

Synthesis of Kaempferol 3-*O*-[2'',3''- and 2'',4''-di-*O*-(*E*)-*p*-coumaroyl]- α -L-rhamnopyranosides

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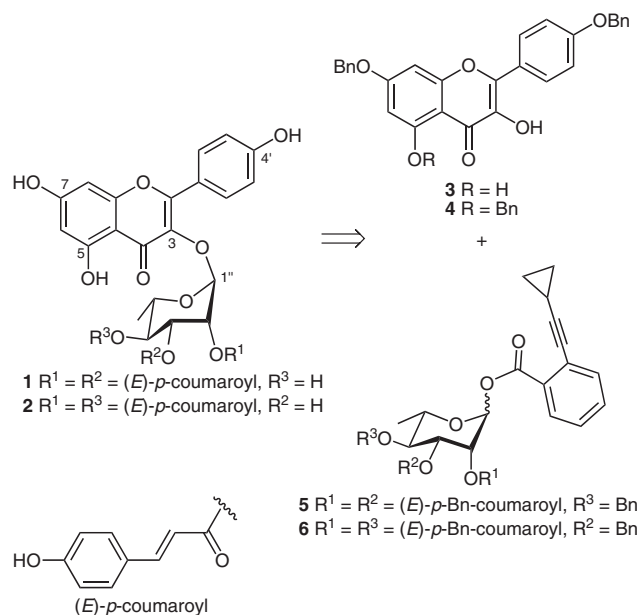
Abstract: Kaempferol 3-*O*-[2'',3''- and 2'',4''-di-*O*-(*E*)-*p*-coumaroyl]- α -L-rhamnopyranoside, two acylated flavonol 3-*O*-glycosides with potent inhibitory activity against methicillin-resistant *Staphylococcus aureus*, were synthesized for the first time, employing glycosyl *o*-cyclopropylethynylbenzoates as donors and Ph₃PAuNTf₂ as a catalyst for the construction of the flavonol glycosidic linkages.

Key words: flavonol glycosides, glycosylation, glycosyl *o*-alkynylbenzoate, antibacterial activity

Kaempferol 3-*O*-[2'',3''-di-*O*-(*E*)-*p*-coumaroyl]- α -L-rhamnopyranoside (**1**), named platanoside, has been isolated from *Platanus acerifolia* (Platanaceae),¹ *P. orientalis*,² *P. occidentalis*,³ *Pentachondra pumila* (Epacridaceae),^{4a} *Faeniculum vulgare* (Apiaceae), and *F. dulce*.^{4b} Recently, this compound was found to have selective and potent activity against methicillin-resistant *Staphylococcus aureus* (MRSA) with an IC₅₀ of 2.0 μ g/mL.⁵ MRSA is a serious pathogen with significant patient mortality.⁵ Hospital-acquired infections of MRSA have shown resistance to multiple antibiotics. Daptomycin is currently the most useful anti-MRSA drug, however, various side effects have been registered, including an increase in blood creatine phosphokinase, rhabdomyolysis, skin exfoliation, and skin ulcers. Thus, development of new anti-MRSA drugs is a matter of great urgency.

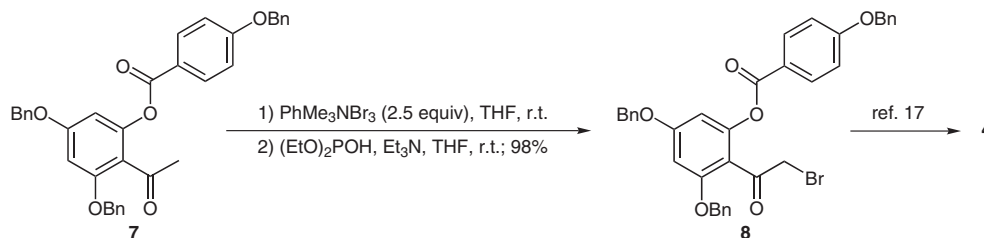
Kaempferol 3-*O*-[2'',4''-di-*O*-(*E*)-*p*-coumaroyl]- α -L-rhamnopyranoside (**2**), the regioisomer of compound **1**, has been identified from *Ocotea vellosiana* (Lauraceae),⁶ *Laurus nobilis* (Lauraceae),⁷ *Mammea longifolia* (Guttiferae),⁸ *Eriobotrya japonica* (Rosaceae),⁹ *Cinnamomum kotoense* (Lauraceae),¹⁰ *Epimedium sagittatum* (Berberidaceae),¹¹ and *Machilus philippinensis* (Lauraceae).¹² Independently, compound **2** was also found to be a potent anti-MRSA agent, with the minimum inhibitory concentrations (MICs) being 0.5–2.0 μ g/mL.¹³ In addition, compound **2** could suppress proliferation of human peripheral blood mononuclear cells induced by phytohemagglutinin, with an IC₅₀ of ca. 6.0 μ M.¹⁰

Compounds **1** and **2** are typical naturally occurring acylated flavonol 3-*O*-glycosides.¹⁴ A key step to synthesize this type of natural metabolites is the construction of the phenolic 3-*O*-glycosidic linkage, which has been mainly resorted to the glycosylation protocol with glycosyl bromides as donors under phase-transfer-catalysis conditions¹⁵ (PTC) or under the action of silver salts.¹⁶ Recently, we disclosed an efficient alternative to the synthesis of this type of linkage with glycosyl *o*-alkynylbenzoates as donors and gold(I) complex as a catalyst.¹⁷ Herein, we report the synthesis of compounds **1** and **2** employing this new protocol for the flavonol 3-*O*-glycoside formation (Scheme 1).



Scheme 1 Acylated flavonol 3-*O*-glycosides **1** and **2** and their retrosynthetic perspective

We have described recently the synthetic routes toward 7,4'-di-*O*-benzyl-kaempferol (**3**) and 5,7,4'-tri-*O*-benzyl-kaempferol (**4**).¹⁷ Worth mentioning is that direct benzylation of kaempferol (2.5 equiv BnBr, K₂CO₃, DMF, r.t.) led to 3,7-di-*O*-benzyl-kaempferol (instead of the desired 7,4'-di-*O*-benzyl-kaempferol) as the major product in ca. 30% yield.¹⁸ In the present synthesis of **4**, the transformation from methyl ketone **7** to bromide **8** was greatly im-



Scheme 2 Improved synthesis of 5,7,4'-tri-*O*-benzyl-kaempferol (**4**)

proved by dibromination (with 2.5 equiv of $\text{PhMe}_3\text{NBr}_3$) first followed by selective debromination [$(\text{EtO})_2\text{POH}$, Et_3N , THF, r.t.],¹⁹ providing bromide **8** in an excellent 98% yield (Scheme 2); previous attempts in direct monobromination of **7** led to **8** in less than 49% yield (1 equiv of $\text{PhMe}_3\text{NBr}_3$, THF, r.t.).¹⁷

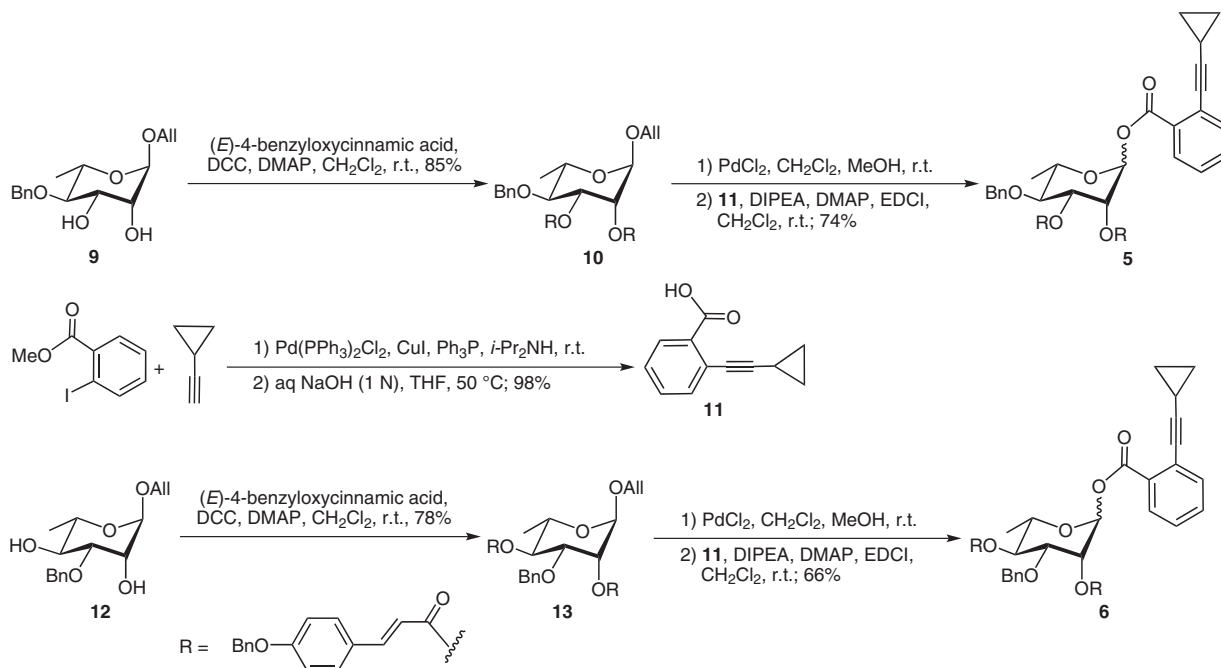
The required rhamnosyl donors **5** and **6** were prepared as shown in Scheme 3. Here we used *o*-cyclopropylethynylbenzoate as the anomeric leaving group instead of the previous *o*-hexynylbenzoate.²⁰ Although the preparation procedure is similar,^{20c} this small variation brings two dividends: the reagent ethynyl cyclopropane is cheaper than 1-hexyne, in addition, *o*-cyclopropylethynylbenzoic acid (**11**) is a crystalline solid (mp 79–82 °C) while *o*-hexynylbenzoic acid is a liquid at room temperature.

Thus, allyl 4-*O*-benzyl- α -L-rhamnopyranoside (**9**)²¹ was condensed with (*E*)-4-benzoyloxycinnamic acid under the action of DCC and DMAP to provide compound **10** (85%). Removal of the anomeric allyl group on **10** was effected with PdCl_2 , the resulting lactol was then directly subjected to condensation with *o*-cyclopropylethynylbenzoic acid (**11**, DIPEA, DMAP, EDCl , CH_2Cl_2 , r.t.)²⁰ to afford the desired rhamnosyl donor **5** in 74% yield (two steps). Employing similar transformations, rhamnosyl do-

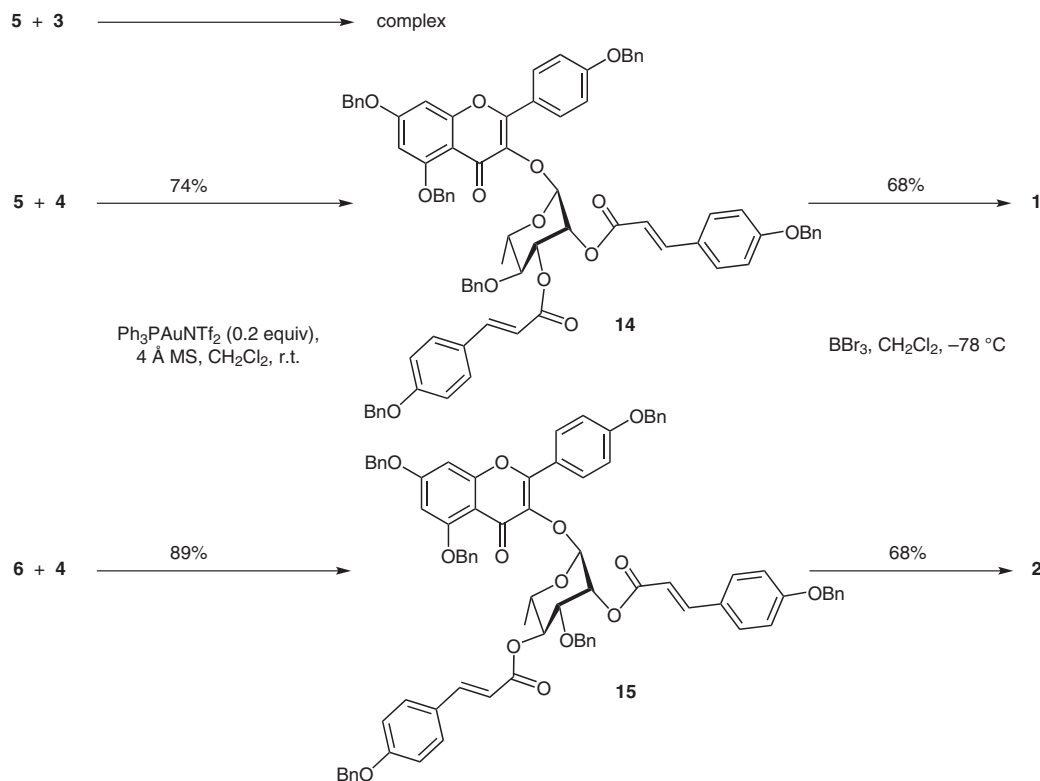
nor **6** was prepared from allyl 3-*O*-benzyl- α -L-rhamnopyranoside (**12**)²² in three steps and 51% overall yield.

The glycosidic coupling of 7,4'-di-*O*-benzyl-kaempferol (**3**) with rhamnosyl *o*-cyclopropylethynylbenzoate (**5**) under the action of $\text{Ph}_3\text{PAuNTf}_2$ led unexpectedly to a complex mixture (Scheme 4). In contrast, the coupling of 5,7,4'-tri-*O*-benzyl-kaempferol (**4**) with **5** under similar conditions (0.2 equiv $\text{Ph}_3\text{PAuNTf}_2$, 4 Å MS, CH_2Cl_2 , r.t.) proceeded smoothly, providing the desired 3-*O*- α -L-rhamnoside (**14**) in a good 74% yield. Similarly, glycosylation of **4** with *o*-cyclopropylethynylbenzoate donor **6** gave α -L-rhamnoside (**15**) in 89% yield. Finally, removal of the six *O*-benzoyl groups on **14/15** was achieved with an excess amount of BBr_3 (18 equiv) in CH_2Cl_2 at low temperature (–78 °C), furnishing the target flavonol glycosides **1** and **2** in 68% yield, respectively. The spectroscopic data of the synthetic compounds are identical to those reported for the natural products.²³

In summary, two acylated flavonol 3-*O*-glycosides **1** and **2** with potent anti-MRSA activities were synthesized for the first time. The synthesis features formation of the flavonol 3-*O*-glycosidic linkages with glycosyl *o*-alkynylbenzoates as donors under the catalysis of a gold(I) complex. This is especially advantageous for the present



Scheme 3 Preparation of rhamnosyl *o*-cyclopropylethynylbenzoates **5** and **6**



Scheme 4 Completion of the synthesis of compounds **1** and **2**

synthesis of the flavonol 3-*O*- α -L-rhamnosides; the previous methods involve unstable rhamnosyl bromides and stoichiometric amount of silver salts.²⁴ Additionally, glycosyl *o*-cyclopropylethynylbenzoates have been used for the first time as glycosyl donors instead of the previous *o*-hexynylbenzoates. The advantage of this slight variation lies in the more convenient and cheaper preparation of the reagent *o*-cyclopropylethynylbenzoic acid.

Acknowledgment

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- (18) **3,7-Di-*O*-benzyl-kaempferol**
¹H NMR (300 MHz, CDCl₃): δ = 12.72 (s, 1 H), 10.29 (s, 1 H), 7.93 (d, *J* = 8.8 Hz, 2 H), 7.47–7.32 (m, 9 H), 6.92 (d, *J* = 8.8 Hz, 2 H), 6.74 (d, *J* = 2.0 Hz, 1 H), 6.42 (d, *J* = 2.0 Hz, 1 H), 5.19 (s, 2 H), 5.03 (s, 2 H). ¹³C NMR (100 MHz, CDCl₃): δ = 178.0, 164.0, 161.0, 160.2, 156.5, 156.1, 136.6, 136.4, 136.0, 130.3, 128.4, 128.3, 128.2, 128.0, 127.7, 120.5, 115.4, 105.2, 98.3, 93.0, 73.3, 69.9, 40.1, 39.9, 39.7, 39.5, 39.3, 39.1, 38.9.
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- (23) **Spectroscopic Data for Synthetic Compounds**
 Compound **1**: [α]_D²⁵ 72.1 (c 0.2, MeOH); lit.⁴ [α] 89.6. ¹H NMR (400 MHz, CD₃OD): δ = 7.87 (d, *J* = 8.8 Hz, 2 H), 7.62 (d, *J* = 15.8 Hz, 1 H), 7.60 (d, *J* = 15.8 Hz, 1 H), 7.46 (d, *J* = 8.6 Hz, 1 H), 7.38 (d, *J* = 8.5 Hz, 1 H), 6.99 (d, *J* = 8.7 Hz, 1 H), 6.81 (d, *J* = 8.6 Hz, 1 H), 6.75 (d, *J* = 8.5 Hz, 1 H), 6.41 (d, *J* = 2.0 Hz, 1 H), 6.38 (d, *J* = 15.8 Hz, 1 H), 6.29 (d, *J* = 15.8 Hz, 1 H), 6.21 (d, *J* = 2.0 Hz, 1 H), 5.82 (dd, *J* = 1.5, 3.2 Hz, 1 H), 5.60 (s, 1 H), 5.28 (dd, *J* = 3.2, 9.5 Hz, 1 H), 3.64 (t, *J* = 9.8 Hz, 1 H), 3.51–3.58 (m, 1 H), 1.05 (d, *J* = 6.0 Hz, 3 H). ¹³C NMR (100 MHz, CD₃OD): δ = 179.4, 168.5, 167.8, 165.9, 163.4, 161.8, 161.5, 161.3, 159.1, 158.6, 147.8, 147.0, 135.5, 131.9, 131.4, 131.2, 127.1 (d), 122.4, 116.9, 116.8 (d), 114.9, 114.4, 105.9, 100.3, 99.9, 94.8, 73.0, 72.2, 71.0, 70.9, 17.8. ESI-HRMS: *m/z* calcd for C₃₉H₃₁O₁₄ [M – H][–]: 723.1719; found: 723.1740.
- Compound **2**: [α]_D²⁵ –36.6 (c 0.3, MeOH); lit.¹⁰ [α] –44.8. ¹H NMR (500 MHz, CD₃OD): δ = 7.81 (d, *J* = 8.7 Hz, 2 H), 7.69 (d, *J* = 15.6 Hz, 1 H), 7.58 (d, *J* = 15.6 Hz, 1 H), 7.53 (d, *J* = 8.7 Hz, 2 H), 7.50 (d, *J* = 8.7 Hz, 2 H), 7.04 (d, *J* = 8.7 Hz, 2 H), 6.84 (d, *J* = 8.7 Hz, 2 H), 6.81 (d, *J* = 8.7 Hz, 2 H), 6.43 (d, *J* = 16.1 Hz, 1 H), 6.40 (d, *J* = 1.4 Hz, 1 H), 6.32 (d, *J* = 15.6 Hz, 1 H), 6.20 (d, *J* = 1.8 Hz, 1 H), 5.76 (s, 1 H), 5.43 (t, *J* = 1.4 Hz, 1 H), 4.98 (t, *J* = 9.9 Hz, 1 H), 4.55 (s, 1 H), 4.16 (dd, *J* = 3.2, 9.7 Hz, 1 H), 3.29–3.26 (m, 1 H), 0.86 (d, *J* = 6.0 Hz, 3 H). ¹³C NMR (125 MHz, CD₃OD): δ = 179.3, 168.5, 168.4, 166.0, 163.3, 161.9, 161.4, 159.5, 158.6, 147.5, 147.0, 134.7, 132.0, 131.4, 131.3, 127.2, 122.5, 116.8, 116.7, 115.0, 114.7, 106.0, 100.0, 99.2, 94.9, 74.7, 73.2, 69.8, 68.5, 17.7.
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