

The Efficient Synthesis of a Complex *O*-Phosphoseryl-containing Peptide Ac-Glu-*PSer*-Leu-*PSer*-*P-Ser*-*PSer*-Glu-Glu-NHMe

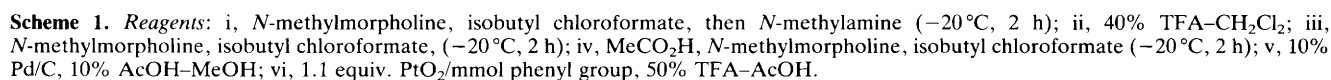
John W. Perich and R. B. Johns*

Department of Organic Chemistry, University of Melbourne, Parkville 3052, Victoria, Australia

The title octapeptide was prepared by the synthesis of the fully protected tetra-Ser(PO₃Ph₂)-octapeptide by incorporation of Boc-Ser(PO₃Ph₂)-OH (Boc = *t*-butoxycarbonyl) in conventional Boc/solution phase peptide synthesis, followed by the complete hydrogenolytic cleavage of the phenyl groups from the Ser(PO₃Ph₂)-octapeptide.

Since 1957, the synthesis of simple *O*-phosphoseryl-containing peptides has generally been accomplished by the 'global' phosphorylation of protected serine-containing peptides using diphenyl or dibenzyl phosphorochloridate-pyridine followed by hydrogenolytic removal of the phenyl or benzyl phosphate protecting groups.¹⁻³ However, as we found this synthetic approach unsuitable for the synthesis of large and/or multi-

PSer-containing peptides,⁴ we developed an alternative Ser(PO₃R₂)-peptide synthetic procedure⁵ which featured (a) the incorporation of Boc-Ser(PO₃Ph₂)-OH⁶ (Boc = *t*-butoxycarbonyl) into conventional Boc/peptide synthesis and (b) the use of modified hydrogenation conditions for the complete removal of the phenyl phosphate groups from Ser(PO₃Ph₂)-peptides. While we have already reported the preparation of



the simple *PSer*-tripeptide Glu-*PSer*-Leu⁶ and the multi *PSer*-tripeptide *PSer-PSer-PSer-NHMe*⁷ using this general synthetic procedure, we now report the straightforward synthesis of the complex tetra-*PSer*-octapeptide, Ac-Glu-*PSer*-Leu-*PSer-PSer-PSer-Glu-Glu-NHMe*. This heavily phosphorylated peptide is of particular biochemical interest since this amino acid sequence, which corresponds to regions 14–21 and 5–12 of bovine and human β -casein, respectively, is known to be a prominent calcium-binding region and is thought to be responsible for maintaining the structural integrity of the casein micelle.

The fully protected Ser(PO₃Ph₂)-octapeptide (**20**) was readily prepared in an overall yield of 61% starting with Boc-Glu(OBzl)-NHMe (all couplings proceeding in over 90% yields) by (a) the use of the mixed anhydride coupling procedure for all amino acid condensations, (b) the incorporation of Boc-Ser(PO₃Ph₂)-OH at the required residue positions, (c) the use of 40% trifluoroacetic acid (TFA)–CH₂Cl₂ for cleavage of the Boc group from all the intermediate Boc-peptides, and (d) the use of the isobutoxycarbonyl mixed anhydride of acetic acid for the *N*-acetylation⁸ of the amino terminus of octapeptide (**19**) (see Scheme 1). The incorporation of the four Ser(PO₃Ph₂)-residues into the octapeptide (**20**) was established from its ³¹P n.m.r. spectrum which displayed four distinct phosphorus resonances at δ –11.0, –12.9, –13.0 and –13.2 p.p.m.

The removal of the glutamyl benzyl groups was effected by the hydrogenation of octapeptide (**20**) in 10% AcOH–MeOH with 10% palladium on charcoal to give the Ser(PO₃Ph₂)-octapeptide (**21**) in quantitative yield. Further hydrogenation of this peptide in 50% TFA–AcOH and 1.1 equiv. PtO₂/mmol phenyl group effected the rapid and complete removal of the phenyl phosphate groups, the reaction being complete after 30 min. C₁₈ Reverse-phase h.p.l.c. purification of the crude product (one major, three minor fractions) using an isocratic

elution of 0.1% aq. TFA–9% acetonitrile gave the target tetra-*PSer*-octapeptide (**1**) [fast atom bombardment (f.a.b.) mass spec. (+ve mode) *m/z* 1242 (MH⁺)] in 53% yield.

To our knowledge, peptide (**1**) represents the largest and most complex multi-*PSer*-peptide that has been reported to date. The simple, straightforward, and high-yielding synthesis of (**1**) dictates that the synthetic strategy described above is the method of choice for the general preparation of *PSer*-peptides and is a significant improvement over the traditional 'global' phosphorylation strategy.

We thank Dr. P. F. Alewood for many useful discussions in the early stages of this research program and Dr. A. L. Chaffee (CSIRO, Division of Energy Chemistry, N.S.W.) for obtaining the f.a.b. mass spectra of (**1**). We also gratefully acknowledge the Australian Dairy Research Committee and (in part) the Australian Wool Corporation for their financial support.

Received, 30th November 1987; Com. 1737

References

- 1 G. Folsch, *Sven. Kem. Tidskr.*, 1966, **79**, 1.
- 2 L. Grehn, B. Fransson, and U. Ragnarsson, *J. Chem. Soc., Perkin Trans. 1*, 1987, 529.
- 3 T. B. Johnson and J. K. Coward, *J. Org. Chem.*, 1987, **52**, 1771.
- 4 P. F. Alewood, R. B. Johns, and J. W. Perich, in 'Peptides: Proceedings of the Seventh American Peptide Symposium,' Pierce Chemical Co., Illinois, 1981, p. 65.
- 5 J. W. Perich, Ph.D. Dissertation, University of Melbourne, 1986.
- 6 J. W. Perich, P. F. Alewood, and R. B. Johns, *Tetrahedron Lett.*, 1986, 1373.
- 7 J. W. Perich and R. B. Johns, *J. Org. Chem.*, 1988, in the press.
- 8 J. W. Perich, P. F. Alewood, and R. B. Johns, *Aust. J. Chem.*, 1987, **40**, 257.