Synthesis of *p*-trifluoroacetamidophenyl 6-deoxy-2-O-{3-O-[2-O-methyl-3-O-(2-O-methyl- α -D-rhamnopyranosyl)- α -L-fucopyranosyl]- α -L-rhamnopyranosyl}- α -Ltalopyranoside: a spacer armed tetrasaccharide glycopeptidolipid antigen of *Mycobacterium avium* serovar 20

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ABSTRACT

The synthesis of the title tetrasaccharide glycoside 38 is reported. p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (4), 3-O-acetyl-2,4-di-O-benzyl- α -L-rhamnopyranosyl trichloroacetimidate (7), methyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- β -L-fucopyranoside (15), 3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl bromide (16), and ethyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- α -D-rhamnopyranoside (33) were prepared as intermediates. Compound 4 was glycosylated with imidate 7 as well as with methyl 3-O-acetyl-2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside (9), affording the same disaccharide derivative 8. Deacetylation of 8 gave crystalline 17. Condensation of 17 with both fucosyl donors 15 and 16 yielded the same trisaccharide derivative 18 stereoselectively. Compound 18 was also prepared by the coupling of 4 with disaccharide glycosyl donor 20. After deacetylation of 18 (\rightarrow 34), methyl triflate-promoted glycosylation with compound 33 resulted in tetrasaccharide 35. Conversion of the p-nitrophenyl group of 35 into the p-trifluoroacetamidophenyl group (\rightarrow 36) and removal of the protecting groups gave the title tetrasaccharide glycoside 38.

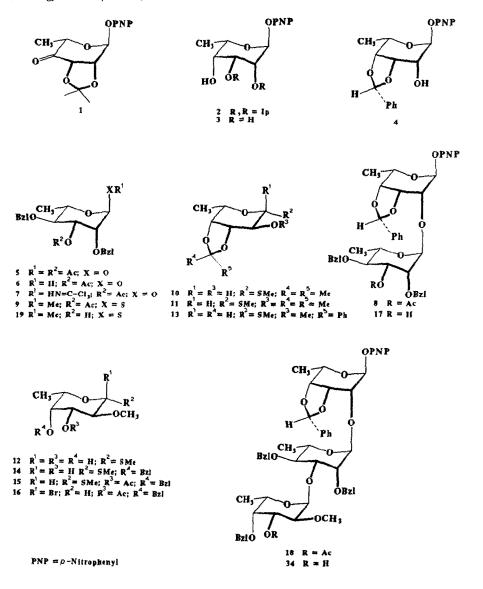
INTRODUCTION

The type-specific surface antigens of the Mycobacterium avium-M. intracellulare-M. scrofulaceum (MAIS) serocomplex are glycopeptidolipids (GPLs) in which the outer mono- or oligo-saccharide residues are responsible for the specific immunoreactions¹. The growing interest in members of the MAIS complex originates mainly from the occurrence of several serovariants as opportunistic pathogens in persons with acquired immunodeficiency syndrome (AIDS)². Recently, a similar observation has also been made in the case of M. kansasii³. Among the structurally established antigens of the MAIS serogroup, the serovar 20 has one of the most complex structures: 2-O-Me- α -D-Rha p-(1 \rightarrow 3)-2-O-Me- α -L-Fuc p-(1 \rightarrow 3)- α -L-Rha p-(1 \rightarrow 2)-6d- α -L-Tal p-(1 \rightarrow Tetrapeptide)⁴. The reducing disaccharide unit $[\alpha$ -L-Rha p-(1 \rightarrow 2)-6d- α -L-Tal p-(1 \rightarrow] is the so-called conservative core-region and its synthesis has been published⁵. We wished to prepare the tetrasaccharide hapten in the form of the glycoside **38** suitable for coupling to proteins. Our strategy was to use benzyl or benzyl-equivalent persistent protecting groups in the synthesis in order to be able to remove them under mild conditions in the last step of the synthetic procedure.

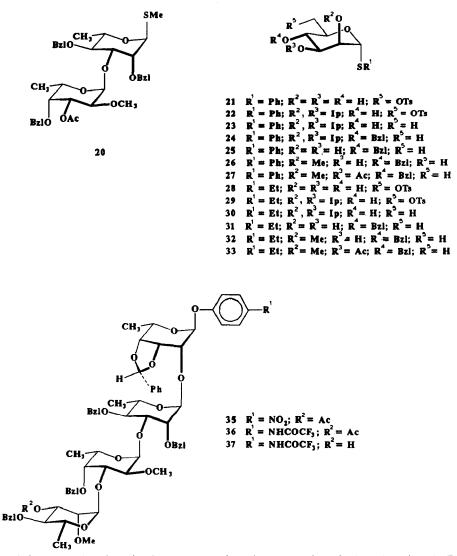
RESULTS AND DISCUSSION

For the synthesis of the title tetrasaccharide glycoside **38**, the following intermediates were chosen to meet our criteria: *p*-nitrophenyl *endo*-3,4-*O*-benzylidene-6deoxy- α -L-talopyranoside (4), 3-*O*-acetyl-2,4-di-*O*-benzyl- α -L-rhamnopyranosyl trichloroacetimidate (7), methyl 3-*O*-acetyl-4-*O*-benzyl-2-*O*-methyl-1-thio- β -Lfucopyranoside (15), 3-*O*-acetyl-4-*O*-benzyl-2-*O*-methyl- α -L-fucopyranosyl bromide (16), and ethyl 3-*O*-acetyl-4-*O*-benzyl-2-*O*-methyl-1-thio- α -D-rhamnopyranoside (33).

p-Nitrophenyl 2,3-*O*-isopropylidene- α -L-rhamnopyranoside^{6,7} was oxidised by pyridinium dichromate to obtain p-nitrophenyl 6-deoxy-2,3-O-isopropylidene- α -L*lyxo*-hexopyranosid-4-ulose (1) in crystalline form, which was reduced by NaBH₄ to *p*-nitrophenyl 6-deoxy-2,3-O-isopropylidene- α -L-talopyranoside (2). The crystalline 2 was hydrolysed and p-nitrophenyl 6-deoxy- α -L-talopyranoside (3) was isolated. Kinetically controlled benzylidenation⁸ of 3 resulted in compound 4 exclusively $(\delta_{\rm H}: 5.78; \delta_{\rm C}: 104.30 \text{ ppm}; \text{ acetalic proton}^9 \text{ and acetalic carbon, respectively). A}$ suitably substituted rhamnosyl donor 7 was prepared from 2,4-di-O-benzyl-Lrhamnose¹⁰ by acetylation, giving 1,3-di-O-acetyl-2,4-di-O-benzyl- α -L-rhamnopyranose (5), followed by the removal of the anomeric acetyl group using hydrazine acetate¹¹, yielding 3-O-acetyl-2,4-di-O-benzyl-L-rhamnose (6), which was converted into 7 by treatment with trichloroacetonitrile in the presence of DBU^{12} . The trimethylsilyl triflate-catalysed glycosylation of 4 with the imidate 7 at low temperature (-50°C) resulted in *p*-nitrophenyl 2-O-(3-O-acetyl-2,4-di-O-benzyl- α -Lrhamnopyranosyl)-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (8). Under these reaction conditions, no isomerisation of the benzylidene ring occurred ($\delta_{\rm H}$: 5.80 and δ_{C} : 104.65 ppm), and only the α -anomeric interglycosidic bond was formed. Compound 8 was also obtained when methyl 3-O-acetyl-2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside¹³ (9) was used as glycosyl donor and activated¹⁴ by methyl triflate. The yield of this latter coupling reaction (65-70%) was comparable with the yield of the imidate procedure. Saponification (Zemplén) of compound 8 afforded crystalline p-nitrophenyl endo-3,40-benzylidene-6-deoxy-2-0-(2,4-di-0benzyl- α -L-rhamnopyranosyl)- α -L-talopyranoside (17), the ideal glycosyl acceptor for the synthesis of higher oligosaccharides containing this conservative core-region. The third synthon, compound 15, was prepared by the following route: methyl 1-thio- β -L-fucopyranoside¹⁵ was converted into the 3,4-O-isopropylidene



derivative¹⁶ (10), and the free HO-2 was methylated to obtain methyl 3,4-O-isopropylidene-2-O-methyl-1-thio- β -L-fucopyranoside (11). Acid hydrolysis of the isopropylidene group gave methyl 2-O-methyl-1-thio- β -L-fucopyranoside (12) which was an excellent derivative for kinetically controlled benzylidenation⁸, giving rise to methyl *endo*-3,4-O-benzylidene-2-O-methyl-1-thio- β -L-fucopyranoside (13). Its reductive ring opening¹⁷ resulted in methyl 4-O-benzyl-2-O-methyl-1-thio- β -Lfucopyranoside (14) exclusively. Conventional acetylation of 14 yielded methyl



38 2-OMe- α -D-Rha *p*-(1 \rightarrow 3)-2-OMe- α -L-Fuc *p*-(1 \rightarrow 3)- α -L-Rha *p*-(1 \rightarrow 2)-6d- α -L-Tal *p*-(1 \rightarrow OPTFAAP) PTFAAP = *p*-Trifluoroacetamidophenyl

3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- β -L-fucopyranoside (15) which was one of our fucosyl donors. Compound 15 was converted into the bromo sugar derivative (16) by bromine¹⁸. Both glycosyl donors (15 and 16) were used for coupling. The reaction of 17 with 15 was promoted by CuBr₂-Bu₄NBr¹⁹ to give *p*-nitrophenyl 2-O-[3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)-2,4-di-O-benzyl- α -L-rhamnopyranosyl]-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (18) in a yield of 63%. Condensation of 17 with 16, using Bu₄NBr as a promoter, resulted in

trisaccharide derivative 18 in a yield of 71%. Under these conditions, no isomerisation of the benzylidene ring occurred and only the 1,2-cis-fucopyranoside derivative was formed. The complete assignment of the ¹³C NMR spectra of the trisaccharide 18 confirmed this finding. Compound 18 was also prepared by block-synthesis. The disaccharide glycosyl donor (20) was obtained in the following way: methyl 2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside¹³ (19) was glycosylated with bromide 16 under the Lemieux²⁰ conditions to give methyl 3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)-2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside (20). The structure of 20 was established by ¹³C NMR spectroscopy. Condensation of compound 4 with disaccharide donor 20, with methyl triflate as promoter, afforded trisaccharide derivative 18. Although the synthesis of 2-Omethyl-D-rhamnose was reported²¹ to give the suitably protected fourth synthon with glycosyl donor as well as glycosyl acceptor ability, we worked out a new procedure. Phenyl 1-thio- α -D-mannopyranoside^{22,23} was converted into phenyl 6-O-(p-toluenesulfonyl)-1-thio- α -D-mannopyranoside (21), which was isopropylidenated with 2,2-dimethoxypropane (\rightarrow 22) and then reduced to phenyl 2,3-O-isopropylidene-1-thio- α -D-rhamnopyranoside (23). Benzylation of 23 gave phenyl 4-Obenzyl-2,3-O-isopropylidene-1-thio- α -p-rhamnopyranoside (24). Acid hydrolysis of the isopropylidene group (\rightarrow 25), followed by a phase-transfer mediated methylation of the axial OH group (\rightarrow 26), and conventional acetylation of 26 resulted in phenyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- α -D-rhamnopyranoside (27). Some pilot experiments showed that glycosylation reactions with 27 proceeded very slowly and yields were low to moderate. To overcome this difficulty, a synthon similar to 27 was prepared, with thioethyl in place of thiophenyl. Ethyl 1-thio- α -Dmannopyranoside^{22,24} was selectively tosylated at position 6. The ethyl 6-O-(ptoluenesulphonyl)-1-thio- α -D-mannopyranoside (28) was isopropylidenated (\rightarrow 29), then reduced to the corresponding *D*-rhamnose derivative 30. Compound 30 was benzylated at position 4 and the isopropylidene group was hydrolysed to give ethyl 4-O-benzyl-1-thio- α -D-rhamnopyranoside (31). Methylation under phase-transfer catalysed conditions²⁵ afforded ethyl 4-O-benzyl-2-O-methyl-1-thio- α -D-rhamnopyranoside (32). Conventional acetylation of 32 gave ethyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- α -D-rhamnopyranoside (33), which proved to be a suitable glycosyl donor for the preparation of our target tetrasaccharide derivative. The glycosyl acceptor was obtained by the removal of the temporary acetyl protecting group from compound 18 (\rightarrow 34). Methyl triflate-promoted glycosylation¹⁴ of 34 with 33 gave the fully protected tetrasaccharide 35. The nitro group in compound 35 was reduced using Adams' catalyst²⁶ and the resulting amino group was acylated with trifluoroacetic anhydride to obtain the *p*-trifluoroacetamidophenyl derivative (36) of the blocked haptenic tetrasaccharide. The acetyl group of 36 was removed (\rightarrow 37) and compound 37 subjected to hydrogenolysis in the presence of 10% Pd-on-carbon catalyst. The smooth removal of the benzyl as well as the benzylidene groups gave the desired target compound 38. The conjugation of these types of glycosides to proteins is well-documented 27 .

EXPERIMENTAL

General methods.—Melting points (uncorrected) were determined on a Kofler apparatus. Optical rotations were measured with a Perkin–Elmer 241 polarimeter. ¹H NMR and ¹³C NMR spectra were recorded with a Bruker WP-200 SY (¹H, 200 MHz) spectrophotometer for solutions in CDCl₃ (internal Me₄Si) or in D₂O (internal, 1,4-dioxane). Reactions were monitored by TLC on Kieselgel G F₂₅₄ (Merck) with detection by charring with H₂SO₄ after examination under UV light. Column chromatography was performed on Kieselgel G (Reanal) or Kieselgel 60 (Merck). All solvents were distilled from appropriate drying agents.

p-Nitrophenyl 6-deoxy-2,3-O-isopropylidene- α -L-lyxo-hexopyranosid-4-ulose (1).—Pyridinium dichromate (16.6 g, 44 mmol) and molecular sieves (4A, 8 g) were added to a stirred solution of *p*-nitrophenyl 2,3-O-isopropylidene- α -L-rhamnopyranoside⁷ (5.0 g, 15.37 mmol) in dry CH₂Cl₂ (60 mL). The reaction mixture was stirred for 1.5 h at room temperature, filtered through a bed of Celite, and evaporated. Column chromatography of the residue resulted in crystalline 1 which was recrystallised from EtOH; 1 (4.03 g, 81%) had mp 101°C; $[\alpha]_D - 182°$ (*c* 0.55, CHCl₃), R_f 0.73 (95:5 CH₂Cl₂-EtOAc); ν_{max}^{KBr} 1740 cm⁻¹ (C=O). NMR data (CDCl₃): ¹H, δ 8.24 and 7.19 (2 m, each 2 H, aromatic), 5.78 (bs, 1 H, H-1) 4.72-4.58 (m, 2 H, H-2,3), 4.19 (q, 1 H, H-5), 1.56 and 1.43 (2 s, each 3 H, CMe₂), 1.4 (d, 3 H, $J_{5,6}$ 7.0 Hz, H-6). Anal. Calcd for C₁₅H₁₇NO₇: C, 55.72; H, 5.30. Found: C, 55.70; H, 5.32.

p-Nitrophenyl 6-deoxy-2,3-O-isopropylidene- α -L-talopyranoside (2).—Compound 1 (3.9 g, 12.06 mmol) was dissolved in 1:1 CH₂Cl₂-MeOH (60 mL), and NaBH₄ (700 mg, 18.5 mmol) was added. After 5 min of stirring, TLC showed the complete disappearance of 1. The mixture was diluted with CH₂Cl₂ (150 mL), washed with water (3 × 50 mL), dried (Na₂SO₄), and evaporated to give a crystalline residue. GLC investigation showed that 2 and the corresponding *rhamno* epimer were produced in a ratio of 98:2. Recrystallisation of the crude product from EtOH yielded 2 (3.32 g, 85%); mp 52-53°C; $[\alpha]_D$ -115° (*c* 1.18, CHCl₃); R_f 0.64 (9:1 CH₂Cl₂-EtOAc). NMR data (CDCl₃): ¹H, δ 8.22 and 7.17 (2 m, each 2 H, aromatic), 5.88 (d, 1 H, $J_{1,2}$ 1.5 Hz, H-1), 4.45-4.29 (m, 2 H, H-2,3), 3.93 (m, 1 H, H-5), 3.71 (m, 1 H, H-4), 2.38 (d, 1 H, OH, disappeared after addition of D₂O), 1.62 and 1.46 (2 s, each 3 H, CMe₂), 1.32 (d, 3 H, H-6). Anal. Calcd for C₁₅H₁₉NO₇: C, 55.38; H, 5.89. Found: C, 55.42; H, 5.85.

p-Nitrophenyl 6-deoxy- α -L-talopyranoside (3).—A solution of 2 (3.0 g, 9.22 mmol) in 6:4 acetic acid-water (60 mL) was kept at 60°C for 2 h, then concentrated. Toluene (3 × 10 mL) was evaporated from the residue, which was crystallised from EtOH to give 3 (2.34 g, 89%), mp 144°C; $[\alpha]_D$ –162° (c 0.89, MeOH); R_f 0.66 (85:15 CH₂Cl₂-MeOH). NMR data (D₂O): ¹³C, δ 99.89 (C-1), 17.13 (CMe). Anal. Calcd for C₁₂H₁₅NO₇: C, 50.53; H, 5.30. Found: C, 50.56; H, 5.32.

p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (4).—p-Toluenesulfonic acid monohydrate (50 mg) was added to a stirred solution of 3 (1.71 g, 5.99 mmol) in α,α -dimethoxytoluene (24 mL) and dry DMF (2.4 mL). After 3 h, TLC revealed one main spot. The mixture was diluted with CH₂Cl₂ (200 mL), and the organic layer was washed with aq NaHCO₃ and water, dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue yielded 4 (1.74 g, 78%); $[\alpha]_D - 106^\circ$ (c 0.68, CHCl₃); R_f 0.39 (1:1 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 8.12–7.00 (m, 9 H, aromatic), 5.78 (s, 1 H, PhCH), 5.51 (d, 1 H, $J_{1,2}$ 5.9 Hz, H-1), 4.55 (dd, 1 H, $J_{3,4}$ 8.2 Hz, H-3), 4.15 (dd, 1 H, $J_{4,5}$ 2 Hz, H-4), 4.02 (m, 1 H, H-2; after deuteration, dd, 1 H, $J_{2,3}$ 3.6 Hz, H-2), 3.91 (m, 1 H, $J_{5,6}$ 6.6 Hz, H-5), 2.75 (d, 1 H, $J_{H,OH}$ 7.65 Hz, OH; after deuteration disappeared), 1.25 (d, 3 H, H-6). For ¹³C NMR data, see Table I. Anal. Calcd for C₁₉H₁₉NO₇: C, 61.12; H, 5.13. Found: C, 61.07; H, 5.10.

3-O-Acetyl-2, 4-di-O-benzyl- α -L-rhamnopyranosyl trichloroacetimidate (7).—2,4-Di-O-benzyl-L-rhamnose¹⁰ (2.87 g, 8.33 mmol) was treated with 1:1 Ac₂O-pyridine (30 mL) overnight. The mixture was concentrated and toluene (3×10 mL) was evaporated from the residue to give 5 (3.3 g, 92%); R_f 0.52 (65:35 hexane-EtOAc). Hydrazine acetate (0.78 g, 8.47 mmol) was added to a stirred solution of this crude product (3.3 g, 7.7 mmol) in DMF (5 mL). The temperature was kept at 50°C for 15 min. When TLC showed that the reaction was complete, the mixture was cooled to room temperature, diluted with EtOAc (150 mL), washed with aq 5% NaCl and water, dried (Na₂SO₄), and concentrated to yield derivative 6 (2.26 g, 76%); R_f 0.35 (65:35 hexane-EtOAc). DBU¹¹ (0.294 mL, 1.96 mmol) was added to a solution of 6 (570 mg, 1.47 mmol) in dry CH₂Cl₂ (6 mL) and trichloroacetonitrile (1.47 mL, 14.7 mmol). The mixture was stirred for 15 min and then concentrated. Column chromatography of the residue gave 7 (0.67 g, 85%); $[\alpha]_{\rm D}$ -5.9° (c 0.75 CHCl₃); R_f 0.68 (7:3 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 8.59 (s, 1 H, = NH), 7.40–7.22 (m, 10 H, aromatic), 6.28 (d, 1 H, $J_{1,2}$ 2 Hz, H-1), 5.23 (dd, 1 H, J_{3,4} 9.4 Hz, H-3), 4.79–4.56 (m, 4 H, 2 PhCH₂), 4.10 (dd, 1 H, J_{2.3} 3.5 Hz, H-2), 4.05–3.92 (m, 1 H, H-5), 3.71 (dd, 1 H, J_{4.5} 9.1 Hz, H-4), 1.98 (s, 3 H, OAc), 1.37 (d, 3 H, J_{5.6} 6 Hz, H-6). Anal. Calcd for C₂₄H₂₆Cl₃NO₆: C, 54.30; H, 4.94. Found: C, 54.43; H, 4.86.

p-Nitrophenyl 2-O-(3-O-acetyl-2,4-di-O-benzyl- α -L-rhamnopyranosyl)-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (8).—Procedure A. A solution of 4 (373 mg, 1 mmol) and 7 (531 mg, 1 mmol) in CH₂Cl₂ (8 mL) containing molecular sieves (4A, 1.5 g) was stirred for 30 min under Ar. A solution of trimethylsilyl triflate (90 μ L, 0.5 mmol) in CH₂Cl₂ (2 mL) was added dropwise at -40°C, and, after 10 min, pyridine was added. The mixture was filtered through Celite, concentrated, and co-concentrated with toluene (3 × 5 mL). Column chromatography (3:2 hexane-EtOAc) of the residue gave 8 (560 mg, 76%); $[\alpha]_D - 96^\circ$ (c 0.66, CHCl₃); R_f 0.59.

Procedure B. A solution of 4 (1.12 g, 3 mmol) and 9^{13} (1.5 g, 3.6 mmol) in dry CH₂Cl₂ (20 mL) containing molecular sieves (4A, 6 g) was stirred for 30 min under Ar, then methyl triflate (2.35 mL, 21.6 mmol) was added. After 1.5 h, pyridine was injected and the mixture was filtered through Celite, concentrated, and co-con-

Carbon	Chemical shifts (ppm)				
	4	8	20	18	38
6-deoxy-α-L-Talp					
1	99.68	98.39		98.76	99.61
2	66.65	71.86		72.13	78.65
3	75.00	73.14		73.26	72.60
1	76.40	76.30		76.39	69.19
5	68.10	68.13		68.73	69.19
6	15.60	15.40		15.49	16.74
C_a^{b}	104.30	104.65		104.75	
J _{C-1,H-1} (Hz)					172.9
а <i>-L-Rha</i> р					
1		95.32	82.38	94.76	103.42
2		76.12	76.03	75.91	70.92
3		71.05	78.33	77.91	78.16
4		78.74	79.24	78.38	72.48
5		67.40	68.72	67.51	70.72
5		17.91	17.91	18.12	17.63
И _{С-1,H-1} (Hz)					172.2
<i>х-L-Fuc</i> р			00.00	00.01	00.00
1			99.39	98.91	99.08
2			79.24	78.38	77.60
3			71.08	71.16	77.30
4			80.26	79.84	72.18
5			66.16	66.38	66.61
5			16.36	16.41	16.18
OCH ₃			59.30	59.30	59.55
J _{C-1,H-1} (Hz)					~ 170
α- <i>D-Rha</i> p 1					99.00
2					81.27
3					71.06
4					73.40
5					67.87
5 6					17.57
					58.80
OCH ₃					171.9
J _{C-1,H-1} (Hz)					171.9

TABLE I

¹³C NMR data of some selected compounds ^a

^a All spectra were recorded in CDCl₃ except that of 38 which was measured in D₂O. ^b C_a = acetalic carbon.

centrated with toluene (4 × 15 mL). Column chromatography (3:2 hexane-EtOAc) of the residue gave 8 (1.56 g, 70%); $[\alpha]_D - 97^\circ$ (c 0.46, CHCl₃); R_f 0.59. NMR data (CDCl₃): ¹H, δ 8.20-7.10 (m, 19 H, aromatic), 5.80 (s, 1 H, PhCH), 5.65 (d, 1 H, $J_{1,2}$ 6.5 Hz, H-1), 5.09 (dd, 1 H, $J_{2',3'}$ 4, $J_{3',4'}$ 9 Hz, H-3'), 4.99 (d, 1 H, $J_{1',2'}$ 2 Hz, H-1'), 3.60 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.5$ Hz, H-4'), 1.92 (s, 3 H, OAc), 1.34 and 1.28 (2 d, each 3 H, H-6,6'). For ¹³C NMR data, see Table I. Anal. Calcd for C₄₁H₄₃NO₁₂: C, 66.38; H, 5.84. Found: C, 66.27; H, 5.80.

p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-(2,4-di-O-benzyl- α -L-rhamnopyranosyl)- α -L-talopyranoside (17).—Sodium methoxide (catalytic amount) was added to a solution of **8** (1.5 g, 2.02 mmol) in dry MeOH (50 mL) which was then stirred overnight. The solution was neutralised with Amberlite IR-120 (H⁺) resin, filtered, and concentrated. The residue was crystallised from EtOAc-hexane to give 17 (1.29 g, 91%); mp 188–190°C; $[\alpha]_D$ –84° (c 0.23, CHCl₃). NMR data (CDCl₃): ¹H, δ 8.20–7.10 (m, 19 H, aromatic), 5.80 (s, 1 H, PhCH), 5.51 (d, 1 H, H-1), 4.98 (bs, 1 H, H-1'), 2.25 (d, 1 H, OH), 1.38 and 1.29 (2 d, each 3 H, H-6,6'). Anal. Calcd for C₃₉H₄₁NO₁₁: C, 66.94; H, 5.91. Found: C, 66.99; H, 5.98.

Methyl 3,4-O-isopropylidene-1-thio- β -L-fucopyranoside (10).—p-Toluenesulfonic acid monohydrate (100 mg) was added to a solution of methyl 1-thio- β -Lfucopyranoside¹⁵ (5.0 g, 21.3 mmol) in 2,2-dimethoxypropane (50 mL). After 30 min, solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂, washed with water, dried (Na₂SO₄), filtered, and concentrated. The residue (5.68 g, 94%) was crystallised from cyclohexane to give 10; mp 69°C; $[\alpha]_D - 44^\circ$ (c 0.86, CHCl₃); R_f 0.50 (9:1 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 4.14 (d, 1 H, J_{1,2} 10.1 Hz, H-1), 4.07–4.01 (m, 2 H, H-3,4), 3.89 (m, 1 H, H-5), 3,56 (m, 1 H, H-2), 2.63 (bs, 1 H, OH), 2.22 (s, 3 H, SMe), 1.54 and 1.37 (2 s, 6 H, CMe₂). Anal. Calcd for C₁₀H₁₈O₄S: C, 51.26; H, 7.74; Found: C, 51.15; H, 7.69.

Methyl 3,4-O-isopropylidene-2-O-methyl-1-thio- β -L-fucopyranoside (11).— Powdered KOH (4 g) was added to a solution of 10 (5.5 g, 22.14 mmol) in dry DMF (25 mL) and the mixture was cooled to 0°C. MeI (3.5 mL) was added and the mixture was stirred for 4 h. The mixture was diluted with CH₂Cl₂ (300 mL), the inorganic salts were filtered off, and the organic layer was washed with water, dried (Na₂SO₄), filtered, and concentrated. The residue (4.98 g, 85%) was crystallised from cyclohexane to yield 11; mp 95°C; $[\alpha]_D - 1.5^\circ$ (c 1.5, CHCl₃); R_f 0.71 (95:5 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 4.19 (d, 1 H, $J_{1,2}$ 10 Hz, H-1), 4.11–3.98 (m, 2 H, H-3,4), 3.78 (m, 1 H, H-5), 3.53 (s, 3 H, OMe), 3.18 (dd, 1 H, H-2), 2.20 (s, 3 H, SMe), 1.52 and 1.34 (2 s, each 3 H, CMe₂), 1.38 (d, 3 H, H-6). Anal. Calcd for C₁₁H₂₀O₄S: C, 53.20; H, 8.12. Found: C, 53.29; H, 8.08.

Methyl 2-O-methyl-1-thio- β -L-fucopyranoside (12).—Compound 11 (2.48 g, 10 mmol) was stirred in 6:4 acetic acid-water (30 mL) for 1 h at 80°C and the mixture was then concentrated and co-concentrated with toluene (4 × 10 mL) and CH₂Cl₂ (4 × 10 mL) to give 12 (1.98 g, 95%); [α]_D + 17° (c 0.86, CHCl₃); R_f 0.30 (3:1 CH₂Cl₂-acetone). NMR data (CDCl₃): ¹H, δ 4.18 (d, 1 H, H-1), 3.59 (s, 3 H, OMe), 2.19 (s, 3 H, SMe), 1.27 (d, 3 H, H-6). Anal. Calcd for C₈H₁₆O₄S: C, 46.14; H, 7.74. Found: C, 46.22; H, 7.70.

Methyl endo-3,4-O-benzylidene-2-O-methyl-1-thio- β -L-fucopyranoside (13).--Compound 12 (2.08 g, 10 mmol) was treated with α, α -dimethoxytoluene, as described for the preparation of compound 4. Purification of the crude product by column chromatography gave 13 (2.25 g, 75.9%); $[\alpha]_D + 17^\circ$ (c 0.99, CHCl₃); R_f 0.74 (9:1 CH₂Cl₂-EtOAc). NMR data (CDCl₃): ¹H, δ 7.58-7.33 (m, 5 H, aromatic), 5.93 (s, 1 H, PhCH), 4.26 (d, 1 H, $J_{1,2}$ 10 Hz, H-1), 4.25 (t, 1 H, $J_{3,4}$ 6 Hz, H-3), 4.13 (dd, 1 H, H-4), 3.90 (m, 1 H, H-5), 3.49 (s, 3 H, OMe), 3.24 (dd, 1 H, $J_{2,3}$ 6 Hz, H-2), 2.20 (s, 3 H, SMe), 1.48 (d, 3 H, H-6). Anal. Calcd for $C_{15}H_{20}O_4S$: C, 60.79; H, 6.80. Found: C, 60.86; H, 6.78.

Methyl 4-O-benzyl-2-O-methyl-1-thio- β -L-fucopyranoside (14).—Compound 13 (880 mg, 2.97 mmol) was hydrogenolysed in 1:1 ether-CH₂Cl₂ (30 mL) with LiAlH₄ (440 mg) and AlCl₃ (880 mg) at room temperature for 30 min. The reaction was terminated by the addition of EtOAc (5 mL) and water (2 mL). The organic layer was decanted and the residue washed with ether (2 × 50 mL). The combined solutions were washed with water (3 × 30 mL), dried (Na₂SO₄), and concentrated. The residue was crystallised from cyclohexane to afford 14 (780 mg, 88%); mp 77°C; [α]_D + 16° (*c* 0.54, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.45–7.28 (m, 5 H aromatic), 4.80 (s, 2 H, PhCH₂), 4.19 (d, 1 H, H-1), 3.65 (s, 3 H, OMe), 3.28 (m, 1 H, H-5), 2.38 (d, 1 H, OH), 2.22 (s, 3 H, SMe), 1.30 (d, 3 H, H-6). Anal. Calcd for C₁₅H₂₂O₄S: C, 60.37; H, 7.43. Found: C, 60.45; H, 7.49.

Methyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- β -L-fucopyranoside (15).—Compound 14 (400 mg, 1.34 mmol) was dissolved in 1:1 pyridine–Ac₂O (12 mL) and left for 3 h. The mixture was concentrated and co-concentrated with toluene. Column chromatography of the residue gave 15 (395 mg, 87%); $[\alpha]_D - 24^\circ$ (c 1.18, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.39–7.23 (m, 5 H, aromatic), 4.84 (dd, 1 H, $J_{2,3}$ 9.7, $H_{3,4}$ 3 Hz, H-3), 4.65 (2 d, 2 H, PhCH₂), 4.26 (d, 1 H, $J_{1,2}$ 9.5 Hz, H-1), 3.54 (s, 3 H, OMe), 2.23 (s, 3 H, SMe), 2.04 (s, 3 H, OAc), 1.23 (d, 3 H, H-6). Anal. Calcd for C₁₇H₂₄O₅S: C, 59.98; H, 7.11. Found: C, 60.07; H, 7.07.

p-Nitrophenyl 2-O-[3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)-2,4-di-O-benzyl- α -L-rhamnopyranosyl]-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (18).—Procedure A. A mixture of 17 (160 mg, 0.23 mmol), 15 (117 mg, 0.34 mmol), CuBr₂ (144 mg, 0.65 mmol), Bu₄NBr (186 mg, 0.58 mmol), and molecular sieves (4A, 1 g) was stirred in 5:1 CH₂Cl₂-DMF (6 mL) for 24 h under Ar. Then the mixture was diluted with CH₂Cl₂, filtered through Celite, washed with aq NaHCO₃ and water, then dried (Na₂SO₄), filtered, and concentrated. Column chromatography (94:6 CH₂Cl₂-EtOAc) yielded 18 (115 mg, 51%).

Procedure B. Bromine (116 μL, 2.26 mmol) was added to a solution of **15** (586 mg, 1.72 mmol) in dry CH₂Cl₂ (8 mL) at 0°C. After stirring the mixture for 10 min at room temperature, the solution was concentrated and co-concentrated with toluene (3×5 mL). The residue (**16**) was dissolved in dry CH₂Cl₂ (8 mL) and this solution was added to a stirred mixture of **17** (600 mg, 0.86 mmol), Bu₄NBr (832 mg, 2.58 mmol), and molecular sieves (4A, 2.4 g) in dry DMF (8 mL). The mixture was stirred for 24 h, diluted with CH₂Cl₂, filtered through Celite, washed with aq NaHCO₃ and water, then dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue afforded **18** (587 mg, 69%); [*α*]_D – 129° (*c* 0.65, CHCl₃). NMR data (CDCl₃): ¹H, δ 8.20–7.10 (m, 24 H, aromatic), 5.84 (s, 1 H, PhCH), 5.63 (d, 1 H, J_{1,2} 6.5 Hz, H-1), 5.22 (dd, 1 H, J_{2",3"} 10.5, J_{3",4"} 3 Hz, H-3), 3.19 (s, 3 H, OMe), 2.04 (s, 3 H, OAc), 1.32, 1.28 and 0.97 (3 d, each 3 H, 3CMe). For ¹³C NMR data, see Table I. Anal. Calcd for C₅₅H₆₁NO₁₆: C, 66.59; H, 6.20. Found: C, 66.48; H, 6.19.

Methyl 3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)-2,4-di-O-benzyl-1-thio- α -L-rhamnopyranoside (20).—Bromine (42 μ L, 0.74 mmol) was added to a solution of 15 (210 mg, 0.62 mmol) in dry CH₂Cl₂ (4 mL) at 0°C. After 10 min, the reaction was worked up as described for 18. The residue (16) was dissolved in dry CH₂Cl₂ (4 mL) and this solution was added to a stirred mixture of 19¹³ (210 mg, 0.62 mmol), Bu₄NBr (189 mg, 0.90 mmol), and molecular sieves (4A, 800 mg) in dry DMF (4 mL). The mixture was stirred for 24 h, then worked up as described for 18 (Procedure B). Column chromatography of the residue gave 20 (228 mg, 61%); [α]_D – 154° (*c* 0.35, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.48–7.22 (m, 15 H, aromatic), 3.29 (s, 3 H, OMe), 2.12 (s, 3 H, SMe), 2.02 (s, 3 H, OAc), 1.31 and 0.93 (2 d, each 3 H, H-6,6'). For ¹³C NMR data, see Table I. Anal. Calcd for C₃₇H₄₆O₉S: C, 66.64; H, 6.95. Found: C, 66.62; H, 6.89.

p-Nitrophenyl 2-O-[3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)-2,4-di-O-benzyl- α -L-rhamnopyranosyl]-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (18).—Procedure C. A solution of 20 (140 mg, 0.21 mmol), and 4 (93 mg, 0.25 mmol) in CH₂Cl₂ (4 mL) containing molecular sieves (4A, 250 mg) was stirred for 30 min under Ar, then methyl triflate (185 μ L, 1.68 mmol) was added. After 4 h, pyridine was injected and the mixture was filtered through Celite, concentrated, and co-concentrated with toluene. Column chromatography of the residue gave 18 (121 mg, 58%).

Phenyl 6-O-(p-toluenesulfonyl)-1-thio-α-D-mannopyranoside (21).—Phenyl 1-thio-α-D-mannopyranoside^{22,23} (9.21 g 33.82 mmol) was treated with *p*-toluene-sulfonyl chloride (9.67 g, 50.72 mmol) in dry pyridine (60 mL) at 0°C overnight. The mixture was concentrated, diluted with CH_2Cl_2 , washed with aq NaHCO₃ and water, then dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue afforded 21 (8.65 g, 60%); $[\alpha]_D$ + 147° (*c* 0.40, CHCl₃); R_f 0.66 (92:8 CH₂Cl₂-MeOH). Anal. Calcd for $C_{19}H_{22}O_7S_2$: C, 53.50; H, 5.20. Found: C, 53.62; H, 5.15.

Phenyl 2,3-O-isopropylidene-6-O-(p-toluenesulfonyl)-1-thio-α-D-mannopyranoside (22).—*p*-Toluenesulfonic acid monohydrate (150 mg) was added to a solution of 21 (8.0 g, 19.30 mmol) in 2,2-dimethoxypropane (30 mL). After 30 min, solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂, washed with water, dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue gave 22 (8.01 g, 89%); $[\alpha]_D$ +99° (*c* 0.9, CHCl₃); R_f 0.51 (6:4 hexane–EtOAc). NMR data (CDCl₃): ¹H, δ 7.75–7.25 (m, 9 H, aromatic), 5.70 (bs, 1 H, H-1), 2.80 (d, 1 H, OH), 2.45 (s, 3 H, ArMe), 1.50 and 1.38 (2 s, each 3 H, CMe₂). Anal. Calcd for C₂₂H₂₆O₇S₂: C, 56.63; H, 5.62. Found: C, 56.71; H, 5.60.

Phenyl 2,3-O-isopropylidene-1-thio- α -D-rhamnopyranoside (23).—Compound 22 (9.17 g, 19.6 mmol) was dissolved in 1:1 benzene-ether (70 mL) and then LiAlH₄ (1.5 g, 39.52 mmol) was added. The mixture was boiled for 10 min under reflux. After cooling, EtOAc was added to decompose the excess of reagent and then water was added to precipitate the aluminium hydroxide. The organic layer was washed with water, dried (Na₂SO₄), filtered, and concentrated. Column chro-

matography of the residue gave a crystalline product that was recrystallised from EtOAc-hexane to afford 23 (4.19 g, 72%); mp 82°C; $[\alpha]_D + 206^\circ$ (c 1.0, CHCl₃); R_f 0.71 (6:4 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 7.51-7.25 (m, 5 H, aromatic), 5.75 (bs, 1 H, H-1), 4.35 (d, 1 H, H-2), 4.20-4.02 (m, 2 H, H-3,5), 3.52-3.40 (m, 1 H, H-4), 2.90 (d, 1 H, OH), 1.54 and 1.37 (2 s, each 3 H, CMe₂), 1.24 (d, 3 H, H-6). Anal. Calcd for C₁₅H₂₀O₄S: C, 60.79; H, 6.80. Found: C, 60.67; H, 6.73.

Phenyl 4-O-benzyl-2,3-O-isopropylidene-1-thio-α-D-rhamnopyranoside (24).— Powdered KOH (3.18 g, 56.67 mmol) and benzyl bromide (4.2 mL) were added to a stirred solution of 23 (4.19 g, 14.1 mmol) in dry DMF (15 mL). After the reaction was complete, MeOH (2 mL) was added to the mixture which was then diluted with CH₂Cl₂. The inorganic salts were filtered off, and the organic layer was washed with water, dried (Na₂SO₄), filtered, and concentrated. The crystalline residue was recrystallised from EtOH to yield 24 (3.71 g, 68%); mp 90°C; $[\alpha]_D$ +220° (c 0.66, CHCl₃); R_f 0.81 (6:4 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 7.50-7.22 (m, 10 H, aromatic), 5.74 (bs, 1 H, H-1), 4.92 and 4.63 (2 d, each 1 H, PhCH₂), 4.39-4.28 (m, 2 H, H-2,3), 4.14 (m, 1 H, H-5), 3.30 (dd, 1 H, H-4), 1.51 and 1.39 (2 s, each 3 H, CMe₂), 1.23 (d, 3 H, H-6). Anal. Calcd for C₂₂H₂₆O₄S: C, 68.36; H, 6.78. Found: C, 68.40; H, 6.70.

Phenyl 4-O-*benzyl-1-thio-α-D-rhamnopyranoside* (25).—Compound 24 (3.71 g, 9.6 mmol) was stirred in 6:4 acetic acid-water (30 mL) for 1 h at 80°C, then concentrated, and co-concentrated with toluene. The crystalline residue was recrystallised from EtOAc-hexane to give 25 (2.69 g, 81%); mp 117°C; $[\alpha]_D$ +232° (*c* 0.90, CHCl₃); R_f 0.33 (6:4 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 7.48-7.25 (m, 10 H, aromatic), 5.47 (d, 1 H, H-1), 4.75 (s, 2 H, PhCH₂), 4.28-4.13 (m, 2 H, H-2,5), 3.98-3.87 (m, 1 H, H-3), 3.42 (dd, 1 H, $J_{3,4} = J_{4,5} = 9.5$ Hz, H-4), 2.81 (bs, 1 H, OH), 2.56 (d, 1 H, OH), 1.35 (d, 3 H, H-6). Anal. Calcd for C₁₉H₂₂O₄S: C, 65.87; H, 6.40. Found: C, 65.97; H, 6.45.

Phenyl 4-O-benzyl-2-O-methyl-1-thio-α-D-rhamnopyranoside (26).—A mixture of 25 (2.69 g, 7.7 mmol), Bu₄NBr (620 mg, 1.92 mmol), MeI (2.5 mL, 38.53 mmol), and aq NaOH (1 mL of a 20% solution) in CH₂Cl₂ (20 mL) was stirred at room temperature for 1 day. Then the mixture was diluted with CH₂Cl₂, washed with water, dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue afforded 26 (1.56 g, 57%); $[\alpha]_D$ +195° (*c* 0.30, CHCl₃); R_f 0.71 (93:7 CH₂Cl₂-EtOAc). NMR data (CDCl₃): ¹H, δ 7.51-7.21 (m, 10 H, aromatic), 5.58 (d, 1 H, $J_{1,2}$ 1.1 Hz, H-1), 4.90 and 4.71 (2 d, each 1 H, PhCH₂), 4.17 (dd, 1 H, $J_{5,6}$ 6 Hz, H-5), 3.95 (dt, 1 H, $J_{3,4}$ 9, $J_{3,OH}$ 8.8 Hz, H-3), 3.75 (dd, 1 H, $J_{2,3}$ 3.5 Hz, H-2), 3.47 (s, 3 H, OMe), 3.34 (dd, 1 H, $J_{4,5}$ 9 Hz, H-4), 2.5 (d, 1 H, OH), 1.42 (d, 3 H, $J_{5,6}$ 6 Hz, H-6). Anal. Calcd for C₂₀H₂₄O₄S: C, 66.64; H, 6.71. Found: C, 66.56; H, 6.73.

Phenyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio- α -D-rhamnopyranoside (27). Compound 26 (560 mg, 1.55 mmol) was dissolved in 1:1 pyridine-Ac₂O (20 mL) and left for 3 h. Then the mixture was concentrated and co-concentrated with toluene. Column chromatography of the residue yielded **27** (575 mg, 92%); $[\alpha]_D$ + 156° (c 0.95, CHCl₃); R_f 0.87 (93:7 CH₂Cl₂-EtOAc). NMR data (CDCl₃): ¹H, δ 7.52-7.23 (m, 10 H, aromatic), 5.52 (d, 1 H, $J_{1,2}$ 1 Hz, H-1), 5.19 (dd, 1 H, $J_{3,4}$ 9 Hz, H-3), 4.70 (ABq, 2 H, PhC H_2), 4.24 (m, 1 H, $J_{5,6}$ 6 Hz, H-5), 3.92 (dd, 1 H, $J_{2,3}$ 3 Hz, H-2), 3.63 (dd, 1 H, $J_{4,5}$ 9 Hz, H-4), 3.43 (s, 3 H, OMe), 2.08 (s, 3 H, OAc), 1.33 (d, 3 H, H-6). Anal. Calcd for C₂₂H₂₆O₅S: C, 65.65; H, 6.51. Found: C, 65.76; H, 6.60.

Ethyl 6-O-(p-toluenesulfonyl)-1-thio-α-D-mannopyranoside (28).—p-Toluenesulfonyl chloride (6.25 g, 32.78 mmol) was added to a solution of ethyl 1-thio-α-Dmannopyranoside^{22,24} (4.9 g, 21.85 mmol) in dry pyridine (30 mL) and kept at 0°C overnight. The mixture was then concentrated, diluted with CH₂Cl₂ (200 mL), washed with aq NaHCO₃ and water, dried (Na₂SO₄), and concentrated. Column chromatography of the residue gave **28** (5.04 g, 61%); $[\alpha]_D$ +115° (c 0.34, MeOH); R_f 0.31 (92:8 CH₂Cl₂-MeOH). NMR data (CDCl₃): ¹H, δ 7.8 and 7.34 (2 d, each 2 H, aromatic), 5.26 (bs, 1 H, $J_{1,2}$ 1 Hz, H-1), 2.52 (m, 2 H, MeCH₂S), 2.42 (s, 3 H, ArMe), 1.21 (t, 3 H, MeCH₂S). Anal. Calcd for C₁₅H₂₂O₇S₂: C, 47.60; H, 5.86. Found: C, 47.51; H, 5.81.

Ethyl 2,3-O-*isopropylidene*-6-O-(p-*toluenesulfonyl*)-1-thio-α-D-mannopyranoside (29).—p-Toluenesulfonic acid monohydrate (100 mg) was added to a solution of 28 (2.6 g, 6.87 mmol) in 2,2-dimethoxypropane (15 mL). After 25 min, solid NaHCO₃ was added. The mixture was diluted with CH₂Cl₂, washed with water, dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue afforded 29 (2.56 g, 89%); $[\alpha]_D$ +86° (c 0.20, CHCl₃), R_f 0.49 (6:4 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 7.79 and 7.34 (2 d, each 2 H, aromatic), 5.49 (bs, 1 H, H-1), 2.51 (m, 2 H, MeCH₂S), 2.44 (s, 3 H, ArMe), 1.49 and 1.33 (2 s, each 3 H, CMe₂), 1.25 (t, 3 H, MeCH₂S). Anal. Calcd for C₁₈H₂₆O₇S₂: C, 51.65; H, 6.26. Found: C, 51.69; H, 6.19.

Ethyl 2,3-O-*isopropylidene-1-thio-α-D-rhamnopyranoside* (30).—Compound 29 (2.4 g, 5.73 mmol) was dissolved in 1:1 benzene–ether (20 mL), and then LiALH₄ (435 mg, 11.46 mmol) was added. The mixture was boiled for 10 min under reflux. After cooling, EtOAc was added to decompose the excess of reagent and then water was added to precipitate the aluminium hydroxide. The organic layer was decanted, then washed with water, dried (Na₂SO₄), filtered, and concentrated. Column chromatography of the residue yielded 30 (1.16 g, 81%); $[\alpha]_D + 178^\circ$ (*c* 1.2, CHCl₃); R_f 0.52 (6:4 hexane–EtOAc). NMR data (CDCl₃): ¹H, δ 5.52 (bs, 1 H, H-1), 4.18 (d, 1 H, H-2), 4.09–3.90 (m, 2 H, H-3,5), 3.43 (m, 1 H, H-4; after addition of D₂O, dd), 2.61 (m, 2 H, MeCH₂S), 1.55 and 1.35 (2 s, each 3 H, CMe₂), 1.31 (t, 3 H, MeCH₂S), 1.29 (d, 3 H, H-6). Anal. Calcd for C₁₁H₂₀O₄S: C, 53.20; H, 8.12. Found: C, 53.11; H, 8.13.

Ethyl 4-O-benzyl-1-thio- α -D-rhamnopyranoside (31).—A solution of 30 (1.04 g, 4.19 mmol) and benzyl bromide (0.75 mL, 6.29 mmol) in dry DMF (10 mL) was added dropwise to NaH (201 mg, 8.38 mmol) and the mixture was stirred for 10 min. Then MeOH was added to decompose the excess of hydride, and the mixture

was diluted with CH_2Cl_2 (150 mL), washed with water, dried (Na_2SO_4), and concentrated. The crude product (R_f 0.87 in 6:4 hexane-EtOAc) was stirred in 6:4 acetic acid-water (20 mL) for 1 h at 80°C, then concentrated, and co-concentrated with toluene (4 × 10 mL). The residue was crystallised from hexanc-EtOAc to afford **31** (910 mg, 73%); mp 94-95°C; $[\alpha]_D$ +185° (*c* 0.60, CHCl₃). Anal. Calcd for $C_{15}H_{22}O_4S$: C, 60.37; H, 7.43. Found: C, 60.43; H, 7.50.

Ethyl 4-O-*benzyl*-2-O-*methyl*-1-*thio*-α-D-*rhamnopyranoside* (32).—A mixture of **31** (800 mg, 2.68 mmol), Bu₄NBr (216 mg, 0.67 mmol), MeI (0.83 mL, 13.4 mmol), and aq NaOH (0.3 mL of a 20% solution) in CH₂Cl₂ (6 mL) was stirred for 24 h, then MeI (0.33 mL, 5.3 mmol) was added to the mixture which was stirred for another 24 h. The mixture was diluted with CH₂Cl₂ (60 mL), washed with water (4 × 10 mL), dried (Na₂SO₄), and concentrated. Column chromatography of the residue gave **32** (461 mg, 55%); $[\alpha]_D$ +126° (*c* 0.40, CHCl₃); R_f 0.58 (9:1 CH₂Cl₂-EtOAc). NMR data (CDCl₃): ¹H, δ 7.40–7.25 (m, 5 H, aromatic, 5.39 (bs, 1 H, $J_{1,2}$ 1 Hz, H-1), 4.91 and 4.67 (2 d, each 1 H, PhCH₂), 4.04 (m, 1 H, H-5), 3.90 (m, 1 H; after addition of D₂O, dd, $J_{3,4}$ 9.5 Hz; H-3), 3.59 (dd, 1 H, $J_{2,3}$ 4 Hz, H-2), 3.49 (s, 3 H, OMe), 3.29 (dd, 1 H, $J_{4,5} \sim J_{3,4}$ 9.5 Hz, H-4), 2.62 (m, 2 H, MeCH₂S), 1.32 (d, 3 H, $J_{5,6}$ 6.5 Hz, H-6), 1.30 (t, 3 H, MeCH₂S). Anal. Calcd for C₁₆H₂₄O₄S: C, 61.51; H, 7.74. Found: C, 61.61; H, 7.68.

Ethyl 3-O-acetyl-4-O-benzyl-2-O-methyl-1-thio-α-D-rhamnopyranoside (33).— Compound 32 (400 mg, 1.28 mmol) was dissolved in 1:1 pyridine-Ac₂O (12 mL) and left for 3 h. The mixture was concentrated using toluene (5 × 5 mL) to give 33 (440 mg, 97%); $[\alpha]_D$ +125° (*c* 0.67, CHCl₃). NMR data (CDCl₃): ¹H, δ 7.37-7.25 (m, 5 H, aromatic), 5.31 (d, 1 H, $J_{1,2}$ 1.5 Hz, H-1), 5.16 (dd, 1 H, $J_{3,4}$ 9.5 Hz, H-3), 4.68 (q, 2 H, PhC H_2), 4.13 (m, 1 H, $J_{5,6}$ 6.5 Hz, H-5), 3.75 (dd, 1 H, $J_{2,3}$ 3 Hz, H-2), 3.57 (dd, 1 H, $J_{4,5}$ 9.5 Hz, H-4), 3.45 (s, 3 H, OMe), 2.63 (m, 2 H, MeC H_2 S), 2.06 (s, 3 H, OAc), 1.32 (d, 3 H, H-6), 1.30 (t, 3 H, MeC H_2 S). Anal. Calcd for $C_{18}H_{26}O_5S$: C, 60.99; H, 7.39. Found: C, 61.10; H, 7.35.

p-Nitrophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-[2,4-di-O-benzyl-3-O-(4-O-benzyl-2-O-methyl- α -L-fucopyranosyl)- α -L-rhamnopyranosyl]- α -L-talopyranoside (34).—Sodium methoxide (catalytic amount) was added to a solution of 18 (280 mg, 0.28 mmol) in dry MeOH (8 mL) and stirred overnight. Then the mixture was neutralised with Amberlite IR-120 (H⁺) resin, filtered, and concentrated to give 34 (257 mg, 96%); $[\alpha]_D - 125^\circ$ (c 0.32, CHCl₃). Anal. Calcd for C₅₃H₅₉NO₁₅: C, 67.00; H, 6.26. Found: C, 67.11; H, 6.31.

p-Nitrophenyl 2-O-{3-O-[3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -D-rhamnopyranosyl)-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl]-2,4-di-O-benzyl- α -L-rhamnopyranosyl}-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (35).—A solution of 34 (190 mg, 0.2 mmol) and 33 (92 mg, 0.26 mmol) in dry CH₂Cl₂ (6 mL) containing molecular sieves (4A, 600 mg) was stirred for 30 min under Ar, then methyl triflate (227 μ L, 2.08 mmol) was added. After 2 h, pyridine was injected, and the mixture was filtered through Celite, concentrated, and co-concentrated with toluene. Column chromatography of the residue yielded 35 (220 mg, 89%); $[\alpha]_{\rm D} - 78^{\circ}$ (c 0.39, CHCl₃); R_f 0.55 (55:45 hexane-EtOAc). NMR data (CDCl₃): ¹H, δ 8.20–7.10 (m, 29 H, aromatic), 5.82 (s, 1 H, PhC*H*), 5.63 (d, 1 H, H-1), 5.29 (dd, 1 H, H-3^{'''}), 3.40 and 3.13 (2 s, each 3 H, 2 × OMe), 2.05 (s, 3 H, OAc), 1.39–1.20 (m, 9 H, 3 × CMe), 1.01 (d, 3 H, CMe); ¹³C, δ 104.72 (PhCH), 98.84, 98.61, 98.35 and 94.44 (anomeric carbons), 58.78 and 58.49 (2 × OMe), 21.07 (*Me*CO), 18.13, 18.11, 16.72, and 15.45 (4 × C*Me*). Anal. Calcd for C₆₉H₇₉NO₂₀: C, 66.71; H, 6.41. Found: C, 66.59; H, 6.33.

p-Trifluoroacetamidophenyl 2-O-{3-O-[3-O-(3-O-acetyl-4-O-benzyl-2-O-methyl- α -D-rhamnopyranosyl)-4-O-benzyl-2-O-methyl- α -L-fucopyranosyl]-2,4-di-O-benzyl- α -L-rhamnopyranosyl}-endo-3,4-O-benzylidene-6-deoxy- α -L-talopyranoside (36).—A solution of 35 (110 mg, 0.09 mmol) in EtOAc (4 mL) was hydrogenated over Adams' catalyst (60 mg) at atmospheric pressure for 1 h. Pyridine (1 mL) and trifluoroacetic anhydride (0.4 mL) were added and the reaction mixture was stirred for 20 min, then filtered, and concentrated. Column chromatography of the residue afforded 36 (83 mg, 72%); $[\alpha]_D - 59^\circ$ (c 0.28, CHCl₃); R_f 0.52 (55:45 hexane-EtOAc).

p-Trifluoroacetamidophenyl endo-3,4-O-benzylidene-6-deoxy-2-O-{2,4-di-O-benzyl-3-O-[4-O-benzyl-3-O-(4-O-benzyl-2-O-methyl- α -D-rhamnopyranosyl)-2-O-methyl- α -L-fucopyranosyl]- α -L-rhamnopyranosyl}- α -L-talopyranoside (37).—Sodium methoxide (catalytic amount) was added to a solution of 36 (70 mg, 0.054 mmol) in dry MeOH (4 mL) and stirred overnight. Then the mixture was neutralised with Amberlite IR-120 (H⁺) resin, filtered, and concentrated. Column chromatography of the residue gave 37 (64 mg, 94%); $[\alpha]_D - 74^\circ$ (c 0.19, CHCl₃); R_f 0.35 (55:45 hexane–EtOAc).

p-Trifluoroacetamidophenyl 6-deoxy-2-O-{3-O-[2-O-methyl-3-O-(2-O-methyl- α -D-rhamnopyranosyl)- α -L-fucopyranosyl]- α -L-rhamnopyranosyl}- α -L-talopyranoside (38).—A solution of 37 (50 mg, 0.04 mmol) in acetic acid (8 mL) was hydrogenated at room temperature over 10% Pd-C (20 mg). When TLC (2:1:1 1-butanol-MeOH-water) showed complete reaction, the mixture was filtered and concentrated to give 38 as a white glass (29 mg, 90%); [α]_D - 21° (c 0.21, MeOH). NMR data (D₂O): ¹H, δ 7.50 and 7.18 (2 d, each 2 H, aromatic), 5.68, 5.16, and 5.08 (3 bs, each 1 H) and 5.43 (d, 1 H), anomeric protons, 3.52 and 3.48 (2 s, each 3H, 2 × OMe), 1.32–1.20 (m, 12 H, 4 × CMe). For ¹³C NMR data, see Table I.

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REFERENCES

¹ D. Chatterjee, G.O. Aspinall, and P.J. Brennan, J. Biol. Chem., 262 (1987) 3528-3533.

² R.C. Good, Annu. Rev. Microbiol., 39 (1985) 347-369.

- 3 G.T. Valainis, L.C. Cordona, and D.L. Greer, J. Acquired Immune Defic. Syndr., 4 (1991) 516-520.
- 4 G.O. Aspinall, A.M. Crane, D.W. Gammon, I.H. Ibrahim, N.K. Khare, D. Chatterjee, B. Rivoire, and P.J. Brennan, *Carbohydr. Res.*, 216 (1991) 337-355; G.O. Aspinall, N.K. Khare, R.K. Sood, D. Chatterjee, B. Rivoire, and P.J. Brennan, *ibid.*, 216 (1992) 357-373.
- 5 G.O. Aspinall and K. Takeo, Carbohydr. Res., 121 (1983) 61-77.
- 6 P.J. Garegg, T. Norberg, P. Konradsson, and S.C.T. Svensson, Carbohydr. Res., 122 (1983) 165-167.
- 7 A. Borbás and A. Lipták, Carbohydr. Res., 241 (1993) 99-116.
- 8 J. Kerékgyártó and A. Lipták, Carbohydr. Res., accepted.
- 9 N. Baggett, K.W. Buck, A.B. Foster, and J.M. Webber, J. Chem. Soc., 21 (1965) 3401-3407.
- 10 P. Fügedi, A. Lipták, P. Nánási, and A. Neszmélyi, Carbohydr. Res., 80 (1980) 233-239.
- 11 G. Excoffier, D. Gagnaire, and J.-P. Utille, Carbohydr. Res., 39 (1975) 368-373.
- R.R. Schmidt, Angew. Chem., 98 (1986) 213-236; Angew. Chem. Int. Ed. Engl., 25 (1986) 212-235;
 S. Sato, Y. Ito, T. Nukada, Y. Nakahara, and T. Ogawa, Carbohydr. Res., 167 (1987) 197-210.
- 13 A. Lipták, L. Szabó, and J. Harangi, J. Carbohydr. Chem., 7 (1988) 687-699.
- 14 H. Lönn, Carbohydr. Res., 139 (1985) 105-113.
- 15 R.K. Jain and K.L. Matta, Carbohydr. Res., 208 (1990) 51-58.
- 16 A. Lipták, J. Imre, and P. Nánási, Carbohydr. Res., 92 (1981) 154-156.
- 17 A. Lipták, Tetrahedron Lett., (1976) 3551-3554.
- 18 F. Weygand, H. Ziemann, and H.J. Bestmann, Chem. Ber., 91 (1958) 2534-2537.
- 19 S. Sato, M. Mori, Y. Ito, and T. Ogawa, Carbohydr. Res., 155 (1986) c6-c10.
- 20 R.U. Lemieux, K.B. Hendricks, R.V. Stick, and K. James, J. Am. Chem. Soc., 97 (1975) 4056-4062.
- 21 A. Lipták, Carbohydr. Res., 107 (1982) 300-302.
- 22 A. Lipták, A. Borbás, J. Kerékgyártó, and I. Bajza, J. Carbohydr. Chem., submitted.
- 23 V. Pozsgay, and H.J. Jennings, J. Org. Chem., 53 (1988) 4042-4052.
- 24 S. Koto, T. Yoshida, K. Takenaka, and S. Zen, Bull. Chem. Soc. Jpn., 55 (1982) 3667-3668.
- 25 V. Pozsgay, Carbohydr. Res., 69 (1979) 284-286.
- 26 G. Ekborg, P.J. Garegg, and B. Gotthammar, Acta Chem. Scand., Ser. B, 29 (1975) 765-771.
- 27 K. Eklind, P.J. Garegg, and B. Gotthammar, Acta Chem. Scand., Ser. B, 30 (1976) 305-308.