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Influence of the C-5 substitution in polysubstituted pyrimidines on inhibition of prostaglandin E_2 production

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Abstract: As a part of a broader structure-activity relationship study of substituted 2aminopyrimidines the influence of the C-5 substitution on inhibition of prostaglandin E_2 (PGE₂) production was studied. Thirty compounds were prepared starting from the corresponding 2amino-4,6-dichloropyrimidines using Suzuki cross-coupling. It was shown previously that 2amino-4,6-dichloropyrimidines with smaller C-5 substituent (hydrogen and methyl) were devoid of significant activity, while 5-butyl derivatives exhibited prominent potency. In this study, on the other hand, both monoaryl- and bisarylpyrimidines were potent inhibitors of PGE₂ production regardless the length of the C-5 substituent (hydrogen, methyl, n-butyl). Moreover, the shorter the C-5 substituent the higher potency to inhibit PGE₂ production was observed. 2-Amino-4,6-diphenylpyrimidine was the best inhibitor of PGE₂ production with IC₅₀ = 3 nM and no cytotoxicity. The most potent inhibitors deserve further preclinical evaluation as potential anti-inflammatory agents.

Keywords: pyrimidines; Suzuki-Miyaura reaction; prostaglandin E2; inhibitor

1. Introduction

Prostaglandins (PGs) are lipid mediators and end products of fatty acid metabolism, that are produced from the key precursor, arachidonic acid (AA), *via* the cyclooxygenase (COX)

pathway: either by constitutively expressed COX-1 or by, in response to cell-specific stimuli, highly inducible COX-2.¹⁻⁴

Prostaglandin E_2 (PGE₂) is a naturally occurring eicosanoid derived from AA, produced by an action of COX enzymes and prostaglandin E synthases (PGES).^{5,6} PGE₂, as a lipid mediator, is known to regulate various important processes including inflammation, pain and pyrexia.⁷ Nonsteroidal anti-inflammatory drugs (NSAIDs)^{1,8-10} are agents that, by inhibiting the activity of COX enzymes (and thus inhibiting PGs production), efficiently reduce pain and fever, prevent blood clots, and at higher doses, decrease inflammation. The traditional (non-selective) NSAIDs belong to the most widely used drugs around the world, but they carry the risk of serious gastrointestinal (GI), as well as cardiovascular (CV) and renal adverse effects.⁸⁻¹⁰ In search for safer medication, COX-2 selective inhibitors (coxibs)^{11,12} were developed with reduced upper GI toxicity, nevertheless still exhibiting CV adverse events.^{10,13} Thus, there is a persisting need for development of novel PGE₂ production inhibitors lacking the negative side effects of currently used therapeutics.

A pyrimidine moiety represents one of the most important medicinal chemistry scaffolds.^{14,15} Numerous compounds based on the pyrimidine scaffold are known to exhibit anti-inflammatory activity,¹⁶⁻¹⁹ as well as properties immunomodulating,²⁰ anti-leishmanial,²¹ antibacterial,²²⁻²⁴ antifungal,²³ analgesic,²⁵ anticonvulsant,²⁶ and anticancer.²⁷⁻³¹

Within previous extensive structure-activity relationship studies, we discovered a series of polysubstituted pyrimidines as potent inhibitors of immune-activated nitric oxide (NO) production,³²⁻³⁶ as well as inhibitors of production of prostaglandin E_2 (PGE₂).³⁴⁻³⁶ It has been demonstrated, that in the case of 2-amino-4,6-dichloropyrimidines, at least a 3-carbon-long aliphatic group in the C-5 position of the pyrimidine moiety was required to inhibit PGE₂ production substantially, while derivatives bearing either hydrogen or methyl in the C-5 position were not PGE₂ production inhibitors.³⁴ Based on these results novel series of compounds was prepared, 2-amino-4,6-di(hetero)arylpyrimidines having a *n*-butyl group in the C-5 position, as an optimal substituent.³⁶ Several compounds in this series were found to be low micromolar dual inhibitors of NO and PGE₂ production.³⁶ Since non-toxic small-molecule inhibitors of PGE₂ production may have great therapeutic potential in treatment of serious inflammatory diseases, we decided to verify the influence of the C-5 substitution in the series of selected 2-amino-4, aryl-6-chloropyrimidines and 2-amino-4, 6-diarylpyrimidines.

In this work, we prepared novel types of symmetrical and unsymmetrical 2-amino-4,6diarylpyrimidines (where aryl is either phenyl, *p*-tolyl or 4-methoxylphenyl) bearing in the C-5 position either hydrogen, methyl or *n*-butyl group. The prepared compounds were subsequently tested for their ability to inhibit PGE_2 production in mouse peritoneal cells.

2. Results and discussion

2.1. Synthesis

Starting 2-amino-4,6-dichloropyrimidines **1a-1c** (Scheme 1) were prepared according to the previously published procedure.³² Compounds **1a-1c** were subsequently treated with commercially available arylboronic acids (phenyl-, *p*-tolyl-, and 4-methoxyphenylboronic acid) under Suzuki-Miyaura cross-coupling conditions.³⁷ Depending on the reaction conditions,³⁶ we were able to prepare both monoarylated products **2-4** (Method A: Pd(PPh₃)₄, Na₂CO₃, toluene/ethanol (3:1), 80 °C, 12 h) or symmetrical diarylated products **5-7** (Method B: Pd(PPh₃)₄, Cs₂CO₃, dioxane/water (4:1), 100 °C, 12 h).

Treatment of 2-amino-4-aryl-6-chloropyrimidines **2-4** with another arylboronic acid using the latter conditions (Method B) afforded unsymmetrical 4,6-diarylpyrimidine analogues **8-10** (Scheme 1). Structures of all prepared compounds are summarized in Table 1.



Scheme 1. General synthesis of target polysubstituted pyrimidines. Ar^1 , Ar^2 – phenyl, *p*-tolyl, 4methoxyphenyl; R – hydrogen, methyl, *n*-butyl. Reaction conditions: a) arylboronic acid, Pd(PPh₃)₄, Na₂CO₃, toluene/ethanol (3:1), 80 °C, 12 h; b) arylboronic acid, Pd(PPh₃)₄, Cs₂CO₃, dioxane/water (4:1), 100 °C, 12 h.

Table 1. List of the prepared compounds. Me, methyl; Bu, *n*-butyl; Ph, phenyl; Tol, *p*-tolyl;MeOPh, 4-methoxyphenyl.

Compound	R	Ar^1	Ar^2
1a	Н	-	-
1b	Me	-	
1c	Bu	-	\geq
2a	Н	Ph	
2b	Me	Ph	-
2c	Bu	Ph	-
3a	Н	Tol	-
3b	Me	Tol	-
3c	Bu	Tol	-
4a	Н	MeOPh	-
4b	Me	MeOPh	-
4c	Bu	MeOPh	-
5a	Н	Ph	-

5b	Me	Ph	-
5c	Bu	Ph	-
6a	Н	Tol	-
6b	Me	Tol	-
6c	Bu	Tol	-
7a	Н	MeOPh	-
7b	Me	MeOPh	-
7c	Bu	MeOPh	-
8a	Н	Tol	Ph
8b	Me	Tol	Ph
8c	Bu	Tol	Ph
9a	Н	MeOPh	Ph
9b	Me	MeOPh	Ph
9c	Bu	MeOPh	Ph
10a	Н	MeOPh	Tol
10b	Me	MeOPh	Tol
10c	Bu	MeOPh	Tol

2.2. Biological evaluation

All prepared polysubstituted pyrimidines were evaluated for their ability to inhibit production of PGE₂ *in vitro*, using C57BL6 mouse peritoneal cells. High output PGE₂ production was activated by lipopolysaccharide (LPS) and assayed at the interval of 5 h of culture. Compounds (as 50 μ M solutions) were applied concomitantly with LPS. The effects of pyrimidines were expressed as a percent change related to the response of LPS-stimulated control cells. With the exception of 2-amino-4,6-dichloropyrimidine analogues bearing hydrogen or methyl at C-5 position (**1a** and **1b**, respectively), all tested compounds were discovered to be potent inhibitors of PGE₂ production (Fig. 1A). The extent of the inhibition of PGE₂ production clearly depended on the character of the C-5 substituent. Contrary to the previously studied 2-amino-4,6dichloropyrimidine analogues,³² the current series of 2-amino-4-arylpyrimidines and 2-amino-4,6-diarylpyrimidines no longer required prolonged aliphatic chain in the C-5 position to exert their biological activity. The most potent pyrimidines were those bearing hydrogen at C-5 position (i.e. compounds **2a**, **3a**, **4a**, **5a**, **6a**, **7a**, **8a**, **9a**, and **10a**), while prolongation of the substituent in this position led to attenuation of the capability to reduce PGE_2 production (Fig. 1B). Namely, analogues bearing methyl in the C-5 position are statistically less efficient to inhibit PGE_2 production compared to those bearing hydrogen at that position and the compounds bearing *n*-butyl group in the C-5 position are even less effective in comparison to the 5-methyl derivatives (Fig. 1B).

For the selected potent compounds, the IC_{50} values were also determined in comparison with two commonly used NSAIDs as control compounds, indomethacin³⁸⁻⁴⁰ and aspirin^{40,41} (Table 2). The compounds were found to be potent at low micromolar to submicromolar levels. The most effective inhibitors of PGE₂ production were compounds **5a**, **6a**, **8a**, and **9a** with IC_{50} s in the range of 3-33 nM (Table 2). Again, all of them were the C-5 unsubstituted analogues, i.e. derivatives with hydrogen at the C-5 position.

Most importantly, the high inhibitory activity of the studied compounds cannot be explained as a result of possible cytotoxic effects, as no changes in viability of cells could be detected in the presence of the compounds (Table 2). Cell death was analyzed in the same cells using the lactate dehydrogenase (LDH) assay which is based on the determination of LDH activity released from the cytosol of damaged cells into the supernatant.



Figure 1. Effects of individual pyrimidine derivatives (50 μ M solutions) (A) and medians of grouped compounds according to the C-5 substituents (B) on *in vitro* production of PGE₂ in mouse peritoneal cells. Bars are means ± S.E.M. that were obtained by averaging results of two to four experiments for each compound.

Previously reported data³⁶ suggested that post-translation mechanism(s) are plausible explanation for the PGE₂-inhibitory action of the pyrimidine derivatives. Their interaction with activity of enzymes involved in biosynthesis of PGE₂ is under current investigation.

Table 2. Concentration of selected pyrimidines inhibiting PGE_2 production in C57BL6 mouse peritoneal cells by 50% (IC₅₀), and effects on cell viability of the same cell type evaluated as a percent change against untreated control cells.

Compound	IC ₅₀ (µM)	Viability (%)
1c	5.34 (4.65-6.12) ^a	99.82±1.33 ^b
3a	2.32 (1.46-3.42)	102.6±0.4
3c	1.86 (1.27-2.74)	101.4±1.7
4a	2.79 (1.98-3.92)	103.2±0.7
4c	2.16 (1.87-2.49)	96.6±1.8
5a	0.003 (0.002-0.004)	97.4±0.9
ба	0.033 (0.024-0.046)	102.8±1.2
7a	0.119 (0.068-0.202)	101.7±0.7
7c	3.71 (2.81-4.90)	99.1±0.6
8a	0.009 (0.007-0.01)	103.1±0.7
8c	2.46 (1.91-3.16)	102.2±0.6
9a	0.009 (0.005-0.016)	99.9±1.7
9c	3.40 (2.55-4.55)	102.0±0.9
indomethacin	0.005 (0.003-0.007)	n.d. ^c
aspirin	4.08 (3.09-5.39)	n.d.
<mark>DMSO</mark>	n.d.	<mark>98.9±1.6</mark>

^a95% limits of confidence; ^b \pm S.E.M. Results represent means of two identical experiments. ^cn.d. = not determined.

3. Conclusion

In summary, we have identified novel potent and non-toxic inhibitors of prostaglandin E_2 (PGE₂) production using C57BL6 mouse peritoneal cells. The selected monoaryl- and diarylsubstituted 2-aminopyrimidines exhibited single-digit micromolar to submicromolar potency to inhibit PGE₂ production. By contrast to the previously studied 2-amino-4,6-dichloropyrimidines, herein reported arylpyrimidines were most potent inhibitors when bearing hydrogen atom in the C-5 position. At least twelve of the prepared compounds were more potent than still popular aspirin. 2-Amino-4,6-diphenylpyrimidine was the most potent inhibitor in the whole series, with IC₅₀ = 3 nM and no observed cytotoxicity. This compound was shown to be

equipotent to indomethacin, a nonsteroidal anti-inflammatory drug commonly used as a prescription medication to treat inflammation symptoms. Indomethacin, as an inhibitor of both COX-1 and COX-2, nevertheless, exhibits many serious adverse effects and superior drugs are needed. Although the exact mode of action of the reported compounds remains to be elucidated, the preliminary findings encourage further preclinical studies of the selected candidates as potential anti-inflammatory agents.

4. Experimental section

4.1. Chemistry

Unless otherwise stated, solvents were evaporated at 40 °C/2 kPa and the compounds were dried over P2O5 at 2 kPa. ¹H NMR and ¹³C NMR were recorded using Bruker Avance 400 MHz spectrometer operating at 9.39 T in DMSO-d6 solution. The ¹H and ¹³C spectra were acquired at 400.00 MHz and 100 MHz, respectively. Chemical shifts (δ) are reported in ppm and interaction constants (J) in Hz. The spectra were referenced to tetramethylsilane (TMS) or to the residual solvent signal. High-resolution mass spectra of the products were obtained using GCT Premier (Waters, USA) spectrometer for electron ionization (EI) and LTQ Orbitrap XL (Thermo Fisher Scientific) spectrometer for electrospray ionization (ESI). Starting 2-amino-4,6dichloropyrimidines were prepared according to the published procedure.³² Purity of the prepared compounds was \geq 95%; the purity was determined by the combination of HPLC (H₂O/CH₃CN,linear gradient), NMR, and HR-MS.

4.1.1. Method A. General procedure for synthesis of 2-amino-4-aryl-6-chloropyrimidines 2a-c, 3a-c, 4a-c. Starting 2-amino-4,6-dichloropyrimidine derivative (1.36 mmol), aryl boronic acid (1.51 mmol), tetrakis(triphenylphosphine) palladium (58 mg, 0.05 mmol) and sodium carbonate (213 mg, 2.01 mmol) were suspended in a toluene/ethanol mixture (3:1, 50 mL). The reaction was heated to 80 °C for 12 h. After cooling to room temperature, the solid was filtered off and the solvents were removed *in vacuo*. The residue was dissolved in ethyl acetate (100 mL) and adsorbed on silica gel. After removal of organic solvent *in vacuo* the product was purified on silica gel column chromatography using a hexane/ethyl acetate mixture (4:1) as the eluent and

subsequently on C-18 reverse chromatography column using water/methanol eluent (0-100% gradient).

4.1.1.1. 4-Chloro-6-phenylpyrimidin-2-amine (2a). Yield 76%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 8.13 – 8.08 (m, 2H), 7.57 – 7.48 (m, 3H), 7.26 (s, 1H), 7.19 (s, 2H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) δ 166.45, 163.99, 161.60, 136.33, 131.57, 129.22, 127.52, 105.15. HRMS (EI) m/z [M]⁺ calcd for C₁₀H₈ClN₃: 205.0407, found: 205.0406.

4.1.1.2. 4-Chloro-5-methyl-6-phenylpyrimidin-2-amine (2b). Yield 50%, white solid. ¹H NMR δ _H (400 MHz, DMSO-*d*₆) 7.56 – 7.42 (5 H, m), 6.89 (2 H, s), 2.11 (3 H, s). ¹³C NMR δ _C (100 MHz, DMSO-*d*₆) 168.27, 161.88, 161.51, 138.74, 129.49, 129.04, 128.52, 113.57, 15.70. **HRMS** (EI) *m*/*z* [M]⁺ calcd for C₁₁H₁₀ClN₃: 219.0563, found: 219.0566.

4.1.1.3. 5-Butyl-4-chloro-6-phenylpyrimidin-2-amine (2c). Yield 50%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.53 – 7.45 (3 H, m), 7.44 – 7.40 (2 H, m), 6.93 (2 H, s), 2.49 – 2.42 (2 H, m), 1.42 – 1.28 (2 H, m), 1.14 (2 H, q, J = 7.2 Hz), 0.72 (3 H, t, J = 7.3 Hz). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 169.08, 161.56, 161.41, 139.00, 129.18, 128.54, 128.42, 118.41, 31.64, 27.96, 22.28, 13.82. **HRMS** (EI) m/z [M]⁺ calcd for C₁₄H₁₆ClN₃: 261.1033, found: 216.1031.

4.1.1.4. 4-Chloro-6-(*p*-tolyl)pyrimidin-2-amine (3a). Yield 41%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-*d*₆) 8.01 (d, *J* = 8.3 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 7.22 (s, 1H), 7.14 (s, 2H), 2.37 (s, 3H). ¹³C NMR NMR $\delta_{\rm C}$ (100 MHz, DMSO-*d*₆) 166.36, 163.96, 161.50, 141.54, 133.56, 129.81, 127.47, 104.77, 21.44. HRMS (EI) *m*/*z* [M]⁺ calcd for C₁₁H₁₀ClN₃: 219.0563, found: 219.0565.

4.1.1.5. 4-Chloro-5-methyl-6-(p-tolyl)pyrimidin-2-amine (3b). Yield 32%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.43 (2 H, d, J = 8.2 Hz), 7.38 – 7.20 (2 H, m), 6.87 (2 H, s), 2.37 (3 H, s), 2.12 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 168.19, 161.50, 161.22, 161.01, 139.07, 135.90, 129.10, 113.48, 21.38, 15.80. HRMS (ESI) m/z [M+H]⁺ calcd for C₁₂H₁₃ClN₃: 234.0793, found: 234.0793.

4.1.1.6. 5-Butyl-4-chloro-6-(p-tolyl)pyrimidin-2-amine (3c). Yield 7%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.33 (2 H, d, J = 8.2 Hz), 7.28 (2 H, d, J = 7.9 Hz), 6.91 (2 H, s), 2.49 – 2.43 (2 H, m), 2.37 (3 H, s), 1.46 – 1.29 (2 H, m), 1.23 – 1.08 (2 H, m), 0.75 (3 H, t, J = 7.3 Hz). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 169.00, 161.57, 161.39, 138.65, 136.18, 129.06, 128.46, 118.38, 31.69, 28.06, 22.33, 21.35, 13.89. **HRMS** (ESI) m/z [M+H]⁺ calcd for C₁₅H₁₉ClN₃: 276.1262, found: 276.1262.

4.1.1.7. 4-Chloro-6-(4-methoxyphenyl)pyrimidin-2-amine (4a). Yield 34%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 8.11 – 8.06 (m, 2H), 7.20 (s, 1H), 7.09 (s, 2H, NH₂), 7.08 – 7.02 (m, 2H), 3.84 (s, 3H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 165.98, 163.89, 162.17, 161.35, 129.21, 128.58, 114.55, 104.25, 55.84. HRMS (EI) m/z [M]⁺ calcd for C₁₁H₁₀ClN₃O: 235.0512, found: 235.0514.

4.1.1.8. 4-Chloro-6-(4-methoxyphenyl)-5-methylpyrimidin-2-amine (4b). Yield 44%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.51 (2 H, d, J = 8.8), 7.03 (2 H, d, J = 8.8), 6.84 (2 H, s), 3.82 (3 H, s), 2.15 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 167.76, 161.86, 161.46, 160.35, 130.95, 130.84, 113.86, 113.30, 55.70, 15.94. HRMS (EI) m/z [M]⁺ calcd for C₁₂H₁₂ClN₃O: 249.0669, found: 249.0673.

4.1.1.9. 5-Butyl-4-chloro-6-(4-methoxyphenyl)pyrimidin-2-amine (4c). Yield 52%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.51 (2 H, d, J = 8.9), 7.12 (2 H, d, J = 8.9), 6.99 (2 H, s), 3.91 (3 H, s), 2.66 – 2.53 (2 H, m), 1.56 – 1.39 (2 H, m), 1.34 – 1.20 (2 H, m), 0.86 (3 H, t, J7.3). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 168.58, 161.60, 161.38, 160.08, 131.29, 130.12, 118.30, 113.89, 55.66, 31.69, 28.11, 22.33, 13.90. **HRMS** (EI) m/z [M]⁺ calcd for C₁₅H₁₈ClN₃O: 291.1138, found: 291.1136.

4.1.2. Method B. General procedure for synthesis of symmetrical 4,6-diarylpyrimidines 5a-c, 6ac, 7a-c. Starting 2-amino-4,6-dichloropyrimidine derivative (1.36 mmol), boronic acid (2.05 mmol), tetrakis(triphenylphosphine)palladium (58 mg, 0.05 mmol) and cesium carbonate (213 mg, 2.01 mmol) were dissolved in a dioxane/water mixture (4:1, 50 mL). The reaction was heated to 100 $^{\circ}$ C for 12 h. After cooling to room temperature the solvents were removed *in* *vacuo*. The residue was dissolved in ethyl acetate (100 mL) and adsorbed on silica gel. After removal of the organic solvent *in vacuo* the product was purified on silica gel column chromatography using a hexane/ethyl acetate mixture (4:1) as the eluent and subsequently on C-18 reverse chromatography column using water/methanol eluent (0-100% gradient).

4.1.2.1. 4,6-Diphenylpyrimidin-2-amine (5a). Yield 81%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 8.26 – 8.19 (m, 4H), 7.71 (s, 1H), 7.53 (p, J = 3.8, 3.3 Hz, 6H), 6.76 (s, 2H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) δ 165.32, 164.48, 137.81, 130.90, 129.07, 127.44, 102.30. HRMS (EI) m/z [M]⁺ calcd for C₁₆H₁₃N₃: 247.1109, found: 247.1108.

4.1.2.2. 5-*Methyl*-4,6-*diphenylpyrimidin*-2-*amine* (5*b*). Yield 44%, white solid. ¹**H** NMR $\delta_{\rm H}$ (400 MHz, DMSO-*d*₆) 7.63 – 7.54 (4 H, m), 7.52 – 7.39 (6 H, m), 6.51 (2 H, s), 2.04 (3 H, s). ¹³**C** NMR $\delta_{\rm C}$ (100 MHz, DMSO-*d*₆) 167.54, 161.90, 139.73, 129.20, 129.04, 128.43, 113.28, 16.72. **HRMS** (EI) *m*/*z* [M]⁺ calcd for C₁₇H₁₅N₃: 261.1266, found: 261.1269.

4.1.2.3. 5-Butyl-4,6-diphenylpyrimidin-2-amine (5c). Yield 69%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.55 – 7.39 (10 H, m), 6.52 (2 H, s), 2.49 – 2.42 (2 H, m), 1.02 – 0.91 (2 H, m), 0.88 – 0.76 (2 H, m), 0.45 (3 H, t, J = 7.3). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 168.08, 161.49, 158.68, 140.10, 128.70, 128.62, 128.45, 118.60, 32.24, 26.92, 22.02, 13.55. HRMS (EI) m/z [M]⁺ calcd for C₂₀H₂₁N₃: 303.1735, found: 303.1740.

4.1.2.4. 4,6-*Di-p-tolylpyrimidin-2-amine* (6*a*). Yield 37%, white solid. ¹**H** NMR $\delta_{\rm H}$ (400 MHz, DMSO-*d*₆) 8.13 (4H, d, *J* = 8.2 Hz), 7.65 (1 H, s), 7.33 (4H, d, *J* = 7.9 Hz), 6.67 (2H, s), 2.39 (s, 6H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-*d*₆) 165.06, 164.40, 140.61, 135.07, 129.65, 127.34, 101.57, 21.43. **HRMS** (EI) *m*/*z* [M]⁺ calcd for C₁₈H₁₇N₃: 275.1422, found: 275.1420.

4.1.2.5. 5-Methyl-4,6-di-p-tolylpyrimidin-2-amine (6b). Yield 40%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.49 (4 H, d, J = 7.8), 7.28 (4 H, d, J = 7.8), 6.43 (2 H, s), 2.37 (6 H, s), 2.05 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 167.39, 161.89, 138.46, 136.93, 129.26, 128.95, 115.46, 113.22, 21.37, 16.99. **HRMS** (EI) m/z [M]⁺ calcd for C₁₉H₁₉N₃: 289.1579, found: 289.1574.

4.1.2.6. 5-Butyl-4,6-di-p-tolylpyrimidin-2-amine (6c). Yield 49%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-*d*₆) 7.38 (4 H, d, *J* 8.2), 7.27 (4 H, d, *J* = 7.7), 6.45 (2 H, s), 2.54 – 2.46 (2 H, m), 2.37 (6 H, s), 1.03 – 0.89 (2 H, m), 0.89 – 0.77 (2 H, m), 0.47 (3 H, t, *J* = 7.3). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-*d*₆) 167.96, 161.48, 137.97, 137.38, 128.97, 128.65, 118.61, 32.22, 27.03, 21.36, 13.66. $\delta_{\rm C}$ (101 MHz, DMSO-*d*₆) 167.96, 161.48, 153.31, 137.98, 137.38, 128.96, 128.65, 118.63, 32.22, 27.03, 22.01, 21.35, 13.66. HRMS (ESI) *m*/*z* [M+H]⁺calcd for C₂₂H₂₆N₃: 332.2121, found: 332.2122.

4.1.2.7. 4,6-*Bis*(4-methoxyphenyl)pyrimidin-2-amine (7a). Yield 25%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d6) 8.22 – 8.17 (m, 4H), 7.60 (s, 1H), 7.09 – 7.04 (m, 4H), 6.58 (s, 2H), 3.85 (s, 6H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d6) 164.55, 164.27, 161.59, 130.23, 128.94, 114.35, 100.71, 55.78. HRMS (EI) *m*/*z* [M]⁺ calcd for C₂₂H₁₇N₃O₂: 307.1321, found: 307.1320.

4.1.2.8. 4,6-Bis(4-methoxyphenyl)-5-methylpyrimidin-2-amine (7b). Yield 33%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.57 (4 H, d, J = 9.0), 7.03 (4 H, d, J = 9.0), 6.38 (2 H, s), 3.82 (6 H, s), 2.10 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 166.94, 161.84, 160.00, 132.06, 130.93, 113.73, 113.00, 55.66, 17.36. HRMS (ESI) m/z [M+H]⁺ calcd for C₁₉H₂₀N₃O₂: 322.1550, found: 322.1551.

4.1.2.9. 5-Butyl-4,6-bis(4-methoxyphenyl)pyrimidin-2-amine (7c). Yield 14%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.45 (4 H, d, J = 8.6), 7.02 (4 H, d, J = 8.8), 6.41 (2 H, s), 3.81 (6 H, s), 2.61 – 2.44 (2 H, m), 1.02 – 0.91 (2 H, m), 0.91 – 0.80 (2 H, m), 0.49 (3 H, t, J = 7.2). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 167.57, 161.48, 159.66, 132.60, 130.22, 118.57, 113.78, 55.61, 32.14, 27.14, 22.02, 13.71. HRMS (ESI) m/z [M+H]⁺ calcd for C₂₂H₂₆N₃O₂: 364.2020, found: 364.2019.

4.1.3. Method C. General procedure for synthesis of unsymmetrical 4,6-diarylpyrimidines 8a-c, 9a-c, 10a-c. Starting 2-amino-4-aryl-6-chloropyrimidine derivative (1.36 mmol), boronic acid (1.51 mmol), tetrakis(triphenylphosphine)palladium (58 mg, 0.05 mmol) and cesium carbonate (213 mg, 2.01 mmol) were dissolved in a dioxane/water mixture (4:1, 50 mL). The reaction was

heated to 100 °C for 12 h. After cooling to room temperature the solvents were removed *in vacuo*. The residue was dissolved in ethyl acetate (100 mL) and adsorbed on silica gel. After removal of the organic solvent *in vacuo* the product was purified on silica gel column chromatography using a hexane/ethyl acetate mixture (4:1) as the eluent and subsequently on C-18 reverse chromatography column using water/methanol eluent (0-100% gradient).

4.1.3.1. 4-Phenyl-6-(*p*-tolyl)pyrimidin-2-amine (8a). Yield 75%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-*d*₆) 8.24 – 8.19 (m, 2H), 8.14 (d, *J* = 8.2 Hz, 2H), 7.68 (s, 1H), 7.55 – 7.50 (m, 3H), 7.33 (d, *J* = 7.7 Hz, 2H), 6.71 (s, 2H), 2.39 (s, 3H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-*d*₆) 165.21, 165.17, 164.44, 140.69, 137.88, 135.01, 130.83, 129.67, 129.05, 127.41, 127.37, 101.93, 21.44. HRMS (EI) *m*/*z* [M]⁺ calcd for C₁₇H₁₅N₃: 261.1266, found: 261.1269.

4.1.3.2. 5-Methyl-4-phenyl-6-(p-tolyl)pyrimidin-2-amine (8b). Yield 59%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.57 (2 H, d, J = 6.1), 7.52 – 7.42 (5 H, m), 7.29 (2 H, d, J = 8.1), 6.47 (2 H, s), 2.37 (3 H, s), 2.04 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 167. 52, 167.47, 161.89, 139.80, 138.51, 136.86, 129.25, 129.20, 129.01, 128.96, 128.41, 113.25, 21.38, 16.86. HRMS (EI) m/z [M]⁺ calcd for C₁₈H₁₇N₃: 275.1422, found: 275.1425.

4.1.3.3. 5-Butyl-4-phenyl-6-(p-tolyl)pyrimidin-2-amine (8c). Yield 52%, white solid. ¹H NMR δ _H (400 MHz, DMSO-d₆) 7.53 – 7.42 (5 H, m), 7.39 (2 H, d, J = 8.1), 7.27 (2 H, d, J = 7.8), 6.49 (2 H, s), 2.49 – 2.45 (2 H, m), 2.37 (3H, s), 1.02 – 0.89 (2 H, m), 0.89 – 0.74 (2 H, m), 0.46 (3 H, t, J = 7.2). ¹³C NMR δ _C (100 MHz, DMSO-d₆) 168.05, 167.99, 161.49, 140.18, 138.03, 137.29, 130.14, 128.98, 128.65, 128.43, 118.61, 115.46, 32.24, 26.98, 22.02, 21.35, 13.60. HRMS (EI) m/z [M]⁺ calcd for C₂₁H₂₃N₃: 317.1892, found: 317.1895.

4.1.3.4. 4-(4-Methoxyphenyl)-6-phenylpyrimidin-2-amine (9a). Yield 47%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d6) 8.23 – 8.18 (m, 4H), 7.66 (s, 1H), 7.55 – 7.47 (m, 3H), 7.09 – 7.03 (m, 2H), 6.68 (s, 2H), 3.84 (s, 3H). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d6) 164.83, 164.68, 161.53, 138.26, 130.57, 130.42, 128.95, 128.92, 127.33, 114.31, 100.21, 55.77. HRMS (EI) m/z [M]⁺ calcd for C₁₇H₁₅N₃O: 277.1215, found: 277.1217.

4.1.3.5. 4-(4-Methoxyphenyl)-5-methyl-6-phenylpyrimidin-2-amine (9b). Yield 49%, white solid.¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.62 – 7.54 (4 H, m), 7.51 – 7.41 (3 H, m), 7.03 (2 H, d, J = 8.8), 6.45 (2 H, s), 3.82 (3 H, s), 2.07 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 167.51, 166.98, 161.87, 160.03, 139.86, 131.92, 130.91, 129.23, 128.98, 128.40, 113.76, 113.14, 55.66, 17.04. HRMS (ESI) m/z [M+H]⁺ calcd for C₁₈H₁₈N₃O: 292.1444, found: 292.1446.

4.1.3.6. 5-Butyl-4-(4-methoxyphenyl)-6-phenylpyrimidin-2-amine (9c). Yield 92%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 7.54 – 7.37 (7 H, m), 7.02 (2 H, d, J = 8.8), 6.48 (2 H, s), 3.81 (3 H, s), 2.57 – 2.46 (2 H, m), 1.07 – 0.92 (2 H, m), 0.90 – 0.74 (2 H, m), 0.47 (3 H, t, J = 7.2). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO-d₆) 168.07, 167.58, 161.49, 159.70, 140.26, 132.43, 130.20, 128.64, 128.42, 118.58, 113.80, 55.61, 32.20, 27.03, 22.02, 13.63. HRMS (ESI) m/z [M+H]⁺ calcd for C₂₁H₂₄N₃O: 334.1914, found: 334.1914.

4.1.3.7. 4-(4-Methoxyphenyl)-6-(p-tolyl)pyrimidin-2-amine (10a). Yield 89%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 8.20 (2 H, d, J = 9.0), 8.12 (2 H, d, J = 8.3), 7.62 (1 H, s), 7.33 (2 H, d, J = 7.8), 7.06 (2 H, d, J = 9.0), 6.61 (2 H, s), 3.85 (3 H, s), 2.39 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 164.90, 164.73, 164.34, 161.65, 140.53, 135.15, 130.16, 129.63, 128.98, 127.32, 114.38, 101.16, 55.79, 21.43. HRMS (ESI) m/z [M+H]⁺ calcd for C₁₈H₁₈N₃O: 292.1444, found: 292.1445.

4.1.3.8. 4-(4-Methoxyphenyl)-5-methyl-6-(p-tolyl)pyrimidin-2-amine (10b). Yield 43%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.57 (2 H, d, J = 8.8), 7.49 (2 H, d, J = 8.3), 7.29 (2 H, d, J = 7.8), 7.03 (2 H, d, J = 8.8), 6.40 (2 H, s), 3.82 (3 H, s), 2.38 (3 H, s), 2.08 (3 H, s). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO- d_6) 167.41, 166.93, 161.86, 160.01, 138.45, 137.00, 132.00, 130.91, 129.28, 128.94, 113.74, 113.11, 55.66, 21.38, 17.18. HRMS (ESI) m/z [M+H]⁺ calcd for C₁₉H₂₀N₃O: 306.1601, found: 306.1602.

4.1.3.9. 5-Butyl-4-(4-methoxyphenyl)-6-(p-tolyl)pyrimidin-2-amine (10c). Yield 36%, white solid. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO- d_6) 7.45 (2 H, d, J = 8.8), 7.38 (2 H, d, J = 8.3), 7.27 (2 H, d, J = 7.6), 7.02 (2 H, d, J = 9.0), 6.42 (2 H, s), 3.81 (3 H, s), 2.56 – 2.48 (2 H, m), 2.37 (3 H, s), 1.02 – 0.91 (2 H, m), 0.90 – 0.79 (2 H, m), 0.49 (3 H, t, J = 7.2). ¹³C NMR $\delta_{\rm C}$ (100 MHz,

DMSO- d_6) 166.75, 161.49, 159.69, 159.25, 137.96, 137.45, 132.54, 130.21, 128.96, 128.67, 118.60, 113.78, 55.62, 32.17, 27.08, 22.01, 21.35, 13.68. **HRMS** (ESI) m/z [M+H]⁺ calcd for C₂₂H₂₆N₃O: 348.2070, found: 348.2071.

4.2. Biology

Stock solutions (200 mM) of pyrimidine analogues were prepared in tissue-culture tested dimethylsulfoxide (DMSO, Sigma-Aldrich). Required working solutions were freshly obtained by the dilution in complete RPMI-1640 culture medium. It contained 10% heat-inactivated foetal bovine serum, 2 mM L-glutamine, 50 μ g/ml gentamicin, and 5 x 10⁻⁵ M 2-mercaptoethanol (all Sigma-Aldrich). Effects of studied pyrimidine analogues on production of prostaglandin E₂ (PGE₂) were evaluated *in vitro*, using mouse (C57BL6, Charles River Deutschland, Sulzfeld, Germany) peritoneal cells. Animals, killed by cervical dislocation, were i.p. injected with 8 ml of sterile saline. Pooled peritoneal cells collected from mice (n = 5-8 in individual experiments) were washed, re-suspended in complete culture medium, and seeded into 96-well round-bottom microplates (Costar) in 100- μ l volumes, 2 x 10⁵ cells/well. Cultures were maintained at 37 °C, 5% CO₂ in humidified incubator (Sanyo Electric Biomedical, Osaka, Japan). Possible effects of the vehiculum (DMSO) were evaluated using the 0.025% dilution corresponding to the highest molarity of tested pyrimidines (i.e. 50 μ M). This amount of DMSO was found to be devoid of any interference with the PGE₂ and viability assays (see Fig. 1A, and Table 2, respectively). Experimental protocols were approved by the institutional ethics committee.

4.2.1. PGE_2 and cytotoxicity assays in mouse C57BL6 peritoneal cells. The cells were cultured in presence of test compounds applied concomitantly with lipopolysaccharide (LPS from *E. coli* 0111:B4, Sigma; 10 ng/ml). Amount of PGE₂ was determined at the interval of 5 h of culture using ELISA kit (R&D Systems), following manufacturer instruction. The effects of compounds were evaluated as a percentage of LPS-induced (control) concentrations of PGE₂.

4.2.3. LDH assay. Viability of cells was analyzed using the lactate dehydrogenase (LDH) assay. It is based on the determination of LDH activity released from the cytosol of damaged cells into the supernatant. The cell supernatants were harvested at the interval of 5 h of culture, diluted 1:1

and mixed with an aliquot of the LDH kit. After 30-min incubation in the dark at ambient temperature, the reaction was stopped with 2 N HCl. Differences between absorbances at 492 and 690 nm were evaluated. The percentage cytotoxicity of test samples was related to the control samples and to the samples with 100% dead cells evoked by 1% Triton, according to the formula: [(exp. value – control value) / (Triton value – control value)] x 100. All control and experimental variants were run in triplicate.

4.2.4. Statistical analysis. Analysis of variance (ANOVA) with subsequent Bonferroni's multiple comparison test, Wilcoxon signed rank test for comparison of medians, and graphical presentation of data were done using the Prism program (GraphPad Software, San Diego, CA).

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Abbreviations

COX, cyclooxygenase; LPS, lipopolysaccharide; NSAIDs, nonsteroidal anti-inflammatory drugs; NO, nitric oxide; PGE₂, prostaglandin E₂.

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Graphical abstract



Highlights

- Synthesis of polysubstituted pyrimidines using Suzuki-Miyaura cross-coupling reaction
- Potent inhibitors of prostaglandin E₂ production
- No cytotoxicity observed at the concentrations tested
- Potential anti-inflammatory agents