# Synthesis and antibacterial evaluation of phosphonic acid analogues of diaminopimelic acid

I van Assche<sup>1</sup>, M Soroka<sup>2</sup>, A Haemers<sup>1\*</sup>, M Hooper<sup>3</sup>, D Blanot<sup>4</sup>, J van Heijenoort<sup>4</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, University of Antwerp, Universiteitsplein 1, B-2610 Antwerp, Belgium;
<sup>2</sup>Institute of Organic and Physical Chemistry, Technical University of Wroclaw, 50-370 Wroclaw, Poland;
<sup>3</sup>Tropical Diseases Chemotherapy Research Unit, Sunderland Polytechnic, SR2 7EE Sunderland, UK;
<sup>4</sup>Laboratory of Bacterial Envelopes and Peptides, CNRS URA 1131, University of Paris-Sud, 91405 Orsay, France

(Received 4 October 1990; accepted 14 February 1991)

Summary — Diaminopimelic acid is an essential amino acid in the peptidoglycan of Gram-negative bacteria, mycobacteria and some Gram-positive bacteria. It can be used as a target for the development of antibacterial agents. In this article the synthesis of a series of phosphonic acid analogues of this diamino-dicarboxylic acid is described: mono- and diphosphonic acid derivatives as well as their homologues, *N*-alkyl analogues and some peptidyl derivatives. A phenylphosphinic derivative was also prepared. All the prepared compounds were tested for their antibacterial activity against a series of Gram-positive, Gram-negative and mycobacteria. They were also tested as possible inhibitors of the UDP-MurNAc-L-Ala- $\gamma$ -D-Glu: *meso*-DAP synthetase. No significant biological activity was found.

**Résumé** — **Synthèse et évaluation antibactérienne d'acides phosphoniques analogues de l'acide diaminopiméliques.** L'acide diaminopimélique est un acide aminé essentiel dans le peptidoglycane des bactéries Gram-, mycobactéries et quelques bactéries Gram+. Il peut être utilisé comme une cible dans le développement d'agents antibactériens. Le présent travail rapporte la synthèse d'une série d'acides phosphoniques analogues de cet acide diaminodicarboxylique: dérivés de l'acide mono- et diphosphonique aussi bien que leurs homologues, analogues N-alkyles et quelques dérivés peptidiques. Un dérivé phénylphosphonique a été également préparé. Tous les composés décrits ont été essayés pour leur activité antibactérienne vis-à-vis d'une série de bactéries Gram+, Gram- et de mycobactéries. Ils ont été également étudiés comme possible inhibiteurs de l'UDP-MurNAc-L-Ala-γ-D-Glu: méso-DAP synthètase. Aucune activité biologique significative n'a été trouvée.

diaminopimelic acid / phosphono- and phosphino amino acids and peptides / diamino (bisphosphonic acids) / diamino (mono-phosphonic) monocarboxylic acids / antibacterial evaluation

# Introduction

Meso-Diaminopimelic acid (meso-2,6-diaminoheptanedioic acid, DAP) **1** is the cross-linking amino acid in the peptidoglycan of the cell wall of most Gram-negative bacteria, mycobacteria and some Gram-positive bacteria [1]. It is biosynthesized from aspartate in the lysine biosynthetic pathway and is built into the peptidoglycan by the UDP-MurNAc-L-Ala- $\gamma$ -D-Glu: meso-DAP synthetase. As DAP has not been found in mammals, it is clear that interference with its biochemistry would not, at least in theory, have any toxic effects in the host. Hence it can be considered as an ideal target in the design of new antibacterial and antimycobacterial agents.

However, only a few studies concerning the target properties of this amino acid in antibacterial drug design have been published. Inhibitors were developed against several enzymes of the biosynthetic pathway, eg, the succinyl-CoA: tetrahydrodipicolinate N-succinyltransferase [2, 3], the L, L-DAP epimerase [4, 5] and the meso-DAP decarboxylase [6, 7]. Some compounds were tested against the peptidoglycan building enzymes [8, 9] and other compounds were prepared as analogues of DAP and their antibacterial activity was determined [10–15]. Mostly, the inhibitory effect found was very weak and was seldom associated with an antibacterial activity in vitro. In the

<sup>\*</sup>Correspondence and reprints

latter case, auxothrophes with a requirement for DAP were nearly always used.

Taking alafosfalin 2 [16] as an example we planned the synthesis of some phosphonic acid analogues of DAP. This dipeptide of L-Ala and the phosphonic acid analogue of L-Ala is a potent antibacterial compound. Its activity is due to the inhibitory activity of this phosphonic acid on both L-Ala racemase and UDP-MurNAc-L-Ala synthetase, which are essential for peptidoglycan biosynthesis. The dipeptide is selectively transported through the bacterial cell membrane, resulting in increased intracellular concentrations of the drug and enhanced *in vivo* antibacterial activity [17].

In this article we report the synthesis, inhibitory activity against the UDP-MurNAc-L-Ala- $\gamma$ -D-Glu: *meso*-DAP synthetase and the antibacterial evaluation of a series of phosphonic acid analogues of DAP: the monophosphonic acid analogue **3** and its lower homologue **4**, the bis(phosphonic acid) analogue **5**, as well as several N-alkyl **6**–**9** and N-acyl (Gly, L-Ala) derivatives **10–11** and homologues **12–15** of this bis(phosphonic) analogue. A phenylphosphinic analogue **16** was also prepared.



# Chemistry

A variety of aminophosphonic acid preparations have been examined for their usefulness in the synthesis of the 1,5-diamino-1,5-pentane bis(phosphonic acid) **5**. Most of the methods used gave disappointing results; this was the case with amination of 2-ketophosphonic acids [19, 20], amidoalkylation of trivalent phosphate esters [21, 22] and the alkylation of a protected aminomethylphosphonic acid with diiodopropane [23–25]. Good results were obtained using amidoalkylation reactions and PCl<sub>3</sub> [26] and addition reactions of dialkyl phosphites to imines.

Using pentanedial 18 as starting aldehyde in the amidoalkylation reaction 2 compounds in  $\approx$  1:1 ratio were obtained: the desired 1,5-diamino-1,5-pentane bis(phosphonic acid) 5 and the cyclic analogue, 2.6piperidine bis(phosphonic acid) 21 (scheme 1). The same reaction was used to prepare the higher homologue, 1,6-diamino-1,6-hexane bis(phosphonic acid) 13. Hexanedial was the starting material and no cyclisation was observed. In an attempted preparation of the lower homologue using butanedial as starting material, the only compound isolated was the cyclic product, 2,5-pyrrolidine bis(phosphonic acid) 22. In an analogous reaction 1,5-diamino-1,5-pentane bis(phenylphosphinic acid) 16 was prepared when dichlorophenylphosphine was used. In this case no cyclisation was observed.



In each of the above mentioned preparations racemic mixtures were obtained. In the case of the linear compound 5 the *meso*-form and the racemate (L, Land D, D-form) were formed in a 1:1 ratio, as seen on the P-NMR spectra. In the case of the piperidine compound 21 the ratio was 1:3 (table I).

The above-mentioned cyclisation prompted us to examine the addition reaction of dialkyl phosphites to imines giving rise, after hydrolysis, to the corresponding 2-aminophosphonic acids. Depending on the amine used, both free amino- and N-substituted aminophosphonic acids can be prepared (scheme 2). An analogous reaction for the preparation of our target compound has been vaguely mentioned in the literature [27]. We used the tritylimine derivative of pentanedial as starting material. This N,N'-ditrityl-1,5-pentanediimine 23, prepared from the dialdehyde with



tritylamine in benzene with  $Na_2SO_4$  and purified by crystallisation, reacted readily with dimethyl phosphite. The *N*,*N*'-ditrityl-1,5-diamino-1,5-pentane bis (phosphonic tetramethylester) **31** was not isolated, but immediately hydrolysed with 6 N HCl to the free aminophosphonic acid **5**.

With dilute acid the trityl groups were selectively removed [28], giving free aminophosphonic ester **38** which can be transformed into the corresponding aminoprotected tripeptides esters **39–40** after reaction with respectively Z-Gly and Z-L-Ala as mixed anhydride. Mild hydrolysis with HBr in AcOH afforded the free tripeptides **10–11**.

N,N'-dialkylsubstituted-1,5-diamino-1,5-pentane bis(phosphonic acids) 6-9 were prepared by addition reactions to N,N'-dialkyl substituted imines 24–26 of pentanedial. The diimine synthesis with pentanedial and simple alkylamines has already been studied by Lubig et al [29]. They found that with alkylamines the predominant compound was the cyclic N-alkyl-1,4dihydropyridine, the N,N'-dialkyl-1,5-pentanediimine being formed in a 2-3% yield. We reinvestigated this reaction and found that with branched amines such as t-butylamine and isopropylamine no cyclisation occurred. In the case of ethylamine we found mixtures of both compounds in a 4:6 ratio as determined by <sup>1</sup>H-NMR. The linear imine can be isolated by distillation. All these N,N'-diimines were, in contrast to the trityl derivative, unstable and were used immediately after distillation. Due to isolation problems with the N,N'-dimethyldiimine, the N,N'-dimethyl bis(phosphonic acid) 6 was prepared from N-methylacetamide by the above-mentioned amidoalkylation reaction.

The above reactions were also used for the preparation of the homologues 12–14, 15. The imines 27–30 were prepared as described above and, after addition of dimethyl phosphite, gave the N,N-dialkyl substituted esters 35–37. Final hydrolysis afforded the aminophosphonic acids. The preparation of the lower homologue failed, possibly due to steric hindrance. The N,N'-ditrityldimine from butanedial 27 was readily prepared but after addition of dimethyl phosphite only starting materials were recovered. In

contrast, the *N*,*N*-diisopropyl derivative **12** was easily prepared by this route. No problems occurred in the synthesis of the higher homologues prepared from hexanedial. The tetraethyl ester of 1,6-diamino-1,6-hexanediphosphonic acid has already been reported. It was prepared from the corresponding diketophosphonate by reductive amination [19].

Finally we also report the synthesis of the monophosphonic analogue **3** and homologue **4** of DAP. We prepared **3** from a protected D, L-2-aminoadipic acid **41** as outlined in scheme 3. The aldehyde in question was prepared by a reduction oxidation reaction of **43** with borane-dimethylsulfide and pyridinium chlorochromate in pyridine [30]. For the preparation of the lower homologue **4** we used ethyl 2-ethoxycarbonyl-4-formyl-2-phthalimidobutanoate **48** as starting material (scheme 4). This was prepared by a 1,4-addition of diethyl phthalimidomalonate **47** to acrolein.

# **Biological activity**

All compounds were tested for their antimicrobial activity *in vitro* against a series of Gram+, Gram- and Mycobacteria: *Staphylococcus aureus, Streptococcus* sp, *Bacillus cereus, Escherichia coli, Klebsiella pneumoniae, Salmonella* sp, *Mycobacterium tuberculosis, Mycobacterium avium, Mycobacterium fortuitum.* The activity against Gram-positive and Gram-negative bacteria was determined by means of the usual dilution method in tryptic soya broth. Mycobacteria were tested by the agar dilution method in Middlebrook 7H9 agar supplemented with 10% oleic acid, albumin and dextrose solution.

In addition to the *in vitro* screening for anti(myco) bacterial activity these DAP analogues were also tested as potential inhibitors of the partially purified UDP-MurNAc-L-Ala- $\gamma$ -D-Glu: *meso*-DAP synthetase from *E coli* [31]. The activity of this enzyme was assayed by following the appearance of UDP-MurNAc-L-Ala-v-D-Glu-*meso*[<sup>14</sup>C]DAP by high-voltage electrophoresis [32]. Compounds were tested at 1 and 10 mM concentrations.

Analogue **3** was tested at 10 mM as a possible substrate for this enzyme. UDP-MurNAc-L-Ala- $\gamma$ -D-[<sup>14</sup>C]Glu [33] was used for this purpose. The formation of a radioactive UDP-MurNAc-tripeptide was checked by reverse-phase HPLC [34]. Even with large amounts of enzyme, no new radioactive peak could be detected. Therefore analogue **3** is not a substrate for the UDP-MurNAc-tripeptide synthetase from *E coli*.

# **Results and Discussion**

All the compounds prepared gave negative results in the *in vitro* antibacterial and antimycobacterial tests

# Table I. Aminophosphonic acids.

n°	R	R'	R"	n	Molecularª Formula	Yield %	IR	$^{l}H NMR$ ( $D_2O + DCl$ )	$^{13}C NMR$ ( $D_2O + DCl$ )	<sup>31</sup> P NMR
3	н	ссодн	OH	1	C6H15N2O5P	32	3600-2300	1.55-2.05(m,6H,	24,42/24,38	13.77-
							1720,1625,1520,	CH2),3.28(m,1H,	$(2d,CH_2,J_{CP}=8Hz)$ ,	13.81
							1175,1060,920	CHP),3.95(t,1H,	31.12( <u>CH</u> 2CHCCOH),	
								CHOOOH)	51.68/51.62(2d,CHP,	
									J <sub>CP</sub> =142.9Hz),	
				•					56.40/56.51(2s,	
									CHCCOH)	
4	н	схан	ŒH	0	$C_5H_{13}N_2O_5P$	45	3600-2400,1720,	1.80-2.20(m,4H,CH <sub>2</sub> )	27.59( <u>CH</u> 2CHP),	-
							1620,1525,1175,	3.3O(m,1H,CHP),	30.13( <u>CH</u> 2CHOOH),	
								4.00(t,1H, <u>CH</u> 000H)	51.33(d,CHP)	
									J <sub>CP</sub> = 143.6Hz,	
								•	56.56( <u>CH</u> CCOH),	
									176.19(COOH)	
5	н	$O=P(OH)_2$	ЮH	1	$C_5H_{16}N_2O_6P_2$	31p	3600-2400,1620,	1.70-2.05(m,6H,	25.36/25.69(2xt,	13.84-
							1540,1150,1040,	CH <sub>2</sub> ), 3.34(m, 2H, CH)	$CH_2, J_{CP}=8.1Hz),$	13.88
						75 <sup>C</sup>	900		31.04/31.25(2s,	
									$\underline{CH_2}CH_2CH_2$ ),	
									51.86/51.63	
									$(2xd,CH,J_{CP}=142Hz)$	
б	CHa	O=P(OH) ⊃	ОН	1	C7H20N2O6P2	25	3600-2200,1620,	1.75(m,2H,CH <sub>2</sub> ),1.94	23.60(CH <sub>2</sub> ),27.33	12.23-
-		( / 2			, 20 2 0 2		1455,1155,1050,	(m,4H, <u>CH</u> 2CH2CH2),	27.57(2s, <u>CH</u> 2CH2	12.26.
							900	2.82(s,6H,CH <sub>3</sub> ),	CH <sub>2</sub> );32.42(CH <sub>3</sub> ),	
								(m,2H,CH)	57.12-57.29	
									(2d,CH,J <sub>CP</sub> =141)	
						_				10.400
7	CH2CH3	3 O=P(OH) <sub>2</sub>	OH	1	C9H24N2O6P2	45	3600-2200,1610,	1.31(t,6H,CH <sub>3</sub> ),	11.74(CH <sub>3</sub> ),23.89	12.42
							900	(m / U OU-OU- )	$(2s, cn_2cn_2cn_2), 42.75$	
		· ·						(111, 411, 012, 012, 012, 012),	$(\underline{un}_2,\underline{un}_3),55.95$	٥
									ца)	2
								<u>un</u> 2un3un)	п <b>2</b> )	

•

# Table I. Continued.

n°	2	R	R'	<i>R</i> "	n	Molecularª Formula	Yield %	IR	${}^{I}H NMR (D_2O + DCl)$	$^{13}C NMR$ $(D_2O + DCl)$	31P NMR
8	CH(	CH3)2	0=P(OH)2	ОН	1	C <sub>11</sub> <u>H</u> 28N2O6P2	55	3600-2400,1615, 1460,1390,1160,	1.35(2d,12H,CH <sub>3</sub> ), 1.70-2.05(m,6H,CH <sub>2</sub> ),	19.28-19.51(CH <sub>3</sub> ), 24.01(CH <sub>2</sub> ),29.09-	12.65 <sup>d</sup>
			•	·		·		1055, <b>9</b> 00	3.30(m,2H,CHP),3.72 (m,2H,CHN)	28.91(2s, <u>CH2</u> CH2 <u>CH2</u> ), 51.69(CHN),54.26 (d,CHP,J <sub>CP</sub> =140.7Hz)	
9	C(	(CH3)3	O=₱(OH) <sub>2</sub>	Он	1	C <sub>13</sub> H <sub>32</sub> N <sub>2</sub> O <sub>6</sub> P <sub>2</sub>	40	3700-2200,1610, 1475,1400,1380,	1.43(s,18H,CH <sub>3</sub> ), 1.70-2.10(m,6H,CH <sub>2</sub> ), 3.28(m 2H (CH)	$25.12(CH_2), 26.22$ (CH <sub>3</sub> ), 31.25( <u>CH_2</u> ) (Ho(CH_2), 52, 59(d, CH	13.50- 13.55
								1100,1000,000	5.20(((),21),21)	J <sub>@</sub> =137.7Hz), 61.03(C)	
10	GL	χe	O=P(OH) <sub>2</sub>	OH	1	C9H22N4O8P2	40	3600-2400,1675, 1550,1170,1060, 930	1.30-1.90(m,6H,CH <sub>2</sub> ) CH <sub>2</sub> CH <sub>2</sub> ),3.85(s,4H, CH <sub>2</sub> );4.02(m,2H <sub>x</sub> CH)	, 23.54(CH <sub>2</sub> ),29.56- 30.14(2s, <u>CH</u> 2CH), 41.47(CH <sub>2</sub> ),48.92-	19.03- 19.17
										49.27(2d,CH,J <sub>CP</sub> = 149.5Hz),167.32 (CO)	
11	ALA	Ąf	0=P(OH) <sub>2</sub>	Он	1	C <sub>11</sub> H <sub>26</sub> N <sub>4</sub> O <sub>8</sub> P <sub>2</sub>	45	3600-2200,1660, 1540,1165,1050, 915	1.30-1.90(m,6H,CH <sub>2</sub> ), CH <sub>2</sub> CH <sub>2</sub> ),1.55(d,6H, CH <sub>3</sub> ),3.90-4.10(m,4H,	. 17.53(CH <sub>3</sub> ),23.54 (CH <sub>2</sub> ),29.21-29.85 . (25, <u>CH</u> 2CH),48.87-	18.77- 18.89
									CH)	49.23(2d,CHP,J <sub>CP</sub> = 149.5Hz),50.14- 171.17(CO)	
12	CH	(CH3)2	O=P(QH) <sub>2</sub>	OH	0	C <sub>10</sub> H <sub>26</sub> N <sub>2</sub> O <sub>6</sub> P <sub>2</sub>	45	3500-2200,1600, 1575,1400,1210, 1160,1055,915	1.38(2d,12H,CH <sub>3</sub> ), 2.05-2.30(m,4H, CH <sub>2</sub> ),3.49(m,2H,CHP), 3.74(m,2H,CHN)	19.22-19.40(CH <sub>3</sub> ), 25.65(CH <sub>2</sub> CH <sub>2</sub> ),52.57 , (CHN),53.15(d,CHP, J <sub>rp</sub> =146.5Hz)	14.22- 14.31
13	J	н	O=P(OH)2	ОH	. 2	С <sub>6</sub> н <sub>18</sub> №206 <sup>р</sup> 2	36	3600-2200,1640, 1580,1525,1250,	1.58(m,4H,CH <sub>2</sub> ), 1.73–2.06(m,4H, <u>CH</u> 2	25.41(CH <sub>2</sub> ),28.22 ( <u>CH<sub>2</sub></u> CH),48.95(d,CHP	16.84 <sup>0</sup>
								1190,1065,950	ΩH),3.46(M,2H,CHP)	J ( <sup>p=152.4HZ</sup> )	

Table	T	Continued

n°	R	R'	R"	n	Molecularª Formula	Yield %	IR	$(D_2O + DCl)$	$(D_2O + DCl)$	<sup>31</sup> P NMR
14 C	H(CH <sub>3</sub> ) <sub>2</sub>	O=P(OH)2	Он	2	C <sub>12</sub> H <sub>30</sub> N <sub>2</sub> O <sub>6</sub> P <sub>2</sub>	50	3600-2200,1600,	1.35(2d,12H,CH <sub>3</sub> ),	19.28-19.46(CH <sub>3</sub> ),	13.03 <sup>d</sup>
							1472,1405,1380,	1.58(m,4H,CH <sub>2</sub> CH <sub>2</sub> ),	$26.06(CH_2CH_2)$ ,	
							11,70,1065,910	1.75-2.20(m,4H,CH <sub>2</sub> )	, 28.04(CH <sub>2</sub> ),52.80	
								3.26(m,2H,CHP),3.71	(CHN),53.38(d,CHP,	
								(m,2H,CHN)	J <sub>CP</sub> =145.2Hz)	
15 C	(CH <sub>3</sub> ) <sub>2</sub>	$O=P(OH)_2$	QН	2	C14H34N2O6P2	40	3600-2200,1600,	1.14(s,18H,CH <sub>3</sub> ),	26.17(CH <sub>3</sub> ),27.17	13.83-
							1475,1405,1380,	1.50-2.10(m,8H,CH <sub>2</sub> ),	(CH <sub>2</sub> CH <sub>2</sub> ), 31.08	13.87
							1170,1065,910	3.24(m,2H,CHP)	(CH <sub>2</sub> ),52.77(d,CH,	
									$J_{CP}=137.7Hz$ ),	×
									60.86(C)	
16	Н	Ͻ=Ρ(C <sub>6</sub> H <sub>5</sub> )(OH)	) С6Н5	1	$C_{17}H_{24}N_2O_4P_2$	30	3600-2400,1625,	1.70-2.30(m,6H,CH <sub>2</sub> )	23.49(CH <sub>2</sub> ),28.33	25.12-
							1540,1440,1180,	3.35(m,2H,CH),7.61	( <u>CH2</u> CH2 <u>CH2</u> ),51.84	25.20
							1130,1040,700	(m,lOH,ArH)	(d,CH,J <sub>CP</sub> =97Hz),	
									129.55-132.35-	
									132.81-134.73	
									(ArC)	
219	-	-	-	-	C5H13NO6P2	33	3600-2500,1600,	1.60-2.20(m,6H,CH <sub>2</sub> )	, 23.25(CH <sub>2</sub> );24.36	12.08-
							1160,1000,935	3.33(m,2H,CH)	(CH <sub>2</sub> C);56.68(d,CH,	12.90
									J =143.6Hz)	
22 <sup>h</sup>	-		-	-	$C_4H_{11}NO_6P_2$	39	3200-2100,1600,	2.05-2.45(m,4H,CH <sub>2</sub> ),	27.57(CH <sub>2</sub> ),57.15	12.58 <sup>d</sup>
							1470,1180,1080,	3.70(m,2H,CH)	(d,CH,J =149.5Hz)	
							920			

<sup>a</sup>Analysis C, H, N, P; <sup>b</sup>amidoalkylation; <sup>c</sup>phosphite addition; <sup>d</sup>insufficient resolution; <sup>e</sup>GLY = Glycyl; <sup>f</sup>ALA = L-alanyl;  $g_{2,6-piperidine}$  bis(phosphonic acid); <sup>h</sup>2,5-pyrrolidine bis(phosphonic acid).

with all MIC-values higher than 100  $\mu$ g/ml. Moreover, they did not show any inhibitory effect in the enzyme assay at 1 mM concentration. Even in a 10 mM solution only 50% inhibition was found with the monophosphonomonocarboxy DAP analogue **3**.

It can be concluded that the replacement of a carboxylic acid function in DAP with a phosphonic acid moiety does not result in an antimetabolite of the parent compound; this is in contrast to other amino acids such as alanine [16], glutamic acid [35] and ornithine [36] (for a review see [37]). The very low

enzyme inhibitory effect of **3** can hardly be considered as important, particularly as this compound does not show any antibacterial activity *in vitro*. The *in vitro* inactivity of the peptides may indicate that at least for the diphosphonic analogues the transport through the cell membrane barrier is possibly not the main reason although transport through the cell membrane has not been studied directly in this work. Moreover, peptides corresponding to the monophosphonic acid analogue were not prepared.



Scheme 2. (a)  $Na_2SO_4$ , ether or benzene; (b) dimethylphosphite; (c) 12 N HCl, H<sup>+</sup> ion exchanger; (d) 1 N HCl in CH<sub>3</sub>OH; Z-Gly or Z-L-Ala, ClCOOC<sub>2</sub>H<sub>5</sub>, triethylamine; (e) HBr-CH<sub>3</sub>COOH, methyloxirane.

# Conclusion

We present the synthesis of some mono- and diphosphonic acid analogues and homologues of diaminopimelic acid. The preparation of some *N*-alkyl and *N*acyl derivatives and a phosphinic acid analogue is also presented. These compounds do not show any antibacterial activity *in vitro*. Moreover, no significant inhibitory effect was found against the enzyme, responsible for the incorporation of DAP into the cell wall peptidoglycan.

### **Experimental protocols**

#### General procedures

A Varian EM 360-A (60 MHz), a Jeol JNF-FX (200 MHz) and a Varian XL (300 MHz) were used for recording <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P-NMR spectra. Chemical shifts are given in ppm ( $\delta$ ), relative to tetramethylsilane (TMS), sodium 3-(trimethylsilyl)



Scheme 3. (a) *N*-carbethoxyphthalimide,  $Na_2CO_3$ ,  $H_2O_3$ ; (b) (CH<sub>3</sub>)SO<sub>2</sub>, triethylamine, CHCl<sub>3</sub>; (c) BH<sub>3</sub>-(CH<sub>3</sub>)<sub>2</sub>S, THF, pyridinium chlorochromate, CH<sub>2</sub>Cl<sub>2</sub>; (d) tritylamine,  $Na_2SO_4$ , CH<sub>2</sub>Cl<sub>2</sub>; (e) dimethylphosphite; (f) HCl, methyloxirane.

propionate (TSP) or 85% orthophosphoric acid as reference. The IR spectra were obtained for KBr discs using a Beckmann-Acculab-4 spectrophotometer ( $v_{max}$  are given in cm<sup>-1</sup>). Mass spectra were determined with a VG 70-SEQ hydrid mass spectrometer, equiped with an Ion Tech saddle field atom gun. Melting points were determined on a Büchi capillary melting point apparatus and are uncorrected.

#### Syntheses

### Pentanedial 18

A commercial solution of pentanedial (25%, 100 ml) was acidified with 12 N HCl (0.25 ml) and stirred for 2 h at room temperature. The solution was then neutralised with NaHCO<sub>3</sub> (2 g), saturated with NaCl (30 g) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 x 70 ml). The organic phases were collected, dried with Na<sub>2</sub>SO<sub>4</sub>, filtered and evaporated. The residue was distilled under reduced pressure. Pentanedial was obtained as a colourless, mobile liquid which solidifies quickly on standing and must be used immediately.

# 1,5-Diamino-1,5-pentanebis(phosphonic acid) 5, and 2,6-piperidinebis(phosphonic acid) **21**- amidoalkylation reaction

Acetylchloride (7.8 g, 0.1 mol) was added dropwise in  $\approx$  10 min, with vigorous stirring and cooling (ice-water bath) to a mixture of freshly distilled pentanedial (5.0 g, 0.05 mol) **18**,



Scheme 4. (a) Acrolein,  $CH_3ONa$ , toluene; (b) tritylamine,  $Na_2SO_4$ ,  $CH_2Cl_2$ ; (c) dimethylphosphite; (d) HCl, methyloxirane.

acetamide (14.7 g, 0.25 mol) and AcOH (50 ml). The solution was stirred overnight at room temperature. PCl<sub>3</sub> (8.8 0.1 mol) was then introduced dropwise to the cooled  $(0^{\circ}C)$ mixture and stirring at 0°C was continued for 15 min with subsequent heating on a boiling water bath, under a reflux condenser, for 1 h. The volatile material was evaporated under reduced pressure on a boiling water bath and the oily residue refluxed overnight with 12 N HCl (100 ml). The hydrolysate was concentrated under reduced pressure on a boiling water bath and the residue extracted with CH<sub>3</sub>OH (60 ml). The precipitated NH<sub>4</sub>Cl was filtered off and washed twice with CH<sub>3</sub>OH (20 ml). The collected filtrates were evaporated under reduced pressure, the combined residues dissolved in a minimum amount of water, placed on a strongly acid ion exchanger (Dowex 50 x 2-100, H+ form) and eluted with H<sub>2</sub>O. The ninhydrin positive fractions were collected and evaporated under reduced pressure. 21 and 5 were eluted consecutively (table I).

N,N'-Dimethyl-1,5-diamino-1,5-pentane bis(phosphonic acid) **6** This compound was prepared as described for **5** by an amido alkylation using N-methylacetamide (table I).

# 1,6-Diamino-1,6-hexane bis(phosphonic acid) 13

This compound was prepared as described for **5** using hexanedial as starting material (table I). Hexanedial was prepared by oxidation of 1,2-cyclohexanediol [38].

# 2,5-Pyrrolidine bis(phosphonic acid) 22

This compound was prepared as described for **21** by an amidoalkylation reaction using butanedial (table I). Butanedial was prepared by hydrolysis of dimethoxytetrahydrofurane [39].

1,5-Diamino-1,5-pentane bis(diphenylphosphinic acid) 16 This compound was prepared as described for 5 by an amidoalkylation reaction using dichlorophenylphosphine. 16 was eluted from the ion exchanger with 0.1 N HCl. The ninhydrin positive fractions were collected and evaporated under reduced pressure (table I).

# N,N'-Ditrityl-1,5-pentanediimine 23

Freshly distilled pentanedial **18** (5.01 g, 0.05 mol) in benzene (25 ml) was added dropwise to a well stirred and cooled (icesalt bath) mixture of tritylamine (25.9 g, 0.10 mol) and anhydrous Na<sub>2</sub>SO<sub>4</sub> (16 g) in benzene (150 ml). The mixture was stirred for 5 h at 0°C and then overnight at room temperature. Na<sub>2</sub>SO<sub>4</sub> was removed by filtration, washed with benzene and the combined filtrates concentrated under reduced pressure. The residue was treated with boiling cyclohexane (100 ml) and left for crystallization (table II).

### *N,N'-Ditritylbutane-1,4-diimine* **27**

This compound was prepared as 23 using butanedial as starting material (table II).

# 1,5-Diamino-1,5-pentane bis(phosphonic acid) 5- phosphite addition

A mixture of N,N'-ditrityl-1,5-pentanediimine **23** (23.6 g, 0.05 mol) and dimethyl phosphite (10 ml; 0.109 mol) was heated at 100°C for 1 h and the excess dimethyl phosphite removed from the clear yellow solution under reduced pressure. The oily residue **31**\* was refluxed for 8 h in 12 N HCl (50 ml). The mixture was extracted with CHCl<sub>3</sub> and the aqueous phase evaporated under reduced pressure. The residue was dissolved in a minimum amount of H<sub>2</sub>O and **5** isolated by ion-exchange chromatography as described in the amidoalkylation reaction. **5** can also be obtained by dissolving the abovementioned residue in CH<sub>3</sub>OH (20 ml) and precipitation with a solution of methyloxirane in CH<sub>3</sub>OH (3 ml in 10 ml) (table I).

## N,N'-Dialkyl- $\alpha$ , $\omega$ -alkanediimines 24–26, 28–30

These imines were prepared by adding a solution of the appropriate dialdehyde (0.02 mol) (pentanedial **18**, butanedial or hexanedial) in ether (20 ml) to a cooled mixture (ice-salt bath) of the amine (0.05 mol) and an excess anhydrous  $Na_2SO_4$  in ether (50 ml). The mixture was stirred for 15 min and then for 1 h at room temperature.  $Na_2SO_4$  was removed by filtration and washed with ether. The combined filtrates were concentrated under reduced pressure and the residue was distilled under reduced pressure. The alkanediimines are unstable liquids and their colour changes to yellow and then red on standing. They were used immediately (table II).

# $\alpha, \omega$ -Diamino- $\alpha, \omega$ -alkane bis(phosphonic acids) and their N,N'-dialkyl analogues 7–9, 12, 14–15

These compounds were prepared by reaction with dimethyl phosphite as described for 5 (table I).

#### 1,5-Diglycylamino-1,5-pentane bis(phosphonic acid) 10

The crude oily residue (0.025 mol tetramethyl 1,5-di (trityl-amino)pentane-1,5-diphosphonate 31) obtained after the addition of dimethyl phosphite to 23 (see preparation of 5) was heated to boiling in 1 N HCl in CH<sub>3</sub>OH (200 ml) for 15 min. The solution was evaporated at room temperature under reduced pressure and the residue treated with dry diethyl ether (100 ml). The ether was decanted and the oily product washed

<sup>\*</sup>This oily residue 31 was also used for the preparation of tripeptides 10 and 11.

with dry diethyl ether (2 x 50 ml) and finally dried under reduced pressure. This crude hydrochloride of tetramethyl 1,5diamino-1,5-pentane bis(phosphonate) was dissolved in dry CHCl<sub>3</sub> (40 ml) and triethylamine (7.5 ml). This solution was added at  $-5^{\circ}$ C to a solution of the mixed anhydride of Zglycine (prepared by dissolving Z-glycine (10.45 g, 0.05 mol) in dry CHCl<sub>3</sub> (100 ml) containing triethylamine (7.5 ml), adding ethyl chloroformate at  $-5^{\circ}$ C (5.0 ml, 0.055 mol) and stirring for 20 min). The mixture was slowly heated to boiling, cooled after 30 min and washed successively with  $H_2O$ (100 ml), 5% HCl (100 ml), H<sub>2</sub>O (100 ml), saturated NaHCO<sub>3</sub> solution (100 ml), and saturated NaCl solution (100 ml). The organic solution was dried with Na2SO4, concentrated under reduced pressure and the oily residue dissolved in a solution of HBr (30%) in AcOH (100 ml) and left at room temperature overnight. The volatile compounds were then removed under reduced pressure and the residue dissolved in  $H_2O$  (60 ml). Benzylbromide was removed by extraction with ether (2 x 40 ml) and the water removed under reduced pressure. The residue was dissolved in CH<sub>3</sub>OH (100 ml) and methyloxirane was added (10 ml). 10 precipitated after a few hours at 0°C. Recrystallization from  $H_2O$ -ÉtOH (table I).

### 1,5-Di-L-alanylamino-1,5-pentane bis(phosphonic acid) 11 This compound was prepared in the same way as 10 using Z-Lalanine (table I).

#### 5-Methoxycarbonyl-5-phthalimidopentanoic acid 43

To a solution of triethylamine (2.43 g, 0.0241 mol) in absolute CHCl<sub>3</sub> (50 ml) was added 2-phthalimidoadipic acid **42** (7 g, 0.0241 mol) and dimethyl sulfate (2.5 ml, 0.0265 mol) and the mixture stirred overnight at room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate (120 ml), washed with H<sub>2</sub>O (3 x 40 ml) and dried over Na<sub>2</sub>SO<sub>4</sub>. After concentration under reduced pressure, the residue was dissolved in a small amount of ether, and the same volume of petroleum ether (bp =  $35-70^{\circ}$ C) was added; and crystallization was induced by cooling and scratching (6.6 g, 78%): mp =  $131^{\circ}$ C; IR 2950, 2910, 1735, 1710, 1470, 1425, 1250, 1170, 720, 910 cm<sup>-1</sup>; <sup>1</sup>H NMR (DMSO-d<sub>6</sub>), 1.55 (m, 2 H, CH<sub>2</sub>), 2.00–2.50 (m, 4 H, CH<sub>2</sub>COOH, CHCH<sub>2</sub>), 3.65 (s, 3 H, COOCH<sub>3</sub>), 4.90 (t, 1 H, CH), 7.89 (m, 4H, ArH). Anal C<sub>15</sub>H<sub>15</sub>NO<sub>6</sub> (C, H, N).

#### Methyl 5-formyl-2-phthalimidopentanoate 44

In a flask provided with a septum inlet and a reflux condenser attached to a connecting tube leading to a mercury bubbler was placed 5-methoxy-carbonyl-5-phthalimidopentanoic acid **43** (5 g, 0.164 mol) and dry THF (30 ml). The mixture was vigorously stirred and boranedimethyl sulfide (1.67 ml) was added dropwise from a syringe. After completion of the addition, the mixture was heated under reflux for 1 h, the solvent was removed under reduced pressure (room temperature), and the residue dissolved in CH<sub>2</sub>Cl<sub>2</sub> (20 ml). This solution was added dropwise to a well-stirred suspension of pyridinium chlorochromate (3.91 g, 0.018 mol) in CH<sub>2</sub>Cl<sub>2</sub> (30 ml) and heated under reflux for 1 h. The mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, passed through a Florisil column and the eluates were collected and concentrated yielding an oil (2.8 g, 60%). Crystallization from different solvent systems failed, but the <sup>1</sup>H NMR spectrum indicated the presence of the aldehyde function (peak at 9.68 ppm). This crude aldehyde was used further without purification.

#### Methyl 2-phthalimido-6-N-trityliminohexanoate 45

Tritylamine (3.37 g, 0.013 mol) was added to a solution of aldehyde 44 (3.76 g, 0.013 mol) in  $CH_2Cl_2$  (20 ml) at 0°C.

Anhydrous  $Na_2SO_4$  (2.8 g) was added and the mixture was stirred overnight. The  $Na_2SO_4$  was removed by filtration and the filtrate concentrated under reduced pressure. The imine was crystallized from ether (4.48 g, 65%): mp = 96°C; IR 2945, 2920, 1740, 1725, 1665, 1495, 1450, 1250, 1215, 760, 720, 705 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>), 1.66 (m, 2 H, CH<sub>2</sub>), 2.30 (m, 2 H, CH<sub>2</sub>CH=N), 2.44 (m, 2 H, CHCH<sub>2</sub>), 3.71 (s, 3 H, CH<sub>3</sub>), 4.88 (t, 1 H, CH), 7.21 (m, 15 H, CH=N – ArH), 7.80 (m, 4H, ArH); <sup>13</sup>C NMR (CDCl<sub>3</sub>), 22.42 (CH<sub>2</sub>), 28.38 (CH<sub>2</sub>CH=N), 35.45 (CHCH<sub>2</sub>), 51.86 (OCH<sub>3</sub>), 52.62 (CH), 123.45 – 134.02 – 145.59 – 167.49 (C<sub>6</sub>H<sub>4</sub>), 126.49 – 127.48 – 129.58 – 131.69 (C<sub>6</sub>H<sub>5</sub>), 159.84 (COOCH<sub>3</sub>). Anal C<sub>34</sub>H<sub>30</sub>N<sub>2</sub>O<sub>4</sub> (C, H, N).

#### 2,6-Diamino-6-phosphonohexanoic acid 3

A mixture of imine 45 (5.3 g, 0.01 mol) and dimethyl phosphite (5.5 g, 0.05 mol) was heated at 100°C for 2 h. The excess dimethyl phosphite was then removed under reduced pressure (100°C) and the residue 46 refluxed overnight in 12 N HCl (20 ml). The acidic solution was filtered and the filtrate was extracted with CHCl<sub>3</sub> (3 x 10 ml). The aqueous phase was concentrated under reduced pressure, the residue dissolved in a minimum amount of H<sub>2</sub>O and placed on an ion exchange resin (Dowex 50X-200, H<sup>+</sup> form). The column was washed with H<sub>2</sub>O and the product eluted with 2 N HCl. The eluate was evaporated under reduced pressure, treated with CH<sub>3</sub>OH (20 ml) and excess methyloxirane. The mixture was kept at 0°C for a few hours and the precipitated 3 isolated and dried (table I).

### Ethyl 2-ethoxycarbonyl-4-formyl-2-phthalimidobutanoate 48

Acrolein (2 ml, 0.03 mol), dissolved in toluene (2 ml) was added dropwise to diethyl phthalimidomalonate **47** (9.16 g, 0.03 mol) in toluene (100 ml), containing a catalytic amount of sodium methoxide and the mixture cooled in a waterbath. The mixture was stirred for 2 h and then concentrated under reduced pressure. The residue was warmed with 2 N H<sub>2</sub>SO<sub>4</sub> (25 ml) for 5 min and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 30 ml). The organic layer was concentrated under reduced pressure and the residue placed on a silicagel 60 (230–340 mesh) column which was eluted with CH<sub>2</sub>Cl<sub>2</sub>. Evaporation afforded **48** as an oil (8.71 g, 83%): <sup>1</sup>H NMR (CDCl<sub>3</sub>), 1.30 (t, 6 H, CH<sub>3</sub>), 2.78 (m, 4 H, CH<sub>2</sub>CH<sub>2</sub>), 4.30 (q, 2H, CH<sub>2</sub>), 7.79 (m, 4H, ArH), 9.65 (t, 1H, CHO).

#### *Ethyl* 2-ethoxycarbonyl-2-phthalimido-5-N-tritylimino-pentanoate **49**

This compound was prepared from **48** in the same way as described for **45**. Crystallization from ether–hexane (4.66 g, 86%): mp = 124°C; IR 1760, 1725, 1665 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>), 1.28 (t, 6 H, CH<sub>3</sub>), 2.60 (m, 2 H, CH<sub>2</sub>CH=N), 2.88 (t, 2H, CCH<sub>2</sub>), 4.32 (q, 4 H, OCH<sub>2</sub>), 7.22 (m, 16 H, ArH - CH=N), 7.80 (m, 4 H, ArH); <sup>13</sup>C NMR (CDCl<sub>3</sub>), 13.78 (CH<sub>3</sub>), 28.85 (CH<sub>2</sub>CH=N), 31.65 (CCH<sub>2</sub>), 62.20 (OCH<sub>2</sub>), 123.45 - 131.40 - 134.20 (C<sub>6</sub>H<sub>4</sub>), 126.49 - 127.48 - 129.58 - 145.53 (C<sub>6</sub>H<sub>5</sub>), 1611.82 (CH=N). Anal C<sub>35</sub>H<sub>30</sub>N<sub>2</sub>O<sub>4</sub> (C, H, N).

### 2,5-Diamino-5-phosphonopentanoic acid 4

This compound was prepared via 50 in the same way as 3 (table I).

#### Mass spectra

FAB mass spectra of the diphosphonic acids from the C5 and C4 series were all characterized by a  $[MH]^+$  and a  $[(MH)-H_3PO_3-NH_2R]^+$  peak where *R* is H 5, alkyl 6–9, 12 or acyl 10, 11. When *R* is ethyl 7, isopropyl 8 or *t*-butyl 9,  $[RNH_2=CH-CH=CH_2]^+$  is also an important peak in the C5 group. In the C6

# 514

# Table II. $\alpha, \omega$ -alkanediimines.

n°	R	n	Molecular Formula	mp℃* bp₁℃	Yield %	IR	<sup>1</sup> H NMR (CDCl₃)	<sup>13</sup> C N <b>MR</b> (CDCl <sub>3</sub> )
23	TRI	1	C43H38N2	134°C*	85	3030,2000,1700,	$1.99(q, 2H, CH_2), 2.48$	21.98(CH <sub>2</sub> ), 36.41
						1653,1600,1500,		
						1450,760,710	).25(m, 5n, ALH, CH=N)	123.01-143.92(ALC),
								105.55((41-4))
24	CH2CH3	1	$C_9H_{18}N_2$	51-53°C	29	-	1.20(t,6H,CH <sub>3</sub> );1.88	-
							(q,2H,CH <sub>2</sub> );2.24(m,4H,	
							<u>CH</u> 2CH);3.4O(q,4H,	
							<u>CH</u> 2CH <sub>3</sub> );7.76(t,2H,	
							N=CH)	
		_						
25	$CH(CH_3)_2$	1	C <sub>11</sub> H <sub>22</sub> N <sub>2</sub>	52-57°C	65	-	1.18(d,12H,CH3);1.90	-
							$(q, 2n, Cn_2); 2.24 (m, 4n, 2n, 2n, 2n, 2n, 2n, 2n, 2n, 2n, 2n, 2$	
							$(H_{CH_2})$ ; 3. 34 (III, 2H, CH);	
							/./4(t,2H,N= <u>CH</u> )	
26	$C(CH_3)_3$	1	$C_{13}H_{26}N_{2}$	68-72°C	60	-	1.22(s,18H,CH <sub>3</sub> );1.84	-
							(q,2H,CH <sub>2</sub> );2.29(m,4H,	
							CH <u>CH</u> 2);7.80(t,2H,N≖	
							<u>(CH</u> )	
27	TRT	0	CapHacNab	205°C*	81	3030,2000,1700,	$2.80(m, 4H, CH_2);$	32.24(CH2),126.49-
		_	-42-50-2			1165.1600.1500.	7.20-7.22(m,30H,ArH),	127.4-129.58-145.64
						1450.760.710	7.38(m.2H.CH=N)	(ArC),162,75(CH=N)
							, , , , , , , , , , , , , , , , , , , ,	· · · · · · · · · · · · · · · · · · ·
28	CH(CH <sub>3</sub> ) <sub>2</sub>	0	C10H20N2	48-50°C	32	-	1.15(d,12H,CH <sub>3</sub> );1.50	-
			20 20 2				(d,4H, <u>CH</u> 2,J <sub>HH</sub> =7.6Hz);	
							3.33(m,2H,CH);7.83	
							(t,2H,N=CH)	
29	CH(CH3)2	2	$C_{12}H_{24}N_2$	58-66°C	85	-	1.18(d,12H,CH <sub>3</sub> );1.68	-
							(m, 4H, CH <sub>2</sub> CH <sub>2</sub> ); 2.26(m, 4	н,
							CH <u>CH</u> <sub>2</sub> );3.10(m,2H,CH);	
							7.82(t,2H,N=CH)	
30	C(CH <sub>3</sub> ) <sub>3</sub>	2	C14H28N2	75-81°C	70	-	1.20(s,18H,CH <sub>3</sub> );1.66	-
			•				(m,4H,CH <sub>2</sub> );2.3O(m,4H,	
							CHCH2);8.20(t,2H,N=CH)	

series **13–15** [MH]<sup>+</sup>, [(MH)-H<sub>3</sub>PO<sub>3</sub>]<sup>+</sup>, [(MH)-H<sub>3</sub>PO<sub>3</sub>-NH<sub>2</sub>R]<sup>+</sup> were found. The cyclic compounds **21**, **22** were characterized by a [MH]<sup>+</sup> and a [(MH)-H<sub>3</sub>PO<sub>3</sub>]<sup>+</sup> peak. The monophosphonomonocarboxylic acids **3**, **4** both showed a [MH]<sup>+</sup> peak. The C5 compound **3** also showed a [(MH)-H<sub>3</sub>PO<sub>3</sub>]<sup>+</sup> and a [(MH)-H<sub>3</sub>PO<sub>3</sub>]<sup>+</sup> and a [(MH)-H<sub>3</sub>PO<sub>3</sub>-NH<sub>3</sub>]<sup>+</sup> peak.

# Acknowledgments

We are grateful to Dr Mannes (Agfa-Gevaert, Antwerp) and L Pieters for the NMR spectra, A Gergely and M Claeys for the mass spectra and S Pattyn and D Vanden Berghe for the microbiological data. This work was supported by a grant from the Belgian National Fund of Scientific Research. We thank The British Council for supporting the co-operation between the University of Antwerp and the Sunderland Polytechnic.

# References

- 1 Ghuysen JM (1980) *Topics in Antibiotic Chemistry* (Sammes PG, ed) Ellis Horwood Ltd, Chichester, UK, vol 5
- 2 Berges DA, De Wolf WE Jr, Dunn GL, Grappel SF, Newman DJ, Taggart JJ, Gilvarg C (1986) J Med Chem 29, 89–95
- 3 Berges DA, De Wolf WE Jr, Dunn GL, Newman DJ, Schmidt SJ, Taggart JJ, Gilvarg C (1986) J Biol Chem 261, 6160–6167
- 4 Baumann RJ, Bohme EH, Wiseman JS, Vaal M, Nichols JS (1988) Antimicrob Agents Chemother 32, 1119–1123
- 5 Higgins W, Tardif C, Richaud C, Krivanek MA, Cardin A (1989) Eur J Biochem 186, 137–143
- 6 Kelland JG, Arnold LD, Palcic MM, Pickard MA, Vederas JC (1986) J Biol Chem 261, 13216–13223
- 7 Girodeau JM, Agouridas C, Masson M, Pineau R, Le Goffic F (1986) J Med Chem 29, 1023–1030
- 8 Abo-Ghalia M, Michaud C, Blanot D, van Heijenoort J (1985) Eur J Biochem 153, 81–87
- 9 Abo-Ghalia M, Flegel M, Blanot D, van Heijenoort J (1988) Int J Pept Protein Res 32, 208–222
- 10 Simmonds DH (1954) Biochem J 58, 520-523
- 11 Cavalleri B, Volpe G, Sartori G, Carniti G, White R (1964) *Il Farmaco Ed Sci* 29, 257–280
- 12 Pedersen E (1970) Dan Kem 51 (4), 53–55; (1970) Chem Abstr 73, 233
- 13 Cavalleri B, Volpe G, Pallanza R (1972) Il Farmaco Ed Sci 27, 829–841

- 14 Hanus J, Tolman V, Veres K (1973) Coll Czech Chem Commun 38, 1212–1220
- 15 Chaloupka J, Strnadova M, Caslavska J, Veres K (1974) Z Allg Mikrobiol 14 (4), 283–296
- 16 Allen JG, Atherton FR, Hall MJ, Hassal CH, Holmes SW, Lambert RW, Nisbet LJ, Ringrose PS (1979) Antimicrob Agents Chemother 15 (5) 684–695
- 17 Atherton FR, Hall MJ, Hassal CH, Lambert RW, Lloyd WJ, Ringrose PS (1979) Antimicrob Agents Chemother 15, 696–705
- 18 Berlin KD, Taylor HA (1964) J Am Chem Soc 86, 3962–3866
- 19 Kowalik J, Kupczyk-Subotkowska L, Mastalerz P (1981) Synthesis 57–58
- 20 Kudzin ZH, Kotynski A (1980) Synthesis 1028-1032
- 21 Birum GH (1974) J Org Chem 39, 209–213
- 22 Oleksyszyn J, Subotkowska L, Mastalerz P (1979) Synthesis 985–986
- 23 Dehnel A, Lavielle G (1978) Bull Soc Chim Fr II 95-96
- 24 Diel PJ, Maier L (1984) Phosphorus Sulfur 20, 313
- 25 Rachon J, Schöllkopf U, Wintel T (1981) Liebigs Ann Chem 709-718
- 26 Soroka M (1990) Liebigs Ann Chem 331-334
- 27 Issleib K, Döpfer KP, Balszuweit A (1983) Phosphorus Sulfur 14, 171–178
- 28 Soroka M, Zygmunt J (1988) Synthesis 370-372
- 29 Lubig R, Kusch P, Röper K, Zahn H (1981) Monatsch Chem 112, 1313–1323
- 30 Brown HC, Gundu Rao C, Kulkarni SU (1979) Synthesis 704–705
- 31 Michaud C, Blanot D, van Heijenoort J (1989) Second Forum on Peptides (Aubry A, Marroud M, Vitoux B, eds) Colloque INSERM/John Libbey Eurotext Ltd, 174, 77-80
- 32 Mengin-Lecreulx D, Flouret B, van Heijenoort J (1982) J Bacteriol 151, 1109–1117
- 33 Mengin-Lecreulx D, Michaud C, Richaud C, Blanot D, van Heijenoort J (1970) *J Bacteriol* 170, 2031–2039
- 34 Flouret B, Mengin-Lecreulx D, van Heijenoort J (1981) J Anal Biochem 114, 59–63
- 35 Lejczak B, Starzemska H, Mastalerz P (1981) *Experientia* 37, 401–402
- 36 Sosnovsky G, Lukszo J, Cravela E, Zuretti MF (1985) J Med Chem 28, 1350–1354
- 37 Kafarski P, Mastalerz P (1984) Beiträge Zur Wirkstofforschung Heft 21. Aminophosphonates, Natural Occurrence, Biochemistry and Biological Properties (Oehme P, ed) Akademie Industrie-Komplex, Berlin-Friedrichsfeld, 1–110
- 38 English J, Barber G (1949) J Am Chem Soc 71, 3312–3313
- 39 Fakstorp J, Raleigh D, Schniepp LE (1950) J Am Chem Soc 72, 872–875