Synthesis of 2,2- and 2,5-Disubstituted 1-Oxyl Pyrrolidine Radicals as New Homobifunctional Cross-Linking Spin Labels

Tamás Kálai,^a József Jekő,^b Wayne L. Hubbell,^c Kálmán Hideg*^a

^a Institute of Organic and Medicinal Chemistry, University of Pécs, 7602 Pécs, P. O. Box 99, Hungary

^b ICN Hungary Ltd., 4440 Tiszavasvári, P. O. Box 1, Hungary

^c Jules Stein Eye Institute and Department of Chemistry and Biochemistry, UCLA, Los Angeles, CA 90095-7008, USA *Received 28 May 2003*

Abstract: The synthesis of 2,2- and 2,5-disubstituted pyrrolidine nitroxide radicals starting from readily available nitrones 1, 11 is described. The stable radicals are acylating (10, 20), alkylating (7, 17) and thiol-specific (8, 18) reagents capable of producing cross-links in proteins over distances in the range of 10–15 Å.

Key words: cross linking reagents, proteins, free radicals, Grignard reactions, nitrones, lithium

Nitroxides have received considerable attention as co-oxidants,¹ spin labels,² MRI agents,³ antioxidants⁴ and double sensor molecules.⁵ Among these uses, spin labeling is the most important and wide-spread application. A major advance in the field of protein spin labeling was the introduction of site directed spin labeling (SDSL). SDSL is proving to be a general strategy for determining the structure of both soluble and membrane proteins of any molecular weight.⁶⁻⁸ Moreover, recent work indicates that protein dynamics, an important determinant of protein function, can be extracted from the motion of nitroxide side chains.9,10 Homo-bifunctional nitroxide reagents, particularly those with reactive functionality toward sulfhydryl groups, promise to significantly extend the capabilities of SDSL. For example, bifunctional reagents that cross-link a pair of cysteine residues spaced by one or two turns along an α -helix are strongly immobilized with respect to the protein backbone, as expected. In this case, motion of the nitroxide group, reflected in the EPR spectrum, is dominated by rigid body motions of the helix. Thus, the EPR spectra of spin labels with dual attachment points may provide a direct and powerful measure of protein dynamics without the usual contributions from the internal motion of the side chain itself.¹⁰ Because the orientation of the nitroxide ring is necessarily fixed by the cross-link, the angular dependence of the EPR spectrum in an oriented system (i.e., membranes) can be used to deduce the orientation of the helical axis with respect to the director of the system. Finally, the rate of cross-link formation, easily monitored by EPR in real time, will provide direct measures of both cysteine proximity and protein flexibility. In our laboratory we have synthesized several heterobifunctional (thiol-specific and photoactivatable)^{11,12} and thiol-specific homobifunctional cross-linking reagents^{13,14} containing reactive substituents only in the 3,4-positions of the pyrroline nitroxide ring. In this paper, we present an extension of the above idea with the synthesis of novel 2,2- and 2,5-disubstituted pyrrolidine nitroxide homobifunctional reagents that are designed to serve the purposes mentioned above.

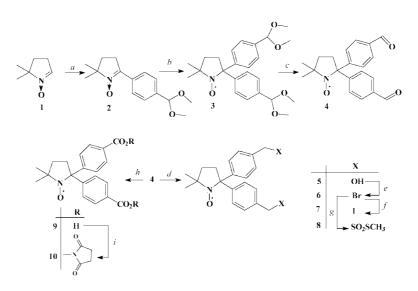
Treatment of 5,5-dimethyl-1-pyrroline N-oxide (DMPO) (1) or 2,5-dimethyl-1-pyrroline N-oxide¹⁵ (11) with 4dimethoxymethylphenylmagnesium bromide yielded trisubstituted nitrones¹⁶ 2 and 12, respectively, after oxidation of the hydroxylamines to the nitrones by activated MnO₂. Further treatment of nitrone 12 with 4dimethoxymethylphenylmagnesium bromide gave trans diphenyl nitroxide **13** analogously to earlier cases.¹⁷ On the other hand, similar treatment of nitrone 2 yielded only the unsubstituted hydroxylamine. Apparently, the second phenyl substituent cannot be easily introduced into this sterically hindered position, as was observed earlier by Iwahashi et al.¹⁸ Treatment of compound **2** with the more reactive 4-dimethoxymethylphenyllithium obtained by translithiation¹⁹ of 4-bromobenzaldehyde dimethyl acetal gave the desired nitroxide 3 after oxidation with activated MnO_2 .

Dimethylacetals 3 and 13 could be deprotected by treatment with aqueous H₂SO₄ in THF to give aldehydes 4 and 14,²⁰ which were in turn reduced with NaBH₄ in ethanol to give alcohols 5 and 15. The alcohols were converted to the corresponding mesylates, which yielded the benzylic bromides 6 and 16 upon treatment with LiBr in acetone.^{21,22} To obtain the more reactive benzylic iodides for use as alkylating agents,²³ compounds 6 and 16 were treated with NaI to give compounds 7 and 17. The thiol-specific bismethanethiosulfonate reagents 8 and 18 were obtained by treatment of bromides 6 and 16 with NaSSO₂CH₃ in aqueous acetone.²⁴ Reagents 8 and 18 are capable of making cross-links between SH of cysteine residues. Dialdehydes 4 and 14 were oxidized to dicarboxylic acids 9 and 19 by Ag₂O in an aq NaOH (10%)-THF mixture.²⁵ The dicarboxylic acids were converted to disuccinimidyl esters²⁶ 10 and 20 by treatment with *N*-hydroxysuccinimide and N,N'-dicyclohexylcarbodiimide (Schemes 1 and 2). The resulting disuccinimides are acylating reagents for amino groups and capable of making cross-link between lysine side chains.

Fax +36(72)536219; E-mail: kalman.hideg@aok.pte.hu

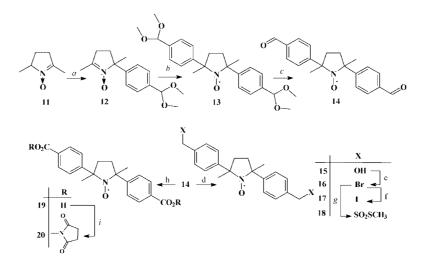
Synthesis 2003, No. 13, Print: 18 09 2003. Web: 22 08 2003. Art Id.1437-210X,E;2003,0,13,2084,2088,ftx,en;P04103SS.pdf. DOI: 10.1055/s-2003-41053

[©] Georg Thieme Verlag Stuttgart · New York



Scheme 1 *Reagents and conditions*: (a) 4-dimethoxymethylphenylmagnesium bromide (1.2 equiv) THF, 0 °C \rightarrow reflux, 3 h, under N₂, then activated MnO₂ (1.0 equiv), O₂, 10 min, CHCl₃, reflux, 1 h, 70%; (b) 4-dimethoxymethylphenyllithium (1.2 equiv), THF, -78 °C, 20 min, then **2**, reflux 5 h, then activated MnO₂, CHCl₃, O₂, 30 min, 65%; (c) THF, aq H₂SO₄ (5%), r.t., 8 h, 92%; (d) NaBH₄ (2.5 equiv) EtOH, 40 °C, 30 min, 78%; (e) MsCl (2.2 equiv), Et₃N (2.2 equiv), CH₂Cl₂, 0 °C \rightarrow r.t., 1 h, then LiBr (4.0 equiv), acetone, reflux, 1 h, 55%; (f) NaI, 4.0 equiv, reflux, 30 min, 68%; (g) NaSSO₂CH₃ (4.0 equiv), acetone–H₂O, 30 min, reflux, 43%; (h) Ag₂O (4.0 equiv), aq NaOH (10%)–THF, 60 °C, 40 min, then H⁺, 45%; (i) *N*-hydroxysuccinimide (2.0 equiv), DCC (2.0 equiv), EtOAc, 0 °C Ø r.t., 12 h, 61%.

In summary, new 2,2- and 2,5-disubstituted stable pyrrolidine radicals were obtained with simple procedures starting from readily available nitrones. These homobifunctional amino-specific and thiol-specific reagents serve as cross-linkers between lysine and cysteine residues, respectively. The general synthetic procedure is readily extendable to include spacer groups other than benzyl to vary the cross-linking distance. Applications of the reagents will be reported separately. Mps were determined with a Boetius micro melting point apparatus and are uncorrected. Elemental analyses (C, H, N, S) were performed on Carlo Erba EA 1110 CHNS elemental analyzer. IR (Specord 75) spectra were in each case consistent with the assigned structure. Mass spectra were recorded on a VG TRIO-2 instrument in the EI mode (70 eV, direct inlet). ESR spectra were obtained from 10^{-5} molar solutions (CHCl₃), using a Bruker ECS-106 spectrometer. All monoradicals exhibited three equally spaced lines with $a_N = 15.1-15.5$ G. Preparative flash column chromatography was performed on Merck Kieselgel 60 (0.040–0.063 mm). Qualitative TLC was carried out on commercially prepared plates



Scheme 2 *Reagents and conditions*: (a) 4-dimethoxymethylphenylmagnesium bromide (1.2 equiv) THF, 0 °C \rightarrow reflux, 3 h, under N₂, then activated MnO₂ (1.0 equiv), O₂, 10 min, CHCl₃, reflux, 1 h, 55%; (b) 4-dimethoxymethylphenylmagnesium bromide (1.2 equiv), THF, 0 °C, then **12**, reflux, 3 h, under N₂, then activated MnO₂, CHCl₃, O₂, 30 min, 60%; (c) THF, aq H₂SO₄ (5%), r.t., 8 h, 85%; (d) NaBH₄ (2.5 equiv) EtOH, 40 °C, 30 min, 70%; (e) MsCl (2.2 equiv), Et₃N (2.2 equiv), CH₂Cl₂, 0 °C \rightarrow r.t., 1 h, then LiBr (4.0 equiv), acetone, reflux, 1 h, 53%; (f) NaI (4.0 equiv), acetone, reflux, 30 min, 77%; (g) NaSSO₂CH₃ (4.0 equiv), acetone–H₂O, 30 min, reflux, 28%; (h) Ag₂O (4.0 equiv), aq NaOH (10%)–THF, 60 °C, 40 min, then H⁺, 40%; (i) *N*-hydroxysuccinimide (2.0 equiv), DCC (2.0 equiv), EtOAc, 0 °C \rightarrow r.t., 12 h, 55%.

PAPER

Table 1New Compounds (2–10, 12–20)Prepared^a

Compound	Yield (%)	Mp (°C)	IR (cm ⁻¹) Neat or Nujol	MS (<i>m</i> / <i>z</i>)
2	70	60–62	1600, 1500 (C=C)	263 (M ⁺ , 27), 248 (5), 232 (100), 216 (20)
3	60	oil	1600, 1500 (C=C)	414 (M ⁺ , 2), 297 (22), 246 (83), 75 (100)
4	92	133–135	1680 (C=O), 1590 (C=C)	322 (M ⁺ , 8), 308 (35), 292 (23), 236 (100)
5	78	107–109	3300 (OH), 1500 (C=C)	326 (M ⁺ , 10), 312 (23), 296 (15), 240 (100)
6	55	174–176	1500 (C=C)	450/452/454 (M ⁺ , 3/6/3), 436/438/440 (8/16/8), 364/366/368 (6/12/6), 285/287 (100/100)
7	68	183–185	1520 (C=C)	546 (M ⁺ , 16), 473 (6), 420 (6), 333 (100)
8	43	115–117	1510 (C=C)	514 (M ⁺ , <1), 484 (2), 404 (2), 65 (100)
9	45	143–145	1680 (C=O), 1600, 1560 (C=C)	354 (M ⁺ , 9), 340 (43), 324 (55), 268 (100)
10	61	103–105	1760, 1720 (C=O), 1550 (C=C)	548 (M ⁺ , <1), 435 (1), 115 (35), 55 (100)
12	55	oil	1600, 1510 (C=C)	263 (M ⁺ , 38), 232 (76), 161 (77), 75 (100)
13	65	55–57	1600, 1500 (C=C)	414 (M ⁺ , 3), 384 (18), 192 (13), 161 (100)
14	85	181–183	1680 (C=O), 1590, 1560 (C=C)	322 (M ⁺ , 5), 308 (16), 292 (23), 146 (100)
15	70	163–165	3300 (OH), 1500 (C=C)	326 (M ⁺ , 4), 312 (9), 296 (14), 148 (100)
16	53	161–162	1500 (C=C)	450/452/454 (M ⁺ , 1/2/1), 420/422/424 (2/4/2), 210/212 (14/14), 131 (100)
17	77	155–157	1510 (C=C)	546 (M ⁺ , 2), 389 (4), 258 (8), 131 (100)
18	28	138–139	1500 (C=C)	514 (M ⁺ , <1), 484 (1), 80 (55), 65 (100)
19	40	242–244	1680 (C=O), 1590 (C=C)	354 (M ⁺ , 2), 340 (6), 310 (20), 148 (100)
20	55	222-224	1760, 1720 (C=O), 1550 (C=C)	548 (M ⁺ , <1), 435 (1), 115 (35), 55 (100)

^a All compounds gave satisfactory microanalyses: C, H, N, S (where appropriate) ± 0.3.

 $(20 \times 20 \times 0.02 \ \text{cm})$ coated with Merck Kieselgel GF₂₅₄. All reagents and 1 were purchased from Sigma-Aldrich, Hungary. Compound 11 was prepared according to published procedures.¹⁵ The physical and spectral data of all new compounds are listed in Table 1.

5-(4-Dimethoxymethylphenyl)-2,2-dimethyl-1-pyrroline *N*-Oxide (2) and 2-(4-Dimethoxymethylphenyl)-2,5-dimethyl-1-pyrroline *N*-Oxide (12)

To a stirred solution of 4-dimethoxymethylphenylmagnesium bromide [made from Mg turnings (1.44 g, 60.0 mmol) and 4-bromobenzaldehyde dimethyl acetal (13.86 g, 60.0 mmol)] in THF (40 mL), nitrone **1** or **11** (5.65 g, 50.0 mmol) was added dropwise at 0 °C and the mixture was stirred and refluxed for 3 h under N₂. After cooling, sat. aq NH₄Cl solution (20 mL) was added, the organic phase was separated, and the aq phase was extracted with CHCl₃ (2 × 20 mL). The combined organic phases were dried (MgSO₄), filtered and evaporated. The residue was dissolved in CHCl₃ (40 mL), activated MnO₂ (4.35 g, 50.0 mmol) was added in one portion, O₂ was bubbled through for 10 min, and the solution was stirred and refluxed for 1 h. The MnO₂ was filtered off, the solvent was evaporated under reduced pressure and the residue was purified by flash column chromatography (CHCl₃–Et₂O) to give nitrone **2** or **12**.

Nitrone 2

Yield: 9.20 g (70%); off-white solid; mp 60–62 °C; $R_{\rm f}$ 0.28 (CHCl_3– Et_2O, 2:1).

Nitrone 12

Yield: 7.23 g (55%); oil; R_f 0.28 (CHCl₃-MeOH, 9:1).

2,2-Bis-(4-dimethoxymethylphenyl)-5,5-dimethylpyrrolidin-1yloxyl Radical (3)

To a stirred solution of 4-dimethoxymethylphenyl bromide (6.93 g, 30.0 mmol) in THF (30 mL), BuLi (2.5 M in hexane; 14 mL, 35 mmol) was added dropwise at -78 °C and the mixture was stirred for 20 min at this temperature. Nitrone **2** (6.57 g, 25.0 mmol) dissolved in THF (20 mL) was added dropwise and the mixture was stirred and refluxed for 5 h under N₂. After cooling, the reaction mixture was quenched with sat. aq NH₄Cl solution (20 mL), the organic phase was separated and the aq phase was extracted with CHCl₃ (2 × 20 mL). The combined organic phases were dried (MgSO₄), activated MnO₂ (870 mg, 10.0 mmol) was added, O₂ was bubbled through for 30 min, and the mixture was filtered and evaporated. The residue was purified by flash column chromatography (hexane–EtOAc) to give compound **3**.

Yield: 6.72 g (65%); red oil; R_f 0.28 (hexane–EtOAc, 2:1).

2,5-*trans*-Bis-(4-dimethoxymethylphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (13)

To a stirred solution of 4-dimethoxymethylphenylmagnesium bromide [made from Mg turnings (720 mg, 30.0 mmol) and 4-bromobenzaldehyde dimethyl acetal (6.93 g, 30.0 mmol)] in THF (25 mL), nitrone **12** (6.57 g, 25.0 mmol) dissolved in THF (20 mL) was added dropwise at 0 °C and the mixture was stirred and refluxed for 3 h under N₂. After cooling, sat. aq NH₄Cl solution (20 mL) was added, the organic phase was separated, and the aq phase was extracted with CHCl₃ (2 × 20 mL). The combined organic phases were dried (MgSO₄), activated MnO₂ (870 mg, 10.0 mmol) was added, O₂ was bubbled through for 30 min, and the mixture was filtered and evaporated. The residue was purified by flash column chromatography (hexane–EtOAc) to give compound **13**.

Yield: 6.21 g (60%); orange solid; mp 55–57 °C; R_f 0.35 (hexane–EtOAc, 2:1).

2,2-Bis-(4-formylphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Radical (4) and 2,5-*trans*-Bis-(4-formylphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (14)

To a stirred solution of bis(dimethyl acetal) 3 or 13 (8.28 g; 20.0 mmol) in THF (40 mL),

aq H_2SO_4 (5%; 10 mL) was added and the mixture was stirred for 8 h at r.t. Brine (30 mL) and Et_2O (30 mL) were then added, the organic phase was separated, dried (MgSO₄), filtered, evaporated and the residue was purified by flash column chromatography (hexane–EtOAc) to give the aldehyde **4** or **14**.

Aldehyde 4

Yield: 5.92 g (92%); yellow solid; mp 133–135 °C; R_f 0.16 (hexane–EtOAc, 2:1).

Aldehyde 14

Yield: 5.47 g (85%); yellow solid; mp 181–183 °C; $R_{\rm f}$ 0.19 (hexane–EtOAc, 2:1).

2,2-Bis-(4-hydroxymethylphenyl)-5,5-dimethylpyrrolidin-1yloxyl Radical (5) and 2,5-*trans*-Bis-(4-hydroxymethylphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (15)

To a stirred solution of aldehyde **4** or **14** (6.44 g, 20.0 mmol) in EtOH (50 mL), NaBH₄ (1.89 g, 50.0 mmol) was added and the mixture was stirred for 30 min at 40 °C. The solvents were evaporated, the residue was dissolved in brine (30 mL), the aq phase was extracted with CHCl₃ (2×20 mL), dried (MgSO₄), filtered and the solvent evaporated. The residue was purified by flash column chromatography (CHCl₃–MeOH) to give compound **5** or **15**.

Compound 5

Yield: 5.08 g (78%); yellow solid; mp 107–109 °C; $R_f 0.26$ (CHCl₃–MeOH, 9:1).

Compound 15

Yield: 4.56 g (70%); yellow solid; mp 163–165 °C; $R_f 0.33$ (CHCl₃–MeOH, 9:1).

2,2-Bis-(4-bromomethylphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Radical (6) and 2,5-*trans*-Bis-(4-bromomethylphenyl)-2,5dimethylpyrrolidin-1-yloxyl Radical (16)

To a solution of bis(benzylic alcohol) **5** or **15** (3.26 g, 10.0 mmol) and Et₃N (2.22 g, 22 mmol) in CH₂Cl₂ (40 mL), methanesulfonyl chloride (2.52 g, 22.0 mmol) was added at 0 °C and the mixture was stirred at r.t. for 1 h. The reaction mixture was then washed with H₂O (30 mL), dried (MgSO₄), filtered, and the solvent evaporated. The residue was dissolved in anhyd acetone (50 mL), LiBr (3.48 g, 40.0 mmol) was added, and the mixture was heated at reflux and stirred for 1 h. After cooling, the acetone was evaporated, the residue was dissolved in Et₂O (40 mL), washed with H₂O (20 mL), the

organic phase was separated, dried ($MgSO_4$), filtered, and the solvent evaporated. The residue was purified by flash column chromatography (hexane–EtOAc) to give compound **6** or **16**.

Compound 6

Yield: 2.48 g (55%); mp 174–176 °C; R_f 0.56 (hexane–EtOAc, 2:1).

Compound 16

Yield: 2.39 g (53%); mp 161–162 °C; R_f 0.60 (hexane–EtOAc, 2:1).

2,2-Bis-(4-iodomethylphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Radical (7) and 2,5-*trans*-Bis-(4-iodomethylphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (17)

A solution of bis(benzylic bromide) **6** or **16** (904 mg, 2.0 mmol) and NaI (1.20 g, 8.0 mmol) in acetone was stirred and refluxed for 30 min. The NaBr was filtered off, and the acetone was evaporated under reduced pressure. The residue was dissolved in EtOAc (20 mL) and washed with aq $Na_2S_2O_3$ solution (5%; 10 mL). The organic phase was separated, dried (MgSO₄), filtered, evaporated under reduced pressure and the residue was purified by flash column chromatography (hexane–Et₂O) to give compound **7** or **17**.

Compound 7

Yield: 742 mg (68%); yellow solid; mp 183–185 °C; $R_{\rm f}$ 0.58 (hexane–EtOAc, 2:1).

Compound 17

Yield: 841 mg (77%); yellow solid; mp 155–157 °C; R_f 0.65 (hexane–EtOAc, 2:1).

2,2-Bis-(4-methanethiosulfonylmethylphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Radical (8) and 2,5-*trans*-Bis-(4-methanethiosulfonylmethylphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (18)

To a solution of dibromo compound **6** or **16** (452 mg, 1.0 mmol) in acetone (10 mL) and H_2O (5 mL), NaSSO₂CH₃ (536 mg, 4.0 mmol) was added and the mixture heated under reflux for 30 min. After cooling, the acetone was evaporated, and the aq phase was extracted with CHCl₃ (2 × 10 mL). The organic phase was separated, dried (MgSO₄), filtered, evaporated and the residue was purified by flash column chromatography (CHCl₃–Et₂O) to give compound **8** or **18**.

Compound 8

Yield: 221 mg (43%); mp 115–117 °C; R_f 0.46 (CHCl₃–Et₂O, 2:1).

Compound 18

Yield: 144 mg (28%); mp 138–139 °C; R_f 0.47 (CHCl₃–Et₂O, 2:1).

2,2-Bis-(4-carboxyphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Radical (9) and 2,5-*trans*-Bis-(4-carboxyphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Radical (19)

To a vigorously stirred suspension of freshly precipitated Ag₂O (9.26 g, 40.0 mmol) in aq NaOH solution (10%; 40 mL), aldehyde **9** or **19** (3.22 g, 10.0 mmol) in THF (15 mL) was added at 60 °C and the mixture was stirred and refluxed for 40 min. After cooling, the solution was filtered through a Celite pad and washed with MeOH (20 mL). The organic solvents were evaporated off, the aq phase was acidified with aq H₂SO₄ (5%) to pH 2, and extracted with CHCl₃ (3 × 20 mL). The organic phase was dried (MgSO₄), filtered and evaporated. The residue was purified by flash column chromatography (CHCl₃–MeOH) to give dicarboxylic acid **9** or **19**.

Dicarboxylic Acid 9

Yield: 1.59 g (45%); mp 143–145 °C; $R_f 0.13$ (CHCl₃–MeOH, 9:1).

Dicarboxylic Acid 19

Yield: 1.41 g (40%); mp 242–244 °C; R_f 0.16 (CHCl₃–MeOH, 9:1).

2,2-Bis-(4-carboxyphenyl)-5,5-dimethylpyrrolidin-1-yloxyl Bis(succinimide Ester) Radical (10) and 2,5-*trans*-Bis-(4-carboxyphenyl)-2,5-dimethylpyrrolidin-1-yloxyl Bis(succinimide Ester) Radical (20)

To a stirred solution of carboxylic acid **9** and **19** (354 mg, 1.0 mmol) and *N*-hydroxysuccinimide (230 mg, 2.0 mmol) in EtOAc (10 mL), DCC (412 mg, 2.0 mmol) dissolved in EtOAc (5 mL) was added dropwise at 0 °C. The mixture was stirred at r.t. overnight, the precipitated dicyclohexyl urea was filtered off, the filtrate was evaporated and purified by flash column chromatography (CHCl₃–Et₂O) to give active ester **10** or **20**.

Ester 10

Yield: 335 mg (61%); mp 103–105 °C; $R_f 0.39$ (CHCl₃–MeOH, 9:1).

Ester 20

Yield: 301 mg (55%); mp 222–224 °C; $R_f 0.35$ (CHCl₃–MeOH, 9:1).

Acknowledgments

This work was supported by grant from Hungarian National Research Foundations (OTKA T34307). A Bolyai fellowship for T. K. from the Hungarian Academy of Sciences is gratefully acknowledged. The authors thank to Mária Balog for technical assistance and Viola H. Csokona for elemental analysis and Mária Szabó for mass spectral measurements (ICN, Hungary).

References

- Formaggio, F.; Bonchio, M.; Crisma, M.; Peggion, C.; Mezzato, S.; Polese, A.; Barazza, A.; Antonello, S.; Maran, F.; Broxterman, Q. B.; Kaptein, B.; Kamphuis, J.; Vitale, R. M.; Saviano, M.; Benedetti, E.; Toniolo, C. *Chem.–Eur. J.* **2002**, *8*, 84.
- (2) Hubbell, W. L.; McConnell, H. M. J. Am. Chem. Soc. 1971, 93, 314.
- (3) He, G.; Samouilov, A.; Kuppusamy, A.; Zweier, J. L. J. Magn. Reson. 2001, 148, 155.
- (4) Mitchell, J. B.; Xavier, S.; DeLuca, A. M.; Showers, A. L.; Cook, J. A.; Krishna, M. C.; Hahn, S. M.; Russo, A. Free Radical Biol. Med. 2003, 34, 93.

Downloaded by: University of Florida. Copyrighted material

- (5) Bilski, P.; Hideg, K.; Kálai, T.; Bilska, M. A.; Chignell, C. F. *Free Radical Biol. Med.* **2003**, *34*, 489.
- (6) Hubbell, W. L.; Mchaourab, H. S.; Altenbach, C.; Lietzow, M. A. *Structure* **1996**, *4*, 779.
- (7) Hubbell, W. L.; Gross, A.; Langen, R.; Lietzow, M. A. Curr. Opin. Struct. Biol. 1998, 8, 649.
- (8) Hubbell, W. L.; Cafiso, D. S.; Altenbach, C. Nat. Struct. Biol. 2000, 7, 735.
- (9) Columbus, L.; Kálai, T.; Jekő, J.; Hideg, K.; Hubbell, W. L. Biochemistry 2001, 40, 3828.
- (10) Columbus, L.; Hubbell, W. L. Trends Biochem. Sci. 2002, 27, 288.
- (11) Kálai, T.; Rozsnyai, B.; Jerkovich, Gy.; Hideg, K. *Synthesis* **1994**, 1079.
- (12) Lösel, R. M.; Philipp, R.; Kálai, T.; Hideg, K.; Trommer, W. E. *Bioconjugate Chem.* **1999**, *10*, 578.
- (13) Kálai, T.; Balog, M.; Jekő, J.; Hideg, K. Synthesis 1999, 973.
- (14) Kálai, T.; Balog, M.; Jekő, J.; Hubbell, W. L.; Hideg, K. Synthesis 2002, 2365.
- (15) Keana, J. F. W. In Spin Labeling II, Theory and Applications; Berliner, L. J., Ed.; Academic Press: New York, **1979**, 115.
- (16) Sár, P. C.; Jekő, J.; Fajer, P.; Hideg, K. Synthesis 1999, 1039.
- (17) Rockenbauer, A.; Korecz, L.; Hideg, K. J. Chem. Soc., Perkin Trans. 2 1993, 2149.
- (18) Iwahashi, H.; Parker, C. E.; Mason, R. P.; Tomer, K. B. *Anal. Chem.* **1992**, *64*, 2244.
- (19) Creary, X.; Aldridge, T. J. Org. Chem. 1991, 56, 4280.
- (20) Kocienski, P. *Protecting Groups*, 2nd ed.; Thieme: Stuttgart, **2000**.
- (21) Hideg, K.; Hankovszky, H. O.; Lex, L.; Kulcsár, Gy. Synthesis **1980**, 911.
- (22) Hankovszky, H. O.; Hideg, K.; Lex, L. Synthesis **1980**, 914.
- (23) Mchaourab, H. S.; Kálai, T.; Hideg, K.; Hubbell, W. L. *Biochemistry* **1999**, *38*, 2947.
- (24) Berliner, L. J.; Grünwald, J.; Hankovszky, H. O.; Hideg, K. Anal. Biochem. 1982, 119, 450.
- (25) Hassner, A.; Stumer, C. In Organic Synthesis Based on Name Reactions and Unnamed Reactions; Pergamon: Oxford, 1994, 89.
- (26) Hill, M.; Bechet, J.-J.; d'Albis, A. FEBS Lett. 1979, 102, 282.