Synthesis of 3,5-Dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl Containing 1,2,3-Thiadiazole Derivatives via Ugi Reaction and Their Biological Activities[†]

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A series of novel 3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl containing 4-methyl-1,2,3-thiadiazole derivatives were designed and synthesized via Ugi reaction. Their structures were confirmed by IR, ¹H NMR, ¹³C NMR and high-resolution mass spectroscopy. The preliminary bioassay results indicated that some title compounds had good fungicide activity at 50 μ g/mL; most of the compounds presented a certain degree of direct inhibition activity, good inactivation and curative activity against tobacco mosaic virus at 500 μ g/mL and 100 μ g/mL; some compounds showed good larvicidal activity against *Plutella xylostella* L. at 200 μ g/mL and excellent larvicidal activities against *Culex pipiens pallens* at 2 μ g/mL.

Keywords 1,2,3-thiadiazole, Ugi reaction, synthesis design, biological activity, multicomponent reactions

Introduction

1,2,3-Thiadiazoles are a versatile class of compounds which present a wide range of biological activities such as antivirus activity,¹ systemic acquired resistance,^{2,3} fungicide activity,^{2,4} insecticide activity,⁵ antitumor activity,⁶ and herbicide activity.^{7,8} Moreover, the properties of easy breakdown of the 1,2,3-thiadiazole ring into low molecular weight compounds through releasing of N₂ favor the use of its derivatives as environmental friendly pesticide candidates with low toxicity.⁹

Combinatorial chemistry has attracted much attention as an efficient tool for drug discovery. The use of combinatorial chemistry or parallel synthesis for the optimization of highly promising lead compounds arising from the drug-discovery processes around privileged structures and their combinations has been successful in many science and development fields.¹⁰ The Ugi reaction is one of the most prominent multiple component coupling reactions (MCRs) due to its wide applications in organic syntheses and medicinal chemistry.¹¹ Ugi reaction is an efficient and convenient green method to combine several active moieties into one molecule, and it is convenient to implement structural modification via alternation of diverse starting materials and is applied in combinatorial chemistry.^{12,13}

Fluorine and chlorine containing compounds have significant applications in drug and pesticide research and development. Introducing F and Cl into target molecules can be a feasible method to improve its biological activities. Xu et al.14a has reported that some fluoroalkoxyl containing S-methyl benzo-1,2,3-thiadiazole-7-carboxylate derivatives exhibited good systemic acquire resistance. The excellent example for novel pesticide development is the successful discovery of fungicide flumorph, which was designed and synthesized by substituting the Cl of the dimethomorph by the F^{14b} Our group^{4c,15} has designed and synthesized series of 1,2,3-thiadiazole derivatives containing 3-chloro-4-methyl-phenyl and 3-fluoro-4methylphenyl via Ugi four-components condensation reaction (U-4CR). The results indicated that some compounds showed broad-spectrum of activities against several fungi tested and excellent potential antivirus activities. Encouraged by these observations, here we introduced 3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl, the active substructure of a commercial insecticide hexaflu-

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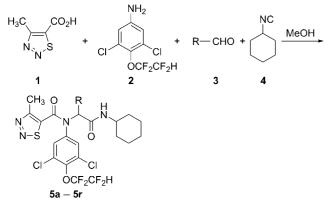
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Received October 7, 2010; revised November 23, 2010; accepted December 21, 2010.

Project supported by the National Natural Science Foundation of China (Nos. 20872071, 20911120069), the Tianjin Natural Science Foundation (No. 10JCZDJC17500), the National Key Project for Basic Research (No. 2010CB126105), the National Key Technology Research and Development Program (Nos. 2011BAE06B02, 2011BAE06B05) and the Foundation of Achievements Transformation and Spreading of Tianjin Agricultural Science and Technology (No. 201002250), the Russian Foundation for Basic Research (Nos. RFBR 08-03-00376 a and RFBR/NNSF 08-03-92208 a). [†] Dedicated to Professor Chuanfan Qian for her 80th birthday.

muron, into the 1,2,3-thiadiazoles via Ugi U-4CR. Like our previous work,^{4c} 4-methyl-1,2,3-thiadiazole-5-carboxylic acid was chosen as the acid component; substituted benzaldehydes were chosen as the carbonyl components; cyclohexyl isocyanide was chosen as the isocyanide component. This will provide useful information for the study of the structure-activity relationship of these new structures for their insecticide activity. The synthetic route was shown in Scheme 1. The structures of title compounds were confirmed by IR, ¹H NMR, ¹³C NMR and high-resolution mass spectroscopy.



Experimental

Materials and instruments

Melting points of all compounds were determined on an X-4 binocular microscope (Gongyi Technical Instrument Co., Henan, China), and the thermometer was not corrected. Proton NMR spectra were obtained using a Bruker AVANCE-400 MHz spectrometer, and chemical shift values (δ) were reported with deuteron chloroform (CDCl₃) as the solvent and tetramethylsilane (TMS) as the internal standard. High resolution mass spectrometry (HRMS) data were obtained on an FTICR-MS Varian 7.0T FTICR-MS instrument. All solvents and liquid reagents were of analytical reagent grade and were dried in advance and distilled before use. Column chromatography purification was carried out by using silica gel. 4-Methyl-1,2,3-thiadiazole-5-carboxylic acid (1) and cyclohexyl isocyanide (4) were synthesized according to Ref. 2 and 4c, respectively.

General procedure for preparation of the title compounds 5a—5r

3,5-Dichloro-4-(1,1,2,2-tetrafluoroethoxy)benzenamine (0.83 g, 3.0 mmol), 4-methyl-1,2,3-thiadiazole-5carboxylic acid (0.43 g, 3.0 mmol), and cyclohexyl isocyanide (0.33 g, 3.0 mmol) were added to a solution of benzaldehyde (0.35 g, 3.0 mmol) in 10 mL of methanol in sequence. The color of the mixture grew darker during this procedure. The solution was then stirred for another 1 h at room temperature; afterward, removal of solvent under reduced pressure and purification by flash column chromatography on silica gel using ethyl acetate and petroleum ether (60—90 °C; 1:3, V:V) as an eluent obtain the products **5a**—**5r**.

N-[2-(Cyclohexylamino)-2-oxo-1-phenylethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4methyl-1,2,3-thiadiazole-5-carboxamide (5a) White solid, yield 40%, m.p. 178—181 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.38—7.05 (m, 7H, ph-H), 6.18 (s, 1H, COCH), 6.12—5.85 (m, 1H, OCF₂CF₂H), 5.37 (d, *J*=7.6 Hz, 1H, CONH), 3.90—3.82 (m, 1H, cyclohexyl-H), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.02—1.04 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.3, 161.1, 142.4, 140.7, 137.7, 132.8, 132.2, 130.8, 130.3, 129.7, 129.1, 65.7, 49.3, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) *v*: 3280, 3080, 2933, 2855, 1647, 1557, 1459, 1353, 1285, 1233, 1142 cm⁻¹; HRMS calcd for C₂₆H₂₄Cl₂F₄N₄O₃S (M+H)⁺ 619.0955, found 619.0959.

N-[1-(4-Chlorophenyl)-2-(cyclohexylamino)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5b) White solid, yield 30%, m.p. 187—189 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.38—7.05 (m, 6H, ph-H), 6.18 (s, 1H, COCH), 6.14—5.87 (m, 1H, OCF₂CF₂H), 5.37 (d, J = 7.6 Hz, 1H, CONH), 3.90 — 3.83 (m, 1H, cyclohexyl-H), 2.87 (s, 3H, thiadiazolyl-CH₃), 2.01— 1.08 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.3, 161.1, 160.8, 149.7, 142.5, 140.8, 137.4, 133.2, 132.1, 131.0, 130.8, 127.4, 125.0, 60.7, 49.6, 32.4, 25.3, 24.8, 24.6, 13.7; IR (KBr) *v*: 3293, 3077, 2937, 2856, 1649, 1552, 1460, 1347, 1287, 1234, 1152 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₃F₄N₄O₃S (M+H)⁺ 653.0565, found 653.0573.

N-[1-(2-Chlorophenyl)-2-(cyclohexylamino)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5c) White solid, yield 62%, m.p. 163—166 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.43—7.02 (m, 6H, ph-H), 6.56 (s, 1H, COCH), 6.10—5.84 (m, 1H, OCF₂CF₂H), 5.52 (d, J = 6.4 Hz, 1H, CONH), 3.91— 3.87 (m, 1H, cyclohexyl-H), 2.87 (s, 3H, thiadiazolyl-CH₃), 2.06— 1.09 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.4, 161.3, 160.9, 149.7, 142.4, 140.7, 137.3, 135.2, 131.3, 131.0, 130.7, 129.8, 127.3, 62.3, 49.5, 32.6, 25.4, 24.8, 24.7, 13.8; IR (KBr) *v*: 3286, 3079, 2934, 2857, 1651, 1561, 1460, 1383, 1341, 1286, 1226, 1159, 1136 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₃F₄N₄O₃S (M+H)⁺ 653.0565, found 653.0570.

N-[1-(3-Chlorophenyl)-2-(cyclohexylamino)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5d) White solid, yield 40%, m.p. 119—123 °C; ¹H NMR

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(CDCl₃, 400 MHz) δ : 7.72—7.00 (m, 6H, ph-H), 6.09 (s, 1H, COCH), 6.13—5.87 (m, 1H, OCF₂CF₂H), 5.43 (d, J = 6.4 Hz, 1H, CONH), 3.89 — 3.83 (m, 1H, cyclohexyl-H), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.02— 1.12 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.9, 161.9, 161.1, 142.6, 140.4, 137.4, 135.1, 134.7, 132.1, 131.0, 130.5, 130.2, 129.8, 128.3, 65.1, 49.5, 32.7, 25.3, 24.8, 24.7, 14.1; IR (KBr) v: 3282, 3075, 2919, 2857, 1654, 1568, 1458, 1380, 1285, 1220, 1151, 1135, 1114 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₃-F₄N₄O₃S (M+H)⁺ 653.0565, found 653.0570.

N-[2-(Cyclohexylamino)-1-(2-nitrophenyl)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5e) White solid, yield 25%, m.p. 189—191 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.97—7.28 (m, 6H, ph-H), 6.57 (s, 1H, COCH), 6.10—5.84 (m, 1H, OCF₂CF₂H), 5.61 (d, J = 6.4 Hz, 1H, CONH), 3.87 — 3.84 (m, 1H, cyclohexyl-H), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.05— 1.09 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.3, 161.1, 160.9, 149.8, 142.5, 140.8, 137.4, 133.2, 132.1, 131.0, 130.8, 127.4, 125.0, 60.6, 49.6, 32.4, 25.3, 24.8, 24.6, 13.7; IR (KBr) v: 3302, 3076, 2936, 2857, 1650, 1561, 1527, 1460, 1349, 1286, 1227,1132 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₂F₄N₅O₅S (M+H)⁺ 664.0806, found 664.0815.

N-[2-(Cyclohexylamino)-1-(3-nitrophenyl)-2-oxoethyl]-N-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5f) White solid, yield 40%, m.p. 214–216 °C; ¹H NMR $(CDCl_3, 400 \text{ MHz}) \delta$: 8.19 (d, J=7.2 Hz, 1H, ph-H), 8.14(s, 1H, ph-H), 7.67-7.31 (m, 4H, ph-H), 6.19 (s, 1H, COCH), 5.86–6.18 (m, 1H, OCF₂CF₂H), 5.53 (d, J=8.4 Hz, 1H, CONH), 3.91-3.86 (m, 1H, cyclohexyl-H), 2.90 (s, 3H, thiadiazolyl-CH₃), 2.04-1.05 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.7, 162.0, 161.1, 148.1, 142.7, 140.3, 137.2, 136.0, 135.1, 132.2, 131.3, 130.0.4, 125.3, 124.2, 64.7, 49.5, 32.6, 25.3, 24.7, 24.6, 14.0; IR (KBr) v: 3264, 3079, 2933, 2855, 1649, 1561, 1536, 1457, 1349, 1286, 1253, 1224, 1125 cm^{-1} ; HRMS calcd for $C_{26}H_{23}C_{12}F_4N_5O_5S$ (M+H)⁺ 664.0806, found 664.0812.

N-[2-(Cyclohexylamino)-1-(4-nitrophenyl)-2-oxoethyl]-N-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5g) White solid, yield 34%, m.p. 179–181 °C; ¹H NMR (CDCl₃, 400 MHz) δ: 8.16 (d, J=7.6 Hz, 2H, ph-H), 7.43 (d, J=8.0 Hz, 2H, ph-H), 7.35 (s, 1H, ph-H), 7.12 (s, 1H, ph-H), 6.15 (s, 1H, COCH), 6.12-5.86 (m, 1H, OCF₂CF₂H), 5.48 (d, J=7.6 Hz, 1H, CONH), 3.88-3.83 (m, 1H, cyclohexyl-H), 2.89 (s, 3H, thiadiazolyl-CH₃), 2.03–1.05 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) *δ*: 166.5, 162.1, 161.1, 148.2, 142.8, 140.1, 140.0, 137.4, 132.0, 131.3, 123.9, 65.0, 49.6, 32.7, 25.3, 24.8, 24.7, 14.1; IR (KBr) v: 3325, 3077, 2932, 2857, 1650, 1548, 1459, 1347, 1280, 1214, 1161, 1133 cm^{-1} ; HRMS calcd for $C_{26}H_{23}C_{12}F_4N_5O_5S$ (M+H)⁺ 664.0806, found 664.0818.

N-[2-(Cyclohexylamino)-2-oxo-1-*m*-tolylethyl]-*N*-(3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl)-4methyl-1,2,3-thiadiazole-5-carboxamide (5h) White solid, yield 24%, m.p. 138—140 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.52—6.88 (m, 6H, ph-H), 6.14 (s, 1H, COCH), 6.12—5.86 (m, 1H, OCF₂CF₂H), 5.37 (d, *J*=8.0 Hz, 1H, CONH), 3.88—3.82 (m, 1H, cyclohexyl-H), 2.89 (s, 3H, thiadiazolyl-CH₃), 2.25 (s, 3H, ph-CH₃), 2.01— 1.10 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.5, 161.5, 161.0, 142.3, 140.8, 139.1, 137.7, 132.6, 132.2, 131.0, 130.6, 130.3, 128.9, 127.4, 65.6, 49.3, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) *v*: 3274, 3078, 2931, 2856, 1651, 1559, 1458, 1385, 1345, 1288, 1225, 1120 cm⁻¹; HRMS calcd for C₂₇H₂₆C₁₂F₄N₄O₃S (M+H)⁺ 633.1112, found 633.1118.

N-[2-(Cyclohexylamino)-2-oxo-1-p-tolylethyl]-N-(3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl)-4-White methyl-1,2,3-thiadiazole-5-carboxamide (5i) solid, yield 38%, m.p. 125-127 °C; ¹H NMR (CDCl₃, 400 MHz) δ: 7.35 (s, 1H, ph-H), 7.12 (s, 1H, ph-H), 7.08 (d, J=8.0 Hz, 2H, ph-H), 6.98 (d, J=8.0 Hz, 2H, ph-H), 6.13 (s, 1H, COCH), 6.13-5.86 (m, 1H, OCF₂CF₂H), 5.36 (d, J=8.0 Hz, 1H, CONH), 3.87-3.83 (m, 1H, cyclohexyl-H), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.30 (s, 3H, ph-CH₃), 2.01-1.01 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.6, 161.5, 161.0, 142.3, 140.9, 139.8, 137.8, 132.2, 130.6, 131.0, 130.2, 129.7, 65.5, 49.3, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) v: 3321, 3379, 2938, 2956, 1653, 1562, 1525, 1460, 1392, 1338, 1289, 1235, 1158, 1126 cm⁻¹; HRMS calcd for $C_{27}H_{26}C_{12}F_4N_4O_3S$ (M + H) + 633.1112, found 633.1118.

N-[2-(Cyclohexylamino)-1-(2-fluorophenyl)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5j) Light yellow oil, yield 36%; ¹H NMR (CDCl₃, 400 MHz) δ: 7.32—6.98 (m, 4H, ph-H), 6.64 (s, 2H, ph-H), 6.42 (s, 1H, COCH), 6.15—5.85 (m, 1H, OCF₂CF₂H), 5.54 (d, *J*=5.2 Hz, 1H, CONH), 3.89—3.84 (m, 1H, cyclohexyl-H), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.25 (s, 3H, ph-CH₃), 2.04—1.06 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ: 167.1, 161.9, 159.5, 146.2, 142.5, 140.7, 137.4, 132.8, 131.9, 131.8, 131.0, 130.8, 130.5, 124.7, 58.9, 49.5, 32.6, 25.3, 24.8, 24.6, 13.8; IR (KBr) *v*: 3292, 3078, 2935, 2858, 1653, 1563, 1460, 1286, 1215, 1166, 1131 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₂F₅N₄O₃S (M+H)⁺ 637.0861, found 637.0866.

N-[2-(Cyclohexylamino)-1-(3-fluorophenyl)-2-oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5k) White solid, yield 36%, m.p. 162—164 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.30—7.25 (m, 3H, ph-H), 7.06— 7.20 (t, *J*=8.0 Hz, 1H, ph-H), 6.94—6.92 (m, 2H, ph-H), 6.10 (s, 1H, COCH), 6.13—5.87 (m, 1H, OCF₂CF₂H), 5.42 (d, *J*=8.0 Hz, 1H, CONH), 3.90—3.81 (m, 1H, cyclohexyl-H), 2.89 (s, 3H, thiadiazolyl-CH₃), 2.02— 1.03 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.0, 163.9, 161.8, 161.5, 161.0, 142.5, 140.5, 137.5, 135.2, 135.1, 132.1, 130.9, 130.7, 130.6, 126.0, 117.4, 117.2, 116.7, 116.5, 65.1, 49.4, 32.7, 25.3, 24.8, 24.6, 14.0; IR (KBr) v: 3303, 3090, 2936, 2854, 1645, 1560, 1460, 1390, 1352, 1288, 1167, 1124 cm⁻¹; HRMS calcd for C₂₆H₂₃C₁₂F₅N₄O₃S (M+H)⁺ 637.0861, found 637.0862.

N-{2-(Cvclohexvlamino)-2-oxo-1-[3-(trifluoromethyl)phenyl]ethyl}-N-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (51) White solid, yield 36%, m.p. 164-166 °C; ¹H NMR (CDCl₃, 400 MHz) δ: 7.60–7.26 (m, 6H, ph-H), 6.20 (s, 1H, COCH), 6.12-5.87 (m, 1H, OCF₂CF₂H), 5.46 (d, J=8.0 Hz, 1H, CONH), 3.92-3.83 (m, 1H, cyclohexyl-H), 2.90 (s, 3H, thiadiazolyl-CH₃), 2.05–1.03 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) *δ*: 166.9, 162.0, 161.0, 142.6, 140.3, 137.2, 134.0, 133.5, 132.2, 131.7, 131.4, 131.1, 129.6, 127.3, 127.2, 126.2, 124.7, 121.9, 65.0, 49.5, 32.7, 32.6, 25.3, 24.7, 24.6, 14.0; IR (KBr) v: 3292, 3081, 2936, 2857, 1651, 1559, 1459, 1332, 1288, 1222, 1171, 1131 cm^{-1} ; HRMS calcd for $C_{27}H_{23}C_{12}F_7N_4O_3S$ (M+H)⁺ 687.0829, found 687.0832.

N-{2-(Cyclohexylamino)-2-oxo-1-[4-(trifluoromethyl)phenyl]ethyl}-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5m) White solid, yield 60%, m.p. 176—178 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.57—6.96 (m, 6H, ph-H), 6.15 (s, 1H, COCH), 6.12—5.86 (m, 1H, OCF₂CF₂H), 5.42 (d, *J*=8.0 Hz, 1H, CONH), 3.91—3.82 (m, 1H, cyclohexyl-H), 2.89 (s, 3H, thiadiazolyl-CH₃), 2.03—1.03 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.8, 161.9, 161.1, 142.6, 140.3, 137.4, 136.9, 132.1, 132.0, 131.7, 131.1, 130.7, 125.9, 125.8, 124.7, 122.0, 65.2, 49.5, 32.7, 25.3, 24.8, 24.7, 14.0; IR (KBr) *v*: 3293, 3082, 2939, 2858, 1649, 1553, 1461, 1328, 1232, 1127,1113 cm⁻¹; HRMS calcd for C₂₇H₂₃C₁₂F₇N₄O₃S (M+H)⁺ 687.0829, found 687.0828.

N-[2-(Cyclohexylamino)-1-(3-hydroxyphenyl)-2oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5n) White solid, yield 39%, m.p. 191—194 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.52—6.65 (m, 6H, ph-H), 6.07 (s, 1H, COCH), 6.13—5.86 (m, 1H, OCF₂CF₂H), 5.47 (d, *J*=8.0 Hz, 1H, CONH), 5.31 (s, 1H, OH), 3.88—3.81 (m, 1H, cyclohexyl-H), 2.87 (s, 3H, thiadiazolyl-CH₃), 2.05—1.03 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.7, 160.8, 159.8, 142.5, 140.6, 140.2, 138.9, 137.7, 134.3, 132.3, 132.0, 130.9, 130.4, 122.5, 65.8, 49.4, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) *v*: 3278, 3150, 2935, 2857, 1645, 1553, 1459, 1355, 1285, 1231, 136 cm⁻¹; HRMS calcd for C₂₆H₂₄C₁₂F₄N₄O₄S (M+H)⁺ 635.0904, found 635.0906.

N-[2-(Cyclohexylamino)-1-(4-hydroxyphenyl)-2oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (50) White solid, yield 24%, m.p. 123—125 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.52—6.65 (m, 6H, ph-H), 6.13 (s, 1H, COCH), 6.13—5.87 (m, 1H, OCF₂CF₂H), 5.63 (s, 1H, OH), 5.44 (d, J=8.0 Hz, 1H, CONH), 3.88—3.81 (m, 1H, cyclohexyl-H), 2.87 (s, 3H, thiadiazolyl-CH₃), 2.00—1.02 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.6, 162.0, 161.1, 148.1, 142.7, 140.3, 137.3, 136.0, 135.1, 132.2, 131.3, 130.0, 125.3, 124.3, 64.8, 49.5, 32.6, 25.3, 24.7, 24.6, 14.0; IR (KBr) v: 3271, 3086, 2933, 2856, 1648, 1560, 1518, 1459, 1384, 1351, 1287, 1216, 1127 cm⁻¹; HRMS calcd for C₂₆H₂₄C₁₂F₄N₄O₄S (M+H)⁺ 635.0904, found 635.0914.

N-[2-(Cyclohexylamino)-1-(2-methoxyphenyl)-2oxoethyl]-*N*-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5p) White solid, yield 33%, m.p. 144—145 °C; ¹H NMR (CDCl₃, 400 MHz) δ : 7.30—6.79 (m, 6H, ph-H), 6.43 (s, 1H, COCH), 6.10—5.84 (m, 1H, OCF₂CF₂H), 5.38 (d, *J*=8.0 Hz, 1H, CONH), 3.93— 3.88 (m, 1H, cyclohexyl-H), 3.80 (s, 3H, OCH₃), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.01—1.02 (m, 10H, cyclohexyl-H); ¹³C NMR (CDCl₃, 400 MHz) δ : 167.8, 157.2, 142.0, 141.2, 137.8, 131.7, 131.3, 131.1, 120.9, 120.8, 110.5, 107.4, 60.3, 55.3, 49.2, 32.8, 25.4, 24.8, 24.7, 13.9; IR (KBr) *v*: 3282, 3074, 2930, 2857, 1654, 1557, 1494, 1461, 1386, 1289, 1253, 1224, 1112 cm⁻¹; HRMS calcd for C₂₇H₂₆C₁₂F₄N₄O₄S (M+H)⁺ 649.1061, found 649.1065.

N-[2-(Cyclohexylamino)-1-(3-methoxyphenyl)-2oxoethyl]-N-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carboxamide (5q) White solid, yield 27%, m.p. 116-118 °C; ¹H NMR (CDCl₃, 400 MHz) δ: 7.52—6.61 (m, 6H, ph-H), 6.13 (s, 1H, COCH), 6.19-5.92 (m, 1H, OCF₂CF₂H), 5.44 (d, J=8.0 Hz, 1H, CONH), 3.89-3.82 (m, 1H, cyclohexyl-H), 3.69 (s, 3H, OCH₃), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.01-1.03 (m, 10H, cyclohexyl-H); 13 C NMR (CDCl₃, 400 MHz) δ : 167.2, 160.0, 142.4, 140.7, 137.7, 134.0, 132.1, 130.8, 130.1, 122.6, 115.8, 115.3, 65.7, 55.3, 49.4, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) v: 3281, 3076, 2931, 2855, 1650, 1585, 1561, 1459, 1381, 1355, 1281, 1228, 1156, 1114 cm⁻¹; HRMS calcd for $C_{27}H_{26}C_{12}F_4N_4O_4S$ (M + H) ⁺ 649.1061, found 649.1063.

N-[2-(Cyclohexylamino)-1-(4-methoxyphenyl)-2oxoethyl]-N-[3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl]-4-methyl-1,2,3-thiadiazole-5-carbox**amide (5r)** White solid, yield 74%, m.p. 152–155 °C; ¹H NMR (CDCl₃, 400 MHz) δ: 7.52–6.77 (m, 6H, ph-H), 6.13 (s, 1H, COCH), 6.13-5.87 (m, 1H, OCF₂CF₂H), 5.38 (d, J=8.0 Hz, 1H, CONH), 3.88-3.3 (m, 1H, cyclohexyl-H), 3.77 (s, 3H, OCH₃), 2.88 (s, 3H, thiadiazolyl-CH₃), 2.01-1.05 (m, 10H, cyclohexyl-H); 13 C NMR (CDCl₃, 400 MHz) δ : 167.7, 161.4, 160.9, 160.4, 142.3, 140.9, 137.8, 134.0, 132.2, 131.7, 130.7, 124.5, 114.3, 107.4, 65.2, 55.3, 49.2, 32.8, 25.4, 24.8, 24.7, 14.0; IR (KBr) v: 3290, 3076, 2935, 2856, 1647, 1555, 1512, 1460, 1350, 1257, 1128 cm⁻¹; HRMS calcd for $C_{27}H_{26}C_{12}F_4N_4O_4S$ (M + H) ⁺ 649.1061, found 649.1060.

Biological screening

Fungicide screening Preliminary screening was conducted by fungi growth inhibition method according to the reference using potato dextrose agar (PDA) as cultivation medium.² A stock solution of each compound was prepared at 500 μ g•mL⁻¹ using sterilized water containing 2 drops of N,N-dimethylformamide (DMF) as a solvent, then 1 mL of the stock solution was transferred into a 10 cm diameter of Petri dish, 9 mL of PDA was then added to prepare the plate containing 50 μ g•mL⁻¹ of the test compound. Before the plate solidification, the PDA was thoroughly mixed by turning around the Petri dish in the sterilized operation desk 5 times to scatter the compound in PDA evenly. Then, 4 mm of diameter of fungi cake was inoculated on the plate and cultured in the culture tank at 24–26 $^{\circ}$ C. The diameter of fungi spread was measured 2 d later. Growth inhibition was then calculated using corresponding control. Azoxystrobin and tricyclazole were used as positive controls. Fungi used in this study included Alternaria solani (AS), Botrytis cinerea (BC), Cercospora arachidicola (CA), Gibberella zeae (GZ), Phytophthora infestans (Mont) de Bary (PI), Physalospora piricola (PP), Pellicularia sasakii (PS), Sclerotinia sclerotiorum (SS), and Rhizoctonia cerealis (RC).

Systemic acquired resistance screening Systemic acquired resistance of the target compounds was detected using tobacco against the tobacco mosaic virus (TMV) system as described in Ref. 2. The induction activity was evaluated using the antivirus inhibition ratio, which was calculated by the average number of viral inflammations on the inoculated leaves with corresponding control accordingly. Tiadinil (TDL) and ribavirin were chosen as positive control and negative control, respectively, and all compounds tested were conducted at 100 and 50 μ g[•] mL⁻¹, respectively. Before the test of induction activity of systemic acquired resistance, the antivirus activity against TMV was conducted by half leaf juice robbing methods according to reference.²

Protective effects of the target compounds against TMV *in vivo* Healthy fresh tobacco leaves growing at six-leaf age were selected for the tests. The compound solution was smeared on the whole leaves, and then the leaves were dried in greenhouse. After 12 h, TMV at a concentration of $5.88 \times 10^{-2} \,\mu \text{g} \cdot \text{mL}^{-1}$ was inoculated on the upper three leaves using the conventional juice-robbing method, and the solvent was smeared on the lower three leaves as a control. The local lesion numbers were then recorded 2—3 d after inoculation. For each compound, three repetitions were conducted. Most compounds tested were conducted at 500 and 100 $\mu \text{g} \cdot \text{mL}^{-1}$, respectively.

Inactivate effect of the target compounds against TMV *in vivo* Healthy fresh tobacco leaves growing at six-leaf age were selected for the tests. TMV virus at a concentration of $5.88 \times 10^{-2} \,\mu\text{g} \cdot \text{mL}^{-1}$ was inhibited by mixing with the target compound solution at the same volume for 30 min. Then the mixture was inoculated on the upper three leaves using the conventional juice-robbing method, and the solvent was smeared on the lower three leaves as a control. The local lesion numbers were then recorded 2-3 d after inoculation. For each compound, three repetitions were conducted. All compounds tested were conducted at 500 and 100 µg/mL, respectively.

Curative effect of target compounds on TMV *in vivo* Healthy fresh tobacco leaves growing at six-leaf age were selected for the tests. TMV at a concentration of $5.88 \times 10^{-2} \ \mu g \cdot m L^{-1}$ was inoculated on the whole leaves using the conventional juice-robbing method. After the leaves were dried in the greenhouse, the compound solution was smeared on the upper three leaves, and the solvent was smeared on the lower three leaves as control. The local lesion numbers were then recorded 2—3 d after inoculation. For each compound, three repetitions were conducted. All compounds tested were conducted at 500 and 100 $\mu g \cdot m L^{-1}$, respectively.

The activities of protection, inactivation, and curative effects against TMV were calculated by the average number of viral inflammations on the inoculated leaves with the corresponding control according to the following equation:

$$Y = \frac{\mathrm{CK} - A}{\mathrm{CK}} \times 100\%$$

where Y is the antivirus inhibition ratio (protection, inactivation, and curative effects *in vivo*) (%), CK is the average number of viral inflammations on the control leaves *in vivo*, and A is the average number of viral inflammations on the target compound treated leaves *in vivo*.

Larvicidal activity against Culex pipiens pallens. The larvicidal activity of target compounds 5a-5r was evaluated against mosquito *Culex pipiens pallens* according to the reported procedure.¹⁶ The test solutions were prepared by dissolving 2 mg of the compounds in 10 mL of acetone and adding distilled water respectively to prepare 200 μ g•mL⁻¹ of mother solution; the 2 $\mu g \cdot mL^{-1}$ working solution was prepared by diluting 1 mL of mother solution with 89 mL of water and 10 mL of feeding solution into a 100 mL breaker, and 10 forth-instar of Culex pipiens pallens larvae were transfered into the beaker. Thereafter, the mosquito larvae in the beakers were cultured in the standard conditioned rooms at 25 °C with humidity of 80% for 8 d. The results are expressed as percentage mortality by comparision with the corresponding CK. For comparative purposes, tebufenozide was tested under the same condition.

Stomach toxicity against *Plutella xylostella* L. The stomach toxicities of the target compounds 5a-5r against *Plutella xylostella* were tested by the leaf dip method using the reported procedure.¹⁷ A stock solution of each test sample was prepared in dimethylformamide at a concentration of 200 mg•L⁻¹ and then diluted to the

required concentration with water containing TW-20. Leaf disks (6 cm \times 2 cm) were cut from fresh cabbage leaves and then were dipped into the test solution for 3 s. After air-drying, the treated leaf disks were placed individually into glass tubes. Each dried treated leaf disk was infested with seven third-instar diamondback moth larvae. Percentage mortalities were evaluated 3 d after treatment. Leaves treated with water and dimethylformamide were provided as controls. Each treatment was repeated for three times.

Results and discussion

Synthesis

This study used U-4CR as the parallel synthesis method, and the target compounds were proved to be easily synthesized and purified. Following the method developed in our previous study,^{4c,15} an amine component and an aldehyde component were first mixed to get an intermediate imine, and then a carbonyl component was added into the mixture for certain time of intervals before the mixing with an isocyanide component. The amount of methanol as solvent added to the reaction mixture was as small as possible, just enough to make the agitation effective. After all components were mixed, products usually precipitated into solvent within 1 h. To obtain high yields, solvent was removed under reduced pressure and then the products were purified by column chromatography on silica gel using ethyl acetate and petroleum ether (60—90 °C) at $1 \div 3$ ($V \div V$) as an eluent.

Biological activities

Fungicidal activity

The newly synthesized compounds 5a-5r were tested for their fungicidal activity against following fungi, which represented the pathogen of typical diseases occurring in the Chinese agricultural ecosystem. AS (Alternaria solani), BC (Botrytis cinerea), CA (Cercospora arachidicola), GZ (Gibberella zeae), PI (Phytophthora infestans (Mont.) de Bary), PP (Physalospora piricola), PS (Pellicularia sasakii), SS (Sclerotinia sclerotiorum) and RC (Rhizoctonia cerealis). The results of Table 1 indicated that, most compounds showed lower fungicide activity at 50 µg/mL compared to the two positive controls azoxystrobin and tricyclazole. Some compounds had good fungicide activity, for example, compounds 5b, 5e and 5m exhibited 72.7%, 72.7% and 63.6% of fungicidal activity against SS, respectively; the activity of compounds 5a, 5d, 5i and 5j against PP were of 64.1%, 71.8%, 61.5% and 71.8%, respectively, which were higher than that of tricyclazole; the inhibition percentages of 5j to RC and AS were 64.5% and 61.9%, respectively.

Table 1 The antifungal activities of title compounds *in vitro* at 50 μ g•mL⁻¹

- F-8									
Compound	GZ	SS	PI	PP	BC	RC	PS	AS	CA
5a	0	45.5	0	64.1	18.2	41.9	14.1	33.3	14.3
5b	0	72.7	23.8	46.2	18.2	41.9	14.1	4.8	21.4
5c	0	54.6	0	33.3	0	32.3	14.1	9.5	0
5d	0	36.4	14.3	71.8	27.3	48.4	14.1	33.3	0
5e	0	72.7	0	20.5	4.6	0	42.3	4.8	0
5f	0	54.6	0	20.5	36.4	22.6	42.3	33.3	0
5g	0	45.5	0	38.5	27.3	32.3	4.2	33.3	14.3
5h	0	36.4	19.1	38.5	0	22.6	42.3	0	14.3
5i	23.8	45.5	23.8	61.5	0	9.7	14.1	47.6	21.4
5ј	0	0	23.8	71.8	27.3	64.5	21.1	61.9	42.9
5k	0	45.5	9.5	28.2	27.3	41.9	0	47.6	0
51	0	36.4	0	53.9	4.6	29.0	14.1	14.3	0
5m	0	63.6	0	33.3	4.6	32.3	42.3	33.3	0
5n	0	27.3	0	7.7	4.6	16.1	14.1	0	0
50	0	18.2	0	33.3	0	48.4	7.0	14.3	0
5р	0	54.6	0	59.0	4.6	0	42.3	42.9	21.4
5q	0	27.3	23.8	28.2	13.6	16.1	0	23.8	0
5r	0	27.3	0	33.3	0	22.6	0	33.3	0
Azoxystrobin	40.0	100	88.9	85.0	21.2	100	69.74	63.1	80.0
Tricyclazole	24.0	100	29.6	25.0	36.8	65.9	29.0	5.3	13.3

Antivirus activity

Tiadinil, ribavirin and ninamycin were used as positive controls. The results of activity screening by half-leaf juice-robbing method against TMV indicated that (Table 2) most compounds showed certain degree of direct inhibition activities against TMV *in vitro* with results no more than 50% except **5j**. However, compounds **5d**, **5k**, **5m** and **5o**, **5j** had direct inhibition activities of 45%, 46%, 43% and 45%, 53% at 100 $\mu g \cdot m L^{-1}$, respectively. These were as twice as that of ribavirin. However, the activities at 500 $\mu g \cdot m L^{-1}$ were lower. Only compound **5j** had high activity at both 500 and 100 $\mu g \cdot m L^{-1}$ with inhibition above 50%, and they are significantly higher than those of the positive control agents ribavirin and tiadinil.

The antivirus activity of all title compounds including protective, inactivate and curative effect, and the inductive activities were also evaluated. The results listed in Table 2 indicated that, most of the compounds had good inactivation and curative activity against tobacco mosaic virus. Among these, compounds **5a**—**5d** and **5o** had inactivation activities of 66%, 87%, 73%, 70% and 71% at 500 µg•mL⁻¹, respectively, and 49%, 59%, 51%, 48% and 52% at 100 µg•mL⁻¹, respectively. These were better than that of the positive control agent ribavirin; compounds **5g**, **5h**, **5j**, **5m** and **5o** had good curative activities of 65%, 59%, 72% and 72% at 500 µg•mL⁻¹, respectively, and 43%, 52%, 47%, 56%

			(Protective			(I J
Compound	Concentration/ $(\mu g \cdot mL^{-1})$	(Half leaf	(Protective	(Inactivate	(Curative	(Inductive
7 -		activity \pm SD)/%	$activity \pm SD)/\%$	activity \pm SD)/%	activity \pm SD)/%	activity \pm SD)/9
5a	500	36 ± 4	45 ± 7	66 ± 5	55 ± 7	65 ± 1^a 57 ± 7^b
7 1.	100	36 ± 5	22 ± 7	49 ± 8	39 ± 3	
5b	500	31 ± 6	48 ± 3	87±3	50 ± 8	38 ± 6^a 7 ± 8^b
-	100	26 ± 5	16 ± 7	59 ± 4	50 ± 3	
5c	500	24 ± 5	22 ± 3	73 ± 8	59 ± 1	30 ± 5^a 64 ± 3^b
- 1	100	27 ± 6	13 ± 6	51 ± 3	41 ± 9	
5d	500	45 ± 2	55 ± 8	70 ± 8	47 ± 6	61 ± 4^a 22 ± 3^b
_	100	45 ± 3	42 ± 8	48 ± 4	14±5	
5e	500	28 ± 5	34 ± 4	58 ± 5	48 ± 8	36 ± 6^a
-0	100	22 ± 2	16 ± 7	48 ± 4	41 ± 9	25 ± 6^b
5f	500	38±3	36±5	49±7	11 ± 4	38 ± 5^a
	100	37 ± 3	29±7	23 ± 6	7±3	5 ± 4^{b}
5g	500	34±5	56±7	55±8	65±9	12 ± 8^a
	100	35 ± 3	27 ± 0	36±7	43±7	6 ± 3^b
5h	500	37 ± 3	49 ± 2	43±4	59±7	64 ± 3^a
	100	33±3	9±9	11 ± 8	52 ± 6	49 ± 8^{b}
5i	500	35 ± 5	58±5	24 ± 6	38 ± 4	57 ± 1^{a}
	100	33 ± 3	45 ± 5	13 ± 2	21 ± 7	56 ± 4^b
5ј	500	51±9	60 ± 4	9±7	59 ± 3	54 ± 10^{a}
	100	53 ± 6	33 ± 8	14 ± 5	47 ± 9	60 ± 3^b
5k	500	37 ± 5	51 ± 2	43 ± 9	30 ± 5	28 ± 4^a
	100	46 ± 1	35 ± 7	12 ± 1	10.37 ± 3	72 ± 4^{b}
51	500	27 ± 1	76 ± 2	15 ± 8	61 ± 5	41 ± 3^a
	100	34 ± 3	59 ± 6	14 ± 5	37 ± 9	45 ± 2^b
5m	500	20 ± 3	58 ± 5	47±7	72 ± 1	75 ± 5^{a}
	100	43±3	51 ± 3	11 ± 7	56 ± 3	75 ± 3^{b}
5n	500	37 ± 3	41 ± 1	50 ± 6	47±7	59 ± 1^{a}
	100	33 ± 3	337 ± 2	10 ± 8	13±5	67 ± 4^b
50	500	37 ± 6	57 ± 3	71 ± 3	72 ± 3	30 ± 4^a
	100	45 ± 3	46 ± 10	52 ± 3	43 ± 8	15 ± 7^{b}
5p	500	43 ± 1	34 ± 6	34 ± 6	37 ± 9	65 ± 5^a
	100	34 ± 8	22 ± 7	11 ± 3	30 ± 3	14 ± 6^{b}
5q	500	16 ± 1	72 ± 4	62 ± 7	34 ± 6	15 ± 2^{a}
	100	29 ± 10	48 ± 5	12 ± 7	16±3	45 ± 8^{b}
5r	500	26 ± 8	69 ± 5	13 ± 6	49 ± 2	83 ± 4^{a}
	100	25 ± 2	59 ± 3	8±5	33±8	67 ± 3^{b}
Ribavirin	500	32 ± 1	65 ± 3	48 ± 4	52±7	20 ± 5^a
	100	20 ± 2	50 ± 8	26 ± 4	30 ± 4	65 ± 8^b
Ninamycin	500	49 ± 4	73 ± 5	63±9	55±8	46 ± 5^a
	100	39±6	56 ± 3	43±9	41 ± 8	63 ± 5^b
Tiadinil	500	32±3	66 ± 3	56±7	50 ± 9	57 ± 2^a
	100	36 ± 4	55±4	47±9	36 ± 6	65 ± 4^{b}

^{*a*} 50 μ g•mL⁻¹ of concentration; ^{*b*} 100 μ g•mL⁻¹ of concentration.

and 43% at 100 µg/mL, respectively. These were also better than the positive control agent ribavirin. As for the protection effect and induction activities, most of them had lower activities as compared with the positive control. However, some compounds stood out, compounds 51, 5q and 5r had good protection activities of 76%, 72% and 69% at 500 μ g•mL⁻¹, respectively, and 58%, 47% and 59% at 100 μ g•mL⁻¹, respectively. Compounds 5m, 5n and 5r had good induction activities of 75%, 59% and 83% at 500 μ g•mL⁻¹, respectively, and 75%, 67% and 67% at 100 μ g•mL⁻¹, respectively. These were approximately equal to or higher than that of the positive controls. We observed no phytotoxicity to tobacco during the course of experiments. No significant structure-activity relationship was observed from these studies. This indicated that the whole molecular structure played an important role in these activities rather than one moiety in the molecule.

Larvicidal activity

Table 3 showed the larvicidal activities of compounds 5a - 5r and positive control tebufenozide against *Plutella xylostella* L. and *Culex pipiens pallens*. The results indicate that most of the compounds exhibited different larvicidal activities against *Plutella xylostella* L. and *Culex pipiens pallens*. On the whole, the larvicidal activities of the *meta*-substituted and *para*-substituted benzoyl against *Plutella xylostella* L. were better than those of *ortho*-substituted and nonsubstituted compounds. For example, compounds **5b**, **5d**, **5f**, **5g**, **5n**, **5o** and **5r** displayed higher larvicidal activities against *Plutella xylostella* L. than compounds **5a**, **5c**, **5e**, **5j** and **5p** at 200 μ g•mL⁻¹. Compounds **5b**, **5f**, **5g**, **5n**, **5o** and **5r** had good larvicidal activity against *Plutella xylostella* L., which were approximately equal to that of the commercialized product tebufenozide at 200 μ g•mL⁻¹. Compounds **5o** and **5q** also exhibited excellent larvicidal activities against *Culex pipiens pallen* among the test compounds, which had 100% mortality at 2 μ g•mL⁻¹.

Conclusion

In summary, a series of new 4-methyl-1,2,3-thiadiazole derivatives containing active substructure of 3,5-dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl were conveniently synthesized and easily purified via 4 component Ugi reaction. Bioassay results indicated that all of these title compounds showed a certain degree of fungicidal activity at 50 μ g•mL⁻¹. The larvicidal activities of the benzoyl meta-substituted or para-substituted compounds against Plutella xylostella L. were better than those of benzoyl ortho-substituted or non-substituted compounds. Compounds 50 and 5g exhibited excellent larvicidal activities against Culex pipiens pallens with 100% mortality at 2 μ g•mL⁻¹. Most of the compounds presented different extent of anti-TMV activity with different mode of action.

Compound -	Plutella xylo	ostella L.	Culex pipiens pallens		
	Concentration/($\mu g \cdot mL^{-1}$)	Larvicidal activity/%	Concentration/($\mu g \cdot mL^{-1}$)	Larvicidal activity/%	
5a	200	7.89	2	20	
5b	200	22.86	2	20	
5c	200	7.69	2	40	
5d	200	17.50	2	20	
5e	200	5.41	2	0	
5f	200	21.05	2	10	
5g	200	23.68	2	0	
5h	200	5.26	2	10	
5i	200	7.89	2	0	
5j	200	15.00	2	20	
5k	200	nd^a	2	30	
51	200	2.63	2	20	
5m	200	15.79	2	20	
5n	200	22.92	2	40	
50	200	21.62	2	100	
5p	200	10.81	2	nd	
5q	200	7.90	2	100	
5r	200	20.00	2	20	
Tebufenozide	200	26.30	2	100	

Table 3 Insecticidal activities of of the title compounds 5a-5r

^{*a*} nd: not detected.

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References

- (a) Zhan, P.; Liu, X. Y.; Fang, Z. J.; Li, Z. Y.; Pannecouque, C.; Clercq, E. D. *Eur. J. Med. Chem.* **2009**, *44*, 4648.
 (b) Zhan, P.; Liu, X. Y.; Li, Z. Y.; Fang, Z. J.; Li, Z.; Wang, D. F.; Pannecouque, C.; Clercq, E. D. *Bioorg. Med. Chem.* **2009**, *17*, 5920.
 - (c) Pannecouque, C.; Szafarowicz, B.; Volkova, N.; Bakulev,V.; Dehaen, W.; Mély, Y.; Daelemans, D. Antimicrob.Agents Chemother. 2010, 54, 1461.
- Fan, Z. J.; Shi, Z. G.; Zhang, H. K.; Liu, X. F.; Bao, L. L.; Ma, L.; Zuo, X.; Zheng, Q. X.; Mi, N. J. Agric. Food Chem. 2009, 57, 4279.
- Fan, Z. J.; Ai, Y. W.; Chen, J. Y.; Wang, H. Y.; Liu, F. L.;
 Bao, L. L.; Shi, Z. G. J. Sichuan Normal Univ. (Nat. Sci.)
 2005, 28, 608 (in Chinese).
- 4 (a) Fan, Z. J.; Yang, Z. K.; Zhang, H. K.; Mi, N.; Wang, H.; Cai, F.; Zuo, X.; Zheng, Q. X.; Song, H. B. J. Agric. Food Chem. 2010, 58, 2630.

(b) Yang, Z. K.; Zhang, H. K., Fan, Z. J.; Mi, N.; Song, H.
B; You, J. M.; Sun, X. J.; Belskaia, N. P.; Bakulev, V. A. *Chin. J. Pestic. Sci.* 2009, *11*(1), 19 (in Chinese).
(c) Zuo, X.; Mi, N.; Fan, Z. J.; Zheng, Q. X.; Zhang, H. K.;

Wang, H.; Yang, Z. K. J. Agric. Food. Chem. 2010, 58, 2755.

(d) Wang, Z. H.; Guo, Y. Z.; Zhang, J.; Ma, L.; Song, H. B.; Fan, Z. J. J. Agric. Food Chem. **2010**, *58*, 2715.

- 5 Nombela, G.; Pascual, S.; Aviles, M.; Guillard, E.; Muniz, M. J. Econ. Entomol. 2005, 98, 2266.
- 6 Wu, M. J.; Sun, Q. M.; Yang, C. H.; Chen, D. D.; Ding, J.; Chen, Y.; Lin, L. P.; Xie, Y. Y. *Bioorg. Med. Chem. Lett.* 2007, 17, 869.
- 7 Chen, Y. F.; Zhang, C. R.; Huang, X. Plant Physiol. Commun. 2006, 42, 127 (in Chinese).

Wang, T. T.; Bing, G. F.; Zhang, X.; Qin, Z. F.; Yu, H. B.; Qin, X.; Dai, H.; Fang, J. X. *ARKIVOC* **2010**, (ii), 330.

8

1982.

- 9 Bakulev, V.; Mokrushin, V. Chem. Heterocycl. Compd. 1986, 22, 811.
- (a) Ojima, I.; Griffen, E.; Clark, J.; Breeden, S.; Summerton, L.; Barker, R.; Iddon, B.; Hall, A.; Hardy, B.; Peltason, L. *Future Med. Chem.* 2009, *1*, 401.
 (b) Zhou, Z. Z.; Huang, W.; Ji, F. Q.; Ding, M. W.; Yang, G. F. *Heteroat. Chem.* 2007, *18*, 381.
 (c) Zhou, Z. Z.; Yang, G. F. *Bioorg. Med. Chem.* 2006, *14*, 8666.
- (a) Akritopoulou-Zanze, I.; Djuric, S. W. *Heterocycles* 2007, *73*, 125.
 (b) Hulme, C.; Dietrich, J. *Mol. Diversity* 2009, *13*, 195.
 (c) Liu, N.; Liu, N. J.; Cao, S.; Shen, L.; Wu, J. J.; Yu, J. L.; Zhang, J.; Li, H.; Qian, X. H. *Tetrahedron Lett.* 2009, *50*,
- 12 Zohreh, N.; Alizadeh, A.; Bijanzadeh, H. R.; Zhu, L. G. J. Comb. Chem. 2010, 12, 497.
- 13 Banfi, L.; Riva, R.; Basso, A. Synlett 2010, 23.
- (a) Xu, Y. F.; Zhao, Z. J.; Qian, X. H.; Qian, Z. G.; Tian, W. H.; Zhong, J. J. J. Agric. Food Chem. 2006, 54, 8793.
 (b) Zhu, S.; Liu, P.; Liu, X.; Li, J.; Yuan, S.; Si, N. Pest Manag. Sci. 2008, 64, 255.
- 15 Zheng, Q. X.; Mi, N.; Fan, Z. J.; Zuo, X.; Zhang, H. K.; Wang, H.; Yang, Z. K. J. Agric. Food Chem. 2010, 58, 7846.
- 16 Chen, L.; Huang, Z. Q.; Wang, Q. M.; Shang, J.; Huang, R. Q.; Bi, F. C. J. Agric. Food Chem. 2007, 55, 2659.
- 17 Sun, R. F.; Zhang, Y. L.; Chen, L.; Li, Y. Q.; Li, Q. S.; Song, H. B.; Huang, R. Q.; Bi, F. C.; Wang, Q. M. J. Agric. Food Chem. 2009, 57, 3661.

(E1010071 Ding, W.; Zheng, G.)