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Preparation and Purification of Desethylamiodarone Hydrochloride

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Abstract: Reaction of the free base of amiodarone with 1-chloroethyl chloroformate in toluene gives desethylamiodarone, which can be easily purified and converted to the hydrochloride salt.

Keywords: Amiodarone, chloroethyl chloroformate, dealkylation, desethylamiodarone, metabolite

INTRODUCTION

Amiodarone [(2-butylbenzofuran-3-yl) (4-[2-(diethylamino) ethoxy])-3,5-diiodophenyl)methanone] is indicated for the treatment of acute and chronic atrial and ventricular arrhythmias in humans^[1] and is seeing a resurgence of use for atrial fibrillation.^[2,3] Both amiodarone and its major metabolite, desethylamiodarone [(2-butylbenzofuran-3-yl)(4-[2-(ethylamino)ethoxy])-3,5-diiodophenyl)methanone],^[4] are pharmacologically active,^[5–7] and both compounds have been implicated in the prolongation of the duration of activation and recovery of the ventricular myocardium (QT interval),^[8–12] which is a major side effect of chronic amiodarone administration. Desethylamiodarone is not commercially available, thereby limiting investigation into the in vivo behavior of both amiodarone and desethylamiodarone. The present work describes a facile method to prepare desethylamiodarone from the commercially available amiodarone hydrochloride salt. The method for de-ethylation of

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amiodarone to give desethylamiodarone is based on the work of Olofson et al. for the dealkylation of *N*-ethylpiperidine, *N,N*-diethylaniline, and various drugs with 1-chloroethyl chloroformate.^[13]

During our evaluation of the optimal reagents and conditions for generation of desethylamiodarone, we evaluated various chloroformic acid derivatives for their ability to dealkylate amiodarone. 1-Chloroethyl chloroformate proved to be the only chloroformic acid derivative of the compounds we studied that provided facile dealkylation. In addition to the work of Olofson et al., this reagent has previously been used to prepare the *N*-demethylated metabolites of tamoxifen,^[14] promazine, levomepromazine, orphenadrine, clomipramine and chlorprothixene,^[15] various erythromycins,^[16] and zopiclone.^[17] The mild reaction conditions involved when using 1-chloroethyl chloroformate for dealkylation of tertiary amines is especially useful for compounds with other acid labile functional groups.

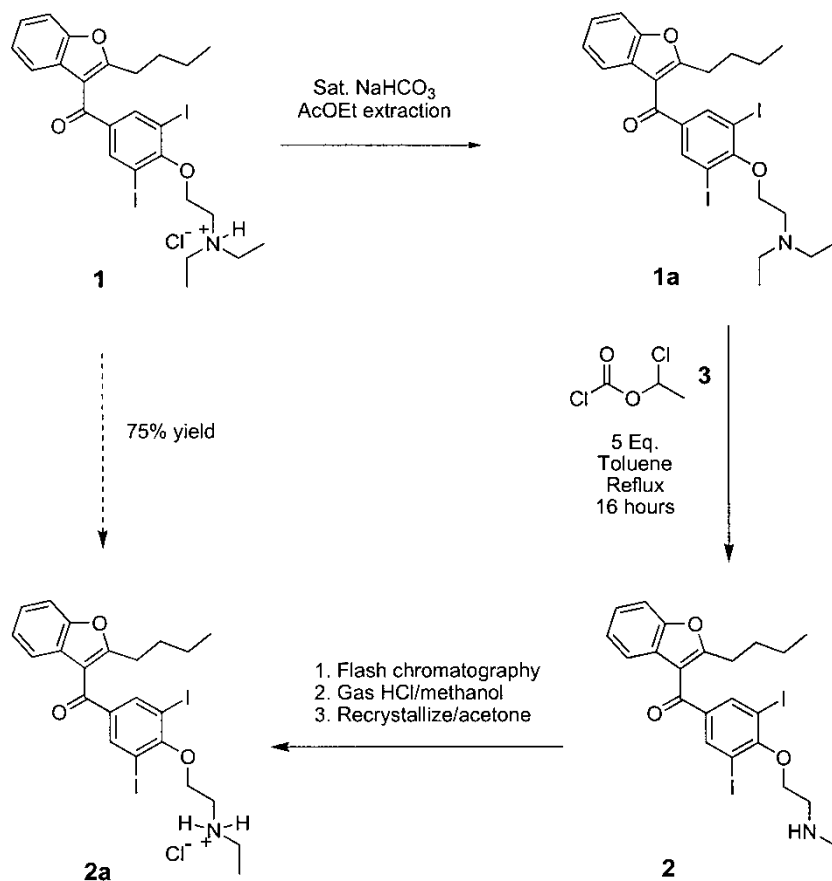
RESULTS AND DISCUSSION

Commercially available amiodarone hydrochloride (**1**) was converted to the free base (**1a**) in saturated aqueous sodium bicarbonate, followed by extraction into ethylacetate Scheme 1. The ethylacetate solution was dried with sodium sulfate and filtered, and **1a** was isolated by rotary evaporation of the solvent. Refluxing **1a** and 1-chloroethyl chloroformate (**3**, 5 equivalents) in toluene for 16 h resulted in nearly quantitative conversion of **1a** to desethylamiodarone (**2**) with minimal side-product formation. The rate of the reaction was enhanced with increased equivalents of **3**; however, the amount of by-products increased also.

Various other chloroformic acid derivatives were also evaluated for conversion of **1a** to **2**. None of the seven other chloroformic acid derivatives attempted (Table 1) resulted in formation of **2**. For those derivatives that provided evidence of a stable tertiary or quaternary carbamate intermediate (liquid chromatography with mass detection, LC/MS analysis) following 16 h at reflux, additional workup in solution with bubbling HCl gas did not convert the carbamates to **2**, and all the intermediates were recovered unchanged.

Because **2a** was to be used in vivo studies and as an analytical standard, having a pure and well-characterized product was important. To obtain a pure product, the crude product **2** was subjected to flash chromatography and converted to **2a** with gas HCl. Compound **2a** was recrystallized twice to give the highly pure product in good yield (75%) based on the original amount of **1**. The identity of **2a** was confirmed with ¹H and ¹³C NMR, elemental analysis, and FAB mass-spectral analysis. The purity of **2a** was determined using thermogravimetric analysis (TGA), differential scanning calorimetry (DSC), Karl Fisher titration, and reverse-phase LC/MS.

The NMR spectra of **2a** were in excellent agreement with the corresponding spectra of **1** and were consistent with the loss of one ethyl group in **2a**. Fast atom bombardment (FAB) positive mass spectral analysis gave a



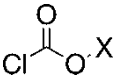


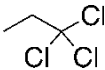





Scheme 1.

major $M+1$ ion peak of 618.1 amu, consistent with the exact mass of **2**. Compound **2a** did not contain residual volatile solvents based on Karl Fisher and TGA analysis. It exhibited a single sharp endotherm on DSC analysis and a single peak on LC/MS analysis with an $M+1$ peak of 618 amu, indicating the high purity of the product. The procedure described provides for the facile preparation of pure **2a** from **1** in high yield.

EXPERIMENTAL

All chemicals were purchased from commercial suppliers and used without further purification. NMR spectra were recorded on a Bruker Avance 400-MHz spectrometer. FAB mass-spectral analysis was performed on a VG (MicroMass) ZAB HS spectrometer in a matrix of nitrobenzyl alcohol in

Table 1. Chloroformic acid derivatives attempted for the de-ethylation of amiodarone

	Desethyl-amiodarone	Stable intermediate	No reaction
	+		
		+	
		+	
		+	
		+	
		+	+
			+
			+

Note: Reaction conditions: 5 eq. chloroformic acid derivative refluxed in toluene for 16 h.

methanol. TGA and DSC analyses were performed on a Perkin Elmer TGA7 gravimetric analyzer and Pyris 1 calorimeter, respectively. Karl Fisher analysis was performed on a Brinkman 652 KF-coulometer. For LC/MS analysis, the sample was fractionated on a Zorbax column (5 μ m, SB-18, 2.1 \times 50 mm) using a modular Shimadzu system consisting of a LCMS-2010 detector in electrospray positive mode, a SIL-10 ADVP auto injector, LC-10 ADVP pumps, DGV-14A degasser, and SL-10AVP system controller, with data collection and analysis performed by LCMS solution ver. 2.04 software. The mobile phase consisted of 0.4:99.6 parts ammonium hydroxide (30%)–methanol at a flow rate of 200 μ L/min.

The free base **1a** was prepared from commercially available **1** by dissolving **1** (3.0 gm, 4.4 mmol) in saturated aqueous sodium bicarbonate (200 mL) and mixing the resulting solution vigorously with ethylacetate (200 mL) for 30 min. The organic layer was isolated, dried with sodium sulfate, filtered, and evaporated to dryness to give a yellow liquid. The residue was dissolved in toluene (10 mL) and 1-chloroethyl chloroformate (5 equivalents; 2.4 mL) was added with stirring. The mixture was refluxed for 16 h, cooled to room

temperature, and evaporated to dryness under vacuum. The resulting residue was dissolved in ethyl acetate and washed with saturated aqueous sodium bicarbonate. The organic phase was isolated, dried with sodium sulfate, filtered, and evaporated to dryness under vacuum. The crude product was purified by flash chromatography on silica gel via elution of impurities with ethyl acetate–hexanes (10:90 V/V) followed by elution of **2** with methanol. Compound **2** was converted to **2a** by bubbling HCl gas through the pooled methanol fractions that were then evaporated to dryness. The product were recrystallized twice in acetone to give 2.17 g of **2a** (75% yield) as a white solid. The ^1H and ^{13}C NMR (400 MHz, CDCl_3) spectra of **2a** were compared to those of **1**, and chemical shifts were within 0.1 ppm. The integration of the ethyl protons (3.47 and 1.58 ppm) corresponds to the loss of one ethyl group in **2a**. FAB positive: $M+1$ peak 618.1 amu; **2** exact mass 616.99 amu. Karl Fisher analysis: 0% water. DSC analysis: single, sharp endotherm at 186.7°C. Anal. calcd. for $\text{C}_{23}\text{H}_{26}\text{Cl}_2\text{NO}_3$: C, 42.26; H, 4.01; N, 2.14. Found: C, 42.47; H, 4.00; N, 2.22.

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