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# **CONCISE ARTICLE**

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Cite this: DOI: 10.1039/ c4md00430b A solid- and solution-phase-based library of 2β-methyl substituted penicillin derivatives and their effects on growth inhibition of tumor cell lines†

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Received 25th September 2014, Accepted 15th December 2014

DOI: 10.1039/c4md00430b

www.rsc.org/medchemcomm

We described the design, synthesis and antiproliferative properties of a series of twenty  $2\beta$ -methyl substituted penicillin derivatives. This analysis includes evaluation against HeLa and MCF-7 human tumor cell lines and LM3 and B16-F0 murine tumor cell lines. The epithelial cell line derived from the normal mammary gland of mice (NMuMG) and the mouse embryo fibroblast cell line (3T3) were used as controls (non-cancer cells).

#### Introduction

The β-lactam ring is an active agent whose efficiency has been proven in a large number of clinically significant therapeutic areas. Apart from being the key structural element of the most broadly prescribed antibiotics, such as penicillins, cephalosporins, carbapenems and monobactams (Fig. 1),2 β-lactams have offered a new panorama with the development of some derivatives as potent cholesterol absorption inhibitors, including commercial ezetimibe (Fig. 1).3 The β-lactam skeleton is also found in prostate specific antigen, thrombin,<sup>4</sup> human cytomegalovirus protein,5 human leukocyte elastase,6 cysteine protease,7 and human fatty acid amide hydrolase8 inhibitors. This class of compounds has also been considered as peptidomimetic species for mimicking certain properties of proteins.9 Their non-antibacterial uses and the constant need for new antibiotics to attack the problem of bacterial resistance to traditional drugs have maintained and even increased the interest in the chemistry of  $\beta$ -lactams. Last but not least,  $\beta$ -lactams are key synthons for the preparation of various heterocyclic compounds of biological importance. 10 For example, substituted hydroxy-β-lactams have been used in the semisynthesis of paclitaxel (Taxol) and related compounds from baccatin III.11

Interestingly, we and others have recently demonstrated that  $\beta$ -lactam derivatives can also be useful in the field of

antiproliferative chemotherapy. In this sense, N-thiolated, <sup>12</sup> 1,4-diaryl <sup>13</sup> 1,3,4-triaryl, <sup>14</sup> and polyaromatic <sup>15</sup> 2-azetidinones have been reported as active against cancer cells. Apart from being considered as prodrugs for the ADEPT strategy, <sup>16</sup> cephalosporin derivatives have also shown anticancer activity. <sup>17</sup> Penicillins are a major class of  $\beta$ -lactams and the oldest used as antibacterial agents. We have recently reported antiproliferative studies on a series of triazolyl aminoacyl(peptidyl) penicillins. <sup>18</sup> These hybrid compounds, prepared by azide–alkyne cycloaddition as the key reaction step, were evaluated against cancer cell lines, and several of them show high selectivity and cytotoxic activity.

As part of our efforts to discover new potential anti-cancer drugs, we have synthesized a solid and solution-phase-based

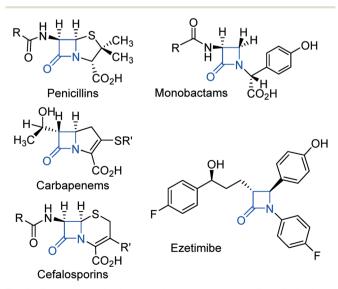


Fig. 1 The  $\beta\text{-lactam}$  ring is present in several commercially available drugs.

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 $<sup>\</sup>dagger$  Electronic supplementary information (ESI) available: Experimental procedures, characterisation data, description of cell lines, culture conditions and cell proliferation assay. See DOI: 10.1039/c4md00430b

library of  $2\beta$ -methyl substituted penicillins and evaluated their *in vitro* antiproliferative activity against four cancer cell lines and two non-cancer cells, and some conclusions on the preliminary structure–activity relationship have been drawn.

## Results and discussion

#### Chemistry

We have developed a convenient solid-phase synthetic sequence for the preparation of 2β-methyl substituted penam derivatives using the commercially available and cost-effective Merrifield resin. 19 The synthesis started with the immobilization of 6,6-dihalo- and 6α-halopenicillanic acids on the resin using standard methodology. Oxidation of the resin-bound penam sulfide (1, Scheme 1) under 1.5 equiv. of m-chloroperbenzoic acid (m-CPBA) leads to the sulfoxides (2). The key step for the generation of the 2β-methyl substituted penicillins is the thermal rearrangement of the corresponding sulfoxides. 20 For the generation of the 2β-chloromethyl penam derivatives, 6,6-dihalo- and  $6\alpha$ -halopenicillanate sulfoxides tethered to the Merrifield resin (2) were treated with 1.5 equiv. of 2-mercaptobenzothiazole (2-MBT) and heated in refluxing benzene, to give the unsymmetrical monobactam disulfides (3). Then, the penam system is reestablished by addition of sulfuryl chloride in dicloromethane at -40 °C, affording the immobilized 2β-chloromethyl penicillanates (4a-c).21 Final products were obtained by treatment with aluminum chloride<sup>21</sup> followed by esterification with diazomethane, to give the methyl 6,6-dihalo- and 6α-halo-2β-chloromethyl penicillanates (5b-c) in good overall isolated yield (see Table 1). Alternatively, sulfone 7a was obtained by oxidation of sulfide 4a with an excess of m-CPBA, followed by cleavage and methylation, according to the procedure described above.

In the case of the preparation of  $2\beta$ -(heterocyclyl)-thiomethyl penam derivatives, immobilized sulfoxides (2)

were treated with excess of heterocyclic thiols (HS-Het) and catalytic amounts of p-toluenesulfonic acid (p-TsOH), in refluxing toluene. <sup>22</sup> Under these conditions and in the absence of an external nucleophile, the initially formed disulfide intermediates (3e–k) underwent an internal rearrangement, assisted by p-TsOH, to yield the 2 $\beta$ -(heterocyclyl)thiomethyl penams (8ae–bk). After cleavage, esterification and purification by column chromatography, the 2 $\beta$ -(heterocyclyl)thiomethyl penicillins (9ae–bk) were isolated, in good overall yields for the entire synthetic sequence (see Table 1). When 9af was treated with tributylphosphine (Bu<sub>3</sub>P) in methanol, <sup>23</sup> the dehalogenated product 9df was obtained. On the other hand, sulfide 9bj was treated with excess of m-CPBA to give the corresponding sulfone 10bj in good yield.

Benzyl ester derivatives were synthesized by a solution-phase strategy (Scheme 2). The corresponding sulfoxide (11a) was treated with heterocyclic thiols (HS-Het) and p-TsOH, under the conditions described above. The benzyl 6,6-dibromo- $2\beta$ -(heterocyclyl)thiomethyl penicillanates (12) were reduced by Bu<sub>3</sub>P to give the dehalogenated derivatives (13de-dk). Otherwise, by treating 13de with excess of m-CPBA, sulfone 14de was obtained. Finally, compound 15a was synthesized from 11a according to a literature procedure. <sup>22</sup>

#### In vitro antiproliferative studies

The *in vitro* antiproliferative activity of a library of twenty  $2\beta$ -methyl substituted penicillin derivatives was initially evaluated at 20  $\mu$ M concentration against epithelial cells derived from the normal mammary gland of mice (NMuMG) and fibroblast cells from mouse embryo (3T3) (Fig. 2). Based on results obtained for these non-neoplastic cells, only those compounds without toxicity towards non-cancer cells (*i.e.*, those that inhibited less than 50% cell proliferation in either NMuMG cells or 3T3 cells at 20  $\mu$ M) were selected for

Scheme 1

#### **Table 1** Library of $2\beta$ -methyl substituted penicillins

Entry	Compound	Y	X	Z	R	n	Yield <sup>a</sup> (%)
1	5 <b>b</b>	Cl	Н	Cl	Me	0	54
2	5c	Br	Н	Cl	Me	0	48
3	7a	Br	Br	Cl	Me	2	37
4	9ae	Br	Br	's-\S	Me	0	52
5	9af	Br	Br	S—NPh	Me	0	70
6	9ag	Br	Br	-s-\(\bigvar_n\)	Me	0	43
7	9ah	Br	Br	N-N N	Me	0	60
8	9ai	Br	Br	/s—(H),N	Me	0	52
9	9be	Cl	Н	S—S	Me	0	40
10	9 <b>b</b> f	Cl	Н	S—Ph	Me	0	46
11	9bh	Cl	Н	-s N	Me	0	50
12	9bj	Cl	Н	s S C P	Me	0	44
13	9bk	Cl	Н	_s_s	Me	0	48
14	9df	Н	Н	S—Ph	Me	0	$32^b$
15	10bj	Cl	Н	s—S—DO-Et	Me	2	$34^c$
16	13de	Н	Н	s—s	Bn	0	$46^d$
17	13df	Н	Н	S—Ph	Bn	0	17 <sup>d</sup>
18	13dk	Н	Н	_ss	Bn	0	19 <sup>d</sup>
19	14de	Н	Н	's-\s\	Bn	2	11 <sup>e</sup>
20	15a	Br	Br	Cl	Bn	0	$32^d$

<sup>&</sup>lt;sup>a</sup> Obtained by solid-phase synthesis, unless otherwise stated. Yield after  $AlCl_3$  cleavage and esterification with  $CH_2N_2$ , and calculated on the basis of the manufacturer's loading of the Merrifield resin. <sup>b</sup> Yield from 9bf (solution-phase synthesis). <sup>c</sup> Yield from 9bj (solution-phase synthesis). <sup>d</sup> Yield from compound 11a (solution-phase synthesis).

Scheme 2

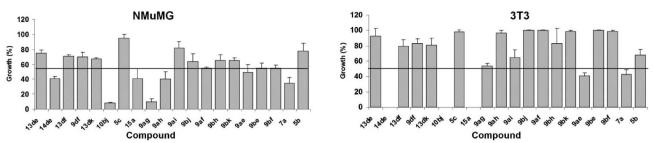


Fig. 2 Effect of synthetic penicillin derivatives on the proliferation of non-neoplastic cell lines.  $2 \times 10^4$  cells per well (NMuMG and 3T3) were incubated in the presence or absence of 20  $\mu$ M of different compounds for 72 h at 37 °C. Cell proliferation was determined by colorimetric determination of hexosaminidase levels.<sup>24</sup> Results are expressed as the percentage of growth obtained in the absence of compounds (control) and represented as mean  $\pm$  S.E.M. of three different experiments. The antiproliferative activity of compounds **14de**, **10bj** and **15a** was not evaluated in 3T3 cells.

further studies. Thus, compounds **14de**, **10bj**, **15a**, **9ag**, **9ah**, **9ae** and **7a** were not tested in tumor cells. The antiproliferative effect of the remaining selected compounds was then determined in two human tumor cell lines (cervix HeLa adenocarcinoma; breast MCF-7 cancer) and two murine cell lines (LM3 mammary adenocarcinoma; B16-F0 melanoma) (Fig. 3). As shown in Fig. 3, the compounds that caused a reduction of  $\geq$ 50% in the growth of tumor cell lines at 20  $\mu$ M

concentration were: 13de, 13df, 9df, 9bj, 9af, 9bk, 9be and 9bf (HeLa cells); 9df and 9af (MCF-7 cells); 9bj, 9af and 9be (B16-F0); 9df, 13dk, 9bj, 9af, 9bk and 9be (LM3).  $IC_{50}$  values, defined as compound concentrations that produce 50% growth inhibition, were determined from dose–response curves only for these derivatives and are summarized in Table 2. Consequently,  $IC_{50}$  values corresponding to compounds that inhibited less than 50% tumor cell proliferation at 20  $\mu$ M were

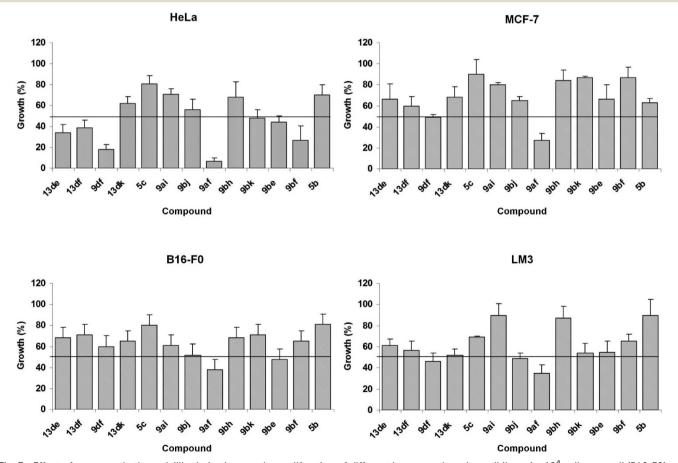


Fig. 3 Effect of some synthetic penicillin derivatives on the proliferation of different human and murine cell lines.  $1 \times 10^4$  cells per well (B16-F0) or  $2 \times 10^4$  cells per well (HeLa, MCF-7, LM3) were incubated in the presence or absence of 20  $\mu$ M of different compounds for 72 h at 37 °C. Cell proliferation was determined by colorimetric determination of hexosaminidase levels.<sup>24</sup> Results are expressed as the percentage of growth obtained in the absence of compounds (control) and represented as mean  $\pm$  S.E.M. of three different experiments.

Table 2 Molecular structures and antiproliferative effect of the compounds tested

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			× (O) ×	Z-	No	Normal cell lines growth (%)	lines	Tumoral	Tumoral cell lines growth (%)	growth (9	(%)	IC <sub>50</sub> valu	$\overline{{ m IC}_{50}}$ values <sup>a</sup> $(\mu M)$				$\mathrm{RP}^b$			
Compound	Y pr	×	Z 0,00-0-R	×	n NN	NMuMG	3T3	HeLa	MCF-7	B16-F0	LM3	$HeLa^d$	MCF-7	$ m B16-F0^d$	LM3	NMuMG	HeLa	MCF-7	B16-F0	LM3
- -	כ		5	Ме			0	70 + 10	63 + 4	81 + 7	90 + 15						E	CN.	E C	Į,
5c	B G		5 5	Me	0 95	95 ± 5		81 ± 8	$90 \pm 14$	-1 +1	69 ± 1	I		I	I	I	2 2	Q Q		2 R
7a	Br			Me				ND	ND	ND	ND	ND	ND	ND	ND	ND	N ON	ND	N Q	ND
9ae	Br	r Br	S S	Me			41 ± 4	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
9af	Br	r Br	y d	Me	0 55	± 11	100 ± 1	7 ± 3	27 ± 7	38 ± 8	35 ± 8	7 ± 2	18 ± 1	11 ± 3	14 ± 3	$31 \pm 1$	4.4	1.7	2.8	2.2
9ag	Br	r Br	£ ( £−z	Me	0 10	+ 4	54 ± 3	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
			Z   S   S																	
9ai	Br	r Br	Z Z	Me	0 82	& +I	65 ± 10	71 ± 5	80±2	61 ± 6	90 ± 11	I	I	I	I	I	ND	ND	NO	ND
9be	CI	Н	z S	Me	0 55	+ 7	$100 \pm 1$	44 ± 6	66 ± 14	$48 \pm 5$	$55 \pm 10$	$18 \pm 2$	1	18 ± 3	$20 \pm 2$	36 ± 6	2	ND	2	1.8
Jq6	ū	Н	y d	Me	0 55	+ 4	99 ± 1	27 ± 14	87 ± 10	65 ± 9	65 ± 7	13 ± 4	I	I	I	32 ± 8	2.5	ND	ND	ND
4q6	CI	Н	Z Z	Me	0 65	& +I	83 ± 20	68 ± 15	84 ± 10	68 ± 2	87 ± 11	I	I	I	I	I	ND	ND	ND	ND
9bj	C	Н	S OEF	it Me	0 64	64 ± 10	100 ± 11	$56 \pm 10$	65 ± 4	52 ± 9	49 ± 5	$24 \pm 6$	I	27 ± 8	24 ± 6	43 ± 4	1.8	ND	1.6	1.8
9bk	C	Н		Me	0 65	+ 4	99 ± 1	48 ± 8	87 ± 1	71 ± 6	54 ± 9	19 ± 4	I	I	$20 \pm 1$	41 ± 1	2.2	ND	N	2
Jp6	H	Н	<u>/</u> >=	Me	0 70 ±	9	83 ± 6	$18 \pm 5$	49 ± 3	60 ± 5	46 ± 8	8 ± 1	$20 \pm 1$	I	$20 \pm 1$	$35 \pm 5$	4.4	1.8	NO	1.8
10bj	CI	н	s s oet	et Me	2 8 +	1	ND	ND	ND	N	ND	ND	ND	ND	N Q	ND	N Q	ND	N Q	ND
13de	H	Н		Bn	0 75	+ 4	93 ± 10	34 ± 8	66 ± 15	68 ± 2	61 ± 6	11 ± 1	I	I	I	>80	7.3°	ND	ND	ND
13df	H	Н	N O N	Bn	0 71	+ 2	8 = 08	39 ± 7	6 = 09	71 ± 5	57 ± 8	$20 \pm 3$	I	I	I	>80	$4^c$	ND	ND	ND
13dk	H	Н	AP S S	Bn	0 67	+ 2 8	81 ± 9	62 ± 7	$68 \pm 10$	65 ± 5	$52 \pm 6$	I	I	I	23 ± 4	>80	ND	ND	N	$3.5^c$
14de	H	Н	N S S	Bn	2 41	±3 I	QN	ND	ND	ND	QN	ND	ND	QN	QN QN	ND	ND	ND	ND	ND
15a	Br	r Br	) CI	Bn	0 41	41 ± 13	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND
-		;	•	-				;			•	,		r	,		;	,		

The symbol (—) indicates the compounds that inhibited less than 50% cell growth at 20  $\mu$ M concentration. ND: not determined. 9ah was not tested in tumor cells (% growth, NMuMG:  $40 \pm 10$ ; 3T3:  $97 \pm 3$ ).  $^a$  The molar drug concentrations required to cause 50% growth inhibition (IC<sub>50</sub>) were determined from dose–response curves. Results are represented as means  $\pm$  SEM of at least three different experiments.  $^b$  The relative potency (RP) of a compound in a tumor cell line was calculated as the ratio of IC<sub>50</sub> NMuMG to IC<sub>50</sub> tumor cells.  $^c$  Minimum value.  $^d$  Reference standards: doxorubicin: 0.68 (HeLa), 8.05 (B16-BL6); cisplatin: 14.31 (HeLa).  $^{25}$ 

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not evaluated. In order to establish the selectivity of the tested compounds, we decided to compare the IC50 values obtained in tumor cells with those obtained in NMuMG cells, the noncancer cell line more susceptible to the cytotoxic effect of penicillin derivatives. Therefore, we defined the relative potency (RP) of each compound in a particular tumor cell line as the ratio of the IC50 obtained in NMuMG to the IC50 value obtained in tumor cells. The higher this value, the greater the antitumor potency and selectivity is exhibited by a compound (Table 2). Based on RP values obtained, the most potent and selective penicillin derivatives (RP ~4-7) were compound 13de followed by compounds 13df, 9df and 9af. Within this group, compounds 13de and 13df are selective for HeLa cells, although 13de showed a higher potency than 13df. Furthermore, compounds 9df and 9af, showing a similar RP to compound 13df, also exerted a slight antiproliferative effect in other tumor cell lines. In addition, compound 13dk was moderately effective in LM3 cells. It should be considered that since compounds 13de, 13df and 13dk could not be tested at concentrations higher than 80 µM due to their lower solubility in the culture medium, the RP values reported correspond to the minimal difference between the efficacy in NMuMG and tumor cells.

Compounds 9bj, 9bk, 9be and 9bf showed a slight and similar antitumor potency (RP ~2). However, although 9bj and 9be were effective in HeLa, B16-F0 and LM3 cells, 9bk was selective for HeLa and LM3 cells, and 9bf only for HeLa.

Compounds 5c, 9ai, 9bh and 5b, which were tested for being not cytotoxic in non-cancer cells (Fig. 2), showed no antiproliferative effect in the tumor cell lines herein studied.

#### Structure-activity relationships

Analysis of the data obtained in this study allowed us to draw some significant conclusions about the structure-activity relationships of this type of molecule, regarding their antiproliferative activity.

As shown in Table 2, derivatives having 2-mercaptobenzothiazole (9be, 9bj and 13de), 2-mercapto-4,5-dihydrothiazole (9bk and 13dk) and 2-(methylthio)-4,5-diphenyloxazole (9af, 9bf, 9df, 13df) at the  $2\beta$ -methyl position of the penam nucleus showed the highest antiproliferative activity against cancer cells. Oxidation of the thiazolidine sulfur atom is directly related to cytotoxicity in normal cells (NMuMG). Thus sulfone derivatives 14de and 10bj affect the growth of normal cells NMuMG (inhibit approximately 60% and 90% growth rate, respectively) to a greater extent than sulfide derivatives 13de and 9bj (25% and 35%, respectively).

We have also noted that the introduction of a chlorine atom at the 2β-methyl position of the penam nucleus is detrimental to antitumor activity since those derivatives were either cytotoxic against normal cells or innocuous to both normal and tumor cells (5c, 15a, 7a and 5b). The presence of the tetrazole heterocycle at the  $2\beta$ -methyl position (9ag and 9ai) is remarkably unfavorable for the antitumor activity of these compounds. Furthermore, the incorporation of a phenyl group at position 3 of the tetrazole ring showed an increase in cytotoxicity against normal cells; 9ag inhibits 90% of cell growth in NMuMG and about 50% in 3T3 cell lines, while 9ai inhibits those cell lines by approximately 20% and 35%, respectively.

Regarding position 6 of a penam nucleus, the presence of the halides appears to have no influence on the cytotoxic activity. Thus, dibromo derivative 9af did not show any significant difference in activity compared to its dehalogenated analogue 9df. No significant differences were observed in cytotoxicity between the two carboxylic acid protective groups of 2β-methyl penicillins, benzyl and methyl esters. This is exemplified by similar activity values obtained for compound 13dc and its analogue 9dc.

### Conclusions

In conclusion, a series of 2β-methyl substituted penicillin derivatives were synthesized and the in vitro cytotoxic activities against four human tumor cell lines and two normal cells were evaluated. Some of the tested compounds showed satisfactory antitumor activity, especially compounds 13de, 13df, 9df and 9af which displayed higher values and good relative potencies. We have explored the structure-activity relationships and some general conclusions have been drawn. 2β-[(Benzothiazol-2-yl)thio]methyl and 2β-[(4,5-diphenyl-oxazol-2-yl)thio methyl have shown to be the most promising penam derivatives of this series.

Our work demonstrates the possibility of obtaining new antitumor leaders by simple chemical transformations from a given molecular framework. The use of 2β-methyl substituted penicillin derivatives may help to discover new related compounds with potent and selective cell growth inhibitory activity which could be further tested as potential therapeutic agents for cancer treatment.

# Acknowledgements

Support from CONICET, ANPCyT, UBA and UNR from Argentina is gratefully acknowledged. P.G.C. thanks CONICET for the fellowship.

### Notes and references

- 1 S. A. Testero, J. F. Fisher and S. Mobashery, Curr. Opin. Microbiol., 2010, 13, 551-557.
- 2 S. A. Testero, J. F. Fisher and S. Mobashery,  $\beta$ -Lactam Antibiotics in Burger's Medicinal Chemistry, Drug Discovery and Development, ed. D. J. Abraham and D. P. Rotella, Wiley and Sons, 2010, vol. 7 (Antiinfectives).
- 3 (a) T. Ritter, L. Kværnø, M. Werder, H. Hauser and E. M. Carreira, Org. Biomol. Chem., 2005, 3, 3514-3523; (b) D. K. Tiwari, A. Y. Shaikh, L. S. Pavase, V. K. Gumaste and A. R. A. S. Deshmukh, Tetrahedron, 2007, 63, 2524-2534; (c) D. A. Burnett, Curr. Med. Chem., 2004, 11, 1873-1887.
- 4 (a) R. M. Adlington, J. E. Baldwin, B. Chen, S. L. Cooper, W. McCoull, G. J. Pritchard, T. J. Howe, G. W. Becker,

R. B. Hermann, A. M. McNulty and B. L. Neubauer, Bioorg. Med. Chem. Lett., 1997, 7, 1689-1694; (b) R. Annunziata, M. Benaglia, M. Cinquini, F. Cozzi and A. Puglisi, Bioorg. Med. Chem., 2002, 10, 1813-1818.

MedChemComm

- D. Borthwick, G. Weingarten, T. M. Haley, M. Tomaszewski, W. Wang, Z. Hu, J. Bedard, H. Jih, L. Yuen and T. S. Mansour, Bioorg. Med. Chem. Lett., 1998, 8, 365-370.
- 6 (a) A. Clemente, A. Domingos, A. P. Grancho, J. Ilev, R. Moreira, J. Neres, N. Palma, A. B. Santana and E. Valente, Bioorg. Med. Chem. Lett., 2001, 11, 1065-1068; (b) K. P. Koteva, A. M. Cantin, W. A. Neugebauer and E. Escher, Can. J. Chem., 2001, 79, 377-387; (c) G. Cainelli, P. Galletti, S. Garbisa, D. Giacomini, L. Sartor and A. Quintavalla, Bioorg. Med. Chem., 2005, 13, 6120-6132.
- 7 (a) E. L. Setti, D. Davis, T. Chung and J. McCarter, Bioorg. Med. Chem. Lett., 2003, 13, 2051-2053; (b) N. E. Zhou, D. Guo, G. Thomas, A. V. N. Reddy, J. Kaleta, E. Purisima, R. Menard, R. G. Micetich and R. Singh, Bioorg. Med. Chem. Lett., 2003, 13, 139-141.
- 8 M. Feledziak, C. Michaux, A. Urbach, G. Labar, G. G. Muccioli, D. M. Lambert and J. Marchand-Brynaert, J. Med. Chem., 2009, 52, 7054-7068.
- 9 (a) E. Alonso, F. López Ortiz, C. del Pozo, E. Peralta, A. Macias and J. González, J. Org. Chem., 2001, 66, 6333-6338; (b) C. Palomo, J. M. Aizpurua, A. Benito, R. Galarza, U. K. Khamrai, J. Vazquez, B. de Pascual-Teresa, P. M. Nieto and A. Linden, Angew. Chem., Int. Ed., 1999, 38, 3056-3058; (c) W. P. Malachowski, Ch. Tie, K. Wang and R. L. Broadrup, J. Org. Chem., 2002, 67, 8962-8969.
- 10 (a) I. Ojima. In The Organic Chemistry of  $\beta$ -Lactams, ed. G. I. Georg, VCH, New York, 1993; (b) C. Palomo, I. M. Aizpurua and I. Ganboa in The Synthesis of  $\alpha$ -Amino Acids and Their Derivatives from  $\beta$ -Lactams, ed. E. Juaristi, Wiley-VCH, New York, 1997; (c) C. Palomo, J. M. Aizpurua, I. Ganboa and M. Oiarbide, Synlett, 2001, 12, 1813-1826; (d) G. S. Singh, Tetrahedron, 2003, 59, 7631-7649.
- 11 H. Ge, J. T. Spletstoser, Y. Yang, M. Kayser and G. I. Georg, J. Org. Chem., 2007, 72, 756-759.
- 12 (a) D. Kuhn, C. Coates, K. Daniel, D. Chen, M. Bhuiyan, A. Kazi, E. Turos and Q. P. Dou, Front. Biosci., 2004, 9, 2605-2617; (b) D. M. Smith, A. Kazi, L. Smith, T. E. Long,

- B. Heldreth, E. Turos and Q. P. Dou, Mol. Pharmacol., 2002, 61, 1348-1358.
- 13 (a) L. Sun, N. I. Vasilevich, J. A. Fuselier, S. J. Hocart and D. H. Coy, Bioorg. Med. Chem. Lett., 2004, 14, 2041-2046; (b) R. Pagliarin, F. Orsini, G. Montano, F. Tripodi, P. Coccetti and P. A. Fusi, Patent Application WO 2013017548A1, 2013.
- 14 N. M. O'Boyle, M. Carr, L. M. Greene, O. Bergin, S. M. Nathwani, T. McCabe, D. G. Lloyd, D. M. Zisterer and M. J. Meegan, J. Med. Chem., 2010, 53, 8569-8584.
- 15 I. Banik, F. F. Becker and B. K. Banik, J. Med. Chem., 2003, 46, 12-15.
- 16 F. A. Harding, A. D. Liu, M. Stickler, O. J. Razo, R. Chin, N. Faravashi, W. Viola, T. Graycar, V. P. Yeung, W. Aehle, D. Meijer, S. Wong, M. H. Rashid, A. M. Valdes and V. Schellenberger, Mol. Cancer Ther., 2005, 4, 1791-1800.
- 17 M. Vorona, G. Veinberg, I. Shestakova, I. Kanepe, I. Potorochina, K. Dikovskaya, R. Bokaldere, M. Petrova, E. Liepinsh and E. Lukevics, Chem. Heterocycl. Compd., 2007, 43, 207-219.
- 18 (a) P. G. Cornier, C. M. L. Delpiccolo, F. C. Mascali, D. B. Boggián, E. G. Mata, M. G. Cárdenas, V. C. Blank and L. P. Roguin, MedChemComm, 2014, 5, 214-218; (b) For a previous cytotoxicity study of penicillin derivatives, including some 2β-methyl substituted penams, see: G. Veinberg, R. Bokaldere, K. Dikovskava, M. Vorona, D. Musel, H. Kazhoka, I. Turovsky, I. Shestakova, I. Kanepe, I. Domrachova and E. Lukevics, Chem. Heterocycl. Compd., 1998, 34, 1266-1267.
- 19 D. B. Boggián and E. G. Mata, Synthesis, 2006, 3397-3404.
- T. Kamiya, T. Tevaji, M. Hashimoto, O. Nakaguchi and T. Oku, Tetrahedron Lett., 1973, 14, 3001-3004.
- 21 E. G. Mata, Tetrahedron Lett., 1997, 38, 6335-6338.
- 22 H. Tanaka, M. Tanaka, S. Yamada, A. Nakai, H. Ohbayashi, T. Terada and S. Torii, Bull. Chem. Soc. Jpn., 1989, 62, 3046-3048.
- 23 A. Ishiwata, L. P. Kotra, K. Miyashita, T. Nagase and S. Mobashery, Org. Lett., 2000, 2, 2889-2892.
- U. Landegren, J. Immunol. Methods, 1984, 67, 379-388.
- 25 (a) F. Li, S. Awale, Y. Tezuka and S. Kadota, Bioorg. Med. Chem., 2008, 16, 5434-5440; (b) S.-L. Zhua, Y. Wua, C.-J. Liuc, C.-Y. Wei, J.-C. Taoc and H.-M. Liua, Eur. J. Med. Chem., 2013, 65, 70-82.