

Tetrahedron Letters 40 (1999) 5983-5986

TETRAHEDRON LETTERS

Conformational Analysis of (R,S)-4-Amido-2,4-dimethyl-butyric Acid Derivatives

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Received 25 May 1999; accepted 21 June 1999

Keywords: Conformation; Hydrocarbon backbone

Abstract: NMR spectra of various N-acyl derivatives of (R,S)-4-amino-2,4-dimethylbutyric acid show these compounds to be biconformational, populating the conformers 4 and 5 of the molecular backbone. Both predominant conformers have the amide group gauche to the main chain. © 1999 Elsevier Science Ltd. All rights reserved.

The molecular backbone of biologically active compounds serves to position functional groups three-dimensionally in space for optimal interaction with the corresponding receptor. It is noteable that nature uses flexible molecular backbones, which may have in many instances a preferred conformation. For example, in polypeptides nature combines α -amino acids as modules into flexible functional molecules with a pre-defined conformation. In search for non-natural analogs, oligomers of β -amino acids¹ have been studied with respect to distinct folding preferences. Recently, oligomers of γ -amino acids have been investigated in the same vein,² revealing distinct preferences for the formation of helical structures. In a few instances nature also relies on γ -amino acids as flexible linker units to confer a particular conformation to biologically active molecules. This is evidenced by the ³J_{H,H}-NMR coupling constants along the backbone of calyculin C, ³ cf. 1, or bleomycin A₂,⁴ cf. 2.



The linker unit in both natural products is a derivative of (R,S)-4-amino-2,4-dimethyl-butyric acid (3). MM3^{*} calculations on 3 (R = CH₃, R' = OMe) suggest that two backbone-conformations 4, and 5 contribute predominantly to the local conformer population, whereas conformer 6 should be of sufficiently higher energy (+ 7 kJ mol⁻¹ relative to 4) to be neglected.

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Conformation 4 corresponds to that deduced for calyculin C³ or bleomycin A_2^{5} in solution from the NMR data. It corresponds as well to that found in the crystalline state for bleomycin A_2^{6} . We became interested to learn, whether this conformational preference is intrinsic to the backbone segment 3, or whether it is imposed to a large extent from the remainder of the molecule, i.e. the flanking groups R and R'. Knowledge of this sort is a prerequisite for the use of compounds 3 as spacer units in a modular aproach to larger flexible molecules with defined shape.⁸

We therefore studied the conformational behaviour of a series of derivatives of 3 and related compounds⁹ by ¹H NMR spectroscopy (cf. table) The relevant vicinal H,H-coupling constants between the hydrogen atoms at C-2, C-3, and C-4 could not be determined in all cases completely, because of signal overlap or line broadening due to the presence of amide rotamers.

				C-3-H (pro- <i>R</i>)			C-3-H (pro- <i>S</i>)		
	\$ M. J. 3 5 5	Solvent	Temp (K)	δ (ppm)	³ J to H-2	³ J to H-4	δ (ppm)	³ J to H-2	³ J to H-4
1 ^{a)}		C ₆ D ₆	298	2.18	1.5	13.0	1.84	12.5	1.5
7 ^{b)}		e CDCl ₃	298	1.94	6.2	9.6	1.68	6.7	n.d.
8		CDCI ₃	298	1.7	7	10	1.4	7 ^{c)}	n.d.
9		CDCI3	333	1.85	7. 2	9.6	1.48	6.7 ^{c)}	4.8
10		CDCl ₃	333	1.87	7.2	9.8	1.48	n.d.	n.d.
CF ₃		CDCI ₃	298	1.89	8.0	9.9	1.49	n.d.	n.d.
B 12		CDCl₃	328	1.86	7.6	10.3	1.49	6.5	3.9
13	FmocNH COO'Bu	C ₆ D ₆	298	1.11	6.6	8.7	1.01	7.7	n.d.

Table: NMR Coupling constants of (R,S)-4-Amido-2,4-dimethyl-Butyric Acid Derivatives

a) Values from ref.³; b) values from ref.⁷; c)Taken from the coupling pattern of the C-2-H signal; n.d. = not determined

In order to get more detailed information on the nature of the preferred conformer, the NMRchemical shifts of the individual diastereotopic protons at C-3 of 8 were assigned by a stereospecific labelling procedure:



Starting from the pyrrolinone $14^{7,10}$ reduction with "copper deuteride"¹¹ provided a (1:1.5)mixture of the C-2-epimeric pyrrolidinones 15, the spectral data of which corresponded to those reported by Koskinen.^{7,10} We presume that the deuterium atom has been incorporated in this reduction <u>anti</u> to the C-4 methyl group of 15. The pyrrolidinones were transformed into the esters 16 and 17 by treatment of 15 with LiOH followed by diazomethane. The esters 16 and 17 could then be separated by preparative HPLC. Comparison of the ¹H-NMR-spectra of 16 and of 8 showed that deuteration has led to the disappearance of the ¹H-NMR-signal at $\delta = 1.4$ ppm and had, moreover, simplified the coupling pattern of the C-3-H NMR signal at $\delta = 1.7$ ppm. This allowed an assignment of chemical shifts to the individual diastereotopic protons at C-3 in 8. This assignment is in line with the one made for the diastereotopic protons in the corresponding segment 1 of calyculin C.³ In view of the consistency between 8 and 1, the chemical shifts of the compounds 9 to 12 have been assigned by analogy.

It is clear from the data in the table, that the conformational preference of the compounds 7 to 13 is lower than that of the corresponding segment 1 in calyculin C. It appears, however, that the same local conformer 4 is preferentially populated, albeit to a lesser extent. The consistently high ${}^{3}J_{H,H}$ -coupling constants between C-4-H and C-3-H_(pro R) indicate that the local conformation around the bond C-3/C-4 is fairly uniform. In turn, there is conformational heterogeneity regarding the C-2/C-3 bond. This is manifest from the low alteration of the values of the two C-2-H to C-3-H coupling constants. We therefore conclude that the 4amino-2,4-dimethyl-butyrate segment in the compounds 7 to 13 is biconformational, with the conformations 4 and 5 populated to a similar extent. The backbone segment 3 present in compounds 7 to 13 has therefore a (moderate) intrinsic preference to populate a distinct conformation. This preference is increased in calyculin C or bleomycin A₂ by an (as not yet determined) effect of the groups flanking the segment 3.

Acknowledgements: This work has been supported by the European Commission through the TMR network ERB FMRX CT 960011.

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- 9 The synthesis of 8 has been described before by Koskinen^{7,10} starting from alanine. Our synthesis was based on the monoester 19,¹² which is readily obtained from meso-2,4-dimethylglutaric anhydride 18 and may easily be resolved,¹³ if required. Curtius degradation of 19¹⁴ in the presence of alcohols led us to the various N-acylated derivatives 8-10 and 13 listed in the table.



The amides 11 and 12 were prepared by attaching the amido acid 10 to 2-chlorotrityl resin and coupling with the second amino acid by standard methods using O-benzotriazolyl-N, N, N', N'-tetramethyluronium tetrafluoroborate (TBTU) as coupling reagent.

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