

An Acid-Inert Fluorescent Probe for the Detection of Nitrite

Meiyi Cai^{1,2} · Xiaoyun Chai² · Xuedong Wang¹ · Ting Wang²

Received: 12 December 2016 / Accepted: 2 March 2017
© Springer Science+Business Media New York 2017

Abstract The fluorescent detection of nitrite ions has been challenging owing to the poor acid-stability of typical fluorescent probes. Herein, an acid-inert rhodamine-based fluorescent probe has been developed for the fast detection of trace amount of nitrite ions with turn-on mode. The detection limit of nitrite ions was determined to be 9.4 nM ($S/N = 3$) and the linear range with high linear correlation was observed between 0.025 to 2.5 μM . A test paper method was developed for rapid visual detection of nitrite ions. Quantitative analysis of two real water samples also confirmed the reliability and sensitivity of the developed fluorescent method.

Keywords Nitrite · Silicon · Rhodamine · Fluorescence enhancement

Introduction

Nitrite (NO_2^-) ion, commonly detected in water and food, has proven to be a great threat to human health [1–4]. Excessive intake of NO_2^- with water or food can lead to a variety of

diseases, such as intrauterine growth restriction, spontaneous abortions, infant methemoglobinemia and birth defects in central nervous system [5–7]. Additionally, in acidic environment, NO_2^- play the role as a reaction substrate for the generation of highly carcinogenic N-nitrosamine compounds [8].

Considering the potential toxicity and hazards of NO_2^- , a simple and sensitive detection of NO_2^- level in water and food is highly demanded. Ion chromatography, capillary electrophoresis, colorimetry and electrochemistry are conventionally used for the detection of NO_2^- [3, 9–13]. However, these aforementioned methods are often limited by complicated procedures or low selectivity. In comparison, design of fluorescent probes for NO_2^- has already attracted great attention owing to the high sensitivity and simplicity. So far, several fluorescent probes have been developed for the detection of nitrite [4, 14–21]. These probes are typically based on the diazotization of amine, which requires strong acidic condition. In particular, rhodamine-based fluorescent probes for NO_2^- are especially preferred because of their remarkable chromogenic and fluorogenic responses [14, 15]. Triggered by NO_2^- , diazotization of amine induces the ring-opening of spiro lactam in the probe, thereby restoring the absorption and emission. However, most of the developed fluorescent probes are sensitive to H^+ , and the rhodamine-based probe is not an exception. In acidic condition, the rhodamine based probes go through a H^+ -induced ring-opening process, giving undesired background signals, which leads to the loss of sensitivity and false positive response to NO_2^- . To eliminate these H^+ -induced side effects, the detections of NO_2^- are usually restricted to low temperature or extra addition of base [14, 15].

Recently, we have found that the spiroing in Si-rhodamine can tolerate a relatively strong acidic condition, existing as the non-fluorescent spirocyclic form over a wide pH range [22–25]. The stability of the spiro lactam suppresses the background signal, thereby providing a promising platform for

Electronic supplementary material The online version of this article (doi:10.1007/s10895-017-2071-9) contains supplementary material, which is available to authorized users.

✉ Xuedong Wang
xdwangwfm@163.com

✉ Ting Wang
wangting1983927@gmail.com

¹ College of Pharmacy, Weifang Medical University, Weifang 261053, China

² Department of Organic Chemistry, College of Pharmacy, Second Military Medical University, Shanghai 200433, China

fabricating acid-inert probes with high sensitivity. Herein, we developed a Si-rhodamine-based probe **SiRB-Nitrite** for the detection of NO_2^- (Scheme 1). Under acidic condition with nitrite ions, probe readily undergoes diazotization, resulting in a ring-opening process with remarkable chromogenic and fluorogenic responses. The acid-stability has been further confirmed by comparing the optical properties with its rhodamine analogous **RB-Nitrite** (Scheme 1). Owing to the selectivity and sensitivity, probe **SiRB-Nitrite** has been applied in monitoring NO_2^- level in water.

Materials and Methods

Materials

All reagents and solvents were of the best grade available. Methanol, phosphorus oxychloride, 1,2-dichloroethane, acetonitrile, ethyl acetate, petroleum ether and triethylamine were supplied by Shanghai Chemical Reagent Co. *o*-Diaminobenzene, rhodamine B, sodium nitrite and all salts used in interference experiment were supplied by Adamas-beta. Unless otherwise stated, all commercial reagents were used without additional purification. Solvents were dried according to standard procedures prior to use.

Instrument

All reactions were monitored by thin-layer chromatography (TLC) on gel F254 plates. Flash chromatography was carried out on silica gel (300–400 mesh). NMR spectra were recorded on a Bruker AC-300P spectrometer (300 MHz for ^1H NMR and at 75 MHz for ^{13}C) or on a Bruker AC-600P spectrometer (600 MHz for ^1H NMR and at 150 MHz for ^{13}C NMR). Spectral data are reported in ppm relative to tetramethylsilane (TMS) as internal standard. High-resolution mass spectra (HRMS) were recorded on an Agilent Technologies 6538 UHD Accurate-Mass Q-TOF MS spectrometer using ESI. All pH measurements were performed with a pH-3c digital pH-meter with a combined glass-calomel electrode. UV-visible spectra were obtained on a Analytikjena Specord 210 PLUS UV-vis spectrophotometer.

Fluorescence spectroscopic studies were performed on a Hitachi F-7000.

Absorption and Fluorescence Analysis

Absorption spectra were obtained with 1.0-cm glass cells. Fluorescence emission spectra were obtained with a Xenon lamp and 1.0-cm quartz cells. The fluorescence intensity was measured at 685 nm for probes **SiRB-Nitrite**. The slit width was 5.0 nm for both excitation and emission of probe. The photomultiplier voltage was 600 V. Phosphate buffer solution at different pH was respectively added into DMSO (1%, v/v) solutions containing probe **SiRB-Nitrite** (250 μM).

Preparation of Colorimetric Test Paper

Colorimetric test paper was prepared by immersing strips of white filter paper (10 mm \times 30 mm) into a methanol solution of **SiRB-Nitrite** (250 μM) for 30 min and then dried gently. The colorimetric test paper was stored in a vacuum desiccator. Before the usage, the colorimetric test paper was exposed to hydrogen chloride gas for 10 s. Then the paper was dipped into the aqueous solution of nitrite ions for 1 s and dried in air. Blue color appears in the dried colorimetric test paper.

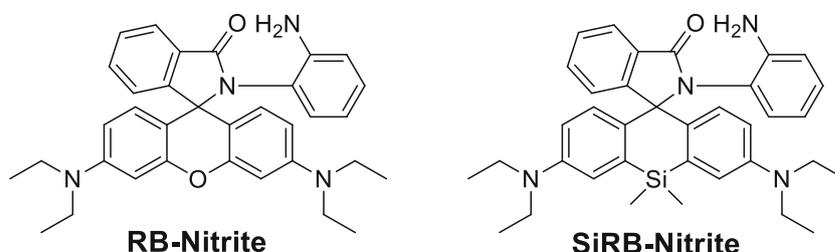
Real Sample Pre-Treatment and Analysis

Water samples were taken from the tap water collected from lab and pond water from the university in Shanghai city. River water and tap water were analyzed after filtration and acidification (pH was adjusted to 2.0), which were spiked with NO_2^- at different concentration for the measurement of recovery.

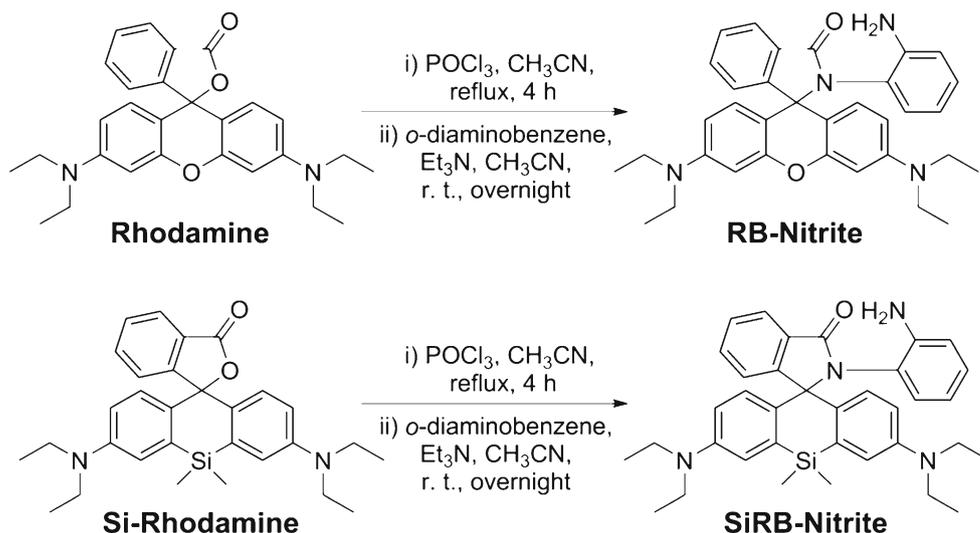
Synthesis of Probes **SiRB-Nitrite** and **RB-Nitrite**

SiRB-Nitrite To a solution of compound **SiRB** (726 mg, 1.5 mmol) in dry 1,2-dichloroethane (10 mL) at room temperature, phosphorus oxychloride (690 mg, 4.5 mmol) was added dropwise over a period of 5 min. After being refluxed for 4 h, the reaction mixture was cooled and concentrated under vacuum to give a blue solid. This blue solid was further dissolved in dry acetonitrile (10.0 mL). The solution was slowly added to a solution of *o*-diaminobenzene (810 mg, 7.5 mmol) in dry

Scheme 1 Chemical structure of probe **SiRB-Nitrite** and its analogous **RB-Nitrite**



Scheme 2 Synthesis of probe **SiRB-Nitrite** and its analogous **RB-Nitrite**



acetonitrile (10 mL) containing triethylamine (5.0 mL). After stirring at room temperature overnight, the mixture was concentrated in vacuo and the crude product was purified by column chromatography on silica gel (ethyl acetate: petroleum ether = 1: 4) to give **SiRB-Nitrite** as a white solid (458 mg, 53% yield). ^1H NMR (600 MHz, CDCl_3): δ 8.03 (dd, $J = 5.6, 2.9$ Hz, 1H), 7.54 (m, 2H), 7.17 (d, $J = 8.1$ Hz, 1H), 6.92–6.86 (m, 1H), 6.51–6.58 (m, 7H), 6.32 (t, $J = 10.0$ Hz, 1H), 5.80 (d, $J = 7.7$ Hz, 1H), 3.35 (d, $J = 1.0$ Hz, 8H), 1.16 (t, $J = 7.0$ Hz, 12H), 0.42 (s, 3H), -0.32 (s, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 167.34, 154.95, 146.17, 145.50, 137.26, 132.69, 132.30, 130.29, 128.87, 127.91, 124.24, 123.53, 122.02, 118.05, 116.76, 114.88, 114.01, 75.04, 53.42, 44.23, 12.45, -0.57 , -1.70 .

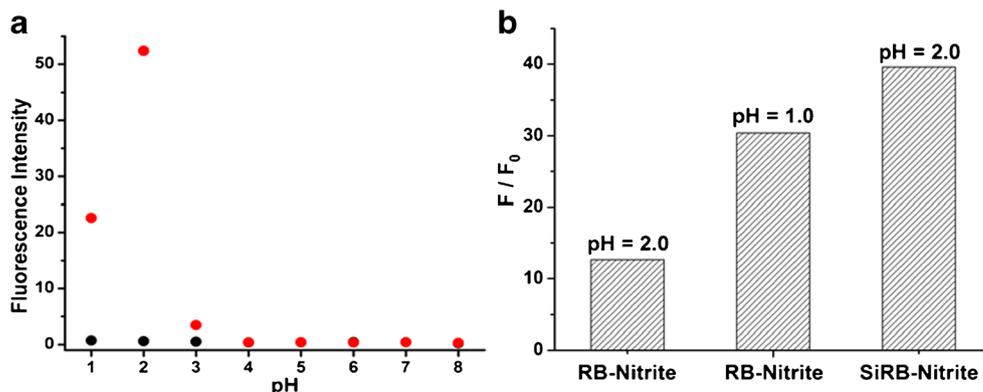
RB-Nitrite To a solution of compound rhodamine B (664 mg, 1.5 mmol) in dry 1,2-dichloroethane (10 mL) at room temperature, phosphorus oxychloride (690 mg, 4.5 mmol) was added dropwise over a period of 5 min. After being refluxed for 4 h, the reaction mixture was cooled and concentrated under vacuum to give a red solid. This blue solid was further dissolved in dry

acetonitrile (10.0 mL). The solution was slowly added to a solution of *o*-diaminobenzene (810 mg, 7.5 mmol) in dry acetonitrile (10 mL) containing triethylamine (5.0 mL). After stirring at room temperature overnight, the mixture was concentrated in vacuo and the crude product was purified by column chromatography on silica gel (ethyl acetate: petroleum ether = 1: 4) to give **SiRB-Nitrite** as a light red solid (447 mg, 56% yield). ^1H NMR (300 MHz, CDCl_3): δ 8.06 (d, $J = 6.9$ Hz, 1H), 7.63–7.54 (m, 2H), 7.27 (s, 1H), 6.98 (t, $J = 7.6$ Hz, 1H), 6.69 (d, $J = 8.5$ Hz, 2H), 6.58 (d, $J = 7.9$ Hz, 1H), 6.53–6.18 (m, 5H), 6.12 (d, $J = 7.8$ Hz, 1H), 3.36 (d, $J = 6.7$ Hz, 8H), 1.18 (t, $J = 6.7$ Hz, 12H). ^{13}C NMR (75 MHz, CDCl_3) δ 166.34, 153.84, 152.22, 148.73, 144.42, 132.60, 131.85, 128.63, 128.71, 128.34, 124.21, 123.40, 122.02, 118.14, 116.93, 107.97, 97.96, 77.18, 67.92, 44.42, 12.38.

Results and Discussion

Probe **SiRB-Nitrite** and its rhodamine analogous **RB-Nitrite** were synthesized according to reported procedures (Scheme 2) [25, 26]. Since the diazotization process requires

Fig. 1 **a** Effect of pH on the probe **SiRB-Nitrite** (2.5 μM , black circle) and its fluorescent responses to 1.0 equiv. of NO_2^- (red circle). **b** The relative fluorescence intensity changes of probes **RB-Nitrite** (2.5 μM) and **SiRB-Nitrite** (2.5 μM) in the absence (F_0) or presence (F) of 1.0 equiv. of NO_2^- at pH = 2.0 and pH = 1.0, respectively



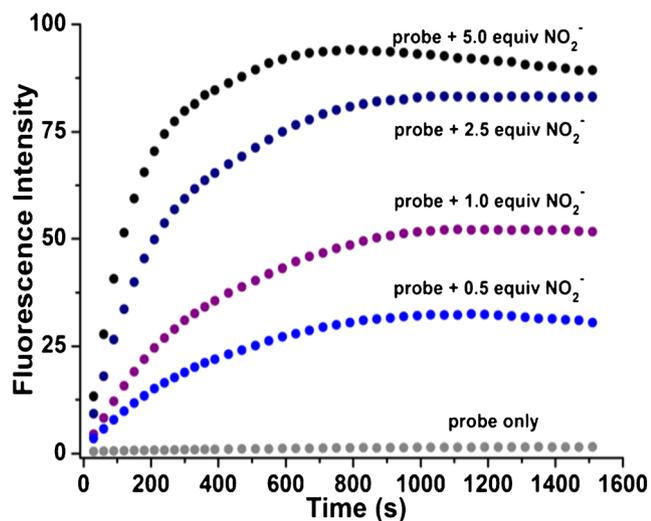


Fig. 2 Time-dependent fluorescence intensity changes of probe **SiRB-Nitrite** ($2.5\mu\text{M}$) upon addition of various concentrations of NO_2^- at $\text{pH} = 2.0$

acidic condition, the effect of pH on the probe **SiRB-Nitrite** and its fluorescent response to NO_2^- were evaluated in the pH range from 1.0 to 8.0. As illustrated in Fig. 1a, **SiRB-Nitrite** displays constant low background fluorescence within a pH range from 8.0 to 1.0, demonstrating that the probe can tolerate a relatively broad range of pH , even under strong acidic condition ($\text{pH} = 1.0$). In sharp contrast, the reference probe **RB-Nitrite** shows pH -dependent fluorescence enhancement under acidic conditions (Fig. S1 and S2, Supporting Information), suggesting

that the spirolactam in probe **RB-Nitrite** is sensitive to H^+ , resulting in relatively high background fluorescence. The fluorescent response of probe **SiRB-Nitrite** to NO_2^- was carefully examined at a series of pH values. Upon the addition of 1.0 equiv. of NO_2^- , probe **SiRB-Nitrite** presented remarkable fluorescence enhancement with the increasing acidity of solution, and exhibited the best response performance to NO_2^- at $\text{pH} = 2.0$. The fluorescence enhancement of the probe started to decrease with further increasing of acidity.

The reference probe **RB-Nitrite** shared similar responses to 1.0 equiv. of NO_2^- at $\text{pH} = 2.0$ (Fig. S1, Supporting Information). However, the fluorescence enhancements in probe **RB-Nitrite** ($\text{pH} = 2.0$ and $\text{pH} = 1.0$) are both lower than that in probe **SiRB-Nitrite** ($\text{pH} = 2.0$) due to the high background fluorescence in acidic environment (Fig. 1b). Comparing with probe **RB-Nitrite**, it is more feasible to use probe **SiRB-Nitrite** to sensitively monitor NO_2^- without extra procedures.

At room temperature, probe **SiRB-Nitrite** can respond to NO_2^- in relatively short time. As shown in Fig. 2, after adding a certain amount of NO_2^- at $\text{pH} = 2.0$, the fluorescence emission at 685 nm remarkably increases within the first 10 min and reaches a plateau in about 15 min.

Under the optimal conditions, the absorption and fluorescence responses of probe **SiRB-Nitrite** with the titration of NO_2^- were examined. Free probe **SiRB-Nitrite** formed a colorless and non-fluorescent solution, indicating the predominant existence of the spirocyclic form. With the addition of

Fig. 3 **a** Absorption change of probe **SiRB-Nitrite** ($2.5\mu\text{M}$) with the increase of NO_2^- (0.0– $25.0\mu\text{M}$). **b** Emission change of probe **SiRB-Nitrite** ($2.5\mu\text{M}$) with the increase of NO_2^- (0.0– $25.0\mu\text{M}$). **c** The relative fluorescence intensity (F/F_0) changes with the increase of NO_2^- (0.0– $25.0\mu\text{M}$). **d** Calibration curve for NO_2^- detection (0.025– $2.5\mu\text{M}$). The excitation and emission wavelengths are 630 and 685 nm. Error bars indicate standard deviation for three replicates

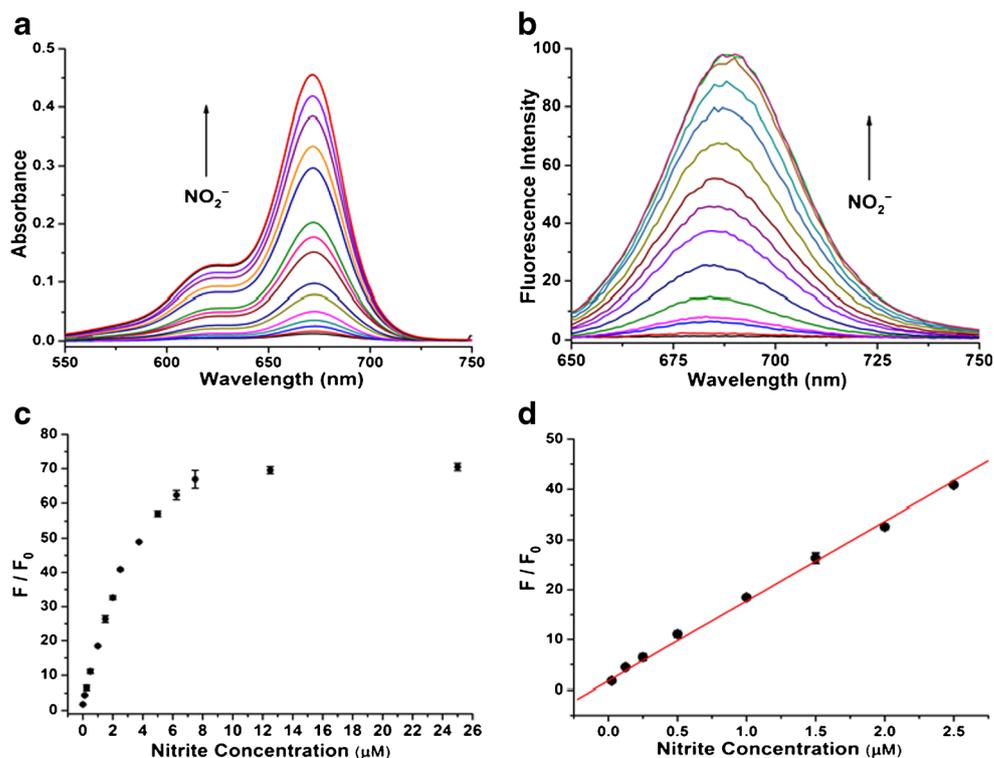
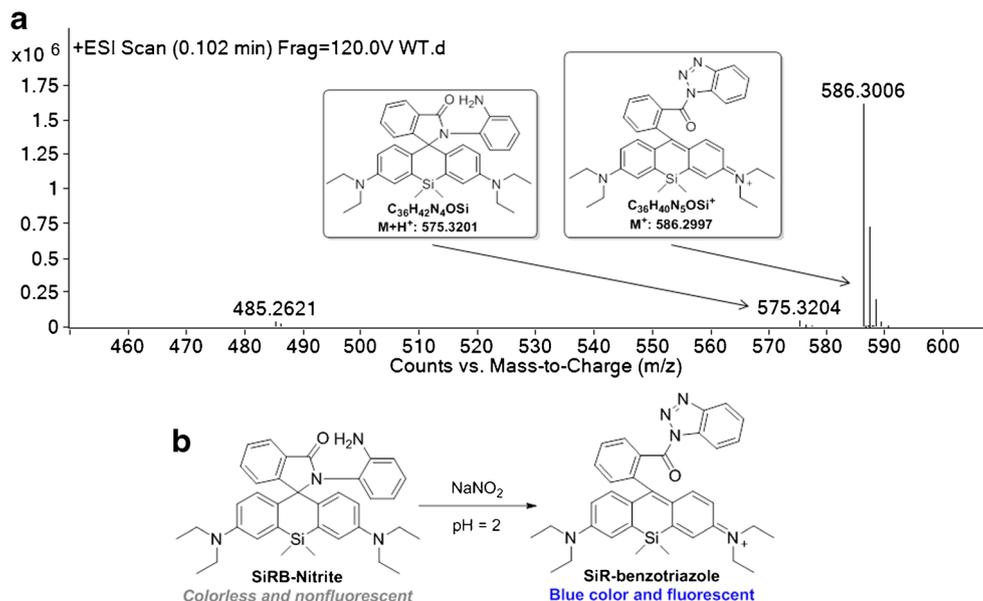


Fig. 4 **a** HRMS analysis of the reaction between probe **SiRB-Nitrite** and nitrite at pH = 2.0. **b** The proposed reaction mechanism of probe **SiRB-Nitrite** with NO_2^-



NO_2^- solution, dramatic color change (colorless to sky blue, Fig. S3, Supporting Information) was observed in the reaction mixture, accompanied with the change in both absorption and fluorescence emission. In absorption spectra, a sharp peak centered at 675 nm increased rapidly and another gently peak centered at 625 nm gradually increased (Fig. 3a). Concomitantly, a gradual enhancement in fluorescence intensity ($\lambda_{\text{em}} = 685 \text{ nm}$) was noted with the increase of NO_2^- (Fig. 3b). The fluorescence enhancement slowed down and reached saturation with 5 equiv. of NO_2^- (Fig. 3c).

A good linear correlation ($R^2 = 0.996$) between the relative intensity of fluorescence enhancement and NO_2^- concentration from 0.025 to 2.5 μM was observed (Fig. 3d). The

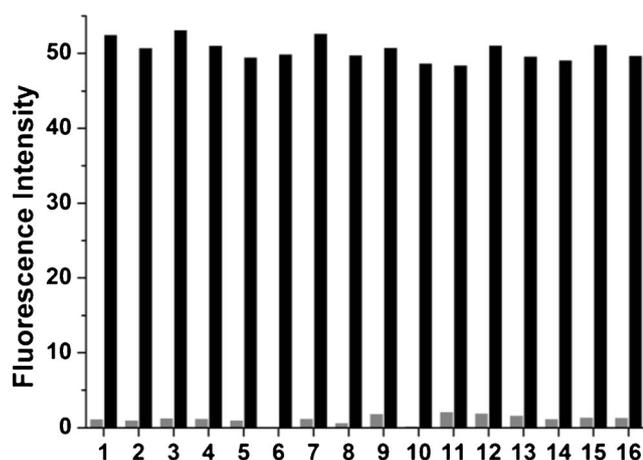


Fig. 5 Fluorescence responses of probe **SiRB-Nitrite** (2.5 μM) to various ions. Gray bars represent the fluorescence response of **SiRB-Nitrite** to the excess ion (250 μM) of interest. Black bars represent the subsequent addition of 2.5 μM NO_2^- to the solution. (1) probe; (2) Na^+ ; (3) K^+ ; (4) Ca^{2+} ; (5) Cu^{2+} ; (6) Fe^{3+} ; (7) Mg^{2+} ; (8) Zn^{2+} ; (9) AcO^- ; (10) NO_3^- ; (11) I^- ; (12) F^- ; (13) Cl^- ; (14) Br^- ; (15) CO_3^{2-} ; (16) $\text{C}_2\text{O}_4^{2-}$

detection limit of probe **SiRB-Nitrite** to NO_2^- was determined to be 9.4 nM ($S/N = 3$), which is more sensitive than the maximum allowable level of contamination of nitrite ions in drinking water (1 ppm, 21.7 μM) defined by Environmental Protection Agency [27].

The reaction mechanism of **SiRB-Nitrite** with NO_2^- was revealed by high-resolution Mass Spectroscopy (HRMS) analysis. In the reaction mixture at pH = 2.0, a unique peak at $m/z = 586.3006$ was clearly found (Fig. 4a). The peak is likely belongs to the highly fluorescent compound **SiR-benzotriazole** (calc. 586.2997), which is the ring-opening product of the probe **SiRB-Nitrite** triggered by NO_2^- in acidic solution (Fig. 4b). We also confirmed the high chemical stability of compound **SiR-benzotriazole** under acidic condition, ensuring the reliability of the analytical application.

Many common cations, including Na^+ , K^+ , Ca^{2+} , Cu^{2+} , Fe^{3+} , Mg^{2+} and Zn^{2+} , and some common anions in water, such as AcO^- , NO_3^- , I^- , F^- , Cl^- , Br^- , CO_3^{2-} and $\text{C}_2\text{O}_4^{2-}$ show negligible interfere in the fluorescent emission even at 100-fold higher concentrations of these ions when compared to NO_2^- . In addition, the enhancement in fluorescence intensity resulting from the addition of NO_2^- was not influenced by the

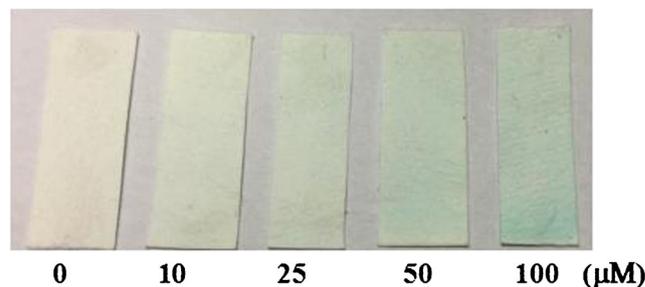


Fig. 6 Photographs of the test paper in the presence of NO_2^-

Table 1 Determination of nitrite ions in water with the proposed method

Samples	NO ₂ ⁻ added (μM)	NO ₂ ⁻ found ^c (μM)	Recovery (%)
Tap water ^a	0	-	
	0.30	0.31 ± 0.01	103.3
	0.60	0.62 ± 0.03	103.3
Pond water ^b	0	0.21 ± 0.02	
	0.30	0.52 ± 0.01	102.0
	0.60	0.79 ± 0.01	97.5

^a Tap water was sampled from our laboratory

^b Pond water was sampled from the lake in our university

^c Average of three determinations

presence of other ions (Fig. 5). These results revealed the high selectivity of **SIRB-Nitrite** and thus is suitable for the detection of NO₂⁻ in complicated real samples.

The fluorescent probe **SIRB-Nitrite** can also work as a turn-on colorimetric probe: visual blue color was observed with the addition of nitrite ions into the solution of the probe (Fig. S3, Supporting Information). Herein, a turn-on colorimetric test paper was prepared for the qualitative analysis of NO₂⁻. With the increasing of nitrite ions from 0 to 100 μM, the color of the test paper has vividly changed from pure white to sky blue (Fig. 6). The darkness of the color correlated to the concentration of NO₂⁻. With the current concentration of probe, the visible detection limit of NO₂⁻ is in the range of 10 μM. This simple and convenient method can be useful for the fast on-site detection of NO₂⁻.

The unique properties of the probe **SIRB-Nitrite**, including rapid response, high selectivity and good sensitivity for NO₂⁻, especially acid-inertance, bring it qualify for complicated practical application. Two water samples, tap water collected from the lab and pond water from the university, were analyzed using this method. The concentration of NO₂⁻ in tap water was not determined and the amount of NO₂⁻ in pond water was quantitatively analyzed. Different known amount of NO₂⁻ solution was directly spiked into the two real samples, and the analysis results were summarized in Table 1. The average recoveries of NO₂⁻ were in the range of 97.5–103.3% for all the spiked samples with low relative standard deviation (1.0–3.0%).

Conclusion

In conclusion, we have developed an acid-inert fluorescent probe **SIRB-Nitrite** for simple, fast, highly sensitive and selective detection of NO₂⁻ (Table S1). A test paper is prepared for qualitative detection of NO₂⁻. Quantitative analysis of two real water samples also confirmed the reliability of the developed fluorescent method.

Acknowledgements This work was supported by the National Natural Science Foundation of China (No. 21205135 and 21602250). We are very grateful for the help and suggestion of Dr. Xiaoyan Cui from East China Normal University.

References

1. Moorcroft MJ, Davis J, Compton RG (2001) Detection and determination of nitrate and nitrite: a review. *Talanta* 54(5):785–803
2. Xiao N, Yu C (2010) Rapid-response and highly sensitive Noncross-linking colorimetric nitrite sensor using 4-Aminothiophenol modified gold Nanorods. *Anal Chem* 82(9):3659–3663
3. Chen Z, Zhang Z, Qu C, Pan D, Chen L (2012) Highly sensitive label-free colorimetric sensing of nitrite based on etching of gold nanorods. *Analyst* 137(22):5197–5200
4. Adarsh N, Shanmugasundaram M, Ramaiah D (2013) Efficient reaction based colorimetric probe for sensitive detection, quantification, and on-site analysis of nitrite ions in natural water resources. *Anal Chem* 85(21):10008–10012
5. Brender JD, Olive JM, Felkner M, Suarez L, Marckwardt W, Hendricks KA (2004) Dietary nitrites and nitrates, nitrosatable drugs, and neural tube defects. *Epidemiology* 15(3):330–336
6. Greer FR, Shannon M (2005) Infant Methemoglobinemia: the role of dietary nitrate in food and water. *Pediatrics* 116(3):784–786
7. Manassaram DM, Backer LC, Moll DM (2006) A review of nitrates in drinking water: maternal exposure and adverse reproductive and developmental outcomes. *Environ Health Perspect* 114(3):320–327
8. Kilfoy BA, Zhang Y, Park Y, Holford TR, Schatzkin A, Hollenbeck A, Ward MH (2011) Dietary nitrate and nitrite and the risk of thyroid cancer in the NIH-AARP diet and health study. *Int J Cancer* 129(1):160–172
9. Kim HJ, Kim YK (1989) Determination of nitrite in drinking water and environmental samples by ion exclusion chromatography with electrochemical detection. *Anal Chem* 61(14):1485–1489
10. Doyle JM, Miller ML, McCord BR, McCollam DA, Mushrush GW (2000) A multicomponent mobile phase for ion chromatography applied to the separation of anions from the residue of low explosives. *Anal Chem* 72(10):2302–2307
11. Chen X, Wang F, Chen Z (2008) An electropolymerized Nile blue sensing film-based nitrite sensor and application in food analysis. *Anal Chim Acta* 623(2):213–220
12. Daniel WL, Han MS, Lee J-S, Mirkin CA (2009) Colorimetric nitrite and nitrate detection with gold nanoparticle probes and kinetic end points. *J Am Chem Soc* 131(18):6362–6363
13. Ning W, Xia C, Xiaolan C, Yanjun X, Lin G (2010) Porous cuprite films: facile solution deposition and their application for nitrite sensing. *Analyst* 135(8):2106–2110
14. Kumar V, Banerjee M, Chatterjee A (2012) A reaction based turn-on type fluorogenic and chromogenic probe for the detection of trace amount of nitrite in water. *Talanta* 99:610–615
15. Xue Z, Wu Z, Han S (2012) A selective fluorogenic sensor for visual detection of nitrite. *Anal Methods* 4(7):2021–2026
16. Strianese M, Milione S, Bertolasi V, Pellecchia C (2013) Iron and manganese pyridoxal-based complexes as fluorescent probes for nitrite and nitrate anions in aqueous solution. *Inorg Chem* 52(20):11778–11786
17. Lu L, Chen C, Zhao D, Yang F, Yang X (2015) A simple and sensitive assay for the determination of nitrite using folic acid as the fluorescent probe. *Anal Methods* 7(4):1543–1548

18. Shen Y, Zhang Q, Qian X, Yang Y (2015) Practical assay for nitrite and Nitrosothiol as an alternative to the Griess assay or the 2,3-Diaminonaphthalene assay. *Anal Chem* 87(2):1274–1280
19. Anuradha LK, Bhosale SV (2016) Selective detection of nitrite ion by an AIE-active tetraphenylethene dye through a reduction step in aqueous media. *RSC Adv* 6(51):45009–45013
20. Gu B, Huang L, Hu J, Liu J, Su W, Duan X, Li H, Yao S (2016) Highly selective and sensitive fluorescent probe for the detection of nitrite. *Talanta* 152:155–161
21. Zhang H, Kang S, Wang G, Zhang Y, Zhao H (2016) Fluorescence determination of nitrite in water using prawn-Shell derived nitrogen-doped carbon Nanodots as fluorophores. *ACS Sensors* 1(7):875–881
22. Wang T, Zhao Q-J, Hu H-G, Yu S-C, Liu X, Liu L, Wu Q-Y (2012) Spirolactonized Si-rhodamine: a novel NIR fluorophore utilized as a platform to construct Si-rhodamine-based probes. *Chem Commun* 48(70):8781–8783
23. Zhu W, Chai X, Wang B, Zou Y, Wang T, Meng Q, Wu Q (2015) Spiroboronate Si-rhodamine as a near-infrared probe for imaging lysosomes based on the reversible ring-opening process. *Chem Commun* 51(47):9608–9611
24. Wang B, Cui X, Zhang Z, Chai X, Ding H, Wu Q, Guo Z, Wang T (2016a) A six-membered-ring incorporated Si-rhodamine for imaging of copper(II) in lysosomes. *Org Biomol Chem* 14(28):6720–6728
25. Wang B, Yu S, Chai X, Li T, Wu Q, Wang T (2016b) A lysosome-compatible near-infrared fluorescent probe for targeted monitoring of nitric oxide. *Chem Eur J* 22(16):5649–5656
26. Zheng H, Shang G-Q, Yang S-Y, Gao X, Xu J-G (2008) Fluorogenic and chromogenic rhodamine Spirolactam based probe for nitric oxide by Spiro ring opening reaction. *Org Lett* 10(12):2357–2360
27. Guo Y-X, Zhang Q-F, Shangguang X, Zhen G (2013) Spectrofluorimetric determination of trace nitrite with o-phenylenediamine enhanced by hydroxypropyl- β -cyclodextrin. *Spectrochim Acta A* 101:107–111