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An orally bioavailable positive allosteric modulator of the mGlu₄ receptor with efficacy in an animal model of motor dysfunction

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ABSTRACT

A high-throughput screening campaign identified 4-((*E*)-styryl)-pyrimidin-2-ylamine (**11**) as a positive allosteric modulator of the metabotropic glutamate (mGlu) receptor subtype 4. An evaluation of the structure–activity relationships (SAR) of **11** is described and the efficacy of this compound in a haloper-idol-induced catalepsy rat model following oral administration is presented.

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The metabotropic glutamate (mGlu) receptors are members of the Group C class of G-protein coupled receptors that bind the excitatory neurotransmitter glutamate. To date, there are eight known mGlu receptors (1-8) and they are sub-divided into three groups according to their amino acid sequence homology, signal transduction pathway and pharmacology (Group I: mGlu 1&5; Group II: mGlu 2&3; Group III: mGlu 4,6,7,8).^{1–4} These receptors are distributed throughout the CNS and they are responsible for the regulation of a variety of physiological processes. Two different approaches to modulate the biological response of the mGlu receptors have been postulated. One approach is to identify ligands that compete with glutamate for binding to the orthosteric site and this can lead to direct activation or inhibition of the receptor. The alternative approach is to identify ligands that do not compete with glutamate but instead bind to an allosteric site. Allosteric ligands can potentially bind in the presence of glutamate and positively or negatively modulate the response of the receptor.

The debilitating side effects associated with the current treatments for patients suffering from Parkinson's disease (PD) coupled with an ageing population indicate that PD is an area of neurological research with a growing unmet medical need.⁵ Stimulation of mGlu₄ is currently regarded as an attractive non-dopaminergic target for the palliative treatment of PD which might provide new drugs without the motor complications (dyskinesias) associated with the existing frontline therapy L-DOPA.^{6,7} Recently orthosteric mGlu₄ agonists such as L-AP-4 or LSP1-2111⁸ and mGlu₄ positive allosteric modulators (PAMs) such as (-)-PHCCC^{9,10} (1) have demonstrated improvement of motor function in pre-clinical rodent models of PD. One of the key advantages of positive allosteric modulation of mGlu₄¹¹ is that the mGlu allosteric sites are likely to be less well conserved across the different mGlu subtypes and hence the specificity for mGlu₄ over other mGlu receptors should be achievable. Additionally positive enhancers do not activate the receptor on their own and thereby minimize interference with the highly regulated neurotransmission. From the medicinal chemistry perspective another advantage of targeting the allosteric site instead of the orthosteric site is that the allosteric ligands will be different to glutamate and so they will have markedly different properties with the potential for improved central nervous system

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(CNS) exposure. Several literature reports of mGlu₄ PAMs that detail structure–activity relationships (SAR) conducted on different chemical series have emerged recently and selected examples of compounds 2,^{12,13} 3,¹⁴ 4,¹⁵ 5,¹⁴ 6,¹⁶ 7,^{17,18} from these publications are illustrated in Figure 1.

Following a high-throughput screening campaign on mGlu₄ we identified compound **8** (Fig. 2) which showed an EC₅₀ ~5 μ M (82% glutamate max) in our mGlu₄ positive modulator assay.¹⁹ Although **8** was a singleton it attracted our attention because of its structural similarity to SIB 1893 **9** and MPEP **10** which have been published previously as mGlu₅ negative allosteric modulators (NAMs) with weak mGlu₄ PAM activity.^{20,21} During the resynthesis of **8** we profiled the parent aminopyrimidine **11** (the product of a two step synthesis from (*E*)-4-phenyl-but-3-en-2-one²²) and found that this compound, and not the oxalate ester **8**, was the source of the mGlu₄ PAM activity with an EC₅₀ ~1 μ M (106% Glu max). With this result in hand we performed an SAR analysis around **11** in an effort to improve the potency and to identify compounds suitable for evaluation in vivo.

For our SAR exploration we divided compound **11** into four parts, the amino group, the pyrimidine, the olefin and the phenyl ring and made conservative modifications in each of these regions of the molecule.

Changes to the amino moiety on the pyrimidine ring are reported in Table 1. Methylation or dimethylation of 11 to provide 12 and 13 was detrimental to activity. Sulfonylation to give 14 also gave a compound that showed no activity at the highest concentration tested. Replacement of the amino group with either a methyl to give the SIB 1893 analogue 15 or a thiol to give 17 provided compounds that were \sim 7–8 μ M but introducing a phenyl ring in place of the amino moiety (compound 16) resulted in loss of activity. The results for 13, 14 and 16 suggest that there might be stringent steric requirements at this region of the binding site. Additionally it could be postulated that an H-bond donor is responsible for improved potency but it is perhaps inconclusive whether the amino group is forming a key hydrogen bond with the receptor. None of the modifications that were made to the amino group represented an advantage over compound **11** in terms of their positive modulatory effect on mGlu₄.

Minor changes to the pyrimidine moiety of **11** were generally well tolerated as shown for compounds **18–28** in Table 2. Addition of a methyl group in the 6-position of compound **11** led to the equipotent compound **18** whereas addition in the 5-position as in compound **19** led to an improvement in potency. Interestingly, when trying to combine this SAR by inserting methyl groups in



Figure 2. mGlu ligands.

Table 1Modifications to the amino group



Compound	R	$mGlu_4$ (h) EC_{50} (μM)	% Glu max ^a
11	NH ₂	1.0	106
12	NHMe	>30	_
13	NMe ₂	>30	_
14	NHSO ₂ Me	>30	_
15	Me	6.9	67
16	Ph	>30	_
17	SH	7.8	115

^a EC₅₀ for potentiation of an EC₂₀ glutamate concentration. All assay signals were normalised to saturating concentrations of (–)-PHCCC (1) which was set to 100% Glu max. Accordingly, the % Glu max of test compounds was referenced to 1 at maximal activation. In the assay described¹⁹ 1 yielded an average EC₅₀ of 5 μ M.

the 5- and 6-position as in compound **20** no synergistic effect could be seen. Complete loss of activity was induced when a typical hydrogen donor group such as the amino or a hydroxy group (compounds **21** and **22**) were attached in position 4 whereas capping the hydroxyl in **22** to give the methoxy derivative **23** regained activity. Introduction of small electron-withdrawing groups (**24**, **25** and **26**) in **11** had minimal effect on the potency in the mGlu₄ positive modulation assay. When a nitrogen was removed from **11** to give aminopyridine **27** or when the pyrimidine was replaced with a pyrazine as shown with compound **28** it had a detrimental effect on the activity suggesting that the nitrogen in the 1-position of **11** might be important for binding to mGlu₄.



Figure 1. Literature mGlu₄ PAMs.



The changes to the olefin moiety in **11** are presented in Table 3. Saturation of the double bond to give **29** resulted in a >30-fold drop in potency. Replacement of the olefin in **11** with an alkyne to give 30 also resulted in a drop in potency although it was less dramatic and the compound was active with an $EC_{50}\,{\sim}20~\mu M$ (86% Glu max). Cyclopropanation of the double bond to give 31 resulted in loss of potency, however, imposing additional conformation constraint on the scaffold as shown with the benzofuranyl derivative 32 was tolerated. When the size of the bicyclic ring was increased to a benzothiophenyl (33) or naphthyl (34), then no activity was observed at 30 µM.

All modifications that were made to the phenyl ring (Table 4) led to at least a twofold drop in activity on mGlu₄. Introduction of fluorine (35-37) in the ortho-, meta-, or para-positions all led to a slight drop in potency but when a cyano group (38-40) was introduced in any position a complete loss of activity was

Table	3	

Modifications to the olefin

Compound	Structure	$mGlu_{4}\left(h\right)EC_{50}\left(\mu M\right)$	% Glu max
29	N NH ₂	>30	-
30	N NH ₂	20	86
31	N N N H ₂	>30	-
32	N N N N N N N H ₂	4.9	58
33	N N N N N N H ₂	>30	-
34	N N NH ₂	>30	_

Table 4 Modifications to the phenyl ring

Compound	Ar	$mGlu_{4}\left(h\right)EC_{50}\left(\mu M\right)$	% Glu max
35	o-F Ph	2.9	101
36	<i>m</i> -F Ph	4.5	99
37	p-F Ph	4.5	106
38	o-CN Ph	>30	_
39	m-CN Ph	>30	_
40	p-CN Ph	>30	_
41	o-Me Ph	>30	_
42	<i>m</i> -Me Ph	>30	_
43	p-Me Ph	>30	_
44	2-Thienyl	4.1	98
45	2-Furyl	9.4	94
46	2-Pyridyl	11.7	92
47	3-Pyridyl	16.4	55
48	4-Pyridyl	>30	-

observed. This was also the case for the methyl analogues (41-43) perhaps indicating steric rather than electronic influences. Replacement of the phenyl ring with isosteric heteroaryl groups (44-48) provided compounds with some activity although the 4-pyridyl (**48**) was inactive up to 30μ M.

The results from our preliminary SAR evaluation of compound 11 suggested that there were limited options for optimisation of the mGlu₄ PAM potency. Whilst several compounds with at least comparable activity had been identified following small changes to the pyrimidine ring, we had observed that mGlu₄ PAM activity was lost when minor point modifications were made to the four regions of the molecule. This observation of limited SAR within

Table 5	
Selectivity profile for compound	11

Species	Receptor	MOA	EC_{50}/IC_{50} (μM)	% Glu max/% antagonism
Human	mGlu4	PAM	1.0	106
Human	mGlu ₄	Agonism ^a	>30	-
Human	mGlu ₅	NAM	>30	_
Human	mGlu ₅	PAM	>10	_
Human	mGlu ₅	Agonism ^a	>10	_
Rat	mGlu ₄	PAM	1.0	110
Rat	mGlu ₅	NAM	>10	-

MOA, mode of action.

^a mGlu assay was run in the absence of glutamate.

the field of mGlu₄ PAMs has been well documented for other chemical series. 13,15

In the absence of a significantly superior compound in terms of potency on mGlu₄ (i.e., >10-fold more potent), we elected to profile 11 in more detail (Table 5). In the rat mGlu₄ PAM in vitro assay the EC_{50} was 1 μ M which is comparable to the human assay. The human mGlu₄ assay was also run in the absence of glutamate and the EC₅₀ was >30 μ M suggesting that **11** does not act as an agonist of mGlu₄. For our selectivity profiling of **11**, we considered mGlu₅ to be the most relevant related target of interest because NAMs of mGlu₅ have been reported to show efficacy in animal models of PD.^{4,23} Hence, **11** was screened in rat and human mGlu₅ assays. In our hands, the IC₅₀ of **11** was >30 μ M on human mGlu₅ and >10 μ M on rat mGlu₅ and so the window of activity between mGlu₅ NAM and mGlu₄ PAM is estimated to be >30-fold and >10-fold in human and rat, respectively. The EC₅₀ for compound 11 in human mGlu₅ PAM or agonist assays was measured as >10 µM. Additionally, 11 was run in a receptor screening panel of 68 targets and no activity was observed at \ge 50% at 10 μ M for any of the receptors.²⁴

Compound **11** was profiled in several in vitro ADMET assays in order to determine whether it was a suitable candidate for in vivo evaluation in pre-clinical PD rodent models. Permeability was assessed in CaCo-2 cells and **11** was found to have good permeability with no apparent efflux issue (A–B 41.9×10^{-6} cm s⁻¹, B–A 19.6×10^{-6} cm s⁻¹). Additionally the plasma protein binding in rats was measured as 90% bound. The metabolic stability of **11** was assessed in rat and human microsomes and found to be 62% and 83% hepatic blood flow (%Qh) in these species.²⁵ The limited stability translated into a high in vivo clearance in rats of 75 mL/min/kg (107% Qh) and **11** had a moderate volume of distribution (2.7 L/kg) with a short mean residence time (0.6 h) when dosed at 2 mg/kg via intravenous injection. Compound **11** was orally bioavailable (*F* = 51%) and 30 min following administration of a



Figure 3. Dose dependent effect of compound 11 in a haloperidol-induced catalepsy rat model.

30 mg/kg dose, the plasma concentration was found to be 11.6 μ M. Encouragingly, **11** was highly CNS penetrant with a concentration in the brain, also at the 30 min time point, measured as \sim 33.8 μ M (brain:plasma ratio \sim 2.9). The CSF levels were measured as \sim 0.7 μ M which is in line with the predicted free fraction based on the plasma protein binding data and in the range of the in vitro mGlu₄ EC₅₀.

Although **11** suffered from limited in vitro mGlu₄ activity together with rapid in vivo clearance, we reasoned that the high CNS levels (CSF levels in the range of the in vitro EC_{50}) that were achieved in rats following oral administration together with a selectivity of >10-fold over mGlu₅ (in the rat assays) qualified **11** as a potential tool compound for an in vivo proof of concept study. We selected the haloperidol-induced catalepsy rat model as our symptomatic model for PD and we administered three doses of **11** (1, 10 and 30 mg/kg po; n = 8 per experimental group) orally 30 min prior to haloperidol (dosed at 2.5 mg/kg ip). Catalepsy was indicated by duration of time (seconds, *y*-axis) that the rat remains with its forelimbs on a bar (height 8 cm). The cut off time was 60 s.²⁶ From the results illustrated in Figure 3 compound **11** showed dose dependent efficacy with a reduction in catalepsy observed at all doses. The ED₅₀ was estimated as ~1 mg/kg.

To conclude we have identified the mGlu₄ PAM 4-((E)-styryl)pyrimidin-2-ylamine **11** from a high-throughput screen. We conducted a preliminary SAR study around **11** and we found that there was only limited scope to modify the structure and improve or indeed maintain the PAM activity on mGlu₄. In vivo pharmacokinetic profiling of **11** revealed it to be highly brain penetrant following oral administration and subsequent examination of **11** in the haloperidol-induced catalepsy model revealed a dose dependent effect thus providing further support for stimulation of mGlu₄ as a potential treatment for PD. To our knowledge compound **11** represents the first disclosure of an orally available mGlu₄ PAM displaying efficacy in a symptomatic PD animal model. Additional in vivo evaluation of the tool compound **11** will be presented in due course.

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- 19 A CHO-K1 cell line that was engineered to over-express human mGlu₄ proteins was used. Activation of mGlu4 in the respective cell line leads to a decrease of intracellular free cAMP due to coupling of these receptors to Gi-proteins and subsequent inhibition of cellular adenylate cyclases. The respective concentration of intracellular cAMP was quantified in order to assess the potency of the compounds of interest. Intracellular cAMP concentrations were determined with the commercially available HitHunter™ cAMP XS+ assay kit (DiscoverRx) with a luminescence read-out. This cAMP assay was performed according to the manufacturer's protocol with slight modifications. Briefly, on the day of the assay culture medium was removed and the cells washed once with Dulbecco's phosphate buffered saline (PBS, PAA). Subsequently, all wash solution was removed and 15 µL of the test compound at the required concentration in an assay 'specific compound solution' (see below) was added to each well of the 384-well assay plate. After incubation for 30 min at 37 °C, 5 µL of XS+ Antibody Reagent and 20 µL of a freshly prepared mixture of XS+ ED/LYSIS/CL Substrate Working Solution were added (all kit components) resulting in a volume of 40 µL. This was followed by an incubation of the assay plate for 1 h at room temperature. Subsequently, 20 µL of XS+ EA Reagent (kit component) were added to each well followed by an incubation of 3 h at room temperature. Finally, luminescence was read on a standard luminescence reader (Novostar, BMG Labtech). In case the potential to positively modulate mGlu₄ (positive modulator assay) was assayed the 'specific compound solution' consisted of compound at the concentration of interest, 5 µM Forskolin and 15 µM Glutamate (equivalent to EC20 in this assay) in PBS. In case the pure stimulation of mGlu₄ was assayed (agonist assay) the 'specific compound solution' consisted of compound at the concentration of interest and 5 µM Forskolin in PBS. Test compounds and positive controls were assayed for dose-response at eight concentrations in triplicate. Serial dilutions in 100% DMSO were made (at 100-fold of final concentration) and starting and final concentrations covered a concentration range of three log units. In order to generate the required assay concentrations in the 'specific compound solutions' the 100% DMSO solutions were diluted in two steps down to 1% DMSO. A valid EC₅₀ value was calculated from a minimum of three independent compound dilutions. Dose-response curves were evaluated according to a 'one binding site' equation in XLfit 4.1 software (Model 204;

 $y = A + (B - A)/(1 + (10^{((C - x) * D))}; C \rightarrow \log(EC_{50})$ value; $D \rightarrow \text{slope factor})$ and values for EC_{50} and Hill slope were calculated. Parameters A (top) and B (bottom) were fixed.

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