.
- 5 G. F. Kaufmann, R. Sartorio, S.-H. Lee, C. J. Rogers, M. M. Meijler, J. A. Moss, B. Clapham, A. P. Brogan, T. J. Dickerson and K. D. Janda, *Proc. Natl. Acad. Sci. U. S. A.*, 2005, **102**, 309; C. A. Lowery, J. Park, C. Gloeckner, M. M. Meijler, R. S. Mueller, H. I. Boshoff, R. L. Ulrich, C. E. Barry, D. H. Bartlett, V. V. Kravchenko, G. F. Kaufmann and K. D. Janda, *J. Am. Chem. Soc.*, 2009, **131**, 14473; C. Ueda, K. Tateda, M. Horikawa, S. Kimura, Y. Ishii, K. Nomura, K. Yamada, T. Suematsu, Y. Inoue, M. Ishiguro, S. Miyairi and K. Yamaguchi, *Antimicrob. Agents Chemother.*, 2010, **54**, 683.
- 6 R. Yendapally, J. G. Hurdle, E. I. Carson, R. B. Lee and R. E. Lee, J. Med. Chem., 2008, 51, 1487–1491; M. G. Gänzle and R. F. Vogel, Appl. Environ. Microbiol., 2003, 69, 1305–1307; M. G. Gänzle, Appl. Microbiol. Biotechnol., 2004, 64, 326–332.
- 7 C. Ueda, K. Tateda, M. Horikawa, S. Kimura, Y. Ishii, K. Nomura, K. Yamada, T. Suematsu, Y. Inoue, M. Ishiguro, S. Miyairi and K. Yamaguchi, *Antimicrob. Agents Chemother.*, 2010, 54, 683–688; J. G. Hurdle, A. E. Heathcott, L. Yang, B. Yan and R. E. Lee, *J. Antimicrob. Chemother.*, 2011, 6, 1773–1776.
- 8 For leading references to our early pre-isoxazole work: R. C. F. Jones and M. Tankard, *J. Chem. Soc., Perkin Trans.* 1, 1991, 250–251; R. C. F. Jones, G. Bhalay, J. M. Patience and P. Patel, *J. Chem. Res.*, 1999, 250–251.
- 9 For our related work on the 3-acyl-4-hydroxypyridin-2-one series: R. C. F. Jones, A. K. Choudhury, J. N. Iley, M. E. Light, G. Loizou and T. A. Pillainayagam, *Beilstein J. Org. Chem.*, 2012, **8**, 308–312, and refs. therein.
- For our 2nd generation approach: (*a*) R. C. F. Jones, C. E. Dawson and M. J. O'Mahony, *Synlett*, 1999, 873–876; (*b*) R. C. F. Jones and T. A. Pillainayagam, *Synlett*, 2004, 2815–2817; (*c*) R. C. F. Jones, C. C. M. Law and M. R. J. Elsegood, *ARKIVOC*, 2013, (iii), 81–97.
- 11 For our 1st generation approach: R. C. F. Jones, G. Bhalay, P. A. Carter, K. A. M. Duller and S. H. Dunn, *J. Chem. Soc., Perkin Trans.* 1, 1999, 765–776, and refs. therein.
- 12 Cf. L. M. Halo, J. M. Marshall, A. A. Yakasai, Z. Song, C. P. Butts, M. P. Crump, M. Heneghan, A. M. Bailey, T. J. Simpson, C. M. Lazarus and R. J. Cox, *ChemBioChem*, 2008, 9, 585–594; L. M. Halo, M. N. Heneghan, A. A. Yakasai, Z. Song, K. Williams, A. M. Bailey, R. J. Cox, C. M. Lazarus and T. J. Simpson, *J. Am. Chem. Soc.*, 2008, 130, 17988–17996.
- 13 For an enantiospecific synthesis of reutericyclin *via* the Dieckmann strategy: R. Böhme, G. Jung and E. Breitmaier, *Helv. Chim. Acta*, 2005, **88**, 2837–2841.
- 14 For a survey of coupling reagents: P. D. Bailey, in *Comprehensive Functional Group Transformations*, ed. A. R. Katritzky and R. J. K. Taylor, Elsevier, Oxford, 2005, vol. 5, pp. 221–225.
- 15 A. L. L. García, Synlett, 2007, 1328–1329; H. Wissman and H.-J. Kleiner, Angew. Chem., Int. Ed. Engl., 1980, 133–134.
- 16 There was no evidence of racemisation during the sequence to form pyrroloisoxazoles **10a–c**; for further discussion of the stereochemical integrity of intermediates in the sequence, see ref. 10*a* and *b*.
- 17 M. Nitta and T. Kobayashi, J. Chem. Soc., Perkin Trans. 1, 1985, 1401-1406.