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Synthesis of 3,3-Dimethylazetidine-2-carboxylic Acid and Some Derivatives

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Abstract : γ -Chloro- α -(N-alkylimino)esters were reduced by sodium cyanoborohydride in methanol in the presence of acetic acid with complete selectivity to give rise to either γ -chloro- α -(N-alkylamino)esters (reaction at 0°C) or 1-alkyl-3,3-dimethylazetidine-2-carboxylic esters (reaction at reflux). The isolable γ -chloro- α -(N-alkylamino)esters are suitable sources for 1-(N-alkylamino)-2,2-dimethylcyclopropane-1-carboxylic esters via base-induced 1,3-dehydrochlorination, while the former substrates as transient species undergo 1,4-dehydrochlorination to the corresponding azetidines. The latter process was used for the synthesis of 3,3-dimethylazetidine-2-carboxylic acid, a new non-proteinogenic sterically hindered α -amino acid, via hydrogenolysis of methyl 1-benzyl-3,3-dimethylazetidine-1-carboxylate and subsequent acidic hydrolysis. Reduction of alkyl 4-chloro-3,3-dimethyl- α -(N-alkylimino)butanoates with lithiumaluminiumhydride in diethyl ether afforded 1-alkyl-3,3-dimethyl-2-(hydroxymethyl)-azetidines. (© 1998 Elsevier Science Ltd. All rights reserved.

INTRODUCTION

Small ring α -amino acids have been the subject of considerable research in the last decade because of the physiological activities associated with these derivatives. Small ring as well as azacyclic α -amino acids received considerable attention in this respect. 1-Aminocyclopropane-1-carboxylic acids 1-3 have been studied intensively in relation to the unsubstituted α -amino acid 1, i.e. 1-aminocyclopropane-1-carboxylic acid (ACC), which is omnipresent in plants and which is converted in plants into ethylene by the ethylene forming enzyme via an oxidative decarboxylation process.^{1,2} Ethylene is a major natural plant growth regular and is responsible for the germination of seeds, flowering, fruit and leaf drop, and ripening of fruits.^{1,2} In general, ethylene is responsible for all factors associated with the senescence of plants. Consequently, a great number of efforts have

been devoted to the synthesis of ACC derivatives because of their potential plant growth regulating activity.³⁻¹³ Additionally, ring substituted ACC derivatives 2 and 3 constitute sterically constrained α -amino acids which



have an interest in peptide chemistry because of their potential to alter conformations of peptides containing these ACC derivatives. Accordingly, such peptides have been studied as potential enzyme inhibitors. The latter research led to the synthesis of peptide mimetics resistant to proteolytic cleavage.¹⁴⁻¹⁸

Azetidine-2-carboxylic acid 4 and its derivatives 5 are frequently observed in a number of plant sources such as the Liliaceae, Solanaceae, Fagaceae, Fabaceae, Chenopodiaceae and red marine algae.^{19,20} Several of these natural derivatives are phytosiderophores, i.e. low molecular weight chelators, which promote uptake and transport of iron in higher plants under iron-deficient conditions.²¹ The antifungal antibiotics of the polyoxin group, for example polyoxine A, contain polyoximic acid, an azetidine-2-carboxylic acid, as structural unit.²² Other azetidine carboxylic acids, also including compounds carrying the carboxyl group at positions different from the α -position, have been shown to display growth regulator activity and male sterilizing activity in cereals,²³⁻³⁰ while cis-azetidine-2,4-dicarboxylic acid has been reported as a neurotransmitter potentiator.^{31, 32}

Cyclopropaneamino acids have become increasingly important as mimics for naturally occurring amino acids. The same is valid for their azetidine based α -amino acids as a result of some special structural features.³³ Peptides containing such small ring amino acids are expected to show improved metabolic stability since tertiary amide bonds are resistant to proteolysis. The reduced conformational space can lead to altered selecti-



vity, while increased ring strain can prove useful for the design of novel peptide-based enzyme inhibitors as exemplified for ACC.³⁴ While now a number of syntheses on cyclopropanecarboxylic acids 2 and 3 have been published in recent years, few synthetic methods are available for azetidine-2-carboxylic acids.^{19,20,35-37}

In the present paper, a suitable access to 3,3-dimethylazetidine-2-carboxylic acid 7, a new small ring unnatural α -amino acid with a sterically hindering geminal dimethyl unit, via selective elaboration of γ -chloro- α -iminocarboxylic esters is disclosed.

RESULTS AND DISCUSSION

β-Chloro imines have been utilized with success in the synthesis of azetidines via addition of a nucleophile across the imino bond and subsequent intramolecular nucleophilic substitution.³⁸⁻⁴⁰ More functionalized β-halo imines, such as the γ-chloro-α-(N-benzyl)iminocarboxylic esters 11a,b, have been shown previously to be excellent substrates for the synthesis of 2,2-disubstituted 1-aminocyclopropanecarboxylic acids 12 by baseinduced 1,5-dehydrochlorination and subsequent hydrolysis (Scheme 1).⁴¹ γ-Chloro-α-iminocarboxylic esters 11 are suitable trifunctional substrates for selective transformations towards cyclopropanes and azetidines. A selective reduction of the imino function of compounds 11 without interference of the halogen and the carboxylic ester moiety provides a γ-chloro-α-aminocarboxylic ester 13 which either can be converted to ACC derivatives 14 by 1,3-dehydrochlorination or can be ring closed in a 1,4-fashion to provide azetidine-2-carboxylic esters 15 (Scheme 2). The latter aspect is now worked out in the following experiments.

 γ -Chloro- α -iminoesters 11 were synthesized from β -chloroketone 8 via a sequence of reactions involving α -monobromination, ethylthiolate substitution, dichlorination, Hg⁺⁺-assisted alcoholysis to give γ -chloro- α -ketoesters 9 and 10, and TiCl₄-mediated imination.⁴¹

In some preliminary experiments, it was noticed that the trifunctional γ -chloro- α -iminoester 11a was inert towards nucleophilic attack across the imino double bond. Both sodium methoxide or sodium borohydride were not capable of inducing any reaction, even under more streneous reaction conditions (Scheme 2). Contrary to the imination of 9, the relative stability of the α -imino ester 11a towards nucleophilic attack can be interpreted in terms of the higher steric shielding of the imine carbon atom combined with a less electrophilic behaviour of it in comparison with the carbonyl precursors 9 or 10. While compound 11a remains unreactive towards sodium borohydride in methanol under reflux for 5h, also a large excess of diborane-dimethyl sulfide complex in dichloromethane under reflux for 18h did not lead to amine 16 (Nu=H) or azetidine 17 (Nu=H). Sodium cyanoborohydride in methanol in the presence of acetic acid proved to be an excellent chemoselective reagent for the reduction of the imino function in compounds 11.⁴¹ In this way, the electrophilic character of the imino carbon atom is enhanced by conversion into an intermediate iminium ion. This reduction reaction turned out to be an interesting transformation as it could be directed selectively to either the functionalized α amino esters 13⁴¹ or the azetidine-2-carboxylic esters 15. Controlling the reaction conditions allowed to stop the reduction reaction of 11 at the level of α -amino esters 13 when performing the reaction at 0°C for 1h. This conversion offered a route to N-alkylated 2,2-disubstituted-1-aminocyclopropanecarboxylic esters 14 by reac-



Scheme 2

tion of γ -chloro- α -imino esters 13a,c with potassium t-butoxide in THF under reflux.⁴¹ The temperature of the reductive conversion of compounds 11 with sodium cyanoborohydride proved to be the key to selective reactions. At elevated temperature, the intermediate α -amino esters 13 cyclized in situ to the azetidine-2-carboxylic esters 18, which were obtained in 75-86% yield. In order to prepare 3,3-dimethylazetidine-2-carboxylic acid 7, methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylate 18a was deprotected at the nitrogen atom by



Scheme 3

hydrogenolysis in methanol in the presence of palladium on carbon. This procedure required the use of the azetidine 18a as the hydrochloride because otherwise the palladium catalyst was poisoned by the N-debenzylated azaheterocycle. This is a common feature for azetidines and is due to the higher basic character compared to other azaheterocycles of different ring size.⁴² In addition, relatively large amounts of the palladium catalyst were required in order to obtain methyl 3,3-dimethylazetidine-2-carboxylate 19 in 76% yield. Another prerequisite for a successful conversion of the N-benzylazetidine 18a into the N-debenzylated derivative 19 concerned the high purity of the starting substrate. A purification of compound 18a by flash chromatography was absolutely necessary, even when the purity of the compound, as judged from ¹H NMR, was higher than 95%. Methyl 3,3-dimethylazetidine-2-carboxylate 19 was hydrolyzed with 10 equiv. 2N hydrogen chloride under reflux for 15h to give 3,3-dimethylazetidine-2-carboxylic acid 20 in virtually quantitative yield.

While the action of relatively strong bases such as potassium t-butoxide on methyl 4-chloro-2-(N-benzylamino)-3,3-dimethylbutanoate 13a gave rise to methyl 1-(N-benzylamino)-2,2-dimethylcyclopropanecarboxylate 14a, the reaction of compound 13a with an excess of potassium cyanide in methanol under reflux for 5h afforded methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylate 18a in 86% yield. Sodium methoxide in methanol (2N; 5 equiv.) under reflux for 1h converted the γ -chloro- α -amino ester 13a into a 82:18 mixture (94% yield) of methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylate 18a and 1-(N-benzylamino)-2,2-dimethylcyclopropanecarboxylate 14a, respectively (Scheme 4). Accordingly, the presence of relatively stronger bases, such as alkoxides, favors the formation of cyclopropane derivatives 14, as compared to the formation of azetidines



Scheme 4

18 in alcoholic medium in the presence of cyanide or cyanoborohydride.

Treatment of γ -chloro- α -amino esters 11a,c with lithium aluminium hydride in ether under reflux for 1h gave rise to 1-alkyl-3,3-dimethyl-2-(hydroxymethyl)azetidines 21a,c in 60-88% yield. Recently, 2-(hydroxymethyl)azetidines have been isolated from marine organisms, i.e. Penares sp.⁴³⁻⁴⁵. These marine natural products, called penaresidins and penazetidins, displayed an ATP-ase activating property and a protein kinase C inhibition.^{42,43}

The methodology, starting with the reductive ring closure of γ -chloro- α -iminocarboxylic esters 11, is certainly applicable to other 3,3-disubstituted azetidine-2-carboxylic acids and its derivatives. In this way γ chloro- α -aminocarboxylic esters 11 are elegant sources for the synthesis of strained α -amino acids, i.e. azetidine-2-carboxylic acids and 1-aminocyclopropanecarboxylic acids, with a potential physiological activity.

EXPERIMENTAL PART

¹H NMR spectra were recorded at 60 MHz, 270 MHz or 360 MHz, while ¹³C NMR spectra were obtained at 67.8 MHz or 90 MHz. IR spectra were measured with a liquid film between NaCl or as a KBr pellet. Mass spectra were recorded at 70 eV, either by the direct inlet mode or via a GC-MS coupling. GC analyses were performed using a capillary column (fused silica, RSL 200, 20 m length, i.d. 0.53 mm, He carrier gas). Flash chromatography was performed using Merck Kieselgel 60 (0.04-0.063 mm) and different solvent combinations, determined via initial tlc analysis (Merck Kieselgel 60 F₂₅₄, precoated). Melting points were measured on a Kofler Hotbench (Reichert Jung) or with a Büchi 535 melting point apparatus.

Preparation of starting materials

Alkyl 4-chloro-3,3-dimethyl-2-oxobutanoates 9 and 10 were prepared starting from β -chloroketone 8 by consecutive bromination, substitution with sodium ethylthiolate, chlorination with sulfuryl chloride and Hg(II)-acetate mediated alcoholysis.⁴¹ The corresponding γ -chloro- α -(N-alkylimino)esters 11 were synthesized by imination of α -ketoesters 9 and 10 with primary amines (benzylamine or isopropylamine) in the presence of stoichiometric amounts of titanium(IV) chloride in diethyl ether.

Synthesis of N-substituted 3,3-dimethylazetidine-2-carboxylic esters 18

The conversion of methyl 4-chloro-2-(N-benzylimino)-3,3-dimethylbutanoate 11a into methyl N-benzyl-3,3dimethylazetidine-2-carboxylate 18a is representative.

To an ice cooled solution of 4-chloro-2-(N-benzyl)imino-3,3-dimethylbutanoate 11a (2.67 g, 0.01 mol) in methanol (40 mL) is added sodium cyanoborohydride (1.24 g, 0.02 mol), followed by 98% acetic acid (0.6 g, 0.01 mol). The reaction mixture was stirred for 1h at reflux, poured into water and extracted with CH₂Cl₂ (3x20 mL). The combined organic extracts were dried (MgSO₄), filtered and evaporated to yield 2.34 g (100%) of methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylic ester 18a. Purification can be performed by means of flash chromatography or distillation ($R_f Et_2O/C_3H_{12}$ 1/1 = 0.49, bp. 113-117°C / 2 mmHg) to yield the pure compound in 86% and 75% yield, respectively. ¹H NMR (CDCl₃, 270 MHz) δ 1.18 and 1.20 (each 3H, each s, Me₂C); 2.69 and 3.10 (2H, d+dd, AX, J=6.35 and 0.73 Hz, CH₂N); 3.45 (1H, d, J=0.73 Hz, CHN); 3.58 and 3.80 (2H, 2xd, AB, J=12.69 Hz, CH₂C₆H₅); 3.64 (3H, s, OMe); 7.29 (5H, m, CH₂C₆H₅). ¹³C NMR (CDCl₃, 67.8 MHz) δ 22.93 and 27.73 (Me₂C); 36.41 (Me₂C); 51.25 (OMe); 61.76 (CH₂C₆H₅), 63.32 (CH₂N); 73.13 (CHN); 127.13 (Cp); 128.16 and 129.20 (Co and Cm); 137.10 (Cq); 171.93 (C=O). IR (NaCl, cm⁻¹); 1728-1752 (C=O). MS (70 eV) m/z (rel. int.) 233 (M⁺, 1); 174(11); 119(8); 118(7); 117(7); 92(7); 91(100); 65(14); 56(8); 55(5); 42(5); 41(8); 40(11). Elemental analysis : Calcd. 72.07% C, found 72.19% C; calcd. 8.21% H, found 8.10% H; calcd. 6.00% N, found 6.19% N.

Methyl N-isopropyl-3,3-dimethylazetidine-2-carboxylic ester 18c :

¹H NMR (CDCl₃, 270 MHz) δ 0.92 and 0.97 (6H, each d, J=6.4 Hz, CH<u>Me</u>₂), 1.18 and 1.22 (6H, each s, <u>Me</u>₂); 2.42 (1H, septet, J=6.4 Hz, C<u>H</u>Me₂); 2.68 and 3.25 (2H, AB, J=6.4 Hz, C<u>H</u>₂N), 3.40 (1H, s, CHN), 3.77 (3H, s, O<u>Me</u>). ¹³C NMR (CDCl₃, 67.8 MHz) δ 172.28 (<u>C</u>=O); 73.46 (<u>C</u>HN); 63.05 (<u>C</u>H₂N); 58.67 (N<u>C</u>H); 51.48 (O<u>Me</u>); 34.55 (Me₂C); 28.18 and 23.09 (<u>Me</u>₂C); 19.91 and 19.38 (CH<u>Me</u>₂). IR (NaCl, cm⁻¹) 1725-1755 (C=O). MS (70 eV) m/z (rel. int.) 186 (M⁺ +1, 2); 185 (M⁺, 18); 184 (M⁺ -1, 2); 170(43); 130(5); 126(100); 115(7); 110(5); 97(8); 88(6); 85(4); 84(51); 83(9); 82(5); 72(6); 71(15); 70(21); 69(9); 68(2); 67(2); 59(5); 57(4); 56(35); 55(12); 54(2); 53(2); 44(4); 43(34); 42(18); 41(18). Elemental analysis : calcd. 64.83% C, found 64.96% C; calcd. 10.34% H, found 10.19% H; calcd. 7.56% N, found 7.50% N.

Preparation of the Hydrochloride of Methyl 1-Benzyl-3,3-dimethylazetidine-2-carboxylic ester 18a

A solution of 1.16 g (0.005 mol) of azetidine 18a in 25 ml of dry ether was treated with an excess of dry hydrogen chloride in ether. The mixture was stirred for 1h at room temperature, after which the precipitated hydrochloride was filtered off and washed with ether. After drying in vacuo, the hydrochloride (1.15 g; yield 85%) was used as such in the next experiment. ¹H NMR (CDCl₃, 270 MHz) : 1.45 and 1.54 (each 3H, each s, <u>Me₂</u>); 3.63 (3H, s, O<u>Me</u>); 3.7-4.7 (5H, m, C<u>H</u>-N-(C<u>H₂)₂); 7.3-7.7 (5H, m, C₆H₅); 12.1 (1H, s, broad, N<u>H</u>).</u>

Debenzylation of methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylic ester 18a to methyl 3,3-dimethylazetidine-2-carboxylate 19

To a solution of methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylic ester hydrochloride 18a.HCl (0.81 g, 3 mmol) in methanol (60 mL) was added 10% Pd on carbon (50% w/w, 0.405 g). The amino ester was hydrogenolyzed under a pressure of 50 psi at room temperature for 20h. After the reaction, the suspen-

sion was filtered and the filtrate was concentrated in vacuo. The residue was poured into water, neutralized with NaHCO₃ and extracted with CH₂Cl₂. The combined organic extracts were dried (MgSO₄), filtered and evaporated. Purification was performed by flash chromatography to yield 0.33 g (76%) of methyl 3,3-dime-thylazetidine-2-carboxylate 19 : R_f MeOH = 0.62. ¹H NMR (CDCl₃, 270 MHz) δ 1.10 and 1.39 (6H, each s, Me₂C); 2.75 (1H, s br, N<u>H</u>); 3.31 (2H, s, C<u>H</u>₂N); 3.76 (3H, s, O<u>Me</u>); 4.02 (1H, s, C<u>H</u>N). ¹³C NMR (CDCl₃, 67.8 MHz) δ 23.29 and 28.05 (Me₂C); 39.73 (Me₂C); 51.70 (O<u>Me</u>); 57.32 (CH₂N); 67.83 (CHN); 173.58 (C=O). IR (NaCl, cm⁻¹) 3320 (NH); 1734 (COOMe). Elemental analysis : calcd. 58.72% C, found 58.59% C; calcd. 9.15% H, found 9.19% H; calcd. 9.78% N, found 9.88% N.

Hydrolysis of Methyl 3,3-dimethylazetidine-2-carboxylate 19 to 3,3-dimethylazetidine-2-carboxylic acid 7

A solution of methyl 3,3-dimethylazetidine-2-carboxylate 19 (0.33 g, 2.3 mmol) in 2N HCl (5 ml, 10 mmol) was stirred at reflux for 15h. After the reaction, the solvent was evaporated in vacuo and the residue was chromatographed on Dowex 50x8 to yield 0.28 g (97%) of pure 3,3-dimethylazetidine-2-carboxylic acid 7 : R_f n-BuOH/AcOH/H₂O 4/1/1 = 0.19, mp. 188°C. ¹H NMR (D₂O, 270 MHz) δ (Ref. <u>CH</u>₃CN = 2.00) 1.17 and 1.33 (6H, each s, <u>Me</u>₂C); 3.57 and 3.77 (2H, AB, J=10.38 Hz, C<u>H</u>₂N); 4.40 (1H, s, C<u>H</u>-N). ¹³C NMR (D₂O, 67,8 MHz) δ (Ref. Trimethylsilylpropanesulfonic acid sodium salt; δ =0); 24.28 and 29.13 (<u>Me</u>₂C); 40.14 (Me₂C); 57.57 (<u>CH</u>₂N); 71.28 (<u>CHN</u>); 174.59 (<u>C</u>=O). IR (NaCl, cm⁻¹) v_{max} = 3420, 3110, 1685, 1610, 1570, 1414 and 1356. MS (70 eV) m/z (rel. int.) no M⁺, 179(20); 157(6); 156(31); 147(4); 110(4); 101(5); 100(9); 86(11); 84(15); 83(13); 82(7); 75(22); 74(9); 73(100); 59(8); 56(5); 55(6); 45(20); 44(6); 43(4); 42(5); 41(6); 40(4). Elemental analysis : calcd. 10.84% N, found 10.70% N.

Synthesis of 2-(hydroxymethyl)-3,3-dimethylazetidines 21

The reduction of methyl 4-chloro-2-(N-isopropyl)imino-3,3-dimethylbutanoate 11c is representative. To an ice cooled solution of 4-chloro-2-(N-isopropyl)imino-3,3-dimethylbutanoate 11c (25.61 g, 117 mmol) in diethyl ether (300 ml) was added lithiumaluminiumhydride (8.9 g, 234 mmol). The reaction mixture was stirred for 2h at reflux and then poured into ice cold water. The organic phase was separated and the aqueous phase was extracted two times additionally (100 mL) with diethyl ether. The combined organic extracts were dried (MgSO₄), filtered and evaporated to yield 11.07 (60%) of 1-isopropyl-2-(hydroxymethyl)-3,3-dimethylaze-tidine 21c. Recrystallization of the reaction mixture in pentane at -20°C yielded pure azetidine 21c as a white solid : mp. 67°C. ¹H NMR (CDCl₃, 360 MHz) δ 0.91 and 0.97 (each 3H, each d, J=6.25 Hz, CHMe₂); 1.09 and 1.23 (each 3H, each s, Me₂); 2.48 (1H, septet, J=6.25 Hz, CHMe₂); 2.71 (1H, d, J_{AB}=7.01 Hz, HCHN); 2.98 (1H, dxd, J_{AX}=5.94 Hz, J_{BX}=2.28 Hz, HCN); 3.16 (1H, d, J_{AB}=7.01 Hz, HCHN); 3.30 (1H, br s, OH); 3.56 and 3.61 (each 1H, ABxd, J_{AB}=10.94 Hz, J_{AX}=5.94 Hz, J_{BX} shows additional broadening, CH₂OH). ¹³C NMR (CDCl₃, 90 MHz) δ 19.95 and 20.84 (CHMe₂); 22.23 and 29.33 (Me₂); 32.06 (CMe₂); 58.08 (CHMe₂);

61.11 (CH₂OH); 63.92 (CH₂N); 72.94 (CHN). IR (KBr, cm⁻¹) 3100 (OH, br). MS (70 eV) m/z (rel. int.) 158 (M^+ +1, 2); 157 (M^+ , 8); 142(12); 127(8); 126(100); 114(4); 101(4); 100(7); 85(6); 84(68); 82(4); 81(4); 72(31); 71(4); 70(58); 69(6); 68(4); 67(3); 60(4); 58(10); 57(7); 56(27); 55(18); 54(3); 53(4); 44(26); 43(88); 42(36); 41(38). Elemental analysis : calcd. 68.74% C, found 68.79% C; calcd. 12.18% H, found 12.03% H, calcd. 8.91% N, found 8.99% N.

1-Benzyl-2-(hydroxymethyl)-3,3-dimethylazetidine 21a

¹H NMR (CDCl₃, 360 MHz) δ 1.10 and 1.21 (each 3H, each s, Me2); 2.60 (1H, br s, OH); 2.73 (1H, d, J=6.83 Hz, <u>H</u>CHN); 3.01 (1H, b, J=4.61 Hz, HCN); 3.15 (1H, d, J=6.83 Hz, HC<u>H</u>N); 3.45 (2H, d, J=4.61 Hz, C<u>H</u>₂OH); 3.60 and 3.72 (each 1H, each d, AB, J_{AB}=12.79 Hz, <u>CH</u>₂C₆H₅); 7.29 (5H, br s, C₆H₅). ¹³C NMR (CDCl₃, 90 MHz) δ 22.24 and 28.79 (Me₂); 33.91 (<u>C</u>Me₂); 60.71 (CH₂N); 62.55 (CH₂OH); 65.46 (<u>C</u>H₂C₆H₅); 74.51 (CHN); 127.20 (Cp); 128.38 and 128.71 (Co and Cm); 138.46 (Cq). IR (NaCl, cm⁻¹) 3300 (OH, br). MS (70 eV) m/z (rel. int.) 205 (M⁺, 3); 175(4); 174(31); 131(3); 120(5); 118(5); 105(2); 92(11); 91(100); 71(3); 65(7); 56(4); 55(3); 42(4); 41(6); 40(21). Elemental analysis : calcd. 76.06% C, found 76.02% C; calcd. 9.33% H, found 9.24% H; calcd. 6.82% N, found 6.93% N.

Reaction of 4-chloro-2-(N-benzylamino)-3,3-dimethylbutanoate 13a with Potassium Cyanide or Sodium Methoxide in Methanol

A solution of 0.54 g (0.002 mol) of $13a^{41}$ in 6 ml methanol was treated with 0.65 g (0.01 mol) of potassium cyanide. After reflux for 5h, the reaction mixture was poured into water and was extracted with dichloromethane to give 0.40 g (86%) of pure methyl 1-benzyl-3,3-dimethylazetidine-2-carboxylic ester 18a (purity > 96%). The same experiment with 0.54 g (0.002 mol) of compound 13a and 5 ml 2N sodium methoxide in methanol (0.01 mol) under reflux for 1h afforded 0.47 g (94%) of an 82:18 mixture of azetidine 18a and methyl cyclopropanecarboxylic ester 14a, respectively, as judged from ¹H NMR and GC analysis (workup as above). The spectral data of compound 18a were given above while the spectral data of compound 14a were identical with data from the literature.⁴¹

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References

- 1. Yang, S.F.; Hoffman, N.E. Ann. Rev. Plant Physiol. 1984, 35, 155-189.
- 2. Yang, S.F. in Plant Nitrogen Metabolism, Chapter 8, pp. 263-287, Ed. Poulton, J.E.; Romeo, J.T. and Conn, E.E. Plenum Press : New York, 1989.
- 3. De Kimpe, N.; Sulmon, P.; Brunet, P. J. Org. Chem. 1990, 55, 5777-5784, and references cited therein.
- 4. Pirrung, M.C.; McGeehan, G.M. Angew. Chem., 1985, 97, 1074-1075.
- 5. Pirrung, M.C.; McGeehan, G.M. J. Org. Chem., 1986, 51, 2103-2106.
- Toshima, H.; Niwayama, Y.; Nagata, H.; Greulich, F.; Ichihara, A. Biosci. Biotech. Biochem. 1993, 57, 1394-1395.
- 7. Hoffman, N.E.; Yang, S.F.; Ichihara, A.; Sakamura, S. Plant Physiol. 1982, 70, 195-199.
- 8. Charette, A.B.; Côté, B. J. Am. Chem. Soc. 1995, 117, 12721-12732.
- 9. Jiménez, J.M.; Rifé, J.; Ortuño, R.M. Tetrahedron Asym. 1996, 7, 537-558.
- 10. Baldwin, J.E.; Adlington, R.M.; Rawlings, B.J. Tetrahedron Lett., 1985, 26, 481-484.
- 11. Burgess, K.; Ho, K.-K.; Moye-Sherman, D. Synlett. 1994, 575-583.
- 12. Wick, L.; Tamm, C.; Boller, T. Helv. Chim. Acta, 1995, 78, 403-410.
- 13. Pirrung, M.C.; Cao, J.; Chen, J. J. Org. Chem., 1995, 60, 5790-5794.
- 14. Stammer, C.H. Tetrahedron 1990, 46, 2231-2254, and references cited therein.
- 15. Burgess, K.; Ho, K.-K.; Pal, B. J. Am. Chem. Soc. 1995, 117, 3808-3819.
- 16. Mendel, D.; Ellman, J.; Schultz, P.G. J. Am. Chem. Soc. 1993, 115, 4359-4360.
- 17. Burgess, K.; Ho, K.-K.; Pettitt, B.M. J. Am. Chem. Soc. 1995, 117, 54-65.
- 18. Burgess, K.; Ke, C.-Y. J. Org. Chem., 1996, 61, 8627-8631.
- 19. Moore, J.A.; Ayers, R.S. in "The Chemistry of Heterocyclic Compounds", Ed. Weissenberger A. and Taylor E.C., J. Wiley, New York, 1983, Vol. 42, "Small Ring Heterocycles", Part 2, Ed. A. Hassner.
- De Kimpe, N. in "Comprehensive Heterocyclic Chemistry Three- and Four-Membered Rings, with All Fused Systems Containing Four-Membered Rings"; Vol. 1B, Chapter 1.18 "Azetidines, Azetines and Azetes : Monocyclic", p. 507-589. Eds. Padwa A., Katritzky A.R., Rees C.W., Scriven E.F.V., Pergamon, Elsevier, Oxford (1996).
- a) Sugiura, Y.; Tanaka, H.; Mino, Y.; Ishida, T.; Ota, N.; Inoue, M.; Nomoto, K.; Yoshioka, H.; Takemoto, T. J. Am. Chem. Soc. 1981, 103, 6979. b) Oida, F.; Ota, N.; Mino, Y.; Nomoto, K.; Sugiura, Y. J. Am. Chem. Soc. 1989, 111, 3436-3437. c) Noma, M.; Noguchi, M.; Tamaki, E. Tetrahedron Lett. 1971, 2017-2020. d) Ohfune, Y.; Tomita, M.; Nomoto, K. J. Am. Chem. Soc. 1981, 103, 2409. e) Hamada, Y.; Shioiri, T. J. Org. Chem. 1986, 51, 5489-5490. f) Matsuura, F.; Hamada, Y.; Shioiri, T. Tetrahedron Lett. 1992, 51, 7917-7920. g) Matsuura, F.; Hamada, Y.; Shioiri, T. Tetrahedron Asymm. 1992, 3, 1069-1074. h) Matsuura, F.; Hamada, Y.; Shioiri, T. Tetrahedron Lett. 1992, 51, 7921-7924.

i) Matsuura, F.; Hamada, Y.; Shioiri, T. Tetrahedron 1994, 50, 265-274. j) Shioiri, T.; Hamada, Y.; Matsuura, F. Tetrahedron 1995, 51, 3939-3958.

- 22. Hanessian, S.; Fu, J.-M.; Chiara, J.-L.; Di Fabio, R. Tetrahedron Lett., 1993, 34, 4157-4160.
- 23. Nielsen, E.; Forlani, G.; Cella, R.; Parisi, B. Plant Sci. 1986, 44, 147; Chem. Abstr. 1986, 105, 112253.
- 24. Fowden, L. Phytochemistry 1991, 30, 3859-3860.
- 25. Devlin, B.R.J. US Pat. 4560401 (1985); Chem. Abstr. 1986, 105, 42626.
- 26. Verbrugge, P.A.; De Waal, J. US Pat. 4812581 (1989); Chem. Abstr. 1989, 111, 57527.
- 27. Wood, D.; Mason, R. Eur. Pat. 131435 (1985); Chem. Abstr. 1985, 103, 195984.
- 28. B.V. Maatschappij (Shell Int. Res.) Jpn. Kokai 6281367 (1987); Chem. Abstr. 1987, 107, 134187.
- 29. De Nie-Sarink, M.; Mason, R. Eur. Pat. 165636 (1985); Chem. Abstr. 1986, 105, 6400.
- 30. De Nie-Sarink, M.; Scholes, G. Eur. Pat. 165637 (1985); Chem. Abstr. 1986, 105, 224786.
- 31. Kozikowski, A.P.; Wroblewski, J.; Costa, E. Eur. Pat. 299513 (1989); Chem. Abstr. 1989, 111, 7211.
- 32. Kozikowski, A.P.; Tuckmantel, W.; Reynolds, I.; Wroblewski, J. J. Med. Chem. 1990, 33, 1561-1571.
- 33. Blythin, D.J.; Green, M.J.; Lauzon, M.J.R.; Shue, H.-J. J. Org. Chem. 1994, 59, 6098-6100 and references cited therein.
- 34. a) Masaki, M.; Uehara, M.; Hirate, K.; Isowa, Y.; Sato, Y.; Nakashima, Y. Eur. Pat. Appl. EP 393934, Al 901024; *Chem. Abstr.* 1990, *114*, 164815. b) Zagari, A.; Nemethy, G.; Scheraga, H.A. *Biopolymers* 1990, *30*, 951. c) Zagari, A.; Nemethy, G.; Scheraga, H.A. *Biopolymers* 1990, *30*, 961. d) Tsai, F.H.; Overberger, C.G.; Zand, R. *Biopolymers* 1990, *30*, 1039.
- 35. Cromwell, N.H.; Phillips, B. Chem. Rev. 1979, 79, 331-358.
- Backes, J. in Organische Stickstoffverbindungen III (Houden-Weyl), Band E16c, pp. 729-909, Ed. Klamonn, D., G. Thieme Verlag, Stuttgart, New York (1992).
- 37. Dureault, A.; Portal, M.; Carreoux, F.; Depezay, J.C. Tetrahedron 1993, 49, 4201-4210.
- 38. Sulmon, P.; De Kimpe, N.; Schamp, N. Tetrahedron 1988, 44, 3653-3670.
- 39. Sulmon, P.; De Kimpe, N.; Schamp, N. Tetrahedron 1989, 45, 2937-2955.
- 40. De Kimpe, N.; Stevens, C. Synthesis 1993, 89-91.
- 41. Boeykens, M.; De Kimpe, N.; Abbaspour Tehrani, K. J. Org. Chem. 1994, 59, 6973-6985.
- 42. Fedor, L.R.; Bruice, T.C.; Kirk, K.L., Meinwald, J. J. Am. Chem. Soc., 1966, 88, 108.
- 43. Kobayashi, J.; Cheng, J.; Ishibashi, M.; Wälchli, M.R.; Yamamura, S.; Ohizumi, Y. J. Chem. Soc. Perkin Trans 1, 1991, 1135-1137.
- 44. Alvi, K.A.; Jaspars, M.; Crews, P.; Strulovici, B.; Oto, E. Bioorg. Med. Chem. Lett. 1994, 4, 2447-2450.
- 45. Kobayashi, J.; Tsuda, M.; Cheng, J.-F.; Ishibashi, M.; Takihawa, H.; Mori, K. Tetrahedron Lett. 1996, 37, 6775-6776.