

Available online at www.sciencedirect.com



Tetrahedron Letters 46 (2005) 4023-4026

Tetrahedron Letters

New methodology for 2-alkylation of 3-furoic acids: application to the synthesis of tethered UC-781/d4T bifunctional HIV reverse-transcriptase inhibitors

Gareth Arnott, Roger Hunter,* Linda Mbeki and Ebrahim Mohamed

Department of Chemistry, University of Cape Town, Rondebosch 7701, South Africa

Received 24 February 2005; accepted 7 April 2005

Abstract—New methodology for 2-alkylation of 3-furoic acids is presented involving Wittig reactions of the 3-methoxycarbonyl-2-furanylmethylphosphonium salt. The methodology has been used to prepare a tethered 2-alkylated-UC-781/d4T conjugate as a potentially new type of HIV reverse-transcriptase inhibitor. © 2005 Elsevier Ltd. All rights reserved.

2,3-Disubstituted furans constitute a widely encountered sub-unit in a range of natural and synthetic products. 2-Alkylation of 3-furoic acids has been a commonly employed strategy for entry¹ into this sub-unit with two carbanionic methodologies standing out as versatile options. Knight was the first person to demonstrate² that treatment of 3-furoic acid with 2 equiv of LDA (THF/-78 °C) regioselectively furnishes the dianion 1 (Fig. 1), which can be alkylated with a range of reactive electrophiles. However, with less reactive electrophiles, for example, ethyl iodide, yields were low. Keay and co-workers subsequently³ demonstrated that 2-methyl-3-furoic acid reacts with 2 equiv of *n*-BuLi at -20 °C to furnish the 2-lithiomethyl dianion **2** which is more stable than **1**, giving higher yields with less reactive electrophiles. Development of **2** followed pioneering work by





Keywords: Furan alkylation; Wittig reaction of 2-furanylphosphonium salt; Alkylated-UC-781; Bifunctional HIV reverse-transcriptase inhibitors.

* Corresponding author. Tel.: +27 021 650 2544; fax: +27 021 689 7499; e-mail: roger@science.uct.ac.za

0040-4039/\$ - see front matter @ 2005 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2005.04.034

Tada et al.⁴ on use of the 2-dianion of 2,4-dimethyl-3-furoic acid 3 in natural product synthesis (Fig. 1).

As part of a programme to synthesise novel, bifunctional HIV reverse-transcriptase inhibitors, we needed access to quantities of 2-alkylated-3-furoates in conjunction with incorporation of the non-nucleoside inhibitor UC-781⁵ into a bifunctional inhibitor. In view of the unattractive prospect of using large quantities of n-BuLi, we embarked on a study to identify an alternative, which we successfully report in this communication. It occurred to us that Wittig methodology based on the 3-methoxycarbonyl-2-furanylmethylphosphonium salt might provide the answer in view of the option of using a mild base to generate the stabilised ylide. Although 2furanylmethylphosphonium salts⁶ have been known and used in synthesis for some time, the corresponding 3-furoates are hitherto unknown. To this end, radical bromination of commercially available methyl 2-methylfuroate using conditions recently reported by Khatuya⁷ furnished methyl 2-bromomethyl furoate in high yield, which, following evaporation of the solvent and addition of triphenylphosphine in toluene furnished (rt, overnight) the desired and novel triphenylphosphonium salt 4 by filtration. Isolation of product involved no chromatography, with a single crystallisation from methanol returning analytically pure material in 80% overall yield.

Pleasingly, reaction of 4 in methanol with sodium methoxide as base (5 M in MeOH; 1.1 equiv) at rt followed by addition of hexanal (1.2 equiv) as a model aldehyde



Scheme 1. Reagents and conditions: (i) NBS, (BzO)₂ (cat), CCl₄, Δ ; (ii) PPh₃, toluene, rt (80% over two steps); (iii) NaOMe (1.1 equiv), MeOH, C₅H₁₁CHO (92%); (iv) H₂, Pd–C, EtOH (80%).

 Table 1. Wittig olefination and hydrogenation of 4 with various aldehydes

R of RCHO	% Yield of 5	% Yield of 6
(a) H	92	80
(b) CH ₃	90	61
(c) C_5H_{11}	92	80
(d) C ₄ H ₉ OBn	91	84

resulted in rapid transformation to the Wittig product **5** in high yield as a mixture (~1:1) of E/Z stereoisomers. Carrying the reaction out in THF using sodium hydride as base gave a significantly lower yield (~50%) of the Wittig product in a higher E/Z ratio. Subsequent hydrogenation (H₂/Pd–C) gave the anticipated 2-alkylated product **6** in high yield (80%). A small percentage (~10%) of the 4,5-dihydro-2-alkylated product⁸ was also obtained, which could be minimised by varying the reaction conditions, but not completely eliminated (Scheme 1).

A range of aldehydes appropriate to producing alkylated side chains were subjected to the olefination/hydrogenation sequence and the results are presented in Table 1. Yields cited are relevant to reactions carried out in 1–10 mmol range. Reactions involving formaldehyde, ethanal and 5-benzyloxypentanal⁹ all underwent smooth Wittig reactions in high yield as with the model reaction and, where appropriate, similarly to products with about an equal E/Z isomer ratio. Subsequent hydrogenation of each one gave a small percentage of the 4,5-dihydro derivative as in the hexanal case, which could be separated from the desired alkylated product by careful silica-gel column chromatography. Hydrogenation of alkene **5d** resulted in concomitant hydrogenolysis of the benzyl ether (Scheme 2).

As part of a programme on the synthesis of new HIV reverse-transcriptase inhibitors, we were able to demonstrate applicability of the methodology to two new C-2 variants of the potent thiocarboxanilide non-nucleoside HIV reverse-transcriptase inhibitor, UC-781⁵ (Fig. 2).





To this end, hydrogenolysis of **6d** to alcohol **7** followed by tosylation and substitution with propargyloxy anion furnished the propynyl ether **8**. Interestingly, the substitution proceeded more cleanly this way round, that is, was superior than propargylation of the alkoxide of **7** with propargyl bromide. Subsequent base-mediated ester hydrolysis, conversion to the acid chloride with thionyl chloride followed by substitution with substituted aniline **9**¹⁰ (Fig. 2) furnished amide **10** in high overall yield from the acid. Finally, thiation of amide **10** with Lawesson's reagent¹¹ produced the C-2 elongated UC-781 derivative **11**¹² for biological probing¹³ of substituent effects in the HIV reverse-transcriptase pocket (Scheme 3).

Alkyne **10** was also subjected to a Sonogashira¹⁴ reaction with the nucleoside reverse-transcriptase inhibitor derivative, 5'-O-benzoyl-5-iodo-d4T,¹⁵ to afford conjugate **12**¹⁶ following benzoyl group deprotection. Conjugate **12** involves a combination of the two antiretroviral drugs d4T and UC-781, albeit with the amide of UC-781 unthiated. Significant interest has been shown recently in conjugates¹⁷ of this type, in view of the fact that the nucleotide substrate binding-site is proximal to the non-nucleoside binding pocket. Compound **12** is the first example of a UC-781-derived conjugate (unthiated) as a result of developing this methodology. Further work on thiation is in progress to produce UC-781 analogues (Scheme 4).

In summary, new methodology applicable to medium to large-scale work has been developed for C-2 alkylation



Scheme 2. Reagents: (i) NaOMe, MeOH, RCHO; (ii) H₂, Pd-C, EtOH.



Scheme 3. Reagents and conditions: (i) *p*-TsCl, NEt₃, CH₂Cl₂, DMAP (cat) (97%); (ii) propargyl alcohol (10 equiv), NaH (10 equiv), THF, Δ ; (iii) KOH, EtOH, (85%, two steps); (iv) SOCl₂, Δ ; (v) RNH₂, Py, (99%, two steps to give amide 10); (vi) Lawesson's reagent, NaHCO₃, toluene, Δ (70%).



Scheme 4. Reagents and conditions: (i) 5'-benzoyl-5-iodo-d4T, Pd(PPh₃)₄ (10%), CuI (50%), NEt₃ (2 equiv), DMF-THF (1:2), rt (65%); (ii) NaOMe, MeOH, rt (52%).

of 3-furoates of interest to both natural product synthesis and medicinal chemistry.

References and notes

- (a) Van Altena, I. A.; Miller, D. A. Aust. J. Chem. 1989, 42, 2181–2190; (b) Buttery, C. D.; Cameron, A. G.; Dell, C. P.; Knight, D. W. J. Chem. Soc., Perkin Trans. 1 1990, 1601–1610; (c) Tada, M.; Yamada, H.; Kanamori, A.; Chiba, K. J. Chem. Soc., Perkin Trans. 1 1993, 239–247; (d) Wang, F.; Chiba, K.; Tada, M. Chem. Lett. 1993, 12, 2117–2120; (e) Perry, P. J.; Pavlidis, V. H.; Hadfield, J. A.; Coutts, I. G. C. J. Chem. Soc., Perkin Trans. 1 1995, 9, 1085–1087; (f) Mal, D.; Bandhyopadhyay, M.; Datta, K.; Murty, K. V. S. N. Tetrahedron 1998, 54, 7525–7538; (g) Kline, T.; Bowman, J.; Iglewski, B. H.; de Kievit, T.; Kakai, Y.; Passador, L. J. Bioorg. Med. Chem. Lett. 1999, 9, 3447–3452.
- (a) Knight, D. W. Tetrahedron Lett. **1979**, 20, 469–472; (b) Knight, D. W.; Nott, A. P. J. Chem. Soc., Perkin Trans. 1 **1981**, 1125–1131.
- 3. Yu, S.; Beese, G.; Keay, B. A. J. Chem. Soc., Perkin Trans. 1 1992, 2729–2731.
- Tada, M.; Sugimoto, Y.; Takahashi, T. Bull. Chem. Soc., Jpn. 1980, 53, 2966–2970.
- (a) De Clercq, E. Farmaco 1999, 26–45; (b) De Clercq, E. Mini-Rev. Med. Chem. 2002, 163–175.
- (a) Schweizer, E. E.; Creasy, W. S.; Light, K. K.; Shaffer, E. T. J. Org. Chem. 1969, 34, 212–218; (b) Cooper, J. A.; Cornwall, P.; Dell, C. P.; Knight, D. W. Tetrahedron Lett. 1988, 29, 2107–2110; (c) Hart, D. J.; Patterson, S.; Urich, J. P. Synlett 2003, 1334–1338.
- 7. Khatuya, H. Tetrahedron Lett. 2001, 42, 2643–2644.
- For an interesting stereoselective Birch reduction of 3methyl-2-furoates as an entry into 2,5-dihydrofurans, see:

Donohoe, T. J.; Raoof, A.; Freestone, G. C.; Linney, I. D.; Cowley, A.; Helliwell, M. *Org. Lett.* **2002**, *4*, 3059–3062.

- Prepared from 1,5-pentanediol via the sequence: (a) NaH, BnBr, THF, Δ (63%); (ii) Swern oxidation (99%).
- Prepared from 2-amino-5-nitrophenol via the following sequence: (i) (a) aq NaNO₂, HCl, NH₂SO₃H (cat); (b) CuCl (69%); (ii) K₂CO₃, 4-bromo-2-methyl-2-butene, 2butanone, rt (94%); (iii) Fe, HCl, EtOH, Δ (82%).
- 11. Jesberger, M.; Davis, T. P.; Barner, L. Synthesis 2003, 13, 1929–1958.
- 12. Data for compound 11 (see numbering in Scheme 3): v_{max} / cm⁻¹ (CHCl₃): 3234m (N-H, thioamide), 2858s (C-H, aliphatic), 2270s (C=C, alkyne), 1618s (C=C, aromatic), 1389s+1162s (C=S stretches); ¹H NMR (400 MHz, C₆D₆): $\delta_{\rm H}$ 1.24–1.46 (8H, m, H-2", 3", 4", 5"), 1.45+1.51 (6H, 2×s, H-10', 11'), 2.02 (1H, t, J 2.4 Hz, H-9"), 3.00 (2H, t, J 7.6 Hz, H-1"), 3.28 (2H, t, J 6.4 Hz, H-6"), 3.83 (2H, d, J 2.4 Hz, H-7"), 4.41 (2H, d, J 6.1 Hz, H-8'), 5.47 (1H, tt, J 6.1, 1.4 Hz, H-9'), 6.13 (1H, d, J 1.2 Hz, H-4), 6.38 (1H, d, J 7.3 Hz, H-7'), 6.82 (1H, d, J 7.6 Hz, H-6'), 7.15 (1H, d, J 1.2 Hz, H-5), 7.98 (1H, s, NH), 8.17 (1H, s, H-3'); ¹³C NMR (75 MHz, C₆D₆): δ_C 17.9 (CH₃), 25.5 (CH₂), 25.9 (CH₃), 27.9 (C-1"+CH₂), 29.1+29.5 (CH₂'s), 57.8 (C-7"), 66.1 (C-8'), 69.8 (C-6"), 73.9 (C-9"), 80.5 (C-8"), 106.1 (C-3'), 108.1 (C-4), 109.4 (C-7'), 115.7 (C-3), 119.7 (C-5'), 120.6 (C-9'), 130.1 (C-5+C-6'), 138.2 (C-2'), 138.7 (C-12'), 141.1 (C-2), 154.8 (C-4'), 160.0 (C-1'); HRMS: m/z (rel int.) 459.16343 $[M^+]$ (9). Calculated for $C_{25}H_{30}O_3NSCI$: 459.16349 [M⁺].
- (a) Esnouf, R. M.; Stuart, D. I.; De Clercq, E.; Schwartz, E.; Balzarini, J. *Biochem. Biophys. Res. Commun.* 1997, 234, 458–464; (b) Goette, M.; Wainberg, M. A. *Infect. Dis. Ther.* 2003, 30, 505–521.
- (a) Agrofoglio, L. A.; Gillaizeau, I.; Saito, T. *Chem. Rev.* **2003**, *103*, 1875–1916; (b) Ciurea, A.; Fossey, C.; Gavriliu,
 D.; Delbederi, Z.; Sugeac, E.; Ladureé, D.; Schmidt, S.;

Laumond, G.; Aubertin, A.-M. J. Enzyme Inhib. Med. Chem. 2004, 19, 511–519.

- Ciurea, A.; Fossey, C.; Benzaria, S.; Gavriliu, D.; Delbederi, Z.; Lelong, B.; Laduree, D.; Aubertin, A. M.; Kirn, A. Nucleosides, Nucleotides Nucleic Acids 2001, 20, 1655– 1670.
- 16. Data for compound **12** (see numbering in Scheme 4): $[\alpha]_D - 15.7$ (*c* 1.03, CHCl₃); v_{max}/cm^{-1} (CHCl₃): 3592br (O–H, free), 2932s (C–H, aliphatic), 2254s (C=C, alkyne), 1720s (C=O, ester), 1693s (C=O, amide), 1599s (N–H and C–N stretching), 1514s+1492s (C=C, aromatic); ¹H NMR (400 MHz, CDCl₃): δ_H 1.24–1.33 (8H, m, H-14, 15, 16, 17), 1.73–1.79 (6H, 2×s, H-33, 34), 2.70 (1H, s, –OH), 3.03 (2H, t, *J* 7.6 Hz, H-18), 3.46 (2H, t, *J* 6.8 Hz, H-13), 3.78 (1H, d, *J* 12.4 Hz, H-1), 3.88 (1H, d, *J* 12.4 Hz, H-1), 4.21 (2H, s, H-12), 4.59 (2H, d, *J* 6.7 Hz, H-30), 4.91 (1H, s, H-2), 5.51 (1H, tt, *J* 6.7, 1.4 Hz, H-31), 5.83 (1H, dt, *J* 5.9, 1.9 Hz, H-3), 6.32 (1H, dt, *J* 5.9, 1.6 Hz, H-4),

6.56 (1H, d, J_{AB} 2.2 Hz, H-21), 6.96 (2H, m, H-5,29), 7.26 (1H, m, H-28), 7.30 (1H, d, J_{AB} 2.2 Hz, H-22), 7.53 (1H, s, H-9), 7.74 (1H, s, NH), 8.05 (1H, s, H-25), 8.23 (1H, s, H-6); ¹³C NMR (75 MHz, CDCl₃): δ_{C} 18.3, 25.6, 25.8, 27.2, 27.7, 28.7, 29.2, 58.7, 63.0, 66.2, 70.2, 76.8 × 2, 87.6, 89.9, 90.3, 99.5, 106.4, 108.2, 112.7, 115.6, 119.1, 125.9, 129.9, 134.9, 137.7, 138.6, 140.6, 144.4, 149.7, 149.7, 154.5, 161.8, 162.2.

 (a) Spence, R. A.; Kati, W. M.; Anderson, K. S.; Johnson, K. A. Science 1995, 267, 988–993; (b) Velazquez, S.; Tunon, V.; Jimeno, M. L.; Chamorro, C.; De Clercq, E.; Balzarini, J.; Camarasa, M. J. J. Med. Chem. 1999, 42, 5188–5196; (c) Ladureé, D.; Sugeac, E.; Fossey, C.; Schmidt, S.; Laumond, G.; Aubertin, A. M. Nucleosides, Nucleotides Nucleic Acids 2003, 22, 873–875; (d) Sugeac, E.; Fossey, C.; Ladureé, D.; Schmidt, S.; Laumond, G.; Aubertin, A.-M. J. Enzyme Inhib. Med. Chem. 2003, 18, 175–186.