Investigations on the 4-Quinolone-3-carboxylic Acid Motif. 2. Synthesis and Structure–Activity Relationship of Potent and Selective Cannabinoid-2 Receptor Agonists Endowed with Analgesic Activity in $Vivo^{\perp}$

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Quinolone-3-carboxamides **11** bearing at position 5, 6, 7, or 8 diverse substituents such as halides, alkyl, aryl, alkoxy, and aryloxy groups differing in their steric/electronic properties, were prepared. The new compounds were tested in vitro for CB1 and CB2 receptor affinity in comparison with the reference compounds rimonabant and SR144528. The tested compounds exhibited CB2 affinity in the range from 55.9 to 0.8 nM and CB1 affinity in the range from >10 000 to 5.3 nM, with selectivity indeces [K_i (CB1)/ K_i (CB2)] varying from >2666.6 to 1.23. On the basis of the structure–selectivity relationship developed, the presence of a substituent at C6/C8 or C7 well accounts for the high or low CB2 selectivity, respectively. Compound **11c**, characterized by high CB2 affinity and selectivity, showed analgesic activity in the formalin test of acute peripheral and inflammatory pain in mice as a result of selective CB2 agonistic activity.

Introduction

The progress achieved over the past 15 years in understanding the action mechanisms of cannabinoids has revived the therapeutic interest in these substances.¹ On the basis of several controlled clinical trials, it is possible to affirm that cannabinoids possess interesting therapeutic potential in a number of pathological conditions, such as pain, immunosuppression, peripheral vascular disease, appetite enhancement or suppression, and locomotor disorders.² Currently, cannabinoid type 1 receptor (CB1) antagonists/inverse agonists are evaluated for obesity, metabolic disorders, smoking cessation, and alcohol abuse.^{3,4} Rimonabant (1) (Chart 1) is a potent and selective antagonist for the CB1 that was launched in Europe in 2006 by Sanofi-Aventis for the treatment of obesity and associated risk factors.^{5,6} On the other hand, selective cannabinoid type 2 receptor (CB2) ligands may be used to treat pain,⁷ inflammation,⁸ osteoporosis,⁹ CB2-expressing malignant gliomas,¹⁰ tumors of immune origin,¹¹ and immunological disorders such as multiple sclerosis.¹² Among the different therapeutic strategies designed to interfere with the deleterious processes involved in the pathology of multiple sclerosis, the use of cannabinoid compounds is currently one of the most promising ones.^{13,14} Growing literature describes a beneficial role of cannabinoids, and more specifically CB2 selective agonists such as 2 (JWH- $(133)^2$ in Alzheimer's disease, the most common form of dementia, by blocking β -amyloid peptide-induced activation of microglial cells.¹⁵ Since CB2 appears to be almost exclusively expressed by microglia, blockade of its activation may be attained by CB2 agonists with no overt psychoactivity. All these findings may set the basis for the use of CB2 agonists as a therapeutic approach for the treatment of multiple sclerosis and the prevention of neurodegenerative disorders, such as Alzheimer's disease. CB2 activation can also produce analgesic effects devoid of psychotropic activity, and importantly, the CB2-specific ligand **3** (GW842166X)¹⁶ is under clinical development for the treatment of inflammatory pain. Other representatives of CB2 selective agonists are, for example, compounds **4** (HU308), **5** (AM1241), **6** (GW405833), and **7** (JWH-015), while the pyrazole derivative **8** (SR144528) is a CB2 selective antagonist/inverse agonist.²

Very recently, 1.8-naphthyridine-4(1H)-on-3-carboxamide derivatives,¹⁷ such as 9, and 4-oxo-1,4-dihydroquinoline-3carboxamide derivatives¹⁸ (for example, **10**) have been described as structurally diverse ligands endowed with high affinity and selectivity toward CB2 (Chart 2). Our interest in the quinolone chemistry¹⁹ led us to choose compound **10** as the prototype for the development of a new family of CB2 selective agonists. This compound is characterized by a 1-adamantyl substituent on the carboxamide nitrogen and a *n*-pentyl substituent at the N-1 position. It was demonstrated that whatever the carboxamide substituent, the *n*-pentyl group is the preferred N-1 side chain.¹⁸ However, the effects on CB2 affinity and/or selectivity of modifications on the benzene portion of the quinolone ring have not been explored in great detail so far, as only few examples of such modified quinolone carboxamides have been recently reported.^{17,20} In light of these considerations and following our previous findings on CB1/CB2 ligands,²¹ we decided to investigate the effect on cannabinoid receptor activity/selectivity of decoration of the quinolone nucleus with various substituents, such as halides, alkyl, aryl, alkoxy, and aryloxy groups differing in their respective steric/electronic properties as well as their position on the condensed benzene ring. The N-1 pentyl chain

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Chart 1. CB1 and CB2 Receptors Selective Ligands



Chart 2. CB2 Selective Quinolones





 $R_5 = 1$ -pentyl, 1-pentenyl, 1-butenyl, allyl

X = 1-adamantyl, (R)-1-phenylethyl

and adamantyl group were either retained or replaced with other lipophilic groups, such as prop-1-en-3-yl, but-1-en-4-yl, pent-1-en-5-yl, or (R)-1-phenylethyl, to obtain the new quinolon-ecarboxamides of general structure **11**.²²

Chemistry

The synthesis of the target compounds 11a - v is highlighted in Schemes 1–4. Initially, in order to achieve in a fast way some chemical diversity, a parallel synthesis approach was performed using a carousel reaction station. Six commercially available or easily accessible anilines²³ were condensed with diethyl ethoxymethylenemalonate (EMME)^{*a*} according to the Gould–Jacobs reaction.²⁴ The intermediate enaminoesters were directly cyclized in refluxing diphenyl ether, and the quinolone esters were hydrolyzed by adding 10% aqueous NaOH to the reaction mixture and by heating at reflux temperature for 4 h. Acids were precipitated by adding HCl to the reaction mixture and collected by filtration. In this way it was possible to prepare six 4-oxo-1,4-dihydroquinoline-3-carboxylic acids (**12a**–**f**) in a single step in good overall yield (23–40%) (Scheme 1, Table 1).

Acids 12a-f were subjected to coupling reaction with 1-aminoadamantane using 1-hydroxybenzotriazole (HOBt) and *O*-benzotriazol-1-yl-*N*,*N*,*N'*,*N'*-tetramethyluronium hexafluorophosphate (HBTU) as coupling reagents in the presence of diisopropylethylamine (DIPEA) in DMF to give the corresponding amides, which in turn were directly reacted with the appropriate alkyl halide and potassium carbonate in the same solvent. The final compounds 11b-h were obtained in a satisfactory overall yield of 32-50%, whereas 11a was obtained in low yield (7%), probably because of some steric hindrance toward the approaching electrophile exerted by the methyl substituent at position 8.

Other final compounds 11i-m were prepared by a linear, step-by-step synthesis (Scheme 2). Thus, esters $13a^{25}$ and $13b^{26}$

^{*a*} Abbreviations: EMME, diethyl ethoxymethylenemalonate; HOBt, 1-hydroxybenzotriazole; HBTU, *O*-benzotriazol-1-yl-*N*,*N*',*N*'-tetramethyluronium hexafluorophosphate; DIPEA, diisopropylethylamine; SI, selectivity index.

Scheme 1^a



11a-h

^{*a*} For R₁–R₅, see Table 1. Reagents and conditions: (i) diethyl ethoxymethylenemalonate, 120 °C, 4 h; (ii) diphenyl ether, reflux, 16 h; (iii) 10% NaOH, reflux, 4 h, then HCl; (iv) 1-aminoadamantane, HOBt, HBTU, DIPEA, DMF, room temp, 20 h; (v) alkyl halide, KI, K₂CO₃, DMF, 90 °C, 20 h.

Scheme 2^a



^{*a*} For R₁–R₅, see Table 1. Reagents and conditions: (i) see (v) in Scheme 1; (ii) 10% NaOH, reflux, 2 h, then HCl; (iii) see (iv) in Scheme 1; (iv) ethyl 3-dimethylaminoacrylate, Et₃N, toluene, 100 °C, 4 h; (v) 1-pentylamine, K₂CO₃, DMF, 100 °C, 2 h; (vi) 6 N HCl, reflux, 7 h.

were first N-alkylated to 14a-c, which were then hydrolyzed to the corresponding acids 15a-c. Finally, coupling reaction of 15a-c with 1-aminoadamantane under the usual conditions provided amides 11i-k.

7-Fluoro-4-oxo-1-(1-pentyl)-1,4-dihydroquinoline-3-carboxylic acid (**15d**) was synthesized in a different way, according to a modified Grohe—Heitzer procedure.²⁷ Treatment of 2,4difluorobenzoyl chloride with 3-dimethylaminopropenoic acid ethyl ester led to the enamino ketone **16** that was subsequently reacted with *n*-pentylamine to yield the quinolone ester **17**. Because of the high reactivity toward nucleophiles exhibited by the fluorine atom at position 7 (para to the carbonyl group), the ester **17** could not be subjected to hydrolysis under basic conditions but was conveniently hydrolyzed using 6 N HCl. The product **15d** so obtained was transformed into amides **111,m** by coupling with 1-aminoadamantane and (*R*)-1-phenylethylamine. Taking advantage of the ease of displacement of their fluorine substituent, amides **111,m** were reacted with different nucleophiles to enhance the chemical diversity within this family of 4-oxoquinoline-3-carboxamides. Accordingly, when treated with pyrrolidine or sodium ethoxide, **11m** afforded **11n** and **11o** in 38% and 61% yield, respectively (Scheme 3). Similarly, amide **111** yielded the alkoxy derivatives **11p** (60%) and **11q** (50%) by reaction with sodium methoxide or sodium ethoxide, respectively. Displacement of fluorine by thiophenol converted **111** into **11r**, which in turn was oxidized using Oxone in 1,4-dioxane/water to give an approximately 1:1 mixture of the corresponding sulfone **11s** and sulfoxide **11t**, which were separated by column chromatography on silica gel.

Finally, the bromoamide **11g** was further modified through Suzuki coupling using phenylboronic acid and *trans*-(4methoxyphenyl)vinylboronic acid under microwave irradia-

Scheme 3^a



^{*a*} Reagents and conditions: (i) pyrrolidine or thiophenol, K₂CO₃, DMF, 100 °C, 6 h; (ii) MeONa or EtONa, THF, 50 °C, 16 h; (iii) Oxone, 1,4-dioxane, H₂O, room temp, 24 h.

tion.²⁸ Compounds **11u** and **11v** were obtained in reasonable yield (30 and 45%, respectively) (Scheme 4).

Biology

The binding affinities (K_i values) of compounds **11** for human recombinant CB1 and CB2 receptors are reported in Table 2. Tested compounds were evaluated in parallel with **8** (Chart 1) and **1** as CB2 and CB1 reference compounds, respectively, as previously described.²⁹

The functional activity of compound **11c** was assessed using the formalin test of acute peripheral and inflammatory pain in mice. Formalin injection induces a biphasic stereotypical nocifensive behavior. Nociceptive responses are divided into an early, short lasting first phase (0–7 min) caused by a primary afferent discharge produced by the stimulus, followed by a quiescent period and then a second, prolonged phase (15–60 min) of tonic pain. Fifteen minutes before injection of formalin, mice received intraperitoneal (ip) administration of vehicle or **11c** (1 or 3 mg/kg), alone or in combination with the selective CB2 antagonist, 6-iodo-2-methyl-1-[2-(4-morpholinyl)ethyl]-1*H*-indol-3-yl](4-methoxyphenyl)methanone (AM630) (3 mg/kg, ip), administered 5 min before the compound. The results are presented in Figure 1.

Results and Discussion

Out of the 22 synthesized quinolone carboxamides 11, compounds 11p and 11q elicited solubility issues that prevented their biological testing as DMSO/water solution in the cannabinoid receptor binding assay, while amides 11a-o,r-v proved to be high affinity CB2 ligands, with K_i values ranging from 55.9 nM (11v) to 0.8 nM (11l) (Table 2). Under the conditions of our assay, compound 10, taken as a representative of

Scheme 4^a



^{*a*} Reagents and conditions: (i) phenylboronic acid or *trans*-2-(4-meth-oxyphenyl)vinylboronic acid, Pd(OAc)₂, Ph₃P, Na₂CO₃, DME, EtOH, microwave, 150 °C, 10 min.

quinolone CB2 agonist, showed $K_i = 4.7$ nM instead of the reported value of 16.4 nM,¹⁸ while the CB2-selective reference ligand **8** showed $K_i = 5.4$ nM. With regard to the CB1 affinity, K_i values spanned 4 orders of magnitude in the range from >10000 nM (**11a**,**j**,**u**) to 5.3 nM (**11r**), with **1** (used as the CB1-selective reference ligand) displaying $K_i = 12.0$ nM. As a consequence of the high variability of affinity for both CB1 and CB2 receptors, the CB2 selectivity index (SI), calculated as $K_i(CB1)/K_i(CB2)$ ratio, for compounds **11a–o,r–v** also varied widely, from >2666.6 to 1.2. In no case was the SI value lower than 1; that is, no compound exhibited a reversed selectivity (higher affinity for CB1 than for CB2).

The nature of the 3-carboxamido substituent seems to affect both receptor affinity and selectivity, since the highly lipophilic adamantylamido substituent confers higher affinity and selectivity for the CB2 receptor with respect to its 1-phenylethylamido counterpart (compare, for instance, **111** and **11m**) and the two completely unselective compounds **11m** and **11o** are both 1-phenylethylamido derivatives. Coming to the effects of the N1 alkyl chain, affinity increased in the order 1-butenyl < 1-pentyl < 1-pentenyl, while no substantial effect on selectivity was noticed (compare **11d**, **11g**, and **11f**). While this result could be expected on the basis of the findings of Stern et al.,¹⁸ no definitive explanation for the better affinity and selectivity profile of the *N*-allyl derivative **11j**, with respect to the corresponding *N*-pentyl analogue **11i**, can yet be given.

To best discuss the effects of substitution on the condensed benzene ring, the issues of CB2 affinity and selectivity should be distinguished. Thus, several substituents with different steric and electronic properties are compatible with high CB2 affinity, since seven compounds (111, 11s, 11r, 11h, 11k, 11t, and 11u) displayed a K_i value lower than that of the unsubstituted compound 10, and even the less active compound 11v was still a nanomolar affinity ligand with a K_i value of 55.9 nM. On the other hand, a preliminary structure-selectivity relationship emerged, with CB2 selectivity depending on the substitution pattern on the aromatic ring. In fact, when the quinolone carboxamides are ranked in order of decreasing SI value, it can be seen that the 12 more selective derivatives 11u, 11j, 11a, **11b**, **11c**, **11h**, **11d**, **11f**, **11g**, **11v**, **11e**, and **11i** (2666.6 \ge SI \geq 39.9) are all substituted at least at C6 or C8, followed by 11k (SI = 38.8) bearing substituents at both C6 and C7, whereas the compounds 111, 11n, 11t, 11s, 11r, 11m, and 11o (38.1 \geq $SI \ge 1.2$) substituted only at C7, along with the unsubstituted compound 10 (SI = 9.0), all belong to the group of the less selective ligands. The effect on selectivity of C5 or C8 substitution cannot be yet defined, since the C5 or C8 substituted compounds 11a, 11i, and 11j also present substituents at other positions on the benzene ring. On the basis of these observations, the possibility for a substituent at the C6/C8 positions on the quinolone ring of 11, regardless of its electronic properties, to occupy defined regions of the space inside CB2 receptor well accounts for the ability of the given ligand to selectively bind to this receptor subtype. That does not mean that substitution at C7 position is detrimental for CB2 affinity, since some of the 7-substituted quinolone carboxamides, such as 111, 11s, 11r, and 11t, are among those with the highest affinity at CB2 receptor. This finding is in line with those reported by Stern et al.,¹⁸ who demonstrated that the introduction of a chlorine atom at position 7 of the quinoline template has a favorable effect on CB2 affinity. On the other hand, moving the substituent from C6 to C7 markedly enhances also CB1 affinity, thus giving rise to compounds endowed with lower selectivity (compare, for instance, 11h and 11r). Such a result is difficult to be compared with Manera's findings, regarding the increased or decreased CB2 selectivity for their 7-chloro or 7-methyl substituted quinolones, respectively. However, it is considered that compounds of general structure 11 here described differ from those reported by Manera et al.¹⁷ (such as 9), being characterized by different substituents at positions 1 and 3 of the bicyclic nucleus.

When tested for functional activity, compound **11c**, one of the most potent and CB2-selective ligands in binding assays, dose-dependently inhibited the second phase of the formalininduced nocifensive response in mice (Figure 1A). The effect was maximal with the 3 mg/kg dose, the action of which was significantly attenuated by the CB2 antagonist AM630 (1 mg/ kg) (Figure 1B). These findings suggest that **11c** is able to exert analgesic activity in vivo by acting as a CB2 agonist.

Conclusions

The results of our preliminary screening program on substituted quinolone-3-carboxamides led to the identification of cannabinoid receptor ligands characterized by high affinity and selectivity. In in vitro assays, some of the new compounds exhibited affinity and selectivity values for the CB2 subtype that are even higher than those of the CB2-specific ligand 8, used as the reference compound. Such a biochemical profile is strongly dependent upon the presence of a substituent at the C6 position of the quinolone ring, while the nature of the substituent itself is far less crucial. The activity shown by derivative **11c** in the formalin-induced nocifensive response in mice suggests that quinolone-3-carboxamides possess agonist properties at the CB2 receptor and might be considered as potential analgesic agents. On the other hand, ligands with high CB1 affinity, though substantially devoid of receptor selectivity, were identified as well, possessing a C7-substituent on the quinolone nucleus. Also in this case, some of the compounds showed CB1 affinity superior to that of 1, used in our tests as the CB1 reference standard.

The structure—affinity/selectivity relationships emerging from our study suggest that the quinolone carboxamide scaffold can be optimized through selected chemical modifications aimed at designing either CB1 or CB2 selective ligands.

Experimental Section

Chemistry. Reagents were purchased from commercial suppliers and used without further purification. Anhydrous reactions were run under (a positive pressure) dry N₂. Merck silica gel 60 was



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compd	R ₁	R ₂	R ₃	R_4	R ₅	R ₆	
11a	Н	Н	Cl	Me	1-pentyl	1-adamantylamino	
11b	Η	CF ₃	Н	Н	1-pentyl	1-adamantylamino	
11c	Η	<i>i</i> -Pr	Н	Н	1-pentyl	1-adamantylamino	
11d	Н	Br	Н	Н	but-1-en-4-yl	1-adamantylamino	
11e	Η	3-Cl-4-F-PhO	Н	Η	1-pentyl	1-adamantylamino	
11f	Η	Br	Н	Η	pent-1-en-5-yl	1-adamantylamino	
11g	Н	Br	Н	Н	1-pentyl	1-adamantylamino	
11h	Н	PhS	Н	Н	1-pentyl	1-adamantylamino	
11i	MeO	Н	Н	MeO	1-pentyl	1-adamantylamino	
11j	MeO	Н	Н	MeO	allyl	1-adamantylamino	
11k	Η	F	pyrrol-1-yl	Η	1-pentyl	1-adamantylamino	
111	Η	Н	F	Н	1-pentyl	1-adamantylamino	
11m	Η	Н	F	Η	1-pentyl	(R)-1-phenylethylamino	
11n	Н	Н	pyrrolidin-1-yl	Н	1-pentyl	(R)-1-phenylethylamino	
110	Н	Н	EtO	Н	1-pentyl	(R)-1-phenylethylamino	
11p	Н	Н	MeO	Η	1-pentyl	1-adamantylamino	
11q	Н	Н	EtO	Н	1-pentyl	1-adamantylamino	
11r	Н	Н	PhS	Н	1-pentyl	1-adamantylamino	
11s	Н	Н	PhSO ₂	Н	1-pentyl	1-adamantylamino	
11t	Н	Н	PhSO	Н	1-pentyl	1-adamantylamino	
11u	Н	Ph	Н	Н	1-pentyl	1-adamantylamino	
11v	Н	(E)-4-MeO-styryl	Н	Н	1-pentyl	1-adamantylamino	
12a	Н	Н	Cl	Me	Н	OH	
12b	Н	CF ₃	Н	Н	Н	OH	
12c	Н	<i>i</i> -Pr	Н	Н	Н	OH	
12d	Н	Br	Н	Н	Н	OH	
12e	Н	3-Cl-4-F-PhO	Н	Н	Н	OH	
12f	Н	PhS	Н	Н	Н	OH	
13a	MeO	Н	Н	MeO	Н	OEt	
13b	Н	F	pyrrol-1-yl	Н	Н	OEt	
14a	MeO	Н	Н	MeO	1-pentyl	OEt	
14b	MeO	Н	Н	MeO	allyl	OEt	
14c	Н	F	pyrrol-1-yl	Н	1-pentyl	OEt	
15a	MeO	Н	Н	MeO	1-pentyl	OH	
15b	MeO	Н	Н	MeO	allyl	OH	
15c	Н	F	pyrrol-1-yl	Η	1-pentyl	OH	
15d	Н	Н	F	Н	1-pentyl	OH	

used for flash chromatography (23–400 mesh). ¹H NMR and ¹³C NMR were recorded at 200 and 50 MHz, respectively, on a Brucker AC200F spectrometer and at 400 and 100 MHz on a Brucker Advance DPX400. Chemical shifts are reported relative to tetramethylsilane at 0.00 ppm. Mass spectral (MS) data were obtained using an Agilent 1100 LC/MSD VL system (G1946C) with a 0.4 mL/min flow rate using a binary solvent system of 95:5 methanol/water. UV detection was monitored at 254 nm. Mass spectra were acquired either in positive mode or in negative mode scanning over the mass range of 105–1500. Melting points were determined on a Gallenkamp apparatus and are uncorrected. Elemental analyses were performed on a Perkin-Elmer PE 2004 elemental analyzer, and the data for C, H, and N are within 0.4% of the theoretical values. Microwave irradiations were conducted using a CEM Discover Synthesis Unit (CEM Corp., Matthews, NC).

Parallel Synthesis of Acid Derivatives 12a–f. Six anilines (1 mmol) were added into six round-bottom flasks of a carousel reactor, and diethyl ethoxymethylemalonate (1 mmol) was added. The reaction mixtures were heated at 120 °C for 4 h and cooled to room temperature. Ph₂O (1 mL) was added to each vessel, and the reaction mixtures were heated at reflux temperature for 16 h. After the mixture was cooled to room temperature, 10% aqueous NaOH was added to each flask and the reaction mixtures were refluxed for 4 h. After the mixture was cooled, concentrated HCl was added to the reaction mixtures, allowing the precipitation of the five acid derivatives, which were collected by filtration, washed with water, then petroleum ether, and recrystallized from DMF.

Example. 4-Oxo-6-(trifluoromethyl)-1,4-dihydroquinoline-3-carboxylic Acid (12b). 12b was prepared by parallel synthesis from 4-(trifluoromethyl)aniline in 25% yield. White solid; mp >280 °C (dec). ¹H NMR (200 MHz, DMSO-*d*₆): δ 17.09 (s, 1H), 8.83 (s, 1H), 8.44 (s, 1H), 7.81–7.75 (m, 2H). MS (ESI): *m/z* 258 [M + H]⁺, 280 [M + Na]⁺. IR (nujol): ν 3406, 1643, 1619 cm⁻¹. Anal. (C₁₁H₆F₃NO₃) C, H, N.

Parallel Synthesis of Amides 11a–h. Acids **12a–f** (1 mmol) and DMF (3 mL) were placed in different vessels of a Syncore reactor under a nitrogen atmosphere. HOBt (1 mmol), HBTU (2 mmol), DIPEA (1.5 mmol), and 1-aminoadamantane (1.2 mmol) were added to the solutions, and the reaction mixtures were stirred at room temperature for 30 min. Further DIPEA (1.5 mmol) was thereafter added, and the reaction mixtures were stirred at room temperature for 20 h. K₂CO₃ (5 mmol), the appropriate alkyl halide (5 mmol), and KI (5 mmol) were added to every mixture, which was heated at 90 °C for 20 h. Each reaction mixture was poured into ice and extracted with AcOEt. The organic layers were washed with brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The crude residues were purified by chromatography using light petroleum ether/AcOEt (5:1 to 1:1) to afford the pure amides.

Example. *N*-(Adamant-1-yl)-7-chloro-8-methyl-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxamide (11a). 11a was prepared by parallel synthesis from 12a in 7% yield. Light-yellow oil. ¹H NMR (200 MHz, CDCl₃): δ 9.29 (s, 1H), 8.51 (s, 1H), 7.88 (d, *J* = 9.0 Hz, 1H), 7.51 (d, *J* = 9.0 Hz, 1H), 4.12 (t, *J* = 6.8 Hz, 2H), 2.8 (s, 3H), 2.15–1.96 (m, 8H), 1.92–1.89 (m, 2H), 1.73–1.63 (m, 7H),

Table 2. CB1 and CB2 Receptor Affinity Values for Compounds $11a-v^a$

	K_{i}^{f} (1	nM)	
compd	$CB1^{b,d}$	$CB2^{c,d}$	SI^e
11a	>10000	25.5	>392.7
11b	2520	11.6	216.7
11c	1220	6.3	194.3
11d	2680	23.8	112.7
11e	739	16.1	46.0
11f	696	9.4	74.4
11g	996	14.3	69.4
11h	640	3.4	189.3
11i	2100	52.6	39.9
11j	>10000	9.2	>1091.7
11k	139.8	3.6	38.8
111	30.5	0.8	38.1
11m	14.3	10.7	1.3
11n	160	17.1	9.4
110	29.7	24.2	1.2
11p	NT^{g}	NT^{g}	
11q	NT^{g}	NT^{g}	
11r	5.3	1.6	3.3
11s	6.1	0.9	6.8
11t	28.0	3.5	8.0
11u	>10000	3.8	>2666.6
11v	2630	55.9	47.0
10^h	42.1	4.7	9.0
$8^{h,i}$	>2820	5.4	> 522.2
$1^{h,j}$	12.0	790	0.015

^{*a*} Data represent mean values for at least three separate experiments performed in duplicate and are expressed as K_i (nM). ^{*b*} CB1: human cannabinoid type 1 receptor. ^{*c*} CB2: human cannabinoid type 2 receptor. ^{*d*} For both receptor binding assays, the new compounds were tested using membranes from HEK cells transfected with either the CB1 or CB2 receptor and [³H]-(-)-*cis*-3-[2-hydroxy-4-(1,1-dimethylheptyl)phenyl]-*trans*-4-(3-hydroxypropyl)cyclohexanol ([³H]CP-55,940). ^{*e*} SI: selectivity index for CB2, calculated as $K_i(CB1)/K_i(CB2)$ ratio. ^{*f*} K_i : "Equilibrium dissociation constant", that is, the concentration of the competing ligand that will bind to half the binding sites at equilibrium in the absence of radioligand or other competitors. ^{*g*} NT: not tested because of solubility problems. ^{*h*} The binding affinities of reference compounds were evaluated in parallel with compounds **11** under the same conditions. ^{*i*} SR144528, CB2 reference compound. ^{*j*} Rimonabant, CB1 reference compound.

1.53-1.35 (m, 4H), 0.96 (t, J = 6.9 Hz, 3H). MS (ESI): m/z 441 [M + H]⁺, 464 [M + Na]⁺. Anal. (C₂₆H₃₃ClN₂O₂) C, H, N.

Synthesis of 4-Oxo-1,4-dihydroquinoline-3-carboxamides 11i-m by Amidation Reaction. General Procedure. To a solution of the appropriate acid derivative 15a-d (1 mmol) in DMF (5 mL), HBTU (2 mmol) was added followed by HOBt (1 mmol), DIPEA (1.5 mmol), and 1-aminoadamantane or (*R*)-1-phenylethylamine (1.2 mmol). The mixture was stirred at room temperature under N₂ for 30 min, and further DIPEA (1.5 mmol) was added. The reaction mixture was stirred at room temperature for 1 h, poured into water, and extracted with CH₂Cl₂. The organic layer was washed with brine, dried over anhydrous Na₂SO₄, and evaporated. The crude residue was recrystallized from EtOH, with the exception of 11k, which was triturated with diethyl ether.

Example. *N*-(Adamant-1-yl)-7-fluoro-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxamide (111). 111 was prepared from 15d and 1-aminoadamantane in 76% yield. Pink solid; mp 197.5–200.5 °C. ¹H NMR (400 MHz, CDCl₃): δ 9.76 (s, 1H), 8.65 (s, 1H), 8.49–8.42 (m, 1H), 7.15–7.07 (m, 2H), 4.09 (t, *J* = 7.5 Hz, 2H), 2.11–2.05 (m, 10H), 1.97–1.78 (m, 2H), 1.74–1.66 (m, 5H), 1.33–1.19 (m, 4H), 0.85 (t, *J* = 6.7 Hz, 3H). MS (ESI): *m/z* 411 [M + H]⁺, 433 [M + Na]⁺. IR (CHCl₃): 1659, 1605 cm⁻¹. Anal. (C₂₅H₃₁FN₂O₂) C, H, N.

Synthesis of 1-Alkyl-4-oxo-1,4-dihydroquinoline Derivatives 14a-c by Alkylation Reaction. General Procedure. To a mixture of the quinolone derivative 13a,b (1 mmol) and K₂CO₃ (2.8 mmol) in dry DMF (2 mL) under a nitrogen atmosphere, the appropriate alkyl iodide (2.8 mmol) or alkyl bromide along with KI (2.8 mmol each) was added. The reaction mixture was heated at 90 °C for 18 h and then poured into ice—water. The resulting *N*-alkylquinolone was collected by filtration and recrystallized from DMF or extracted in CH_2Cl_2 and purified by flash chromatography on silica gel eluting with $CH_2Cl_2/MeOH$ (98:2).

Example. Ethyl 5,8-Dimethoxy-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxylate (14a). 14a was prepared in 41% yield from **13a**. White solid, mp 121–122 °C. ¹H NMR (200 MHz, CDCl₃): δ 8.17 (s, 1H), 7.05 (d, J = 8.9 Hz, 1H), 6.76 (d, J = 8.9 Hz, 1H), 4.38–4.28 (m, 4H), 3.87 (s, 3H), 3.85 (s, 3H), 1.77–1.63 (m, 2H), 1.35 (t, J = 7.0 Hz, 3H), 1.28–1.20 (m, 4H), 0.86 (t, J = 6.2 Hz, 3H). MS (ESI): m/z 348 [M + H]⁺, 370 [M + Na]⁺. IR (nujol): ν 1656, 1615 cm⁻¹. Anal. (C₁₉H₂₅NO₅) C, H, N.

Synthesis of 7-Substituted-4-oxo-1,4-dihydroquinoline-3-carboxamides 11n and 11r. General Procedure. K_2CO_3 (3 mmol) and pyrrolidine (7.6 mmol) or thiophenol (3 mmol) were added to a solution of 11m or 11l (1 mmol), respectively, in DMF (8 mL) under a nitrogen atmosphere. The reaction mixture was heated at 100 °C for 6 h, cooled to room temperature, and poured into water. The solid precipitate was collected by filtration, washed with light petroleum ether, and recrystallized from EtOH.

Example. *N*-(Adamant-1-yl)-4-oxo-1-pentyl-7-(phenylthio)-1,4dihydroquinoline-3-carboxamide (11r). 11r was prepared from 111 and thiophenol in 58% yield. White solid; mp 164.5 °C. ¹H NMR (200 MHz, CDCl₃): δ 9.86 (s, 1H), 8.60 (s, 1H), 8.33 (d, *J* = 8.6 Hz, 1H), 7.57–7.52 (m, 2H), 7.50–7.41 (m, 3H), 7.24–7.23 (m, 1H), 7.19 (s, 1H), 3.92 (t, *J* = 7.7 Hz, 2H), 2.14–2.08 (m, 8H), 1.76–1.57 (m, 9H), 1.32–1.10 (m, 4H), 0.86 (t, *J* = 6.8 Hz, 3H). MS (ESI): *m*/*z* 501 [M + H]⁺, 523 [M + Na]⁺. IR (nujol): ν 1659, 1602 cm⁻¹. Anal. (C₃₁H₃₆N₂O₂S) C, H, N.

Synthesis of 7-Substituted-4-oxo-1,4-dihydroquinoline-3-carboxamides 110–q. General Procedure. A solution of 111 or 11m (0.14 mmol) in dry THF (3 mL) was added to a solution of freshly prepared MeONa or EtONa (0.28 mmol) under a nitrogen atmosphere. The reaction mixture was heated at 50 °C for 16 h and neutralized using 1 N HCl. The mixture was concentrated under reduced pressure and extracted with CH_2Cl_2 . The organic layer was washed with brine, dried over Na_2SO_4 , and evaporated to dryness. The crude residue was purified by chromatography using light petroleum ether:AcOEt (1:1) as eluent.

Example. (*R*)-7-Ethoxy-4-oxo-1-pentyl-*N*-(1-phenylethyl)-1,4-dihydroquinoline-3-carboxamide (110). 110 was prepared from 11m and EtONa in 61% yield. White solid; mp 128.2 °C (from EtOH). ¹H NMR (400 MHz CDCl₃): δ 10.50 (d, *J* = 6.8 Hz, 1H), 8.61 (s, 1H), 8.37 (d, *J* = 9.1 Hz, 1H), 7.36–7.14 (m, 5H), 7.0–6.97 (m, 1H), 6.76 (s, 1H), 5.24 (t, *J* = 7.0 Hz, 1H), 4.12–4.05 (m, 4H), 1.80–1.73 (m, 2H), 1.53 (d, *J* = 6.8 Hz, 3H), 1.42 (t, *J* = 6.8 Hz, 3H), 1.31–1.27 (m, 4H), 0.87–0.78 (m, 3H). MS (ESI): *m/z* 407 [M + H]⁺, 429 [M + Na]⁺. IR (nujol): ν 1667, 1603 cm⁻¹. Anal. (C₂₅H₃₀N₂O₃) C, H, N.

Synthesis of Compounds 11s and 11t. To a solution of Oxone (1.1 g, 1.8 mmol) in H₂O (2 mL), a solution of 3r (28 mg, 0.06 mmol) in 1,4-dioxane (2 mL) was added. The reaction mixture was stirred at room temperature for 20 h and extracted with CH_2Cl_2 . The organic layers were washed with brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The crude residue was purified by chromatography using light petroleum ether/AcOEt (2:1) as eluent to provide compounds 11s (9.5 mg, 33% yield) and 11t (11 mg, 37% yield) as white solids.

N-(Adamant-1-yl)-4-oxo-1-pentyl-7-(phenylsulfonyl)-1,4-dihydroquinoline-3-carboxamide (11s). Mp 215.6 °C. ¹H NMR (200 MHz, CDCl₃): δ 9.36 (s, 1H), 8.78 (s, 1H), 8.60 (d, J = 8.7 Hz, 1H), 8.17 (s, 1H), 7.99–7.96 (m, 2H), 7.82 (d, J = 8.7 Hz, 1H), 7.57–7.54 (m, 3H), 4.26 (t, J = 7.5 Hz, 2H), 2.14–2.02 (m, 10H), 1.88–1.81 (m, 2H), 1.80–1.70 (m, 5H), 1.37 (m, 4H), 0.92–0.84 (m, 3H). MS (ESI): m/z 555 [M + Na]⁺. IR (CHCl₃): ν 1716, 1659 1598 cm⁻¹. Anal. (C₃₁H₃₆N₂O₄S) C, H, N.

N-(Adamant-1-yl)-4-oxo-1-pentyl-7-(phenylsulfinyl)-1,4-dihydroquinoline-3-carboxamide (11t). Mp 102-103 °C. ¹H NMR (200 MHz, CDCl₃): δ 9.73 (s, 1H), 8.78 (s, 1H), 8.50 (d, J = 8.1 Hz, 1H), 8.02 (s, 1H), 7.68-7.34 (m, 6H), 4.27 (t, J = 7.2 Hz, 2H), 2.14-2.09 (m, 10H), 1.90-1.81 (m, 2H), 1.80-1.71 (m, 5H), 1.37



Figure 1. (A) Analgesic effect of 11c in the formalin test in mice. Each point represents the mean \pm standard error of the mean (SEM) of 8–10 animals per group. Asterisks denote values that were significantly different from vehicle (P < 0.05), as assessed using two-way ANOVA followed by the Bonferroni's test. (B) Effect of the CB2 antagonist 6-iodo-2-methyl-1-[2-(4-morpholinyl)ethyl]-1*H*-indol-3-y l](4-methoxyphenyl)methanone (AM630) on the analgesic effect of 11c. Asterisks denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05), whereas circles denote values that were significantly different from vehicle (P < 0.05).

(m, 4H), 0.92–0.86 (m, 3H). MS (ESI): m/z 517 [M + 1]⁺, 539 [M + Na]⁺. IR (CHCl₃): ν 1722, 1660, 1598 cm⁻¹. Anal. (C₃₁H₃₆N₂O₃S) C, H, N.

Synthesis of Compounds 11u and 11v by Suzuki Reaction. General Procedure. To a solution of 11g (1 mmol) in DME (4 mL), $Pd(OAc)_2$ (0.1 mmol), PPh_3 (0.3 mmol), the appropriate boronic acid (0.5 mmol), EtOH (1 mL), and 1 N Na₂CO₃ (2 mL) were added. The mixture was irradiated with microwaves at 150 °C for 10 min, then filtered through a plug of Celite. The filtrate was washed with H₂O, brine, dried over anhydrous Na₂SO₄, and evaporated to dryness. The solid residue was purified by flash chromatography using CH₂Cl₂ as eluent.

Example. *N*-(Adamant-1-yl)-4-oxo-1-pentyl-6-phenyl-1,4-dihydroquinoline-3-carboxamide (11u). 11u was prepared from 11g and phenylboronic acid in 45% yield. Yellow oil. ¹H NMR (200 MHz, CDCl₃): δ 9.97 (s, 1H), 8.76–8.74 (m, 2H), 7.96 (dd, $J_1 = 1.8$ Hz, $J_2 = 8.8$ Hz, 1H), 7.72–7.67 (m, 2H), 7.57 (d, J = 8.9 Hz, 1H), 7.51–7.32 (m, 3H), 4.23 (t, J = 7.4 Hz, 2H), 2.18–2.10 (m, 8H), 1.95–1.78 (m, 2H), 1.72–1.62 (m, 7H), 1.40–1.27 (m, 4H), 0.91 (t, J = 6.8 Hz, 3H). MS (ESI): m/z 469 [M + H]⁺, 491 [M + Na]⁺. IR (CHCl₃): ν 3248, 1657, 1601 cm⁻¹. Anal. (C₃₁H₃₆N₂O₂) C, H, N.

Synthesis of 1-Alkyl-4-oxo-1,4-dihydroquinoline-3-carboxylic Acids 15a-c by Basic Hydrolysis. General Procedure. A suspension of ester 14a-c (1 mmol) in 10% aqueous NaOH (10 mL) was refluxed for 2 h (until the suspension became a solution). After cooling at room temperature, the reaction mixture was acidified using concentrated HCl. The resulting precipitate was filtered and washed with water to give the corresponding quinolone-3-carboxylic acid, which was recrystallized from EtOH.

Example. 5,8-Dimethoxy-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxylic Acid (15a). 15a was prepared from **14a** in 86% yield. White solid; mp 209.7 °C. ¹H NMR (200 MHz, DMSO-*d*₆): δ 15.73 (s, 1H), 8.59 (s, 1H), 7.40 (d, J = 9.0 Hz, 1H), 6.97 (d, J = 9.0 Hz, 1H), 4.54 (t, J = 7.5 Hz, 2H), 3.87 (s, 3H), 3.78 (s, 3H), 1.68–1.50 (m, 2H), 1.03 (m, 4H), 0.81 (t, J = 6.8 Hz, 3H). MS (ESI): *m/z* 320 [M + H]⁺, 342 [M + Na]⁺. IR (nujol): ν 3378, 1728, 1630 cm⁻¹. Anal. (C₁₇H₂₁NO₅) C, H, N. **7-Fluoro-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxylic Acid** (**15d**). A suspension of **17** (305 mg, 1 mmol) in EtOH (3.5 mL) and 6 N HCl (3.5 mL) was refluxed for 7 h. After the mixture was cooled to room temperature, the solid precipitate was filtered and recrystallized from EtOH to furnish **15d** (260 mg, 94%) as a light-pink solid; mp 148.3 °C. ¹H NMR (200 MHz, CDCl₃): δ 14.7 (b, 1H), 8.73 (s, 1H), 8.61–8.53 (dd, J_1 = 6.6 Hz, J_2 = 8.8 Hz, 1H), 7.33–7.21 (m, 2H), 4.23 (t, J = 7.3 Hz, 2H), 1.95–1.88 (m, 2H), 1.55–1.39 (m, 4H), 0.96–0.89 (m, 3H). MS (ESI): m/z 278 [M + H]⁺, 300 [M + Na]⁺. IR (CHCl₃): ν 1722, 1615 cm⁻¹. Anal. (C₁₅H₁₆FNO₃) C, H, N.

Ethyl 2-(2,4-Difluorobenzoyl)-3-(dimethylamino)acrylate (16). To a solution of 2,4-difluorobenzoyl chloride (2.8 mL, 22.7 mmol) and Et₃N (4.8 mL, 34.1 mmol) in toluene, ethyl (dimethylamino)acrylate (3.2 g, 22.7 mmol) was added, and the reaction mixture was heated at 90 °C for 4 h. The solvent was removed at reduced pressure, and the crude solid was purified by chromatography using AcOEt/light petroleum ether (1:1 to 2:1) to afford 16 (4.29 g, 67% yield) as a yellow solid, which was recrystallized from EtOH. Mp >280 °C (dec). ¹H NMR (200 MHz CDCl₃): δ 7.73 (s, 1H), 7.66–7.55 (m, 1H), 6.92–6.82 (m, 1H), 6.79–6.69 (m, 1H), 3.97 (q, *J* = 7.1 Hz, 2H), 3.05 (b, 6H), 0.94 (t, *J* = 7.1 Hz, 3H). MS (ESI): *m/z* 284 [M + H]⁺, 306 [M + Na]⁺. IR (CHCl₃): ν 1725, 1612, 1234, 1220 cm⁻¹. Anal. (C₁₄H₁₅F₂NO₃) C, H, N.

Ethyl 7-Fluoro-4-oxo-1-pentyl-1,4-dihydroquinoline-3-carboxylate (17). *n*-Pentylamine (1.7 mL, 15.2 mmol) was added dropwise to a solution of **16** (4.29 g, 15.2 mmol) in Et₂O (160 mL) and EtOH (69 mL). The reaction mixture was stirred at room temperature for 25 min, and the solvent was removed under reduced pressure. The residue was washed with 0.1 N HCl, dried under anhydrous Na₂SO₄, and evaporated to dryness. DMF (20 mL) and K₂CO₃ (3.5 g, 25.6 mmol) were added to the solid residue, and the mixture was heated at 90 °C for 1 h, then poured into ice—water. The precipitated solid was collected by filtration and recrystallized from EtOH to give **17** (3.88 g, 89% yield) as a yellow solid; mp 104–106.3 °C. ¹H NMR (200 MHz, CDCl₃): δ 8.57–8.49 (m, 1H), 8.43 (s,1H), 7.16–7.04 (m, 2H), 4.39 (q, *J* = 7.2 Hz, 2H), 4.08 (t, *J* = 7.5 Hz, 2H), 1.90–1.80 (m, 2H), 1.43–1.34 (m, 7H), 0.92 (t, *J* = 6.6 Hz, 3H). MS (ESI): m/z 306 [M + H]⁺, 328 [M + Na]⁺. IR (CHCl₃): ν 1726, 1689 cm⁻¹. Anal. (C₁₇H₂₀FNO₃) C, H, N.

CB1 and CB2 Receptors Binding Assays. The new compounds were evaluated in CB1 and CB2 receptors binding assays using membranes from HEK cells transfected with either the CB1 or CB2 receptor and [³H]-(-)-*cis*-3-[2-hydroxy-4-(1,1-dimethylheptyl)phe-nyl]-*trans*-4-(3-hydroxypropyl)cyclohexanol ([³H]CP-55,940; $K_d = 0.18$ nM for CB1R and $K_d = 0.31$ nM for CB2R) as the high affinity ligand, as described by the manufacturer (Perkin-Elmer, Italy).²⁹ Stock solutions of compounds were daily prepared in DMSO with final DMSO concentration less than 0.1%. Displacement curves were generated by incubating drugs with [³H]CP-55,940 (0.14 nM for CB1R and 0.084 nM for CB2R binding assay). In all cases, K_i values were calculated by applying the Cheng–Prusoff equation³⁰ to the IC₅₀ values (obtained by GraphPad) for the displacement of the bound radioligand by increasing concentrations of the test compounds.

Formalin Test in Mice. Mice received formalin (1.25%) in the dorsal surface of one side of the hind paw. Each mouse was randomly assigned to one of the experimental groups (n = 8-10) and placed in a Plexiglas cage and allowed to move freely for 15-20 min. A mirror was placed at a 45° angle under the cage to allow full view of the hind paws. Lifting, favoring, licking, shaking, and flinching of the injected paw were recorded as nociceptive responses. Fifteen minutes before injection of formalin, mice received intraperitoneal vehicle (10% DMSO in 0.9% NaCl, 50 μ L) or **11c** (1 or 3 mg/kg, ip) in the same volume solution, alone or in combination with the selective CB2 antagonist, AM630 (3 mg/kg, ip), administered 5 min before the compound. The total time of the nociceptive response was measured every 5 min and expressed as the total time of the nociceptive responses in min (mean \pm SEM). Recording of nociceptive behavior commenced immediately after formalin injection and was continued for 60 min.

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Supporting Information Available: Synthetic and spectral data for compounds 11b-k,m,n,p,q,v, 12a,c-f, 14b,c, 15b,c and elemental analyses for compounds 11–17. This material is available free of charge via the Internet at http://pubs.acs.org.

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