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Thiol-based SAHA analogues as potent histone deacetylase inhibitors

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Abstract—In order to find novel nonhydroxamate histone deacetylase (HDAC) inhibitors, a series of thiol-based compounds modeled after suberoylanilide hydroxamic acid (SAHA) was synthesized, and their inhibitory effect on HDACs was evaluated. Compound **6**, in which the hydroxamic acid of SAHA was replaced by a thiol, was found to be as potent as SAHA, and optimization of this series led to the identification of HDAC inhibitors more potent than SAHA. © 2004 Elsevier Ltd. All rights reserved.

The reversible acetylation of the side chain of specific histone lysine residues by histone deacetylases (HDACs) and histone acetyl transferases (HATs) is an important regulator of gene expression.¹ Histone hyperacetylation by HDAC inhibition neutralizes the positive charge of the lysine side chain, and is thought to be associated with change of the chromatin structure and the consequential transcriptional activation of a number of genes.² One important outcome of histone hyperacetylation is induction of the cyclin-dependent kinase inhibitory protein $p21^{Waf1/Cip1}$, which causes cell cycle arrest.³ Indeed, HDAC inhibitors such as trichostatin A (TSA) and suberoylanilide hydroxamic acid (SAHA) (Fig. 1) have been reported to inhibit cell growth,⁴⁻⁶ induce terminal differentiation in tumor cells,^{4,5} and prevent the formation of tumors in mice.⁷ Therefore, HDACs have been viewed as attractive targets for anticancer drug development. A number of structurally diverse HDAC inhibitors have been reported⁸ and most of them belong to hydroxamic acid derivatives, typified by TSA and SAHA, which chelate the zinc ion in the active site in a bidentate fashion through its CO and OH groups.⁹ Although hydroxamic acids are frequently employed as zinc-binding groups (ZBGs), they often present metabolic and pharmacokinetic problems such as glucuron-

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Figure 1. HDAC inhibitors

idation, sulfation, and enzymatic hydrolysis that result in a short in vivo half-life.^{10,11} Because of such concerns with the metabolic stability associated with hydroxamic acids, it has become increasingly desirable to find replacement groups that possess strong inhibitory action against HDACs. Thus far, *o*-aminoanilides,^{12–14} electrophilic ketones,^{15–17} bromoacetamides,¹⁸ semicarbazides¹⁸ (Fig. 2), and *N*-formyl hydroxylamines¹⁹ have been reported as small molecule nonhydroxamate HDAC inhibitors. However, they have reduced potency as compared to hydroxamate inhibitors, and unfortunately, electrophilic ketones have a metabolic disadvantage in that they are readily reduced to inactive alcohols.^{15–17} We therefore initiated a search for

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Figure 2. SAHA-based nonhydroxamate HDAC inhibitors.

replacement groups for hydroxamic acid with the goal of drug discovery as well as finding new tools for biological research.

In the search for a suitable hydroxamic acid replacement, thiols seemed to be reasonable targets, because they have been reported to inhibit zinc-dependent enzymes such as angiotensin converting enzyme²⁰ and matrix metalloproteinase.²¹ Recently, Furumai et al. demonstrated that the disulfide bond of FK228 (Fig. 1), a cyclic peptide HDAC inhibitor, is reduced in the cellular environment, releasing the free thiol analogue as the active species,²² and Nishino et al. reported that cyclic tetrapeptides bearing disulfide group such as **1** (Fig. 1) inhibit HDACs under reductive conditions.²³ These reports prompted us to report on the synthesis and HDAC inhibition of thiol-based small molecule analogues.

The compounds prepared for this study are shown in Tables 1–3. Syntheses were accomplished as illustrated in Schemes 1–3. Compounds 6–8, 10–12, 15–21, and 22 were synthesized from the corresponding acid chlorides 27a–d by the route shown in Scheme 1. The amino group of aromatic amines 28 was acylated with an appropriate acid chloride 27 to give the amides 29a–f and 29h–m. Suzuki coupling²⁴ of bromobenzene 29f with phenylboronic acid provided the biphenyl 29g. Bromides 29 were treated with potassium thioacetate to give compounds 7 and 30a–k, after which hydrolysis of the thioacetates under alkaline conditions gave the desired compounds 6, 10–12, 15–21, and 22. Sulfide 8 was obtained by the alkylation of sodium methanethiolate with bromide 29b.

Synthesis of aminoethanethiol 9 was accomplished via aldehyde 34 (Scheme 2). The condensation of dicarboxylic acid 31 with an equivalent amount of aniline gave mono-anilide 32. The carboxylic acid 32 was converted to Weinreb amide²⁵ 33 in the presence of EDCI and HOBt. Compound 33 was allowed to react with lithium aluminum hydride at 0° C to give aldehyde 34 and subsequent reductive amination afforded amino-ethanethiol 9.

Compounds 13, 14, 23–25, and 26 were prepared from alcohol 35 or 36 by the procedure outlined in Scheme 3. Treatment of bromide 35 with phenol in the presence of K_2CO_3 gave ether 38a, and condensation of amine 36 with an appropriate aromatic carboxylic acid 37 affor-



Scheme 1. Reagents and conditions: (a) $ArNH_2$ (28), Et_3N , CH_2Cl_2 , rt, 25–99%; (b) PhB(OH)₂, Pd(PPh₃)₄, NaHCO₃, 1-methyl-2-pyrrolidinone, H₂O, 80 °C, 18%; (c) AcSK, EtOH, 60 °C, 84–99%; (d) 2 N aq NaOH, EtOH, THF, rt, 47–99%; (e) 15% aq NaSMe, EtOH, rt, 99%.

ded amides **38b–f**. Alcohols **38a–f** were converted to thiols **13**, **14**, **23–25**, and **26** in a three-step sequence: conversion of the alcohols to bromides, treatment of the bromides with potassium thioacetate, and hydrolysis of the resulting thioacetates.

The compounds synthesized in this study were tested with an in vitro assay using a HeLa nuclear extract rich in HDAC activity.²⁶ The results are summarized in Tables 1–3 as IC_{50} values.



Scheme 2. Reagents and conditions: (a) Aniline, $180 \circ C$, 43%; (b) *N*,*O*-dimethylhydroxylamine hydrochloride, Et₃N, EDCI, HOBt, DMF, rt, 94%; (c) LiAlH₄, THF, $0 \circ C$, 72%; (d) 2-aminoethanol, NaBH(OAc)₃, THF, AcOH, rt; (e) (Boc)₂O, Et₃N, rt; (f) TFA, CH₂Cl₂, rt, 44% (three steps).



Scheme 3. Reagents and conditions: (a) Phenol, K_2CO_3 , DMF, 80 °C, 96%; (b) ArCOOH (37), EDCI, HOBt, DMF, rt, 61–96%; (c) CBr₄, PPh₃, CH₂Cl₂, 0 °C, 25–99%; (d) AcSK, EtOH, 60 °C, 47–99%; (e) 2 N aq NaOH, EtOH, THF, rt, 28–74%.

As seen in Table 1, the IC₅₀ values of SAHA, *o*-aminoanilide **2**, bromoacetamide **4**, and semicarbazide **5** were 0.28, 120, 14, and 150 μ M, respectively (entries 1, 2, 4, and 5). Trifluoromethyl ketone **3** was reported previously to inhibit HDACs with an IC₅₀ of 6.7 μ M (entry 3).¹⁵ In our study, changing the hydroxamic acid of SAHA to thiol yielded fruitful results. A pronounced inhibitory effect (IC₅₀ = 0.21 μ M) was observed with thiol **6**, which was about 30–700-fold more active than the previously reported nonhydroxamates, and as potent as SAHA (entry 6). To confirm that the thiol group plays an important role in anti-HDAC activity, thioacetate **7**, and sulfide **8** were tested. As expected, thiol transformation into thioacetate and sulfide led to an

 Table 1. HDAC inhibition data for SAHA and SAHA-based non-hydroxamates^a

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Entry	Compd	n	R	$IC_{50} \ (\mu M)$			
1	SAHA ^b	6	-CONHOH	0.28			
2	2 ^c	6	$\overset{H}{\underset{O}{}}\overset{NH_{2}}{\underset{O}{}}$	120			
3	3	6	-COCF ₃	6.7 ^d			
4	4 ^e	6	-NHCOCH2Br	14			
5	5 ^e	5	-NHCONHNH ₂	150			
6	6	6	-SH	0.21			
7	7	6	–SAc	7.1			
8	8	6	-SMe	>100			
9	9 ^f	6	-NHCH2CH2SH	>100			

^a Values are means of at least three experiments.

^b Prepared as described in Ref. 31.

^c Prepared as described in Ref. 13.

^d Data taken from the literature (Ref. 15).

^e Prepared as described in Ref. 18.

^fTrifluoroacetic acid salt.

 Table 2. Effect of linker variation on HDAC inhibitory activity of thiols^a

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Entry	Compd	Х	п	IC_{50} (μM)			
1	6	-NHCO-	6	0.21			
2	10	-NHCO-	7	1.5			
3	11	-NHCO-	5	0.37			
4	12	-NHCO-	4	6.2			
5	13	-0-	6	11			
6	14	-CONH-	6	0.36			

^a Values are means of at least three experiments.

inhibitor that was about 30-fold less potent and a compound devoid of anti-HDAC activity, respectively (entries 7 and 8). These results suggest that although the inherent affinity of a monodentate thiol ZBG is less than that of bidentate ZBGs such as hydroxamate and hydrated electrophilic ketone, its ease of ionization makes thiol **6** as powerful as SAHA.²⁷ In order to find more potent ZBGs, we examined the activity of aminoethanethiol **9**, which was expected to coordinate zinc ion in a bidentate fashion. Although aminoethanethiols have been reported to inhibit zinc proteins,²⁸ this functional group led to a loss of HDAC inhibitory activity (entry 9).

We next examined the effect of linker parts of thiol 6. The results are shown in Table 2. HDAC inhibition was distinctly dependent on chain length, with n = 7 (10) and n = 4 (12) resulting in less potent inhibitors. However, compound 11, in which n = 5, proved to be equally effective to 6, in which n = 6 (entries 1–4). The similar structure–activity relationship (SAR) between thiols and hydroxamates, with n = 6 optimal,^{29,30} indicates that thiols inhibit HDACs in a binding mode similar to that of hydroxamates. As for the group attaching the phenyl moiety, ether 13 displayed a moderate activity, whereas the activity of the reversed amide 14 was maintained (entries 5 and 6).

Having investigated the requirements for the ZBGs and linker parts, we next turned our attention to aromatic groups (Table 3). In the amide-linked series (entries 1-9), 4-substituted phenyl compounds tended to reduce the potency. Specifically, compounds 15 (Ar = 4-NMe₂-Ph), 16 (Ar = 4-biphenyl), and 18 (Ar = 4-PhO-Ph) showed about a 3-10-fold decrease in potency when compared to the parent thiol 6 (entries 2, 3, and 5). In contrast, when a phenyl group was introduced at the 3-position of the phenyl group of 6, the IC_{50} of compound 17 was improved and reached 0.075 µM (entry 4). In addition, 3-phenoxy compound 19 was equipotent with compound 6 (entry 6). Next, we investigated the effect of the replacement of the phenyl group of compound 6 with heteroaryl rings (entries 7-9). While pyridine 20 and phenylthiazole 22 retained the potency of compound 6, quinoline 21 was about 3-fold more active than compound 6 (IC₅₀ = $0.072 \,\mu$ M). The reversed amide-linked series (entries 11-14) exhibited potencies similar to or Table 3. Effect of aromatic group variation on HDAC inhibitory activity of ${\rm thiols}^{\rm a}$

Entry	Compd	Ar	Х	IC ₅₀ (µM)
1	6	-Ph	-NHCO-	0.21
2	15		-NHCO-	1.2
3	16	— Ph	-NHCO-	1.1
4	17	Ph-Ph	-NHCO-	0.075
5	18		-NHCO-	0.62
6	19	~-OPh	-NHCO-	0.21
7	20	$-\!$	-NHCO-	0.11
8	21		-NHCO-	0.072
9	22		-NHCO-	0.17
10	14	–Ph	-CONH-	0.36
11	23		-CONH-	0.61
12	24		-CONH-	0.085
13	25		-CONH-	0.079
14	26	- N H	-CONH-	0.1

Ar X CSH

^a Values are means of at least three experiments.

greater than the parent thiol 14, except for 23 (Ar = 4-NMe₂-Ph), which resulted in a slightly less potent inhibitor. In particular, the reversed amides 24 with a naphthalene substituent and 25 with a benzofuran substituent showed stronger inhibition of HDACs with IC₅₀s of 0.085 and 0.079 μ M, respectively. As a result, IC₅₀s in the double-digit nanomolar range were observed with 3-biphenyl 17, quinoline 21, naphthalene 24, and benzofuran 25, which were approximately 3–4-fold more potent than SAHA.

In summary, in order to find novel nonhydroxamate HDAC inhibitors, we designed and prepared a series of thiol-based SAHA analogues, and evaluated their inhibitory effect on HDACs. Compound 6, in which the hydroxamic acid of SAHA was replaced by a thiol, was found to be as potent as SAHA. We have shown that the potency is related to chain length, with n = 6 optimal, and the amide and reversed amide were preferred as the group attaching the phenyl moiety. The conversion of the phenyl group of compound 6 to other aromatic groups led to the identification of inhibitors more potent than SAHA. The SAR results within the thiol series indicate that thiols inhibit HDACs in a manner similar to that of hydroxamates. As far as we could determine, this is the first report of nonmacrocyclic thiol inhibitors of HDACs. These small molecule thiols may be useful as tools for biological research and as orally bioavailable anticancer drugs. Currently, further detailed SAR studies and the next stage of evaluations are under way.

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