Synthesis and Biological Evaluation of New Oral Carbapenems with 1-Methyl-5-oxopyrrolidin-3-ylthio Moiety

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The synthesis and biological properties of 1β -methylcarbapenems with 1-methyl-5-oxopyrrolidin-3-ylthio group at the C-2 position were studied. The sodium (1R,5S,6S)-6-[(R)-1-hydroxyethyl]-1-methyl-2-[(R)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate and its (S)-isomer at the 2-position show potent and well-balanced antibacterial activity. The pharmacokinetic parameters of the pivaloyloxymethyl esters of these two carbapenems were compared in mice. The *in vivo* potency of these carbapenems was compared with that of cefdinir. Good *in vivo* efficacy of these ester prodrugs reflected the high and prolonged blood levels in parent drugs achieved after oral administration to mice.

Carbapenems are the most potent β -lactam antibiotics, displaying broad antibacterial spectrum and have potent bactericidal activity against both Gram-positive and Gramnegative organisms. 1,2) Three parenteral carbapenems, imipenem/cilastatin,^{3,4)} panipenem/betamipron,^{5~7)} meropenem,8) have already been launched. However, oral carbapenems are not yet available although oral cephalosporins such as cefpodoxime proxetil and cefdinir have been used clinically. 9~11) In the course of structureactivity relationship studies of CS-834, 12,131 our attention was focused on the 1-methylated compound of the 5oxopyrrolidin-3-ylthio group at the C-2 position. We were especially interested in increasing the activity and improving the pharmacokinetics of oral carbapenems when compared with oral cephalosporins. For this purpose, we synthesized sodium (1R,5S,6S)-6-[(R)-1-hydroxyethyl]-1methyl-2-[(R)-1-methyl-5-oxopyrrolidin-3-ylthio]-1carbapen-2-em-3-carboxylate and its (S)-isomer at the 2position, and evaluated these derivatives for their antibacterial activity and pharmacokinetics after intravenous administration in mice. Also, the pivaloyloxymethyl esters of these two carbapenems were synthesized and the pharmacokinetics and in vivo efficacy were evaluated after oral administration in mice. Both carbapenem esters showed good oral absorption and the prolonged half-lives in mice. The half-lives (T_{1/2}) in both

parent carboxylic acids were superior compared with that of cefdinir. The results showed that the favorable pharmacokinetics of these compounds have good *in vivo* efficacy against experimental infections in mice.

Chemistry

 1β -Methyl carbapenems with (R)- and (S)-1-methyl-5oxopyrrolidin-3-ylthio groups (11a and 11b) were synthesized from (S)-4-hydroxy-2-oxopyrrolidine 1 via the routes as shown in Scheme 1 and 2, respecively. Optically active (R)-3-mercapto-1-methyl-5-oxopyrrolidine (5) was prepared from 1. The hydroxy group of 1 was protected with t-butyldimethylsilyl chloride (TBSCl) and imidazole to give silyl compound 2 in quantitative yield. N-Methylation of 2 was carried out by using 60% NaH and methyl iodide to afford N-methylated compound in 57% yield. Deprotection of the t-butyldimethylsilyl group of the N-methyl compound with 1 N-HCl-MeOH afforded (S)-4hydroxy-1-methyl-2-oxopyrrolidine (3) in 97% yield. Mesylation of 3 with methanesulfonyl chloride (MsCl) followed by thioacetylation with potassium thioacetate (AcSK) afforded thioacetate (4) in 67% yield. Hydrolysis of the thioacetyl group of 4 afforded mercaptan 5 in quantitative yield. Condensation of 1β -methylcarbapenem-2-yl diphenylphosphate (9) with 5 gave the carbapenem 10a

Scheme 1.

Reagents: (a) TBSCl, imidazole; (b) NaH, then CH₃I; (c) 1 N-HCl; (d) MsCl, Et₃N; (e) AcSK; (f) 1 N-NaOMe; (g) Ph₃P, DEAD, 4-nitrobenzoic acid; (h) K₂CO₃.

Scheme 2.

Reagents: (a) 5 or 8, (iPr)₂NEt; (b) H₂, 10% Pd-C; (c) ICH₂OCOC(CH₃)₃.

in 75% yield. Deprotection of the 4-nitrobenzyl (PNB) group of **10a** was carried out by hydrogenation in the presence of 10% Pd-C. Purification of the product by reversed phase column chromatography and lyophilization afforded the parent carbapenem **11a** in 80% yield. Esterification of **11a** with pivaloyloxymethyl iodide in *N*, *N*-dimethylformamide furnished carbapenem ester **12a** in 94% yield. Alternatively, (*R*)-4-hydroxy-1-methyl-2-oxopyrrolidine (**6**) was prepared in 59% yield from **3** by Mitsunobu reaction with triphenylphosphine, dietyl azodicarboxylate (DEAD) and 4-nitrobenzoic acid followed by ester hydrolysis. Optically active (*S*)-3-mercapto-1-methyl-5-oxopyrrolidine (**8**) was prepared *via* **7** from **6** in a

similar manner to **5**. The carbapenem **11b** was synthesized by condensation of **9** with **8** followed by deprotection of the PNB group. Esterification of **11b** with pivaloyloxymethyl iodide afforded the carbapenem ester **12b** in the same way.

Biological Properties

The antimicrobial activities (MICs) of the parent carbapenems 11a and 11b are shown in Table 1. The activities of both carbapenems are compared with that of cefdinir which is a representative oral cephalosporins. The carbapenems 11a and 11b showed a particularly broad spectrum and more potent activity than cefdinir against

Table 1. Antibacterial activity (MIC, μ g/ml)^a of carbapenems 11a and 11b, and cefdinir.

Organism	11a	11b	Cefdinir
Staphylococcus aureus 209P	0.05	0.05	0.02
S. aureus 56R	0.05	0.05	0.05
S. aureus 535 (MRSA)	12.5	6.2	100
Enterococcus faecalis 681	1.5	1.5	3.1
Escherichia coli NIHJ	≤0.01	≤0.01	0.2
E, coli 609	0.05	0.05	0.4
Salmonella enteritidis	≤0.01	0.02	0.2
Klebsiella pneumoniae 806	≤0.01	≤0.01	0.1
K. pneumoniae 846	0.1	0.05	3.1
Enterobacter cloacae 903	0.8	0.8	3.1
Serratia marcescens 1184	0.02	0.02	0.2
Proteus vulgaris 1420	0.05	0.05	0.1
Morganella morganii 1510	0.2	0.2	12.5
Pseudomonas aeruginosa 1001	25	50	>200

^a MIC was determined by the agar dilution method with an inoculum of 10⁷ cfu/ml.

Table 2. Pharmacokinetic parameters of oral carbapenems 12a and 12b in mice.

	12a	12ь	
dose (administration route)	50 mg/kg (po) ^a	50 mg/kg (po) ^a	
Cmax (µg/ml) ^b	24.60	33.5	
Tmax (hour) ^c	0.50	0.50	
$T_{1/2}(hour)^d$	1.08	1.01	
AUC (μg·h/ml) ^e	23.37	31.26	
Absolute bioavailability (%) ^f	44.95	55.34	

^a Esters 12a and 12b were orally administered at a dose of 50 mg/kg as 11a and 11b, respectively.

Gram-positive and -negative bacteria. Both the carbapenems showed moderately potent activity against methicillin-resistant *S. aureus* (MRSA) and *P. aeruginosa*, which are resistant to cefdinir. The antibacterial activity of

both the carbapenems against MRSA and P. aeruginosa was only slightly affected by the stereochemistry at the 3-position of the oxopyrrolidine side chain. The (R)-isomer 11a showed slightly inferior activity against MRSA, but

^b Maximum plasma concentration.

^c Time at which Cmax is achieved.

^d Harmonic mean apparent terminal disposition half-life.

^e The area under the concentration-time curve.

^f Absolute oral bioavailability assuming linear pharmacokinetics = $[(AUC/dose)_{po} / (AUC/dose)_{iv}] \times 100$. The AUCs of **11a** and **11b** were 52.01 and 60.57 µg·h/ml, respectively, after intravenous administration (50 mg/kg).

Table 3. Protective effect of oral carbapenems **12a** and **12b**, and cefdinir against experimental infection in mice.

Organism		ED ₅₀ (mg/kg) ^a	
	12a	12b	Cefdinir
S. aureus Smith ^b	1.35	1.21	5.94
E. coli 704	0.70	0.46	14.1
K. pneumoniae 866 ^b	1.06	2.01	9.46

^a 50% effective po dose.

showed slightly more potent activity against P. aeruginosa. In order to estimate possible differences in oral absorption, the pharmacokinetics in mice were compared with each other (Table 2). The carbapenems 12a and 12b showed good oral bioavailability and prolonged half-lives in the parent carboxylic acids which were 1.08 and 1.01 hours, respectively. These values are longer than that of cefdinir; 0.39 hours in mice as previously reported. 14) The Cmax, AUC and absolute bioavailablity of 12b were slightly better than those of 12a. In order to clarify advantages in the in vivo activity, the protective effects of 12a and 12b against experimental infections in mice were compared with those of cefdinir as shown in Table 3. Although a significant difference of in vivo activity between 12a and 12b was not observed, both the carbapenems showed good protection against S. aureus Smith, E. coli 704 and K. pneumoniae 866. These carbapenems exhibited approximately 4 to 30 times greater in vivo efficacy against these 3 organisms compared with cefdinir. Clearly, these protective effects reflect the pharmakokinetics of both carbapenems.

Conclusion

New oral active 1β -methylcarbapenems with (R)- and (S)-1-methyl-5-oxopyrrolidin-3-ylthio moieties at the 2-position were synthesized. As expected, the parent carbapenems **11a** and **11b** showed more potent *in vitro* activity against Gram-positive and Gram-negative bacteria when compared with cefdinir. The respective carbapenem esters **12a** and **12b** showed prolonged half-lives after oral

administration in mice. Also, both the carbapenems had higher *in vivo* potency against *S. aureus* Smith, *E. coli* 704 and *K. pneumoniae* 866 compared with cefdinir. On the whole, both carbapenems are superior to cefdinir, but no major difference of *in vitro* and *in vivo* activities between 12a and 12b based on the stereochemistry of (*R*)- and (*S*)-1-methyl-5-oxopyrrolidin-3-ylthio group was observed. Consequently, the pharmacokinetic evaluation of both carbapenems using other experimental animals are of interest in future research.

Experimental

General Methods

IR spectra were recorded on a Nicolet NIC FT-IR (5SXC) spectrometer. NMR spectra were determined on a Jeol GX-270 (270 MHz) or GX-400 (400 MHz) spectrometer using tetramethylsilane (TMS) or sodium 3-(trimethylsilyl)-propionate- d_4 (TSP) as an internal standard. The mp was determined using a Yanagimoto micro-melting point apparatus and was not corrected. Optical rotations were obtained with a Jasco DIP-370 polarimeter. UV spectra were recorded on a Shimadzu UV-3100 spectrometer. Column chromatography was carried out on Silica gel 60 (230~400 mesh, Art. 9385, Merck), Cosmosil 75C₁₈ PREP (75 μ m, Nacalai Tesque, Inc.).

^b Challenged with 5% mucin.

Preparation of (R)-3-Mercapto-1-methyl-5-oxopyrrolidine (5)

i) (S)-4-t-Buthyldimethylsilyloxy-2-oxopyrrolidine (2)

To a solution of (*S*)-4-hydroxy-2-oxopyrrolidine **1** (10.1 g, 0.1 mol) in DMF (150 ml) were added imidazole (10.2 g, 0.15 mol) and *t*-butyldimethylsilyl chloride (18.1 g, 0.12 mol) at room temperature, and then the mixture was stirred for 3 hours. The mixture was diluted with EtOAc and washed with water and brine. The EtOAc layer was dried (Na₂SO₄) and concentrated by evaporation under reduced pressure to give compound **2** as a colorless powder (22.28 g): ¹H NMR (270 MHz, CDCl₃, TMS) δ 0.04 (3H, s), 0.08 (3H, s), 0.12 (9H, s), 2.20~2.36 (1H, m), 2.48~2.64 (1H, m), 3.18~3.32 (1H, m), 3.55~3.67 (1H, m), 4.51~4.65 (1H, m), 5.75 (1H, br s).

ii) (S)-4-Hydroxy-1-methyl-2-oxopyrrolidine (3)

To a solution of 2 (4.31 g, 20 mmol) in DMF (45 ml) was added sodium hydride (0.96 g, 24 mmol, 60% w/w dispersion in mineral oil) under ice-cooling, and then the mixture was stirred at the same temperature for 30 minutes. To this mixture was added methyl iodide (2.49 ml) under ice-cooling and then the mixture was stirred at room temperture for 4 hours. The mixture was poured into icewater and extracted with EtOAc. The extract was washed with brine, dried (MgSO₄), and concentrated by evaporation under reduced pressure. The residue was purified by silica gel column chromatography (hexane - EtOAc, 1:4) to give (S)-4-t-buthyldimethylsilyloxy-1-methyl-2-oxopyrrolidine as a pale-brown powder (2.6 g, 57% from 1): ¹H NMR (270 MHz, CDCl₃, TMS) δ 0.07 (6H, s), 0.88 (9H, s), 2.33 (1H, dd, J=17.2, 4.0 Hz), 2.61 (1H, dd, J=17.2, 7.3 Hz), 2.85 (3H, s), 3.23 (1H, dd, J=10.5, 3.0 Hz), 3.58 (1H, dd, $J=10.5, 6.6 \text{ Hz}), 4.42\sim4.52 \text{ (1H, m)}.$

To a solution of (*S*)-4-*t*-buthyldimethylsilyloxy-1-methyl-2-oxopyrrolidine (9.52 g, 41.5 mmol) in MeOH (55 ml) was added 1 N hydrochloric acid (33.2 ml, 33.2 mmol) under ice-cooling, and the mixture was stirred at room temperature for 15 minutes. To the mixture was added NaHCO₃ (3.53 g, 42 mmol) and concentrated by evaporation under reduced pressure and dried *in vacuo*. The residue was purified by silica gel column chromatography (CH₂Cl₂ - EtOH, 9:1) to afford the title compound 3 as colorless crystals (4.64 g, 97.0%): mp <50°C; $[\alpha]_D^{24} = -34.0^\circ$ (c = 0.67, H₂O); IR (KBr) cm⁻¹ 1681, 1494, 1471, 1460, 1441, 1417, 1407, 1382, 1361, 1319, 1298, 1269, 1253, 1219, 1171, 1112; ¹H NMR (270 MHz, D₂O, TSP) δ 2.32 (1H, dd, J = 18.1 Hz), 2.82 (1H, dd, J = 18.1, 6.6 Hz), 2.85 (3H, S), 3.36 (1H, dd, J = 11.5, 5.3 Hz), 4.54 (1H, m).

iii) (R)-3-Acetylthio-1-methyl-5-oxopyrrolidine (4)

To a solution of 3 (4.6 g, 40 mmol) in CH₂Cl₂ (90 ml) were added methane sulfonyl chloride (3.71 ml, 48 mmol) and triethyl amine (6.65 ml, 48 mmol) under ice-cooling, and then the mixture was stirred at the same temperature for 40 minutes. After addition of saturated aqueous NaHCO₃ (2 ml), the mixture was concentrated by evaporation under reduced pressure. To the residue was added CH₂Cl₂-AcOEt (1:1, 250 ml) which was then filtered. The filtrate was concentrated by evaporation under reduced pressure. The residue was purified by silica gel column chromatography (CH₂Cl₂-EtOAc-MeOH, 47:47:6) to give (S)-4-methanesulfonyloxy-1-methyl-2oxopyrrolidine as a colorless powder (7.45 g, 96.5%): IR (KBr) cm⁻¹ 1702, 1683, 1498, 1460, 1439, 1404, 1380, 1337, 1306, 1263, 1214, 1182, 1173, 1161, 1104; ¹H NMR (270 MHz, CDCl₃, TMS) δ 2.65 (1H, dd, J=17.8, 2.6 Hz), 2.82 (1H, dd, J=17.8, 7.9 Hz), 2.90 (3H, s), 3.07 (3H, s), 3.64 (1H, dd, J=11.9, 2.0 Hz), 3.80 (1H, 11.9, 5.9 Hz), 5.30~5.37 (1H, m).

To a solution of (*S*)-4-methanesulfonyloxy-1-methyl-2-oxopyrrolidine (7.45 g, 39 mmol) in CH₃CN (260 ml) was added potassium thioacetate (6.16 g, 54 mmol) at room temperature, and the mixture was refluxed for 4.5 hours. The mixture was diluted with AcOEt (160 ml) and filtered. The filtrate was concentrated by evaporation under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc) to give the title compound **4** as a pale-brown oil (5.83 g, 87.3%): IR (liquid film) cm⁻¹ 1683, 1560, 1499, 1440, 1404, 1301, 1264, 1199, 1110; ¹H NMR (270 MHz, CDCl₃, TMS) δ 2.35 (3H, s), 2.35 (1H, dd, J=17.2, 6.6 Hz), 2.85 (1H, dd, J=17.2, 5.9 Hz), 2.86 (3H, s), 3.29 (1H, dd, J=10.6, 5.3 Hz), 3.88 (1H, dd, J=10.6, 7.3 Hz), 4.02~4.13 (1H, m).

iv) (R)-3-Mercapto-1-methyl-5-oxopyrrolidine (5)

To a solution of 4 (5.80 g, 33.5 mmol) in MeOH (60 ml) was added 28% sodium methoxide in MeOH (6.8 ml, 33.5 mmol) for 5 minutes under ice-cooling, and the mixture was stirred at the same temperature for 10 minutes. To this solution was added 1 N hydrochloric acid (35.5 ml, 33.5 mmol) and the mixture was concentrated by evaporation under reduced pressure and dried *in vacuo*. To the residue was added AcOEt (200 ml) which was then filtered. The filtrate was concentrated by evaporation under reduced pressure and dried to give the title mercaptan 5 as a pale-brown oil (4.13 g, 94.1%): IR (liquid film) cm⁻¹ 1690, 1499, 1435, 1403, 1357, 1302, 1264, 1127; ¹H NMR (270 MHz, CDCl₃, TMS) δ 1.93 (1H, d, J=6.6 Hz), 2.36 (1H, dd, J=17.2, 6.6 Hz), 2.75~2.29 (1H, m), 2.87 (3H, s),

3.29 (1H, dd, J=10.6, 5.3 Hz), 3.50 \sim 3.66 (1H, m), 3.77 (1H, dd, J=10.6, 7.3 Hz).

Preparation of (S)-3-Mercapto-1-methyl-5-oxopyrrolidine (8)

i) (R)-4-Hydroxy-1-methyl-2-oxopyrrolidine (6)

To a solution of tripenylphosphine (11.4 g, 43.4 mmol) in dry THF (120 ml) was added diethyl azodicarboxylate (6.83 ml) under ice-cooling and then nitrobenzoic acid was added. The mixture was stirred at the same temperature for 1 hour and was cooled to $-25\sim-20^{\circ}$ C. To this mixture was added 3 (2.5 g, 21.7 mmol) in THF (50 ml) at the same temperature, and then the mixture was warmed to room temperature and was allowed to stand overnight. The mixture was concentrated by evaporation under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc-MeOH, 9:1) to give (R)-4-(4nitrobenzoyloxy)-1-methyl-2-oxopyrrolidine as a colorless powder (3.95 g, 68.9%): IR (KBr) cm⁻¹ 1722, 1687, 1645, 1611, 1600, 1526, 1492, 1438, 1406, 1399, 1353, 1321, 1307, 1271, 1222, 1183, 1122, 1109; ¹H NMR (270 MHz, CDCl₃, TMS) δ 2.66 (1H, dd, J=17.8, 1.9 Hz), 2.92 (1H, dd, J=17.8, 6.3 Hz), 2.93 (3H, S), 3.54 (1H, dd, J=11.8, 1.9 Hz), 3.91 (1H, dd, J=11.8, 6.1 Hz), 5.57~5.66 (1H, m), 8.21 (2H, d, J=9.1 Hz), 8.31 (2H, d, J=9.1 Hz).

To a solution of (R)-4-(4-nitrobenzoyloxy)-1-methyl-2oxopyrrolidine (5.1 g, 19.3 mmol) in MeOH (100 ml) was added K₂CO₃ (0.7 g, 5.1 mmol) under ice-cooling and the mixture was stirred at the same temperature for 20 minutes. To the mixture was added ammonium chloride (1.1 g, 20.5 mmol) and the mixture was stirred for 5 minutes and filtered. The filtrate was concentrated by evaporation under reduced pressure and the residue was purified by silica gel column chromatography (EtOAc-MeOH, 9:1) to afford the title compound 6 as an oil (1.90 g): $\left[\alpha\right]_{D}^{24} = +34.4^{\circ}$ $(c=1.23, H_2O)$, IR (liquid) cm⁻¹ 1667, 1503, 1446, 1406, 1339, 1305, 1267, 1220, 1112; ¹H NMR (270 MHz, CDCl₃, TMS) δ 2.37 (1H, dd, J=17.2, 2.3 Hz), 2.69 (1H, dd, J=17.2, 6.6 Hz), 2.87 (3H, s), 3.03 (1H, br s), 3.32 (1H, dd, J=10.9, 2.0 Hz), 3.66 (1H, dd, J=10.9, 5.7 Hz), 4.48~4.57 (1H, m).

ii) (S)-3-Mercapto-1-methyl-5-oxopyrrolidine (10)

The title compound 8 was prepared as an oil from 6 by a similar manner as that described for the preparation of 5.

Synthesis of Pivaloyloxymethyl (1R,5S,6S)-6-[(R)-1-Hydoxyethyl]-1-methyl-2-[(R)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (12a)

i) 4-Nitrobenzyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydoxyethyl]-1-methyl-2-[(*R*)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**10a**)

To a solution of 4-nitrobenzyl (1R,5S,6S)-2-diphenylphosphoryloxy-6-[(R)-1-hydoxyethyl]-1-methyl-1carbapenam-3-carboxylate (18.58 g, 31.3 mmol)) acetonitrile (260 ml) were added a solution of 3 in acetonitrile (20 ml) and N,N-diisopropylethylamine (5.46 ml) under ice-cooling and the mixture was stirred for 30 minutes and was allowed to stand in a refrigerator overnight. The mixture was concentrated by evaporation under reduced pressure and to the residue were added water (100 ml) and acetonitrile (150 ml). The precipitated crystals were collected by filtration, washed with acetonitrile - water (9:1), and dried to give the title compound 11a as palebrown crystals (12.8 g, 86.0%): mp 169°C; IR (KBr) cm⁻¹ 1772, 1702, 1692, 1667, 1608, 1546, 1518, 1501, 1467, 1455, 1420, 1410, 1401, 1383, 1342, 1322, 1308, 1281, 1267, 1256, 1229, 1206, 1183, 1138, 1101; ¹H NMR (400 MHz, DMSO- d_6 , TMS) δ 1.16 (3H, d, J=6.5 Hz), 1.17 (3H, d, J=7.5 Hz), 2.24 (1H, dd, J=17.1, 3.8 Hz), 2.74 (3H, s), 2.89 (1H, dd, J=17.1, 7.9 Hz), 3.22 (1H, dd, J=11.0, 3.7 Hz), 3.29 (1H, dd, J=6.3, 2.6 Hz), 3.42~3.50 (1H, m), 3.81 (1H, dd, J=11.0, 7.5 Hz), 3.94~4.05 (2H, m), 4.25 (1H, dd, J=9.5, 2.7 Hz), 5.09 (1H, d, J=5.1 Hz), 5.29, 5.45 (2H, AB, $J=14.0\,\mathrm{Hz}$), 7.71 (2H, d, $J=8.7\,\mathrm{Hz}$), 8.23 (2H, d, J=8.7 Hz).

ii) Sodium (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydoxyethyl]-1-methyl-2-[(*R*)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**11a**)

To a solution of 10a (6.18 g, 13 mmol) in THF (240 ml) and aqueous NaHCO₃ (1.09 g, 13 mmol; water 120 ml) was added 7.5% Pd-charcoal (10 g, wet, 52.4% water, Kawaken Fine Chemicals Co., Ltd.) and the mixture was stirred rigorously at 30°C in an atomosphere of hydrogen for 2 hours. The catalyst was removed by filtration from the reaction mixture, and the filtrate was washed twice with diethyl ether. The aqueous layer was then concentrated by evaporation under reduced pressure and the residue was purified by a reverse phase column chromatography (Nacalai Tesque, Inc., Cosmosil $75C_{18}$ -PREP, water). The desired fraction was concentrated and then lyophilized to give the title compound 11a as a colorless powder (3.46 g, 73.4%):

UV (H₂O) λ_{max} nm 300; IR (KBr) cm⁻¹ 1752, 1678, 1603, 1504, 1450, 1398, 1306, 1267, 1226, 1183, 1147, 1110; ¹H NMR (270 MHz, D₂O, TSP) δ 1.22 (3H, d, J= 7.3 Hz), 1.30 (3H, d, J=6.6 Hz), 2.49 (1H, dd, J=17.8, 3.3 Hz), 2.86 (3H, s), 3.02 (1H, dd, J=17.8, 7.6 Hz), 3.28 \sim 3.47 (3H, m), 3.92 \sim 4.05 (2H, m), 4.21 \sim 4.30 (2H, m).

iii) Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydoxyethyl]-1-methyl-2-[(*R*)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**12a**)

To a solution of **11a** (2.3 g, 6.34 mmol) in *N*,*N*-dimethylacetamide (25 ml) was added pivaloyloxymethyl iodide (1.14 ml, 7.61 mmol) under ice-cooling, and then the mixture was stirred for 1 hour. The mixture was diluted with AcOEt (200 ml) and was washed with 2% sodium thiosulfate, water and brine. The organic layer was dried (MgSO₄) and was concentrated by evaporation under reduced pressure. Crystals were formed during the evaporation. The resulting crystals were collected, filtered, and dried to give the title compound **12a** (2.70 g, 93.7% yield): mp 192°C;

Anal Calcd for $C_{21}H_{30}N_2O_7S$: C 55.49, H 6.65, N 6.16, S 7.05. Found: C 55.93, H 6.57, N 6.21, S 6.96.

[α]_D²³=+24.6° (c=1.0, MeOH); UV (CH₃CN) λ _{max} nm 323; IR (KBr) cm⁻¹ 1787, 1757, 1709, 1664; ¹H NMR (400 MHz, DMSO-d₆, TMS) δ 1.13~1.17 (15H, m), 2.22 (1H, dd, J=17.1, 3.9 Hz), 2.73 (3H, s), 2.88 (1H, dd, J=17.1, 7.9 Hz), 3.20 (1H, dd, J=11.0, 3.9 Hz), 3.26 (1H, dd, J=6.4, 2.7 Hz), 3.41~3.49 (1H, m), 3.81 (1H, dd, J=11.0, 7.8 Hz), 3.94~4.05 (2H, m), 4.23 (1H, dd, J=9.5, 2.7 Hz), 5.08 (1H, d, J=5.1 Hz), 5.71 (1H, d, J=5.9 Hz).

Synthesis of Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydoxyethyl]-1-methyl-2-[(*S*)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (12b)

i) 4-Nitrobenzyl (1R,5S,6S)-6-[(R)-1-hydoxyethyl]-1-methyl-2-[(S)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**10b**)

The title compound **10b** was prepared in 85% yield as a powder from **8** by a similar manner as that described for the preparation of **10a**: ¹H NMR (400 MHz, DMSO- d_6 , TMS) δ 1.16 (3H, d, J=6.4 Hz), 1.18 (3H, d, J=7.6 Hz), 2.14 (1H, dd, J=17.2, 4.7 Hz), 2.71 (3H, s), 3.28 \sim 3.31 (2H, m), 3.41 \sim 3.48 (1H, m), 3.85 (1H, dd, J=10.4, 6.7 Hz), 3.96 \sim 4.06 (2H, m), 4.24 (1H, dd, J=9.3, 2.9 Hz), 5.09 (1H, d, J=5.2 Hz), 5.29, 5.46 (2H, AB, J=14.1 Hz), 7.72 (2H, d, J=

 $8.7 \,\mathrm{Hz}$), $8.23 \,(2\mathrm{H}, \,\mathrm{d}, \,J = 8.7 \,\mathrm{Hz})$.

ii) Sodium (1R,5S,6S)-6-[(R)-1-Hydoxyethyl]-1-methyl-2-[(S)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (11b)

The title compound 11b was prepared in 80% yield as a powder from 10b by the similar manner as that described for the preparation of 11a:

Anal Calcd for C₁₅H₁₉N₂O₅SNa · H₂O:

C 47.36, H 5.57, N 7.36, S 8.43.

Found: C 46.84, H 5.71, N 7.24, S 8.45.

UV ($\rm H_2O$) $\lambda_{\rm max}$ nm 300; IR (KBr) cm⁻¹ 1744, 1673, 1640, 1597, 1553, 1504, 1451, 1403, 1309, 1266, 1230, 1183, 1165, 1149, 1111; ¹H NMR (270 MHz, $\rm D_2O$, TPS) δ 1.23 (3H, d, $\it J$ =7.3 Hz), 1.30 (3H, d, $\it J$ =6.6 Hz), 2.37 (1H, dd, $\it J$ =17.2, 3.3 Hz), 2.84 (3H, s), 2.99 (1H, dd, $\it J$ =17.2, 8.3 Hz), 3.27~3.49 (3H, m), 3.92~4.03 (2H, m), 4.21~4.31 (m).

iii) Pivaloyloxymethyl (1*R*,5*S*,6*S*)-6-[(*R*)-1-Hydoxyethyl]-1-methyl-2-[(*S*)-1-methyl-5-oxopyrrolidin-3-ylthio]-1-carbapen-2-em-3-carboxylate (**12b**)

The title compound 12b was prepared in 80% yield as a powder from 11b by a similar manner as that described for the preparation of 12a: mp 137°C;

Anal Calcd for C₂₁H₃₀N₂O₇S: C 55.49, H 6.65, N 6.16, S 7.05. Found: C 55.11, H 6.66, N 5.99, S 6.96.

 $[\alpha]_{\rm D}^{23} = -3.4^{\circ} \ (c=1.0, {\rm MeOH}); \ {\rm UV} \ ({\rm CH_3CN}) \ \lambda_{\rm max} \ {\rm nm}$ 323; IR (KBr) cm⁻¹ 1784, 1754, 1725, 1685, 1667; $^{1}{\rm H}$ NMR (400 MHz, DMSO- d_6 , TMS) δ 1.14~1.18 (15H, m), 2.12 (1H, dd, J=17.3, 4.7 Hz), 2.70 (3H, s), 2.81 (1H, dd, J=17.3, 8.8 Hz), 3.24~3.28 (2H, m), 3.41~3.48 (1H, m), 3.85 (1H, dd, J=10.6, 6.7 Hz), 3.94~4.02 (2H, m), 4.22 (1H, dd, J=9.5, 2.6 Hz), 5.08 (1H, d, J=5.2 Hz), 5.73 (1H, d, J=5.9 Hz), 5.88 (1H, d, J=5.9 Hz).

Measurement of Antibacterial Activity

MICs were measured on Nutrient agar (Eiken Chemical Co., Ltd.) by the two-fold dilution method. The inoculum size of the bacteria was one-loopful of 10^7 cfu/ml.

Pharmacokinetics

Plasma levels of carbapenems 11a and 11b were determined by HPLC, after oral administration of 12a and 12b (50 mg/kg as 11a and 11b, respectively) as a 0.2 ml suspension of 0.5% tragacanth or intravenous administration of 11a and 11b (50 mg/kg, respectively) in 0.2 ml saline to SPF male ddY mice (n=3).

Therapeutic Effect on Systemic Infection in Mice

Overnight cultures of organisms grown at 37° C in Trypto-soy broth (Eiken Chemical Co., Ltd.) were diluted according to their virulence. The diluted cultures, if necessary, were mixed with the same amount of gastric mucin (Tokyo Kasei Kogyo Co., Ltd.). Seven male SPF ddY mice in each group were infected intraperitoneally with 0.2 ml portions of these bacterial mixtures. β -Lactam antibiotics (12a, 12b and cefdinir) were administered orally as a 0.2 ml suspension of 0.5% sodium carboxymethyl cellose (Kanto Chemical Co., Inc.) immediately and 4 hours after infection. The ED₅₀s of the mice were calculated by the probit method from the survival rates on the 5th day after infection.

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References

- KAHAN, J. S.; F. M. KAHAN, R. GOEGELMAN, S. A. CURRIE, M. JCKSON, E. O. STAPLEY, T. W. MILLER, A. K. MILLER, D. HENDLIN, S. MOCHALES, S. HERNANDEZ, H. B. WOODRUFF & J. BIRNBAUM: Thienamycin, a new β-lactam antibiotic. I. Discovery, taxonomy, isolation and physical properties. J. Antibiotics 32: 1~12, 1979
- KROPP, H.; J. G. SUNDELOF, R. HAJDU & F. M. KAHAN: Metabolism of thienamycin and related carbapenem antibiotics by the renal dipeptidase, dehydropeptidase-I. Antimicrob. Agents Chemother. 22: 62~70, 1982
- LEANZA, W. J.; K. J. WILDONGER, T. W. MILLER & B. G. CHRISTENSEN: N-Acetimidoyl and N-formimidoylthienamycin derivatives: Antipseudomonal β-lactam antibiotics. J. Med. Chem. 22: 1435~1436, 1979
- 4) BIRNBAUM, J.: F. M. KAHAN, H. KROPP & J. S. MACDONALD: Carbapenems, a new class of betalactam antibiotics: Discovery and development of

- imipenem/cilastatin. Am. J. Med. 78 (Suppl. 6A): $3\sim21$, 1985
- MIYADERA, T.; Y. SUGIMURA, T. HASHIMOTO, T. TANAKA, K. IINO, T. SHIBATA & S. SUGAWARA: Synthesis and in vitro activity of a new carbapenem, RS-533. J. Antibiotics 36: 1034~1039, 1983
- 6) INOUE, K.; Y. HAMANA & S. MITSUHASHI: Antibacterial activity of panipenem, a new carbapenem antibiotic. Chemotherapy (Tokyo) 39 (Suppl. 3): 1~13, 1991
- 7) NAGANUMA, H.; H. TOKIWA, Y. HIROUCHI, Y. KAWAHARA, J. FUKUSHIGE, M. FUKAMI, K. HIROTA, S. MURAMATSU, H. TAKAHAGI, K. INUI, Y. TANIGAWARA, M. YASUHARA, R. HORI & S. KUWAHARA: Nephroprotective effect and its mechanism of betamipron (1): Relationships of renal transport. Chemotherapy (Tokyo) 39 (Suppl. 3): 166~177, 1991
- 8) SUNAGAWA, M.; H. MATSUMURA, T. INOUE, M. FUKASAWA & M. KATO: A novel carbapenem antibiotic, SM-7338: Structure-activity relationships. J. Antibiotics 43: 519~532, 1990
- 9) KAWAMOTO, I.: 1β -Methylcarbapenem antibiotics. Drugs Fut. 23: $181 \sim 189$, 1998
- 10) Fujimoto, K.; S. Ishihara, H. Yanagisawa, J. Ide, E. Nakayama, H. Nakao, S. Sugawara & M. Iwata: Studies on orally active cephalosporin esters. J. Antibiotics 40: 370~384, 1987
- 11) MINE, Y.; T. KAMIMURA, H. SAKAMOTO, S. TAWARA, K. HATANO, Y. WATANABE & S. KUWAHARA: *In vitro* antibacterial activity of cefdinir, a new orally active cephalosporin. Chemotherapy (Tokyo) 37 (Suppl. 2): 100~121, 1989
- 12) MIYAUCHI, M.; R. ENDO, M. HISAOKA, H. YASUDA & I. KAWAMOTO: Synthesis and structure-activity relationships of a novel oral carbapenem, CS-834. J. Antibiotics 50: 429~439, 1997
- 13) MIYAUCHI, M.; O. KANNO & I. KAWAMOTO: A novel oral carbapenem CS-834: Chemical stability of pivaloyloxymethyl esters of carbapenems and cephalosporins in phosphate buffer solution. J. Antibiotics 50: 794~796, 1997
- 14) SAKAMOTO, H.; T. HIROSE, S. NAKAMOTO, K. HATANO, Y. MINE & S. KUWAHARA: Pharmacokinetics of cefdinir, a new oral cephalosporin, in experimental animals. Chemotherapy (Tokyo) 37 (Suppl. 2): 165~178, 1989