

Accepted Manuscript

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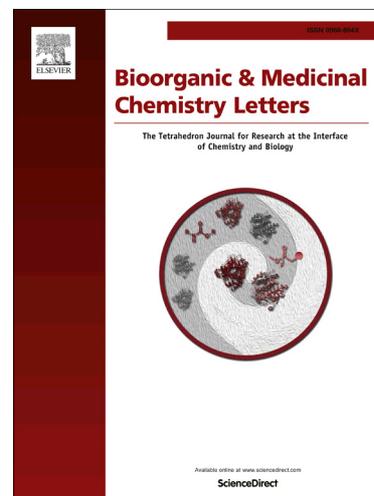
PII: S0960-894X(15)00546-6
DOI: <http://dx.doi.org/10.1016/j.bmcl.2015.05.072>
Reference: BMCL 22762

To appear in: *Bioorganic & Medicinal Chemistry Letters*

Received Date: 16 March 2015
Revised Date: 19 May 2015
Accepted Date: 21 May 2015

Please cite this article as: Passalacqua, T.G., Dutra, L.A., de Almeida, L., Arenas Velásquez, A.M., Esteves Torres, F.A., Yamasaki, P.R., dos Santos, M.B., Regasini, L.O., Michels, P.A.M., da SilvaBolzani, V., Graminha, M.A.S., Synthesis and Evaluation of Novel Prenylated Chalcone Derivatives as Anti-leishmanial and Anti-trypanosomal Compounds, *Bioorganic & Medicinal Chemistry Letters* (2015), doi: <http://dx.doi.org/10.1016/j.bmcl.2015.05.072>

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1 Synthesis and Evaluation of Novel Prenylated Chalcone 2 Derivatives as Anti-leishmanial and Anti-trypanosomal 3 Compounds

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19 Received: / Accepted: / Published:
20

21 **Abstract:** Chalcones form a class of compounds that belong to the flavonoid family and
22 are widely distributed in plants. Their simple structure and the ease of preparation make
23 chalcones attractive scaffolds for the synthesis of a large number of derivatives enabling
24 the evaluation of the effects of different functional groups on biological activities. In this
25 paper, we report the successful synthesis of a series of novel prenylated chalcones via
26 Claisen – Schmidt condensation and the evaluation of their effect on the viability of the
27 Trypanosomatidae parasites *Leishmania amazonensis*, *Leishmania infantum* and
28 *Trypanosoma cruzi*.

29
30 **Keywords:** Prenylated chalcone; leishmanicidal activity; trypanocidal activity; *Leishmania*
31 *amazonensis*; *Leishmania infantum*; *Trypanosoma cruzi*; drug discovery
32

33 Neglected Tropical Diseases (NTDs) have a higher prevalence in tropical and subtropical regions
34 and affect more than one billion people worldwide¹. A list of 17 NTDs² includes the insect vector-
35 borne diseases leishmaniasis and Chagas' disease. Leishmaniasis is a widespread disease caused by
36 parasites belonging to more than 20 species of *Leishmania*, which are transmitted by phlebotomine
37 sandflies after injection of promastigote forms into mammals during feeding. These infective stages

38 are phagocytized by macrophages and other types of mononuclear phagocytic cells. The internalized
39 promastigotes transform into the tissue-stage amastigotes, which multiply by simple division and
40 proceed to infect other macrophages³. The disease affects 12 million people around the world with
41 about 1–2 million estimated new cases occurring every year⁴. About 8-10 million people are currently
42 infected with *Trypanosoma cruzi*, the parasite that causes Chagas' disease, mostly in Latin America⁵.
43 Chagas' disease is transmitted by triatomine insects, which release trypomastigote forms in their feces
44 on the skin of a host. The trypomastigotes enter the host through the bite wound made by the blood-
45 feeding insect or mucous membranes. Inside the host, the trypomastigotes invade cells near the site of
46 entrance, where they differentiate into proliferating amastigotes and then again to trypomastigotes
47 which are released back into the bloodstream to infect other cells. The ingested trypomastigotes
48 transform into epimastigotes in the vector's midgut where the parasites multiply and differentiate into
49 metacyclic trypomastigotes^{6,7}.

50 The current treatment of leishmaniasis is based on chemotherapy and includes pentavalent
51 antimonials (sodium stibogluconate or meglumine antimoniate), the polyene amphotericin B (AmpB)
52 (as the deoxycholate salt or its liposomal formulation), the alkylphosphocholine miltefosine and the
53 aminoglycoside paromomycin⁸. The chemotherapy for Chagas' disease involves two drugs, nifurtimox
54 (nitrofurane) or benznidazole (nitroimidazole), which are efficient only during the acute and initial period
55 of the subsequent chronic phase of the disease. Unfortunately, the toxicity of all currently available
56 drugs to treat these Trypanosomatidae-caused diseases, the inconvenience of parenteral administration,
57 the lack of a guaranteed drug supply and the increasing incidence of treatment failure, make the
58 development of new therapies against them urgent⁹.

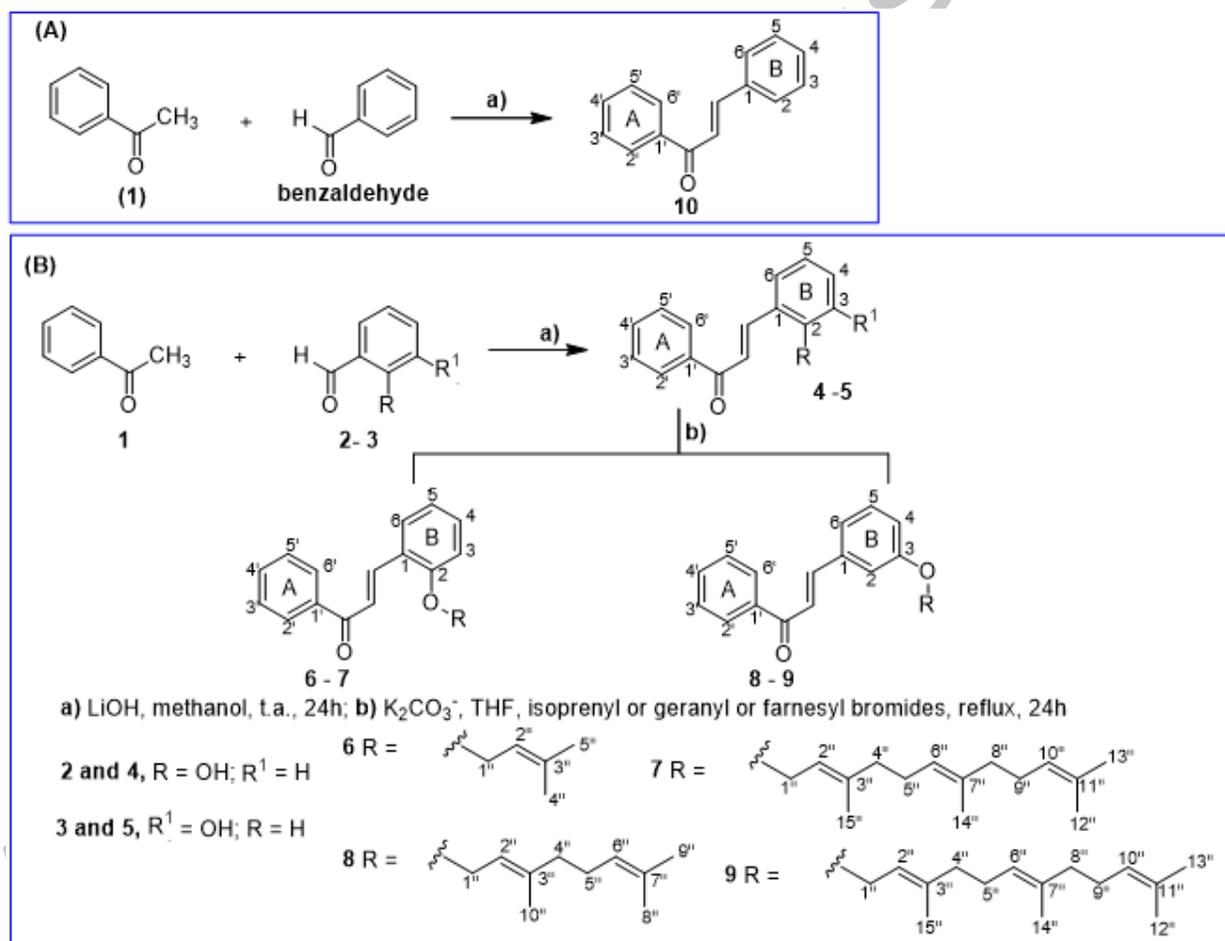
59 Chalcones are molecules consist of open-chain flavonoids in which the two aromatic rings are
60 joined by a three-carbon α , β -unsaturated carbonyl system¹⁰. Their simple structure and the ease of
61 preparation make chalcones attractive scaffolds for the synthesis of a large number of derivatives
62 enabling the evaluation of the effect of different functional groups on biological activities¹¹. Indeed,
63 several synthetic chalcones have shown a number of biological effects such as anti-inflammatory,
64 antibacterial, antifungal, antiviral and antiprotozoal activities¹⁰. Some natural prenylated chalcones
65 such as bartericina, medicanegina and licochalcone are promising compounds for designing new drugs
66 against neglected diseases^{12,14}. Here, we present the effect of new synthetic prenylated chalcones on
67 the viability of the Trypanosomatidae parasites *L. amazonensis*, *L. infantum* and *T. cruzi*.

68 The chalcones **4**, **5** and **10** were synthesized by Claisen-Schmidt condensation between
69 acetophenone (**1**) and benzaldehyde derivatives (**2** and **3**)^{15,16}; these compounds were used as
70 precursors for the synthesis of four novel prenylated chalcones, **6-9** as shown in figure 1. For the
71 preparation of prenylated chalcones **6** and **7**, *O*-isoprenyl and *O*-farnesyl groups were respectively
72 added at position C-2, whereas for **8** and **9**, *O*-geranyl or *O*-farnesyl were respectively added at
73 position C-3. Synthetic reactions in order to produce chalcones and their prenylated derivatives were
74 followed by Nuclear Magnetic Resonance spectroscopy analysis (NMR). The ¹H-NMR spectra
75 displayed a coupling constant range (*J*) for H- α and H- β of 15.0 Hz, corresponding to an (*E*)-chalcone
76 scaffold¹³. The data obtained by NMR analyses for molecules **4** - **9** are provided as Supplementary
77 Material, which confirmed the identity of prenylated chalcones.

78 The novel synthetic prenylated chalcones **6** - **9** as well as their precursor molecules, were used in
 79 screens of *in vitro* growth inhibition assays of promastigote and amastigote forms of *L. amazonensis*
 80 and *L. infantum*, as well as epimastigotes of *T. cruzi*. The cytotoxicity of all molecules for peritoneal
 81 murine macrophages was also tested, allowing the determination of the selectivity index (SI)
 82 parameter. The inhibitory activity of these compounds was evaluated at several concentrations ranging
 83 from 1.0 to 250 μ M. The results expressed as the compound concentrations corresponding to 50 % of
 84 parasite growth inhibition and 50 % macrophage cytotoxicity values (EC_{50} = effective concentration;
 85 the concentration giving 50 % effect) are summarized in Table 1. Since the *O*-prenylated chalcones **6-9**
 86 are novel, no reports of the leishmanicidal activities of such compounds have been published yet (see
 87 methodology in Supplementary Material).

88
 89

Figure 1. General scheme for the synthesis of chalcones.



90

91

Table 1. Leishmanicidal, trypanocidal and cytotoxic activities of chalcone and its prenylated derivatives in μM .

Cpds	<i>L. amazonensis</i> (EC ₅₀)		<i>L. infantum</i> (EC ₅₀)		<i>T. cruzi</i> (EC ₅₀)	Swiss (EC ₅₀)
	Promastigotes	Amastigotes	Promastigotes	Amastigotes	Epimastigotes	Macrophages
4	333.70±23.60 (0.43)	n.d.	91.74±6.70 (1.56)	n.d.	75.89±1.88 (1.88)	143.00±0.16
5	411.60±17.70 (0.35)	n.d.	184.82±13.88 (0.96)	n.d.	68.30±0.63 (2.09)	143.00±0.63
6	4.38±0.07 (25.41)	19.93±1.34 (5.58)	4.69±0.14 (23.73)	10.41±0.99 (10.69)	21.58±1.23 (5.16)	111.30±0.34
7	6.47±0.05 (21.74)	2.90±0.19 (48.50)	10.21±0.19 (13.78)	2.24±0.86 (62.79)	69.37±4.25 (2.03)	140.65±3.62
8	8.92±0.25 (9.18)	25.81±5.05 (3.17)	7.81±0.14 (10.49)	63.89±5.89 (1.28)	26.25±3.11 (3.12)	81.94±8.11
9	9.25±0.47 (6.00)	21.89±0.47 (2.52)	5.98±0.02 (9.22)	32.55±1.89 (1.69)	23.79±1.89 (2.32)	55.14±5.60
10	13.2±1.21 (0.70)	n.d.	8.89±1.91 (1.06)	n.d.	102.64±6.11 (0.09)	9.50±0.03
Pent	10.19±0.85 (3.50)	6.25±0.58 (5.71)	67.71±8.11 (0.53)	19.77±0.52 (1.81)	n.d.	35.69±6.84
AmpB	3.22±0.03 (7.17)	4.92±0.14 (4.70)	0.92±0.01 (25.11)	2.98±0.38 (7.75)	n.d.	23.10±2.52
Benz	n.d.	n.d.	n.d.	n.d.	4.07± 0.31	n.d.

The selectivity indices (SI) are shown in parentheses

n.d. not determined

92

93 Although chalcone (**10**) presented a mild leishmanicidal activity, it showed high cytotoxicity with
94 SI values of 0.7 and 1.06 for *L. amazonensis* and *L. infantum*, respectively. Thus, structural changes
95 were considered in order to obtain more effective and less cytotoxic chalcone derivatives. The
96 intermediate molecules **4** and **5**, which present hydroxyl groups at positions C-2 and C-3, respectively,
97 did not show antileishmanial activity and their cytotoxicity appeared to be very high. In contrast, the
98 prenylation of **4** and **5** resulting in the derivatives **6** - **9** exhibited variable *in vitro* activity against the
99 evaluated parasites. For both *Leishmania* species tested, *L. amazonensis* and *L. infantum*, compounds **6**
100 - **9** were very active on the promastigote forms, with **6** and **7** being the most selective ones to the
101 parasites (SI values of 25.41 and 21.74, respectively) (Table 1).

102 For the amastigote form of *Leishmania*, the clinically most relevant stage of these parasites,
103 compound **7** was the most active and selective prenylated chalcone, for both *L. amazonensis* ($EC_{50} =$
104 $2.90 \mu\text{M} \pm 0.19$; SI= 48.50) and *L. infantum* ($EC_{50} = 2.24 \mu\text{M} \pm 0.86$; SI= 62.79). Compound **7** showed
105 the most outstanding results for *L. infantum* when compared to pentamidine, with respect to both its
106 anti-amastigote activity and its selectivity to the parasite; these values were about 9- and 35-fold higher
107 than those of the reference drug. Regarding *L. amazonensis*, despite its low activity (only two-fold
108 higher than pentamidine), this compound (**7**) still displayed an eight times higher selectivity index.

109 A reasonable explanation for the good antileishmanial activity of **6** - **9** over **4** - **5** and **10** could be
110 the presence of prenyl moieties influencing the lipophilicity of the compounds. Indeed, there is some
111 evidence in the literature that prenylation increases lipophilicity and, consequently, causes an
112 improvement of the biological activity. It has been reported that prenylated flavonoids presented
113 enhanced effects against bacteria, fungi and viruses¹⁷. The prenylated chalcones evaluated in this work
114 contain *O*-isoprenyl (**6**) and *O*-farnesyl (**7**) at C-2 or *O*-farnesyl (**9**) and *O*-geranyl (**8**) at the C-3
115 positions; thus these prenyl moieties may result in an increased lipophilicity of the compounds that is
116 responsible for the augmented leishmanicidal activity by facilitating the passage of the molecules
117 through the cell-membrane barriers of the macrophages and parasites. A close relation between
118 lipophilicity and leishmanicidal activity was also reported by Maciel-Rezende and coworkers (2013)¹⁸,
119 when they evaluated the leishmanicidal activity of a series of *O*-alkyl substituted benzophenones. With
120 regard to other prenylated chalcones, Dal Picolo and coworkers showed that adunchalcone, a
121 prenylated dihydrochalcone (possessing an isoprenyl chain), displayed good antileishmanial activity
122 against promastigote forms of *L. amazonensis*, but lacked any effect against its intracellular
123 amastigotes¹⁹. Gupta and co-workers reported that geranyl chalcones were more potent than isoprenyl
124 chalcones against amastigote forms of *L. donovani*²⁰; these findings could be attributed to a better drug
125 delivery into host cells as a result of the size of the prenyl moiety that affects the potency of the
126 molecules on intact cells.

127 In fact, for *L. infantum* and *L. amazonensis*, **7** was 4.65 and 6.87 more active than **6**, respectively.
128 This effect should possibly be attributed to the presence of the 15 carbons prenyl chain of *O*-farnesyl in
129 **7** compared to the 5 carbons prenyl chain of *O*-isoprenyl in **6**. The long carbon skeleton of *O*-farnesyl
130 contributes to an increased lipophilicity, as confirmed by the calculated ClogP values (see table 2 in
131 Supplementary Material); the ClogP of **7** was found to be 7.75 whereas for **6** it was 4.69. The partition
132 coefficient logP is an important chemical parameter that determines the tendency of a given compound
133 to be distributed in a biphasic system composed of two phases (octanol/water) determining its

134 lipophilicity / hydrophilicity^{21,22,23}. All prenylated chalcones **6** – **9** presented calculated ClogP values
135 higher than those obtained for **4**, **5** and **10**. Indeed, the anti-amastigote activity of **7** is at least five times
136 higher than that of **6**, in agreement with its ClogP, which reinforces the hypothesis that the larger the
137 lipophilicity, the better the antileishmanial activity.

138 It was also observed that the presence of the *O*-farnesyl group at position C-2 in the chalcone
139 skeleton rendered compound **7** more active than both reference drugs tested; furthermore, considering
140 the observed SI values, compound **7** exhibited at least eight times less cytotoxicity than the standard
141 drug Amp B. On the other hand, the *O*-farnesyl group at position C-3 (compound **9**) resulted in a
142 strong loss of leishmanicidal activity as well as an increased cytotoxicity profile suggesting that the
143 position of the *O*-farnesyl group (C-2 or C-3) influences both the leishmanicidal and cytotoxicity
144 activities. These observations suggest that the position of prenyl moieties is important for the correct
145 interaction of the compound with its potential target molecule in the parasite, which supports the
146 notion that the position of the prenyl groups as well as the presence or absence of other substituents in
147 the B ring are important for the leishmanicidal activity.

148 For *T. cruzi*, compounds **6** ($EC_{50} = 21.58 \mu\text{M} \pm 1.23$; $SI = 5.16$), **8** ($EC_{50} = 26.25 \mu\text{M} \pm 3.11$; $SI =$
149 3.12) and **9** ($EC_{50} = 23.79 \mu\text{M} \pm 1.89$; $SI = 2.32$) presented similar potency ranging from 21 to 26 μM ,
150 five times less active than benznidazol; moreover, their low SI values rendered these molecules
151 unattractive for further evaluation against these parasites, at least the available epimastigote stage.

152 Four prenylated chalcone derivatives (**6** – **9**) were synthesized and exhibited good antipromastigote
153 activity as well as good selectivity for both *L. amazonensis* and *L. infantum*. The most promising
154 results were obtained for compound **7** that showed the best anti-amastigote activity, the clinically most
155 relevant parasite form. This compound showed the best relationship between the leishmanicidal effect
156 and the cytotoxicity activity against murine macrophages, which represents a good indication for its
157 therapeutic index for further drug development. Variation in lipophilicity of the evaluated compounds
158 seems to be an important requisite for leishmanicidal activity. Further studies should be conducted to
159 better understand the mechanism of action of these molecules. Thus, these series of prenylated
160 chalcones could be further explored as potential new leishmanicidal drugs.

161 Acknowledgments

162 The authors thank Isabel Martinez for her technical support. They are also grateful to the Fundação de
163 Amparo à Pesquisa do Estado de São Paulo (FAPESP: n° 2010/52327-5), Conselho Nacional de
164 Desenvolvimento Científico e Tecnológico (CNPq: n° 407588/2013-2), Fundação Coordenação de
165 Aperfeiçoamento de Pessoal de Nível Superior (CAPES) and PROPe (Pro-Reitoria de Pesquisa da UNESP) and
166 the FUNDUNESP for financial support.

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168 Author Contributions:

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170 Conceived and designed the experiments: LOR, VSB, MASG. Performed the experiments: TGP, LAD, LA,
171 AMAV, FAET, MBS. Analyzed the data: PRY, MASG. Contributed reagents/materials/analysis tools: LOR,
172 VSB, MASG. Wrote the paper: TGP, LAD, LA, PAMM, MASG.

173 **Supplementary Material**

174 Supplementary material associated with this article can be found, in the online version, at doi:

175 **Conflicts of Interest**

176 The authors declare no conflict of interest.

177 **References**

- 178 1. World Health Organization. Why are some tropical diseases called "neglected"?
179 <http://www.who.int/features/qa/58/en/>, 2013 [accessed 04.12.13].
- 180 2. World Health Organization. Working to overcome the global impact of neglected tropical
181 diseases. First WHO report on neglected tropical diseases.
182 http://whqlibdoc.who.int/publications/2010/9789241564090_eng.pdf, 2010 [accessed 15.08.13].
- 183 3. Kedzierski, L. *Human Vaccines*. **2011**, 7, 1204.
- 184 4. Alvar, J.; Ivan, D.; Velez, C. B.; Merce, H.; Desjeux, P.; den Boer, M. *Plos One*. **2012**, 7, e35671.
- 185 5. Rassi, A. Jr.; Rassi, A.; Rezende, M. J. *Infect. Dis. Clin. North. Am.* **2012**, 26, 275.
- 186 6. Schmunis, G. A.; Yadon, Z. E. *Acta Trop.* **2010**, 115, 14.
- 187 7. Noireau, F.; Diosque, P.; Jansen, A.M. *Vet. Res.* 2009, 40, 1.
- 188 8. Seifert, K. *Open Clin. Chem. J.* **2011**, 5, 31.
- 189 9. Castro, J.A.; de Mecca, M.M.; Bartel, L.C. *Hum. Exp. Toxicol.* **2006**, 25, 471.
- 190 10. Nowakowska, Z. *Eur. J. Med. Chem.* **2007**, 42, 125.
- 191 11. Romagnoli, R.; Baraldia, P. G.; Carrion, M. D.; Cruz-Lopez, O.; Cara, C. L.; Balzarini, J.;
192 Hamel, E.; Canella, A.; Fabbri, E.; Gambari, R.; Basso, G.; Viola, G. *Bioorg. Med. Chem. Lett.*
193 **2009**, 19, 2022.
- 194 12. Ngameni, B.; Watchueng, J.; Boyom, F. F.; Keumedjio, F.; Ngadjui, B. T.; Gut, J.; Abegaz, B. B.;
195 Rosenthal, P. J. *Arkivoc.* **2007**, 13, 116.
- 196 13. Narender, T.; Tanvir, K.; Rao, M. S.; Srivastava, K.; Puri, S. K. *Bioorg. Med. Chem. Lett.* **2005**,
197 10, 2453.
- 198 14. Chen, M.; Christensen, S. B.; Blom, J.; Lemmich, E.; Nadelmann, L.; Fivvh, K.; Theander, T. G.;
199 Kharazmi, A. *Antimicrob. Agents Chemother.* **1993**, 37, 2550.
- 200 15. Zeraik, M.L.; Ximenes, V.F.; Regasini, L.O.; Dutra, L.A.; Silva, D.H.S.; Fonseca, L.M.; Coelho,
201 D.; Machado, S.A.S.; Bolzani, V.S. *Curr. Med. Chem.* **2012**, 19, 5405.
- 202 16. Vogel, S.; Heilmann, J. *J. Nat. Prod.* **2008**, 71, 1237.
- 203 17. Chen, X.; Mukwaya, E.; Wong, M.S.; Zhang, Y. *Pharm. Biol.* **2014**, 52, 655.
- 204 18. Maciel-Rezende, C. M.; de Almeida, L.; Costa, É. D.; Pires, F. R.; Alves, K. F.; Junior, C. V.;
205 Diase, D. F.; Doriguetto, A. C.; Marques, M. J.; dos Santos, M. H. *Bioorg. Med. Chem. Lett.*
206 **2013**, 21, 3114.
- 207 19. Dal Picolo, C. R.; Bezerra, M. P.; Gomes, K. S.; Passero, L. F. D.; Laurenti, M. D.; Martins, E. G.
208 A.; Sartorelli, P.; Lago, J. H. G. *Fitote.* **2014**, 97, 28.
- 209 20. Gupta, S.; Shivahare, R.; Korthikunta, V.; Singh, R.; Gupta, S.; Tadigoppula, N. *Eur. J. Med.*
210 *Chem.* **2014**, 81, 359.

211 21. Leo, A.; Hansch, C.; Elkins, D. *Chem. Rev.* **1971**, *71*, 525.

212 22. Liao, Q.; Yao, J.; Yuan, S. *Mol. Divers.* **2006**, *10*, 301.

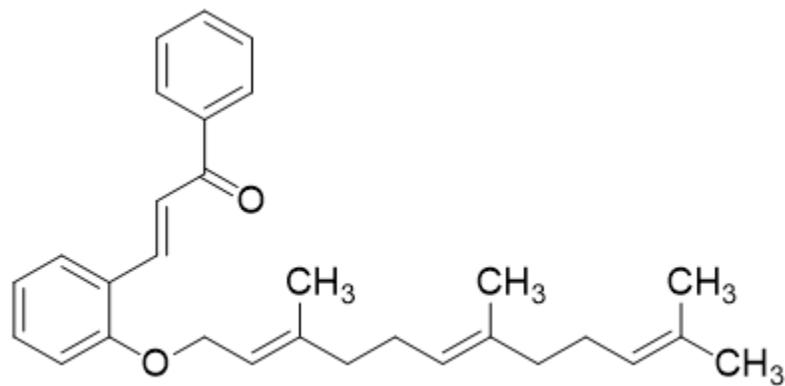
213 23. Scherrer, R.A.; Howard, S.M. *J. Med. Chem.* **1977**, *20*, 53.

214 *Sample Availability:* Samples of the compounds 6-9 are available from the authors.

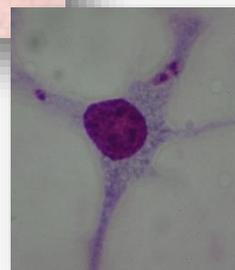
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(*E*)-2-*O*-farnesyl-chalcone (**7**)



Antileishmanial
activity