A Practical Synthesis of [(1*S*,3*S*)-3-Aminocyclohexyl]methanol and 2-[(1*S*,3*S*)-3-Aminocyclohexyl]propan-2-ol, Useful Intermediates for the Preparation of Novel *m*PGES-1 Inhibitors

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Abstract: Microsomal prostaglandin E2 synthase-1 (*m*PGES-1) is a novel therapeutic target for the treatment of inflammation and pain. During the course of studies aimed at the identification of a suitable *m*PGES-1 inhibitor for clinical development, a need arose for preparing enantiomerically enriched amino alcohols (*S*,*S*)-**2** and (*S*,*S*)-**3**. Described herein, a concise synthesis of (*S*,*S*)-**2** and (*S*,*S*)-**3** has been developed wherein both amino alcohols are derived from a commercially available, low-cost starting material.

Key words: carbocycles, heterocycles, anti-inflammatory agents, chiral resolution, rearrangement

Inducible microsomal prostaglandin E_2 synthase-1 (*m*PG-ES-1), the predominate synthase involved in cyclooxygenase-2 (COX-2) mediated PGE₂ production, is a novel therapeutic target of considerable interest in the treatment of inflammation and pain.¹ We have been interested in preparing substituted benzoxazoles of type **1** as *m*PGES-1 inhibitors (Figure 1).² During the course of studies aimed at the identification of a suitable inhibitor for clinical development, a need arose for obtaining hundreds of grams of enantiomerically enriched amino alcohols (*S*,*S*)-**2** and (*S*,*S*)-**3**. Due to the *trans*-1,3-relationship between the substituents on (*S*,*S*)-**2** and (*S*,*S*)-**3**, one of the substituents on the cyclohexyl ring occupies a presumably thermodynamically unfavorable axial orientation, which could make the synthesis more challenging.



Figure 1 General structure of novel *m*PGES-1 inhibitors (1) and new synthetic targets (S,S)-2 and (S,S)-3

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While no prior studies on the synthesis of either (S,S)-2 or (S,S)-3 have appeared in the literature, Zhu et al. prepared the related carboxylic acid (S,S)-4 in six steps from enantiomerically enriched (97% ee) lactone (S,S)-5 (Scheme 1);³ the latter compound being prepared in four steps from commercially available material.⁴ While (S,S)-4 presumably could be converted into desired (S,S)-2, the existing synthesis of (S,S)-4 is lengthy, and the starting dienophile is expensive. Based on prior work by Murahashi,⁵ Zhu et al. attempted to shorten their route via a direct $S_N 2$ opening of lactone (*S*,*S*)-**5** with sodium azide. Unfortunately, partial racemization occurred during the reaction, presumably due to ring opening via a competing $S_N 2'$ pathway.³ We detail below a concise synthesis of (S,S)-2 and (S,S)-3, of which both compounds are derived from a low cost starting material.



Scheme 1 Zhu's synthesis of (S,S)-4 based on Trost's enantiomerically enriched lactone (S,S)-5. *Reagents and conditions*: (a) Na₂CO₃, MeOH, r.t.; (b) H₂, Pd/C, MeOH; (c) TsCl, py, CH₂Cl₂; (d) NaN₃, DMF, 80 °C; (e) LiOH, THF–MeOH–H₂O, 40 °C; acidic workup.

Our initial API requirements for (S,S)-2 were met via the synthetic sequence outlined in Scheme 2. Thus, resolution of Cbz-protected amino alcohol *rac*-6, which was purchased from a vendor,⁶ using chiral super critical fluid chromatography (SFC)⁷ afforded enantiomerically enriched carbamate (S,S)-6⁸ (98% ee). Hydrogenolysis of the Cbz-group in (S,S)-6 gave rise (98%) to the desired amine (S,S)-2.^{9,10} HATU-mediated coupling of amine (S,S)-2 with carboxylic acid 7² afforded amide (S,S)-8.¹¹ Compound (S,S)-8 was shown to be a potent inhibitor in our *m*PGES-1 enzyme assay (Scheme 2). Conversely, enantiomer (R,R)-8¹² was 75-fold less active, displaying only weak activity. The absolute configuration at C(1) and C(3) on the cyclohexyl ring in (S,S)-8 was unambiguously

established as (1S,3S) via X-ray crystallography (Figure 2).¹³

The option of purchasing carbamate *rac*-**6** was convenient as it provided access to gram quantities of enantiomerical-



Scheme 2 Reagents and conditions: (a) resolution, SFC, Chiral Pak AD-H 250 column (EtOH–CO₂, 1:3, 40 °C); (b) H₂, 10% Pd/C (cat.), MeOH, 50 psi, 16 h; (c) carboxylic acid 7 (1.0 equiv), amine 2 (1.2 equiv), HATU (1.2 equiv), *i*-Pr₂NEt (5.0 equiv), DMF, r.t., 16 h. ^a mPGES-1 enzyme inhibition assay. Numbers indicate IC₅₀ values generated from 10-point concentration response relationships in duplicate, *n* values in parentheses denotes number of iterations. For enzyme assay conditions, see ref. 14.



Figure 2 ORTEP diagram of compound (*S*,*S*)-8



Scheme 3 Reagents and conditions: (a) EtOH (1.0 equiv), concd HCl (cat.), 80 °C, 4 h; (b) DPPA (1.0 equiv), Et₃N (1.1 equiv), toluene, 70 °C, 1 h; BnOH (1.05 equiv), Et₃N (1.1 equiv), 80 °C, 5 h; (c) LiOH·H₂O (5.0 equiv), THF–MeOH–H₂O (6:3:1), r.t., 5 h; (d) BH₃·THF (2.2 equiv), THF, -5 °C, 4 h.

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ly pure amino alcohol (S,S)-2; however, this tactic did not represent a long-term solution for meeting our API needs. The high cost of *rac*-6, coupled with long lead times, meant sourcing this material was no longer practical; hence, we sought to develop a practical synthesis of *rac*-6, capable of delivering hundreds of grams of API.

Our synthesis of rac-6 commenced from dicarboxylic acid 9, a low cost, commercially available mixture of cisand *trans*-isomers (Scheme 3).¹⁵ The *trans*-isomer can be efficiently isolated in high purity using the procedure of Skita and Rossler.¹⁶ Monoesterification of *trans*-isomer 10 afforded ester 11.¹⁷ Under these conditions, no epimerization to the corresponding cis-isomer was seen. Curtius rearrangement of carboxylic acid 11 and trapping of the intermediate isocyanate with benzyl alcohol led to the corresponding carbamate. Subsequent saponification of the ester with lithium hydroxide gave rise (72%) to carboxylic acid 12. Again, under these conditions, no epimerization to the *cis*-isomer occurred. It should be mentioned that in a prior study Hewgill and Jefferies utilized a Schmidt reaction to convert dicarboxylic acid 10 into the related amino acid rac-4.¹⁸ Unfortunately, this procedure utilizes the highly toxic and potentially explosive hydrazoic acid, which was seen as a significant drawback to our API needs.

Desired alcohol *rac*-**6** was realized in 72% yield via borane reduction of carboxylic acid **12** (Scheme 3). The spectral properties of *rac*-**6**, prepared according to Scheme 3, were identical to an authentic sample of *rac*-**6**. Over 100 grams of *rac*-**6** was prepared according to Scheme 3.

Enantiomerically enriched amino alcohol (S,S)-3 was also prepared from commercial dicarboxylic acid 9 (Scheme 4). Toward this end, treatment of monoester 11 with methyl magnesium bromide, followed by subsequent Curtius rearrangement of the carboxylic acid led to tertiary alcohol rac-13. Resolution of rac-13 using SFC affordenriched alcohol (S,S)-13.¹⁹ enantiomerically ed Hydrogenolysis of the benzyloxycarbonyl group in (S,S)-13 gave rise (90%) to the desired amino alcohol [(S,S)-**3**].^{9,20} Over 100 grams of *rac*-**13** was prepared via this synthetic route. HBTU-Mediated coupling of amine (S,S)-**3** with carboxylic acid **14**² gave rise to amide (S,S)-**15**.²¹ Compound (S,S)-15 was a potent inhibitor of mPGES-1 in the enzyme assay (Scheme 3); the corresponding enantiomer [(R,R)-15] was ca. 50-fold less potent.²²

The absolute configuration of (S,S)-3 was established based on the synthesis shown in Scheme 5. Thus, condensation of enantiomerically enriched amine (S,S)-2 with acetyl acetone afforded dimethylpyrrole 16. Oxidation (TPAP, NMO) of the alcohol gave the corresponding aldehyde. Treatment of the aldehyde with methyl magnesium bromide led to secondary alcohol 17. The addition to the aldehyde was essentially nonselective under these conditions (dr = 55:45). Oxidation of the alcohol with TPAP and NMO gave rise (76%) to the corresponding methyl ketone. Subjection of the ketone to methyl Grig-



Scheme 4 Reagents and conditions: (a) MeMgBr (3.0 M solution in Et₂O, 3.15 equiv), THF, 0 °C \rightarrow r.t., 16 h; (b) DPPA (1.0 equiv), Et₃N (1.1 equiv), toluene, 70 °C, 1 h; BnOH (1.05 equiv), Et₃N (1.1 equiv), 80 °C, 5 h; (c) resolution, SFC chromatography, Chiral Pak IA 250 column (EtOH–*i*-PrOH-CO₂, 1:1:6, 40 °C); (d) H₂, 10% Pd/C, MeOH, 50 psi, r.t., 18 h; (e) carboxylic acid **14** (1.0 equiv), amine **3** (1.2 equiv), HBTU (1.2 equiv), Et₃N (1.5 equiv), DMF, r.t., 16 h. ^a mPGES-1 enzyme inhibition assay. Numbers indicate IC₅₀ values generated from 10-point concentration response relationships in duplicate, *n* values in parentheses denotes number of iterations. For enzyme assay conditions, see ref. 14.



Scheme 5 Reagents and conditions: (a) acetyl acetone (1.1 equiv), AcOH (cat.), toluene, reflux, Dean–Stark, 2 h; (b) TPAP (cat.), NMO (1.5 equiv), 4 Å mol. sieves, CH_2Cl_2 , 2 h; (c) MeMgBr (3.0 M soln in Et₂O, 1.2 equiv), Et₂O, 0 °C, 30 min; (d) NH₂OH-HCl (20 equiv), Et₃N (10 equiv), *i*-PrOH–H₂O (2.5 mL, 4:1), reflux, 6 h; NaOH (4.0 equiv), 0 °C, 16 h; (e) carboxylic acid **14** (1.0 equiv), amine (*S*,*S*)-**3** (1.2 equiv), HBTU (1.2 equiv), Et₃N (1.5 equiv), DMF, r.t., 16 h.

nard afforded the tertiary alcohol in 83% yield. Subsequent hydrolysis of the dimethyl pyrrole gave amine (S,S)-3 in low yield. Coupling (S,S)-3 with carboxylic acid 14 afforded (S,S)-15. Using chiral SFC conditions similar to above, (S,S)-15, prepared according to Scheme 5, coeluted with (S,S)-15 prepared according to Scheme 4. In addition, the biological activity of (S,S)-15, prepared according to Scheme 5, cording to Scheme 5, was identical to the activity of (S,S)-15 prepared according to Scheme 4. In addition, the Scheme 5, was identical to the activity of (S,S)-15 prepared according to Scheme 4. Based on these data,

it is highly likely that the absolute configuration at C(1) and C(3) on the cyclohexyl ring of (S,S)-15 is (1S,3S).

In summary, we have described a concise racemic synthesis of two amino alcohols, which were resolved by chiral chromatography to afford enantiomerically enriched (S,S)-2 and (S,S)-3. Those key intermediates were used for the preparation of potent *m*PGES-1 inhibitors. Both amino alcohols were derived from the same low cost starting material. Over 100 grams of (S,S)-2 and (S,S)-3 were prepared utilizing the described chemistry.

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References and Notes

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- (8) Analytical data for (*S*,*S*)-**6**: Peak 2; white solid; mp 69–71 °C; $[\alpha]^{25}_{D}$ 0.2 (*c* = 1.5, MeOH). ¹H NMR (500 MHz, MeOH*d*₄): δ = 7.26–7.40 (m, 5 H), 5.10 (s, 2 H), 3.77–3.85 (m, 1 H), 3.41 (d, *J* = 6.4 Hz, 2 H), 1.76–1.83 (m, 1 H), 1.64–1.75 (m, 2 H), 1.53–1.64 (m, 4 H), 1.38–1.45 (m, 1 H), 1.14–1.23 (m, 1 H). ¹³C NMR (125 MHz, MeOH-*d*₄): δ = 158.3, 138.6, 129.6, 129.1, 129.0, 67.37, 67.34, 47.67, 36.37, 34.74, 32.37, 29.34, 21.42. LRMS (ESI): *m/z* = 264 [M + H]⁺.
- (9) Amines 2 and 3 were found to absorb carbon dioxide and darken with age. We found it more convenient to store these amines as the corresponding carbamates, 6 and 13, which were shelf stable for more than a year, and convert them into 2 and 3 on an add-need basis.
- (10) Analytical data for (*S*,*S*)-**2**: ¹H NMR (400 MHz, MeOH-*d*₄): $\delta = 3.43$ (d, *J* = 6.7 Hz, 2 H), 3.02–3.09 (m, 1 H), 1.79–1.90 (m, 1 H), 1.49–1.70 (m, 7 H), 1.19–1.30 (m, 1 H). ¹³C NMR (100 MHz, MeOH-*d*₄): $\delta = 66.54$, 47.42, 35.99, 35.42, 33.33, 28.87, 20.83. LRMS (ESI): *m*/*z* = 130.1 [M + H]⁺.
- (11) Analytical data for (*S*,*S*)-**8**: 98% ee (analytical chiral HPLC); [α]²⁵_D -6.4 (*c* = 1.8, MeOH). ¹H NMR (400 MHz, MeOH *d*₄): δ = 7.78 (br d, *J* = 7.4 Hz, 1 H), 7.27 (d, *J* = 8.4 Hz, 1 H), 7.23 (d, *J* = 1.96 Hz, 1 H), 7.02 (dd, *J* = 8.2, 1.95 Hz, 1 H), 4.26–4.32 (m, 2 H), 3.96–4.03 (m, 1 H), 3.43 (d, *J* = 6.6 Hz, 2 H), 3.20 (td, *J* = 13.3, 3.1 Hz, 2 H), 2.57 (tt, *J* = 11.3, 3.9 Hz, 1 H), 1.72–1.91 (m, 5 H), 1.63–1.72 (m, 2 H), 1.53–1.62 (m, 4 H), 1.37–1.46 (m, 1 H), 1.17–1.26 (m, 1 H). ¹³C NMR (100 MHz, MeOH-*d*₄): δ = 175.3, 163.2, 147.4, 144.2, 129.4, 120.5, 115.3, 109.5, 65.81, 45.16, 44.80, 44.70, 42.11, 42.06, 35.24, 33.03, 30.79, 28.04, 27.93, 20.17. LRMS (ESI): *m/z* = 392 [M + H]⁺.

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- (12) Compound (R,R)-8 (98% ee) was prepared using similar conditions to that described for the preparation of (S,S)-8 except (R,R)-6 (98% ee) was used in place of (S,S)-6.
- (13) Data for compound (*S*,*S*)-**8** (crystals from MeCN) were collected on a Bruker APEX diffractometer at Pfizer Groton laboratories, and all crystallographic calculations were facilitated by the SHELXIL system: $C_{20}H_{26}N_3O_3Cl\cdotMeCN\cdot H_2O$; FW = 450.96; monoclinic; space group P2 (1); unit cell dimensions: a = 5.0339 (3) Å, b = 12.3675 (5) Å, c = 18.4354 (9) Å; volume = 1145.14 (10) Å³; Z = 2; $D_{calcd} = 1.308$ Mg/m³; absorption coefficient = 1.772 mm⁻¹; F(000) = 480; GOF on F2 = 1.029; final *R* indices [$I > 2\sigma(I)$]: $R_1 = 0.0393$, $wR_2 = 0.1026$.
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- (19) Analytical data for (*S*,*S*)-**13**: clear oil; Peak 2 (98% ee); $[\alpha]^{25}_{D}$ -10 (*c* = 1.0, MeOH). ¹H NMR (500 MHz, MeOH-*d*₄): δ = 7.26-7.38 (m, 5 H), 6.92 (br d, *J* = 6.59 Hz, 1 H), 5.07 (ABq, *J*_{AB} = 15.0 Hz, Δv_{AB} = 12.0 Hz, 2 H), 3.92-3.98 (m, 1 H), 1.91-1.97 (m, 1 H), 1.82 (d, *J* = 12.7 Hz, 1 H), 1.74 (d,

 $J = 12.9 \text{ Hz}, 1 \text{ H}), 1.58-1.64 \text{ (m, 1 H)}, 1.47-1.58 \text{ (m, 2 H)}, 1.43 \text{ (tt, } J = 13.2, 3.4 \text{ Hz}, 1 \text{ H}), 1.27 \text{ (td, } J = 13.2, 3.5 \text{ Hz}, 1 \text{ H}), 1.11 \text{ (s, 3 H)}, 1.10 \text{ (s, 3 H)}, 1.02 \text{ (qd, } J = 12.3, 3.8 \text{ Hz}, 1 \text{ H}). 1^3\text{C} \text{ NMR} (125 \text{ MHz}, \text{MeOH-}d_4): \delta = 158.4, 138.6, 129.6, 129.1, 129.0, 73.24, 67.37, 43.92, 32.65, 31.35, 28.19, 27.03, 26.93, 25.41, 22.09. LRMS (ESI):$ *m*/*z*= 313.9 [M + Na]⁺.

- (20) Analytical data for (*S*,*S*)-3: clear oil. ¹H NMR (400 MHz, DMSO-*d*₆): δ = 3.80–4.25 (br s, 2 H), 3.24–3.28 (m, 1 H), 1.65–1.76 (m, 2 H), 1.48–1.61 (m, 3 H), 1.40–1.48 (m, 1 H), 1.34 (tt, *J* = 13.7, 3.9 Hz, 1 H), 1.18 (td, *J* = 12.9, 3.5 Hz, 1 H), 1.00 (s, 6 H), 0.89 (qd, *J* = 12.1, 3.5 Hz, 1 H). LRMS (ESI): *m/z* = 158.1 [M + H]⁺.
- (21) Analytical data for (*S*,*S*)-**15**: white solid; 98% ee (analytical chiral HPLC). ¹H NMR (400 MHz, DMSO-*d*₆): δ = 7.57 (br d, *J* = 7.69 Hz, 1 H), 7.39 (s, 1 H), 7.29 (s, 1 H), 4.11–4.15 (m, 2 H), 3.99–4.03 (m, 1 H), 3.92 (s, 1 H), 3.09–3.15 (m, 2 H), 2.53–2.57 (m, 1 H), 2.34 (s, 3 H), 1.71–1.79 (m, 4 H), 1.54–1.64 (m, 3 H), 1.46–1.53 (m, 3 H), 1.26–1.34 (m, 1 H), 1.08–1.15 (m, 1 H), 1.02 (s, 3 H), 1.01 (s, 3 H), 0.87–0.95 (m, 1 H). ¹³C NMR (125 MHz, DMSO-*d*₆): δ = 173.1, 162.4, 147.3, 142.5, 128.1, 126.9, 115.4, 110.8, 70.46, 44.97, 44.08, 42.00, 40.69, 30.89, 29.64, 27.86, 27.76, 27.21, 26.81, 26.49, 20.52, 19.83. LRMS (ESI): *m*/*z* = 434 [M + H]⁺.
- (22) Compound (R,R)-15 (98% ee) was prepared using similar conditions to that described for the preparation of (S,S)-15 except (R,R)-13 (98% ee) was used in place of (S,S)-13.

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