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## Article

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# **Divergent Total Syntheses of Rhodomyrtosones A and B**

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**Abstract:** Herein, we report total syntheses of the tetramethyldihydroxanthene natural product rhodomyrtosone B and the related *bis*-furan  $\beta$ -triketone natural product rhodomyrtosone A. Nickel-(II)-catalyzed 1,4-conjugate addition of an  $\alpha$ -alkylidene- $\beta$ -dicarbonyl substrate was developed to access the congener rhodomyrtosone B and oxygenation of the same monoalkylidene derivative followed by cyclization was employed to obtain the *bis*-furan natural product rhodomyrtosone A.

### INTRODUCTION



Figure 1. Rhodomyrtone A and Related Natural Products

Rhodomyrtone A (1) and its isomer rhodomyrtosone B (2) are derived from the plant *Rhodomyrtus Tomentosa* found in Thailand (**Figure 1**).<sup>1,2</sup> These natural products present interesting synthetic challenges due to the presence of a highly oxygenated  $\beta$ -triketone moiety fused to an acylphloroglucinol which is present in both isomers. In rhodomyrtone A (1), the ether linkage is *para* to the acyl group, while in rhodomyrtosone B (2) it is *ortho*. Rhodomyrtosone A (3) was also isolated from the *Rhodomyrtus* genus and possesses an intriguing *bis*-furan acylphloroglucinol core.<sup>2,3</sup> Recently, the related natural product watsonianone B (4) was isolated from the plant

*Corymbia watsonia.*<sup>4</sup> Rhodomyrtone A **1** and rhodomyrtosone B (**2**) were found to be potent antibiotics against gram positive bacteria including *Staphylococcus Aureus*, methicillin-resistant *Staphylococcus Aureus* (MRSA), and several

*Streptococcus* strains (MIC = 4 and 16 µg/mL, respectively).<sup>5</sup> Watsonianone B (**4**) possesses anti-malarial properties by inhibiting the growth of chloroquine sensitive (3D7) and resistant (Dd2) strains of *Plasmodium falciparum* displaying IC<sub>50</sub> values of 0.44 and 0.29 µM, respectively.<sup>4</sup> Accordingly, these interesting structures and highly relevant biological activities make rhodomyrtosones A (**3**) and B (**2**) appealing synthetic targets. The Maier laboratory<sup>6</sup> recently achieved syntheses of compounds **1** and **2** adopting a similar strategy developed by Jauch and coworkers for the synthesis of myrtucommulone A.<sup>7</sup> In this paper, we report a strategy involving nickel (II)-catalyzed 1,4-conjugate addition to an  $\alpha$ -alkylidene- $\beta$ -dicarbonyl substrate to selectively access rhodomyrtosone B (**2**) and oxygenation of the same monoalkylidene derivative to obtain the *bis*-furan congener rhodomyrtosone A (**3**).





**RESULTS AND DISCUSSION** 

In our retrosynthetic analysis, we envisioned that rhodomyrtone A (1), rhodomyrtosone B (2), and rhodomyrtosone (3) could be synthesized from a common starting material in a divergent manner (**Figure 2**). Selective dehydrative cyclizations of intermediate **5** could be used to access both rhodomyrtone A (1) and rhodomyrtosone B (2). Intermediate **5** may arise from conjugate addition of the known acylphloroglucinol  $7^8$  to monoalkylidene **6**. In accordance with the proposed biosynthesis for rhodomyrtosone A,<sup>2</sup> natural product **3** may be obtained from acylphloroglucinol **7** and endoperoxide **8** after *bis*-furan formation. Endoperoxide **8** may arise from [4+2] cycloaddition of oxygen with a dienol intermediate that may be obtained

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via photoenolization of monoalkylidene 6 (Figure 2).<sup>9</sup>

Our synthetic efforts began with the synthesis of monoalkylidene reaction partner **6** (**Scheme 1**). Treatment of syncarpic **Scheme 1.** Synthesis of a Monoalkylidene Reaction Partner



acid  $10^6$ , isovaleraldehyde, and pyrrolidine (diethyl ether, 0 °C) afforded the Mannich product 9 (90 %). Acid-mediated elimination of 9 cleanly afforded monoalkylidene 6 (75% yield). We next examined a range of catalysts for 1,4-conjugate addition<sup>10</sup> of acylphloroglucinol 7 to enone 6. A reaction conducted without catalyst provided a 9 % yield of the Michael adduct 5 along with a significant amount of the endoperoxide byproduct 8 (stereochemistry unassigned) (Table 1, entry 1). The latter compound may be derived from [4+2] cycloaddition between the dienol tautomer of **6** and triplet oxygen (**Table 1**, entry 1) (vide infra).<sup>11,12,13</sup> In the presence of YbCl<sub>3</sub> or FeCl<sub>3</sub>, a moderate amount of **5** was isolated, albeit in low selectivity (Table 1, entries 2 and 3). Rare-earth metal triflate catalysts were also investigated; however, these reactions resulted in low yields and selectivity (**Table 1**, entries 4-6).<sup>10a</sup> We also investigated metal perchlorate salts, which have previously been used in 1,4-conjugate additions.<sup>14</sup> Cu(ClO<sub>4</sub>)<sub>2</sub>, Mg(ClO<sub>4</sub>)<sub>2</sub>, and Zn(ClO<sub>4</sub>)<sub>2</sub> did not result in significant amounts of adduct 5 (Table 1, entries 7-9). Additionally, it was found that Pd (II) catalysis was not effective for this transformation (Table 1, entry 10). Ni(ClO<sub>4</sub>)<sub>2</sub> proved to be an effective catalyst, affording a 4.6:1 ratio of compounds 5:8 in 74 % isolated yield (Table 1, entry 11). Using a 5:1 mixture of CH<sub>2</sub>Cl<sub>2</sub> and acetic acid as solvent led to an increased yield of the desired adduct 5 (Table 1, entry 12). Moreover, thorough degassing of solvent eliminated the production of endoperoxide 8 (Table 1, entry 13) and provided adduct 5 in 90 % yield. A control screen using the same catalysts as listed in Table 1 with degassed CH<sub>2</sub>Cl<sub>2</sub> was also conducted. Although degassing the solvent alleviated the formation of 8, the isolated yields remained low (Table 1, entries 2, 6-8) except when Ni(ClO<sub>4</sub>)<sub>2</sub>•6H<sub>2</sub>O was employed as catalyst. Based on the results highlighted in **Table 1**, Ni(ClO<sub>4</sub>)<sub>2</sub>•6H<sub>2</sub>O was identified as a promising catalyst for conjugate addition.<sup>15</sup>

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#### 4Å M.S. *i-*Bu OH OH OH 0 Lewis acid (0.1 equiv) CH<sub>2</sub>Cl<sub>2</sub> ΩН Ĥ rt to 40°C, 17 h OH 7 5 8 Entrv<sup>[a]</sup> 5:8<sup>[c]</sup> 5 (%) <sup>[b]</sup> Lewis-acid 1:2.7 no catalyst 1 9 2 YbCl<sub>3</sub> 1.15:1 38 3 FeCl<sub>3</sub> 1.3:1 36 4 Yb(OTf)<sub>3</sub> 1:3.3 16 5 Gd(OTf)<sub>3</sub> 1:2.4 29 6 $Lu(OTf)_3$ 1:1.7 17 7 $Cu(ClO_4)_2$ 1:4.3 6 8 $Mg(ClO_4)_2$ N/A 0 9 $Zn(ClO_4)_2$ N/A 0 10 PdCl<sub>2</sub>(PhCN)<sub>2</sub> N/A 19 74 11 Ni(ClO<sub>4</sub>)<sub>2</sub>•6H<sub>2</sub>O 4.6:1 12 Ni(ClO<sub>4</sub>)<sub>2</sub>.•6H<sub>2</sub>O (7 80 2.8:1mol%) CH<sub>2</sub>Cl<sub>2</sub>:AcOH (6:1) 13<sup>[d]</sup> 1:0 90 Ni(ClO<sub>4</sub>)<sub>2</sub>.•6H<sub>2</sub>O (7 mol%) CH2Cl2:AcOH (6:1)

 Table 1. Evaluation of the Conjugate Addition under Lewis Acidic Conditions

<sup>[a]</sup>Reactions conducted with monoalkylidene **6** (1 equiv) and acylphloroglucinol **7** (1.5 equiv) in 1 mL of CH<sub>2</sub>Cl<sub>2</sub>. <sup>[b]</sup>Yields reported after isolation by silica gel column chromatography. <sup>[c]</sup>Compound **8** was the only byproduct observed after complete consumption of starting materials. Ratio was determined after isolating both products. <sup>[d]</sup>Solvents were thoroughly degassed using the freeze-pump-thaw method.

Upon characterization by <sup>1</sup>H NMR analysis in CDCl<sub>3</sub>, we observed that the adduct **5** existed as a mixture of two atropisomers in a 2.4:1 ratio at room temperature. The tight hydrogen bond network existing within the structure may hinder free rotation generating two rotational conformers in equilibrium at room temperature (**Figure 3**).<sup>16</sup> Variable temperature NMR studies were performed to determine the coalescence temperature for **5** which was found to be 60 °C which corresponds to a rotational barrier of 15 kcal/mol.<sup>17</sup> A single X-ray crystal structure analysis of compound **5** was obtained which further highlights the strong internal hydrogen bond network between the acylphloroglucinol and vinylogous acid segments (**Figure 3**).





Scheme 2. Dehydrative Cyclization of 5 to Rhodomyrtosone B (2)



We anticipated that adduct 5 could be utilized achieve dehydrative to cyclizations to afford natural products 1 (vida supra). Under acidic and conditions. we observed exclusive formation of rhodomyrtosone B 2 via dehydrative cyclization of the phenol ortho to the acyl group (Scheme 2).<sup>6</sup> After considerable optimization, it was found that treatment of 5 in the carbocationstabilizing protic solvent hexafluoroisopropanol (HFIP)<sup>18</sup> with added trifluoroacetic acid (60 °C, 12 h) afforded rhodomyrtosone B (2) in 42 % yield (Scheme

**3**). Our proposed mechanism for selective dehydrative cyclization leading to rhodomyrtosone B (2) is shown in **Scheme 3**. Protonation of vinylogous acid **5** leads to the vinyl oxocarbenium intermediate **11** which may exist in equilibrium with its atropisomer **12**. We believe that hydrogen bonding between the *ortho*-phenol and the ketone within the acyl substituent renders the *ortho*-phenol more nucleophilic by increasing the electron density of the phenolic oxygen.<sup>19</sup> Cyclization of **12** to hemiacetal **13** followed by dehydration affords rhodomyrtosone B (2). Hemiacetal **13** is likely a relevant intermediate in the proposed mechanism as this structural motif exists in closely related natural products including myrtucommulone D (**14**) (**Scheme 4**).<sup>20,21</sup>





In line with recent literature describing asymmetric annulations between quinone methides and cyclic 1,3dicarbonyls,<sup>22,23</sup> we also explored an alternative three-component coupling approach toward the synthesis of rhodomyrtosone B (2) (Scheme 4). We found that treatment of a mixture of acylphloroglucinol 7, dimethyl acetal 15, and syncarpic acid 10 with 20 mol% of scandium(III) triflate and camphorsulfonic acid (CSA) in refluxing isopropanol generated the natural product 2 in a single step. While the reaction still proceeds if only a Lewis acid or a Brønsted acid was used, a modest increase in yield was observed by using both in a cooperative manner. A plausible reaction mechanism is proposed in Scheme 5. Acylphloroglucinol 7 first condenses with dimethyl acetal 15 derived from isovaleraldehyde to generate the benzyl methyl ether 16. In the presence of acid and heat, the methyl ether may be extruded to produce quinone methides 17 / 18. We believe quinone methide 18, likely activated by complexation with Sc(OTf)<sub>3</sub> is a more relevant tautomeric form as its increased conjugation affords greater stability relative to 17. Quinone methide 18 then engages with vinylogous ester 19 (derived from the condensation of syncarpic acid 10 and isopropanol)<sup>24</sup> thereby forming the tricyclic acetal 20 which subsequently loses isopropanol to afford 2. It is possible that the annulations proceeds *via* cycloaddition; however, literature reports on quinone methide annulations also propose stepwise mechanisms.<sup>26,27</sup>

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Scheme 5. Proposed Mechanism for Formation of 2 via Quinone Methides 17/18



Alternatively, we considered that syncarpic acid (10) may be acting as the relevant nucleophile and may condense with the oxocarbenium to form monoalkylidene 6, similar to our previous approach (*cf.* Table 1). Using the reaction conditions shown in Scheme 4, we tested this possibility by running two control experiments: one without the acylphloroglucinol 7 and one without syncarpic acid (10) (Scheme 6). We found that in the absence of 7, syncarpic acid (10) was quickly converted to vinylogous ester 19, but minimal condensation was observed. On the other hand, in the absence of syncarpic acid (10), acylphloroglucinol 7 completely reacted with dimethyl acetal 15 to form the quinone methide (*cf.* 17 in Scheme 5). The quinone methide was subsequently reduced<sup>25</sup> in the presence of Lewis acid and isopropanol, cleanly affording the alkylated phenol 21 which could be isolated in excellent yield. The latter reaction conditions represent a promising and mild reductive transformation for alkylation of electron-rich arenes. These control experiments strongly support a mechanism for the 3-component coupling (*cf.* Scheme 4) in which quinone methide formation precedes the annulation step.



Scheme 6. Control Experiments Probing the Mechanism of the Three-Component Coupling

After successfully synthesizing rhodomyrtosone B (2), we next turned our attention to the synthesis of rhodomyrtosone A (3). Specifically, we wished to prepare endoperoxide 8 as the penultimate intermediate in the synthesis of 3 (Scheme 7). This was achieved using flow photochemistry<sup>26</sup> employing monoalkylidene **6** using oxygen-saturated methanol as solvent to provide a 50% yield of endoperoxide<sup>27</sup>  $\mathbf{8}$  as a mixture of diastereomers. The stereochemistry of endoperoxide  $\mathbf{8}$  was not assigned due to its labile nature and as its configuration was inconsequential for the subsequent cyclization step. Endoperoxide 8 and acylphloroglucinol 7 were subsequently thermolyzed in acetic acid (100 °C) which afforded the bisfuran rhodomyrtosone (3) in % vield. А proposed mechanism for the acid-mediated А formation of 3 is shown in Scheme 8. Treatment of peroxy acetal 8 with acid promotes the formation of the endocyclic peroxycarbenium ion  $24^{28}$  via protonated intermediate 23. Interception of the putative peroxocarbenium intermediate 23 with acylphloroglucinol 7 may generate endoperoxide adduct 25. The peroxide may then undergo a Kornblum–DeLaMare<sup>29</sup>-type opening to 1,4-diketone 26 which can further cyclize to bis-furan 3 under acidic conditions. It should be noted that the reaction sequence may also formally be reversed in which endocyclic peroxide cleavage may occur prior to nucleophilic addition of the acylphloroglucinol.

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Scheme 7. Synthesis of Rhodomyrtosone A (3) via Endoperoxide 8



Scheme 8. Proposed Mechanism for the Formation of Rhodomyrtosone A



### CONCLUSION

In summary, we have developed a Ni (II)-mediated 1,4-addition of acylphloroglucinols to a syncarpic acid-derived alkylidene as a Michael acceptor to access the natural product rhodomyrtosone B after selective dehydrative cyclization. A three-component coupling strategy using a quinone methide intermediate was also found to be effective. In addition, oxygenation of a dienol derived from the same alkylidene scaffold permitted access to an endoperoxide intermediate which was utilized in a short synthesis of the *bis*-furan natural product rhodomytosone A. Further studies on the synthesis and biological properties of rhodomyrtone A and related molecules are currently in progress and will be reported in due course.

### EXPERIMENTAL SECTION

General Methods. <sup>1</sup>H NMR spectra were recorded at either at 400 MHz or 500 MHz (as noted) at ambient temperature with CDCl<sub>3</sub> as the solvent unless otherwise stated. <sup>13</sup>C NMR spectra were recorded either at 100.0 MHz or 125.0 MHz (as noted) at ambient temperature with complete proton decoupling using CDCl<sub>3</sub> as the solvent unless otherwise stated. Infrared spectra were recorded on a FT-IR spectrophotometer. High-resolution mass spectra (HRMS) was carried out by electron impact (EI) using a Q-TOF mass spectrometer. Melting points were recorded on a Mel-temp apparatus. Analytical thin layer chromatography was performed using 0.25 mm silica gel 60-F plates. Preparative TLC was conducted with glass backed 1000  $\mu$ m silica gel 60-F plates. Flash chromatography was performed using 200-400 mesh silica gel. Photochemistry experiments were performed using a Hanovia 450 W medium pressure mercury lamp housed in quartz immersion cooler with a system circulator. Analytical LC-MS was performed on a UPLC (Ultra Performance Liquid Chromatography) with a Binary solvent manager, SQ mass spectrometer, PhotoDiode Array detector, and ELSD (Evaporative Light Scattering Detector). An Acquity UPLC BEH C18 1.7 µm column was used for analytical UPLC-MS. Yields refer to chromatographically and spectroscopically pure materials, unless otherwise stated. All reactions were carried out in flame-dried glassware under an argon atmosphere unless otherwise noted. Syncarpic acid (10)<sup>30</sup> and acylphloroglucinol 7<sup>31,32</sup> were prepared using literature procedures.

**5-Hydroxy-2,2,6,6-tetramethyl-4-(3-methyl-1-(pyrrolidin-1-yl)butyl)cyclohex-4-ene-1,3-dione (9):** To a flask was added syncarpic acid **10**, (500 mg, 3 mmol, 1 equiv) and anhydrous diethyl ether (45 mL). The resulting solution was cooled to 0°C and pyrrolidine (275  $\mu$ L, 3.29 mmol, 1.2 equiv) was added dropwise followed by the addition of isovaleraldehyde (370  $\mu$ L, 3.43 mmol, 1.25 equiv). The solution became slightly cloudy. The reaction was stirred at 0 °C until a white precipitate was formed (about 30 min). The white solid was filtered and washed with cold ether and dried *in vacuo* to yield **9** as a white powder (800 mg, 90 %). **R**<sub>f</sub>: 0.44 hexanes:EtOAc:MeOH (1:1:0.1) **Mp:** 165-168 °C (diethyl ether), **IR (thin film):** 2974.01, 1698.89, 1583.04, 1470.19, 1455.19, 1410.05, 1366.04, 1296.99, 1215.98 cm<sup>-1</sup>, <sup>1</sup>**H NMR (CDCl<sub>3</sub>, 500 MHz):**  $\delta$  4.43 (d, *J*=9.9 Hz, 1 H); 3.66-3.51 (ddd, *J*=2.70, 5.98, 10.44, 1H); 3.07 – 2.88 (m, 1 H); 2.89 – 2.71 (m, 1H); 2.23 – 2.08 (m, 1H); 2.08 – 1.86 (m, 5H); 1.53 – 1.34 (m, 2H); 1.34 – 1.22 (m, 12 H); 0.96 – 0.87 (m, 4 H); 0.87 – 0.75 (m, 3H) <sup>13</sup>C **NMR** (**CDCl<sub>3</sub>, 125.67 MHz):**  $\delta$  216.9, 98.7, 69.4, 54.1, 48.9, 32., 25.3, 24.5, 22.6, 20.4, 17.5 ppm **HRMS-ESI (m/z):** [M+H]<sup>+</sup> calculated for C<sub>19</sub>H<sub>32</sub>NO<sub>3</sub>, 322.2382; found, 322.2384

**2,2,4,4-Tetramethyl-6-(3-methylbutylidene)cyclohexane-1,3,5-trione (6):** To a flask was added adduct **9** (200 mg, 0.62 mmol) then methylene chloride was added (15 mL, 0.04 M). A 1M solution of hydrochloric acid in water was prepared and

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saturated with ammonium chloride. This solution was added to the reaction mixture. The mixture was vigorously stirred at room temperature for 1h. The organic layer was washed with saturated brine (3 x) and the organic fractions were dried over anhydrous sodium sulfate. Solvents were concentrated *in vacuo* to yield compound **6** (116 mg, 75 %) as a pale yellow oil. **R**<sub>r</sub> : 0.6 hexanes:EtOAc (3:1), **IR (thin film):** 2965.63, 1695.43, 1606.38, 1465.51, 1383.58,1296.07, 1215.98. 1132.50, 1039.52 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  7.52 (t, *J* = 7.6 Hz, 1H), 2.61 (t, *J* = 7.3 Hz, 2H), 1.89 (ddt, *J* = 13.5, 10.8, 6.8 Hz, 1H), 1.31 – 1.23 (m, 12 H); 0.97 (d, *J* = 6.7 Hz, 6H) <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125.67 MHz):  $\delta$  208.9, 199.5, 196.4, 159.1, 133.1, 58.5, 57.9, 38.9, 35.6, 28.7, 22.5, 22.3, 21.9, HRMS-ESI (m/z): [M+H]<sup>+</sup> calculated for C<sub>15</sub>H<sub>23</sub>O<sub>3</sub>, 251.1647; found, 251.1647

5-Hydroxy-2,2,6,6-tetramethyl-4-(3-methyl-1-(2,4,6-trihydroxy-3-(3-methylbutanoyl)phenyl)butyl)cyclohex-4-ene-1,3dione (5): The dichloromethane and acetic acid used in this reaction were degassed three times using the freeze-pump-thaw method. To the acyl phloroglucinol 7 (25 mg, 0.12 mmol, 1.5 equiv) was added Ni(ClO<sub>4</sub>)<sub>2</sub>.6H<sub>2</sub>O (2.9 mg, 0.0080 mmol, 0.1 equiv) and methylene chloride (2 mL) at room temperature under argon followed by addition of 4Å MS (20 mg). Next, a solution of monoalkylidene 6 (20 mg, 0.08 mmol. 1 equiv) in dichloromethane (1 mL) was added to the reaction mixture followed by acetic acid (0.5 mL). The reaction was stirred at room temperature for 5 h and heated to 40 °C for 12 h. The reaction was quenched with water and a solution of 1M KHSO<sub>4</sub> at 0 °C until reaching a pH  $\approx$  2. The reaction mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> and washed with saturated brine. Organic fractions were gathered and dried over anhydrous sodium sulfate. Solvents were evaporated in vacuo yielding a yellow oil. Column chromatography purification on silica gel with a gradient of CH<sub>2</sub>Cl<sub>2</sub>: MeOH (90:1 to 20:1) provided 29 mg (0.06 mmol) of compound 5 in 80% yield. Mp: 51-54 °C (hexanes, MeOH), IR (thin film): 2958.19, 2872.19, 1716.58, 1622.77, 1594.68, 1467.34, 1383.88, 1367.29, 1300.61, 1215.23, 1118.62, 754.18 cm<sup>-1</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta$  0.83 (q, J= 5 Hz, 6H), 0.97 (d, J= 5 Hz, 6H), 1.23 (s, 3H), 1.31 (d, J= 5 Hz, 3H), 1.36 (d, J=5 Hz, 3H), 1.42 (broad m, 1H), 1.47 (s, 3H), 1.75 (m, J= 10 Hz, 1H), 2.06 (m, J= 10 Hz, 1H), 1H), 2.25 (m, J=10Hz, 1H), 2.95 (d, J= 5Hz, 2H), 4.34 (t, J= 10 Hz, 1H), 5.86 (s, 2/3H) and 5.92 (s, 1/3H), 10.28 (s, 2/3H) and 10.55 (s, 1/3H); 11.15 (s, 1/3H) and 11.57 (s, 2/3H), <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125.67 MHz): δ 22.5, 22.8, 23.1, 24.5, 25.5, 26.4, 27.1, 28.2, 29.6, 29.9, 38.4, 48.9, 52.4, 55.3, 98.2, 109.5, 114.9, 158.6, 176.8, 203.4, 206.4, 212.5, HRMS-ESI (m/z):  $[M+H]^+$  calculated for C<sub>26</sub>H<sub>37</sub>O<sub>7</sub>, 461.2539; found, 461.2534.

**Rhodomyrtosone B (2) from compound 5:** To a solution of compound **5** (20 mg, 0.043 mmol, 1 equiv) in hexafluoroisopropanol (0.2 mL) was added trifluoroacetic acid (0.05 mL, 14 equiv.) at room temperature. The reaction was stirred at 60°C for 12h. Then, the reaction was diluted with water and saturated brine and the mixture was extracted with

ethyl acetate (3 x 1 mL). The organic fractions were dried over anhydrous sodium sulfate and solvents were evaporated *in vacuo* to yield a yellow solid. Purification using column chromatography on silica gel with a hexane:acetone gradient (12:1 to 4:1) provided 8 mg (0.018 mmol) of rhodomyrtosone B **2** (42% yield). **R**<sub>f</sub>: 0.4 hexanes : acetone (3:1), **Mp**: 58-62 °C **IR** (**thin film**): 3359.86, 2959.21, 2925.37, 2869.81, 1719.44, 1656.43, 1625.75, 1597.94, 1504.38, 1469.46, 1431.80, 1388.20, 1368.12, 1255.28, 1161.24, 1121.96, 1041.55, 1015.89 cm<sup>-1</sup>, <sup>1</sup>H **NMR** (**CDCl**<sub>3</sub>, **500 MHz**):  $\delta$  13.46 (s, OH), 6.26 (s, OH), 4.30 (t, *J* = 6.1 Hz, 1H), 3.24 – 2.89 (m, 2H), 2.36 (dd, *J* = 13.2, 6.6 Hz, 1H), 1.64 (s, 3H), 1.47 (s, 3H), 1.43 (s, 3H), 1.39 (s, 3H), 1.39 – 1.31 (m, 4H), 1.02 (dd, *J* = 14.3, 6.6 Hz, 6H), 0.87 (dd, *J* = 8.5, 6.4 Hz, 6H), <sup>13</sup>C **NMR** (**CDCl**<sub>3</sub>, **125.67 MHz**):  $\delta$  211.7, 203.9, 198.3, 167.2, 164.2, 159.5, 153.1, 114.5, 105.9, 105.5, 100.2, 56.1, 53.4, 47.2, 46.9, 25.3, 25.0, 24.7, 24.7, 24.5, 24.4, 24.2, 23.4, 23.1, 22.9, 22.6, **HRMS-ESI (m/z)**: [M+H]<sup>+</sup> calculated for C<sub>26</sub>H<sub>35</sub>O<sub>6</sub> 443.2434; found, 443.2427

**Rhodomyrtosone B (2) (three-component reaction):** Acylphloroglucinol **7** (25 mg, 0.119 mmol), dimethyl acetal **15** (16 mg, 0.119 mmol), and syncarpic acid **10** (22 mg, 0.119 mmol) were suspended in dry isopropanol (1 mL). Camphorsulfonic acid (6 mg, 0.2 equiv.) was added followed by the addition of scandium (III) triflate (12 mg, 0.2 equiv.). The mixture was heated to 80°C for 6 h and was then cooled to room temperature. The reaction was quenched with saturated sodium bicarbonate solution (3 mL) and diluted with ethyl acetate (5 mL). The organic phase was removed and the aqueous phase was extracted with ethyl acetate (3 x 5 mL). The combined organic layers were washed with brine and dried over sodium sulfate before being concentrated under vacuum. The resulting crude oil was then purified by silica gel chromatography eluting with 10% ethyl acetate in hexanes to provide compound **2** (23 mg, 42% yield).

**5-Isopropoxy-2,2,6,6-tetramethylcyclohex-4-ene-1,3-dione (19):** Syncarpic acid **10** (22 mg, 0.119 mmol) and dimethyl acetal **15** (16 mg, 0.119 mmol) were suspended in dry isopropanol (1 mL). Camphorsulfonic acid (6 mg, 0.2 equiv.) was added followed by the addition of scandium (III) triflate (12 mg, 0.2 equiv.). The solution was then heated to 80°C for 12 h. The reaction mixture was then cooled to room temperature and quenched with sodium bicarbonate solution (3 mL) and diluted with ethyl acetate (5 mL). The organic phase was removed and the aqueous phase was extracted with ethyl acetate (3 x 5 mL). The combined organic layers were washed with brine and dried over sodium sulfate before being concentrated under vacuum. The crude oil was purified by silica gel chromatography yielding compound **19** as a clear liquid (12 mg, 44% yield). **IR (thin film):** 2980.8, 2928.6, 1714.6, 1653.8, 1604.1, 1471.0, 1378.2, 1236.1, 1178.4, 1106.4,1042.7, 926.3 cm<sup>-1</sup>; <sup>1</sup>**H NMR (CDCl<sub>3</sub>, 500 MHz**):  $\delta$  5.41(s, 1H), 4.47(sep, *J* = 6.1 Hz, 1H), 1.34-1.26 (m, 18H); <sup>13</sup>**C NMR (CDCl<sub>3</sub>, 100 MHz**):  $\delta$  213.4, 199.3, 176.4, 99.6, 71.3, 55.0, 48.0, 24.9, 24.2, 21.1 ; **HRMS-ESI (m/z):** [M+H]<sup>+</sup> calculated for C<sub>13</sub>H<sub>21</sub>O<sub>3</sub>; 225.1485 found, 225.1483

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**3-Methyl-1-(2,4,6-trihydroxy-3-isopentylphenyl)butan-1-one (21):** Acylphloroglucinol **7** (25 mg, 0.119 mmol) and dimethyl acetal **15** (16 mg, 0.119 mmol) were suspended in dry isopropanol (1 mL). Camphorsulfonic acid (6 mg, 0.2 equiv.) was added followed by the addition of scandium (III) triflate (12 mg, 0.2 equiv.). The mixture was heated to 80°C for 6 h and was then cooled to room temperature. The reaction was then quenched with saturated sodium bicarbonate solution (3 mL) and diluted with ethyl acetate (5 mL). The organic phase was removed and the aqueous phase was extracted with ethyl acetate (3 x 5 mL). The combined organic layers were washed with brine and dried over sodium sulfate before being concentrated under vacuum. The resulting crude oil was then purified on silica using 10% ethyl acetate in hexanes to provide compound **21**, which was a white oil (32 mg, 95% yield). **IR(thin film):** 2955.8, 2560 (br), 1609.4, 1579.8, 1522.9, 1301.6, 1214.9, 1094.9 cm<sup>-1</sup>; <sup>1</sup>H NMR (MeOD, 500 MHz):  $\delta$  5.90 (s, 1H), 2.94 (d, *J* = 6.8 Hz, 2H), 2.56 – 2.49 (m, 2H), 2.24 (dt, *J* = 13.4, 6.7 Hz, 1H), 1.57 (dt, *J* = 13.2, 6.6 Hz, 1 H), 1.40 – 1.32 (m, 2H), 0.97 (dd, *J* = 12.2, 6.6 Hz, 13 H); <sup>13</sup>C NMR (MeOD, 100 MHz):  $\delta$  207.9, 166.0, 164.6, 162.1, 110.0, 106.3, 95.7, 54.7, 40.2, 30.3, 27.7, 24.1, 24.0, 22.0 ; HRMS-ESI (m/z): [M+H]<sup>+</sup> calculated for C<sub>16</sub>H<sub>25</sub>O<sub>4</sub>; 281.1747 found, 281.1759

**8a-Hydroxy-3-isopropy1-6,6,8,8-tetramethy1-8,8a-dihydrobenzo[c][1,2]dioxine-5,7(3H,6H)-dione (8):** A 0.01M solution of alkylidene **6** (80 mg, 0.32 mmol) was prepared in 32 mL of methanol in a flamed dried flask under argon at room temperature. A balloon of oxygen was bubbled through the solution for 30 minutes. The reaction flask with a balloon of oxygen bubbling through was connected to the flow setup. This setup employed Idex Health Science PFA (perfluoroalkoxyalkane) tubing (ID 0.062 in, OD 0.125 in, 500 psi max pressure) looped tightly around the Hanovia lamp which was then placed in a chiller. The reactor was 1.614 meters in length and the total volume was 3.1 mL. A recirculating chiller maintained the flow reactor at -10 °C and a fan insured proper air circulation in the photobox to maintain the overall temperature of the reaction mixture in the system at 23 °C. The optimal flow rate determined and used was 0.5 mL/min. After the entire solution was consumed, the system was flushed with an additional 5 mL of methanol and the receiving flask was replaced by a waste container. Irradiation was stopped and the entire flow setup was flushed with benzene. The solution contained in the receiving flask was collected and the solvent was evaporated. Purification using column chromatography over silica gel using a gradient of hexanes:acetone (20:1 to 10:1) provided 45 mg (50%) of endoperoxides **8** as a 1:1 mixture diasteromers (stereochemistry unassigned).

Diastereomer 1: **Rf** : 0.67 hexanes:EtOAc (2:1), **Mp**: 115-120 °C (CH<sub>2</sub>Cl<sub>2</sub>, MeOH), **IR (thin film):** 3408.97, 2975.64, 1726.15, 1690.59, 1633.86, 1469.19, 1377.38, 1286.22, 1216.22, 1159.82.11, 1100.16 cm-1, <sup>1</sup>**H NMR** (500 MHz, Chloroform-d)  $\delta$  7.29 (d, J = 1.6 Hz, 1H), 4.74 (dd, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 2.03 (dt, J = 13.4, 6.8 Hz, 1H), 1.38 (d, J = 1.6 Hz, 1H), 4.74 (dd, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 2.03 (dt, J = 13.4, 6.8 Hz, 1H), 1.38 (d, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 2.03 (dt, J = 13.4, 6.8 Hz, 1H), 1.38 (d, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 2.03 (dt, J = 13.4, 6.8 Hz, 1H), 1.38 (d, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 2.03 (dt, J = 13.4, 6.8 Hz, 1H), 1.38 (d, J = 5.9, 1.6 Hz, 1H), 3.51 (s, 1H), 3.51

J = 8.1 Hz, 7H), 1.31 (s, 3H), 1.06 (dd, J = 6.8, 1.4 Hz, 7H), 1.03 (s, 3H).<sup>13</sup>C NMR (126 MHz, cdcl3)  $\delta$  210.5, 197.5, 137.9, 134.3, 97.9, 83.5, 54.9, 51.7, 30.5, 26.6, 24.1, 20.7, 18.2, 17.9, 15.1., HRMS-ESI (m/z): [M+H]+ calculated for C<sub>15</sub>H<sub>23</sub>O<sub>5</sub>, 283.1545; found, 265.1440 [M+H-H<sub>2</sub>O]+

Diastereomer 2: **Rf** : 0.38 hexanes:acetone (2:1), **IR** (**thin film**): 3000.50, 2933.43, 2872.53, 1693.60, 1638.03, 1470.98, 1375.00, 1262.55, 1187.51, 1131.25, 1100.01 cm-1, <sup>1</sup>**H NMR** (500 MHz, Chloroform-d):  $\delta$  7.43 (d, J = 4.1 Hz, 1H), 4.13 (dd, J = 8.3, 4.1 Hz, 1H), 3.62 (s, 1H), 2.19 – 2.09 (m, 1H), 1.43 (s, 3H), 1.39 (d, J = 5.4 Hz, 6H), 1.33 (d, J = 3.4 Hz, 3H), 0.97 (d, J = 6.7 Hz, 6H), <sup>13</sup>**C NMR** (126 MHz, cdcl3):  $\delta$  207.1, 197.9, 137.9, 97.6, 85.0, 32.2, 31.1, 30.1, 29.9, 26.8, 24.2, 21.1, 20.0, 19.2, 15.3., **HRMS-ESI (m/z):** [M+H]+ calculated for C<sub>15</sub>H<sub>23</sub>O<sub>5</sub>, 283.1545; found, 283.1547

**Rhodomyrtosone A (3):** A solution of endoperoxides **8** (1:1 mixture of diastereomers) (6 mg, 0.021 mmol) in acetic acid 1 mL was transfered in a sealed tube and heated at 100 °C for 12 h. Then the reaction mixture was diluted with water and quenched with a saturated sodium bicarbonate solution. The pH was kept slightly acidic. The reaction mixture was extracted with ethyl acetate (3 x 5 mL) and the organic fractions were gathered and dried over anhydrous sodium sulfate. The solvents were evaporated *in vacuo* to yield a dark yellow oil. Purification by column chromatography over silica gel provided 6 mg (60 %) of **3** as a pale yellow gum. **R**<sub>f</sub>: 0.7 hexanes:EtOAc 1:1, **IR (thin film):** 2960.88, 2930.37, 2873.99, 1720.40, 1649.79, 122.93, 1469.93, 1420.62, 1384.14, 1305.44, 1215.98, 1171.78, 1052.58, 921.63 cm<sup>-1</sup>, <sup>1</sup>**H NMR (500 MHz, CDCl<sub>3</sub>)**:  $\delta$  13.27 (s, 1H), 9.79 (s, 1H), 6.11 (s, 1H), 4.49 (s, 1H), 2.96 (dd, *J* = 14.7, 6.6 Hz, 1H), 2.76 (dd, *J* = 14.7, 7.2 Hz, 1H), 2.40 (d, *J* = 6.8 Hz, 1H), 2.17(m, J=6.6 Hz, 1H) 1.52 (s, 3H), 1.42 (d, *J* = 5.3 Hz, 6H), 1.34 (s, 3H), 1.10 (dd, *J* = 11.8, 6.8 Hz, 6H), 1.00 (dd, *J* = 10.8, 6.7 Hz, 6H), <sup>13</sup>**C NMR (126 MHz, CDCl<sub>3</sub>)**:  $\delta$  211.3, 203.8, 198.5, 179.8, 166.8, 159.9, 159.8, 129.5, 113.3, 104.3, 101.8, 99.7, 55.3, 51.69, 45.8, 45.1, 35.5, 26.1, 24.5, 24.3, 23.3, 22.9, 22.9, 15.9, 15.9. **HRMS-ESI (m/z):** [M+H]<sup>+</sup> calculated for C<sub>26</sub>H<sub>32</sub>O<sub>7</sub>; 457.2226 found, 457.2218

**Supporting Information Available** Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra for all new compounds, and X-ray crystal data for compound **5** (CIF). This material is available free of charge *via* the Internet at http://pubs.acs.org.

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