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Highly regioselective synthesis of N-3 organophosphorous derivatives of 3,4-dihydropyrimidin-2(1H)-ones and their calcium channel binding studies

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HIGHLIGHTS

G R A P H I C A L A B S T R A C T

- Novel 3,4-dihydropyrimidinones are obtained in highly regioselective manner.
- Organophosphorous compounds are valuable owing to therapeutic potential.
- N3-substituted dihydropyrimidinones are prized for different biological effects.
- Calcium channel binding study has been conducted on new compounds.
- X-Ray shows structure similar to receptor bound conformation similar to nifedipine.

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1. Introduction

1,4-dihydropyridine (DHP) derivative nifedipine **1** is a wellknown calcium channel blocker (CCB) used for the clinical

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ABSTRACT

A series of novel N-3 substituted 3,4-dihydropyrimidin-2(1*H*)-ones derivatives bearing diaminophosphinyl, phosphonate and phosphorous containing heterocycles were obtained from 3,4dihydropyrimidinones (DHPMs) in a regioselective manner through an efficient reaction protocol, tolerant to substitutional variation at the key diversity positions around the DHPM core. None of the representative compounds screened for calcium channel blocking activity was found to have significant activity compared to nifedipine.

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treatment of cardiovascular diseases such as hypertension, cardiac arrhythmias and angina pectoris [1,2]. Recently, structurally related 3,4-dihydropyrimidin-2(1*H*)-ones (DHPMs), also known as Biginelli compounds [3] have been identified as efficient calcium channel blockers [4,5] as well as α_{1a} -adrenergic receptor antagonists, mitotic kinesin inhibitors, and hepatitis B virus replication suppressors. Several marine natural products containing the DHPM core [6,7] are potent HIV gp-120CD inhibitors.

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Fig. 1. Biologically important 3,4-dihydropyrimidin-2(1H)-ones derivatives.

Over a century old acid catalyzed three-component Biginelli condensation [3] as well as a plethora of its variants furnish DHPMs in moderate to excellent yields, although rationally designed tailormade diversification of six (N1, C2, N3, C4, C5 and C6) positions around the DHPM core [8-12] gave access to those DHPMs which could not be procured through the three-component route owing to either inaccessibility of the latter route or non-availability of one of these components. Most of the potent DHPM based calcium channel blockers for example SQ 32926 **2** and SQ 32547 **3** or α_{1a} adrenergic receptor like SNAP 6201 4 [5] are substituted at N3 position (Fig. 1). A series of 1,4-dihydropyridine-5-cyclic phosphonate derivatives 5 (Fig. 1) were found to have activities superior or comparable to that of nifedipine **1** [13]. Recently phosphorous heterocycles have received widespread attention due to their ubiquity in biological systems [14,15] and their potential to serve as novel pharmaceuticals [16-19]. In continuation of our interest in N-3 substitution of DHPMs [11,12], we wondered, if an organophosphorus group or heterocyclic moiety at N3 could yield a beneficial cardiovascular effect as well as furnish hitherto unknown DHPM derivatives. In this paper we report a useful and highly

regioselective reaction giving access to DHPMs bearing a diaminophosphinyl, phosphonate as well as phosphorous heterocyclic unit at the N-3 position as well as report their calcium channel binding activity.

2. Chemistry

Reaction of DHPM lacking an N3 substituent with phosphorous oxychloride yielding a DHPM bearing a reactive phosphorous oxychloride group at the more basic N3 presented itself as a method of choice owing to ease in availability of reagents as well as operational ease of this as well as its further reactions with various nucleophiles. Thus, when DHPM **6** was treated with POCl₃ at 105 °C (Scheme 1), N3-elaborated **7** was obtained in 90–95% yield. Lower stability of **7** at ambient temperature meant that it could only be used in subsequent reaction without purification. Thus, treating **7** *in situ* with ammonia gas dissolved in dry THF for 45 min at room temperature, led to the smooth formation of N3-diaminophosphinyl DHPM **8** in excellent yields, after recrystallization from methanol (Table 1). This fairly simple and efficacious



Scheme 1. Synthesis of N3-substituted 3,4-dihydropyrimidin-2(1H)-ones.

K. Singh et al. / European Journal of Medicinal Chemistry 54 (2012) 397-402

Table T				
Synthesis	of N3-substituted	3,4-dihydropy	yrimidin-2(1H)-ones.

Entry	Product	<i>R</i> ¹	R ²	<i>R</i> ³	R^4/X	Yield (%)
1.	8a	C ₆ H ₅	C ₂ H ₅	Н	_	95
2.	8b	4-OMeC ₆ H ₄	C_2H_5	Н	_	95
3.	8c	$3-NO_2C_6H_4$	(CH ₃) ₂ CH	Н	_	95
4.	8d	C ₆ H ₅	CH ₃	Н	_	93
5.	8e	Н	C_2H_5	Н	_	85
6.	8f	CH ₃	C_2H_5	Н	_	92
7.	8g	CH ₃	CH ₃	Н	_	83
8.	9a	C ₆ H ₅	C_2H_5	Н	C ₄ H ₉ /NH	66
9.	9b	C ₆ H ₅	C_2H_5	Н	C ₆ H ₅ /NH	60
10.	9c	C ₆ H ₅	C_2H_5	Н	C_2H_5/O	70
11.	10a	C ₆ H ₅	C_2H_5	Н	-/NH	72
12.	10b	CH ₃	C_2H_5	Н	-/NH	65
13.	10c	C ₆ H ₅	C_2H_5	Н	-/O	64
14.	10d	C ₆ H ₅	C_2H_5	CH_3	-/O	65

reaction served as a general protocol for obtaining a range of regioselectively N-3 substituted DHPMs tolerant to substitutional variation around DHPM core as shown in Table 1. In addition to ³¹P NMR and spectral data the characteristic feature of the ¹H NMR spectrum of C-4 aryl derivatives (**8a**–**8d**) is the presence of doublet corresponding to C4–H, due to P–H coupling.

In analogy with the reactions of 7 with ammonia, reactions of various amines with 7 led to the formation of N3-dialkylaminophosphinyl substituted DHPMs 9. Thus, when a solution of appropriate N3-dichlorophosphinyl appended DHPM in dry dioxane was treated with amines such as *n*-butylamine and benzylamine or ethanol (80 °C) (Scheme 1) corresponding N-3 substituted DHPMs 9a–9c were obtained in good yield after purification with column chromatography (entries 8-10, Table 1). Similar reactions with diamines and amino alcohols in dry dioxane at room temperature furnished DHPMs derivatives 10a-10d The scope and limitations of this reaction protocol has been explored by varying substituents at C-4 position as well as ring size of phosphorous containing heterocyclic ring substituted at N-3 position (Table 1). All these novel compounds have been well characterized using spectroscopic techniques and presence of phosphorous in these compounds has been ascertained by recording their ³¹P NMR spectrum.

3. Result and discussion

3.1. Structural correlation with receptor bound conformation

In order to unambiguously establish the incorporation of the diaminophosphinyl group at N-3 positions of **6** and to establish correlation with proposed receptor bound confirmation with nifedipine as well as DHPM type calcium channel blockers, structure of **8b** was additionally confirmed by single crystal X-ray analysis (Fig. 2) [20].

The solid state structure of **8b** (Fig. 2) shows a half-boat type conformation of the dihydropyrimidinone ring. The C-4 aryl ring is perpendicular to the molecular plane and bisects the half-boat conformation of the DHPM ring. The C-5 ester is in *cis* orientation with respect to the C5, C6 double bond of the ring and coplanar with the DHPM ring, so is the N3–P=O link, the P=O oxygen pointing in the direction of C4-H of the DHPM unit. Such conformations drawn close analogy with the proposed receptor bound conformation of DHPMs resulting in the observed calcium channel blocking activity (calcium antagonists) [21].

3.2. Calcium channel binding studies

Representative compounds **8–10** were screened for calcium blocking activity based on their ability to relax a membrane



Fig. 2. X-ray crystal structure of **8b** showing the stereoview of the molecule and the numbering scheme used in the structure analysis. (N2, C8, C13 positions may be read as N3, C4 and C6, respectively, following IUPAC nomenclature scheme).

depolarization induced contraction of vascular smooth muscle. For this purpose, swine carotid arteries were obtained from a slaughterhouse and transported to the laboratory in an ice-cold physiological salt solution (PSS) buffered (pH 7.4) with 3-[N-morpholino] propane sulfonic acid (MOPS). The PSS contained, in mM: 140 NaCl, 4.7 KCl, 1.2 MgSO₄, 1.6 CaCl₂, 1.2 Na₂HPO₄, MOPS, 5 D-glucose, and 0.02 Na₂-ethylenediaminetetraacetic acid (EDTA). Arteries were cleaned of connective tissue, and then dissected free of both intima and adventitia leaving a thin medial layer for experimentation. Intact medial strips of swine carotid artery (7 \times 0.7 mm) were suspended between a Grass FT.03 force transducer and a stationary clip in water jacketed organ baths. The strips were equilibrated in PSS at 37 °C, pH 7.4 and bubbled with 100% O₂ for 90-120 min. A passive force of ~ 2 g was applied to all tissues. The passive force sets the muscles at a length that approximates L_o, the length at which maximal active force is generated. During the equilibration period, tissues were maximally contracted with 110 mM KCl (equimolar substitution for NaCl) several times until similar levels of force were attained.

The equilibrated vascular strips were contracted with 110 mM KCl containing PSS, allowed to achieve a stable level of force and then subjected to the cumulative addition of calcium channel blockers (0.001 µM-10 µM), nifedipine (0.001 µM-10 µM) or DMSO as a vehicle control. Data are presented (Fig. 3) as a percent of the initial maximal response to 110 mM KCl at each dose of compound. The calcium channel blockers were compared against nifedipine 1 for their ability to relax a membrane depolarization induced contraction which is almost exclusively dependent on the influx of extra-cellular calcium [22]. The novel compounds and nifedipine were tested across a concentration range of 0.001 µM-10 µM. Nifedipine completely relaxed the KCl-induced contractions with an IC₅₀ of approximately 0.01 µM against 110 mM KCl. In contrast the compounds 8a-g, 9a-c and 10a-c maximally relaxed the KCl-induced contractions by only 0-25% with the relaxation only significant at 10 µM.



Fig. 3. Medial strips from the swine carotid artery were contracted with 110 mM KCl–PSS and then subjected to the cumulative addition of calcium channel blocker to determine their potential for relaxation. Nifedipine and 13 novel calcium channel blockers (**8a–g**, **9a–c** and **10a–c**) were tested.

4. Conclusion

Thus an efficient protocol using neutral reaction conditions for the regioselective synthesis of N3-substituted DHPM derivatives has been developed. DHPM derivatives bearing a diaminophosphinyl, phosphonate as well as phosphorous containing heterocycles at N-3 position has been synthesized. Among the novel compounds tested the calcium channel blocking activity was less compared to nifedipine in a screen using relaxation of a membrane depolarized vascular tissue as the end-point.

5. Experimental section

5.1. General

All liquid reagents were dried/purified following recommended drying agents and/or distilled over 4 Å molecular sieves. ¹H NMR (400 MHz and 300 MHz), 13 C NMR (100 MHz and 75 MHz) and 31 P (162 MHz and 121 MHz) spectra were recorded in DMSO- d_6 and CDCl₃ on a multinuclear Bruker-400 MHz and Jeol FT-AL-300 instruments with chemical shifts being reported in parts per million (δ) relative to internal tetramethylsilane (TMS, δ 0.0, ¹H NMR or CDCl₃, δ 77.0, ¹³C NMR). Mass spectra were recorded from Indian Institute of Integrative Medicine (CSIR), Jammu, under electron impact at 70 eV on a Bruker Daltonics Esquire 3000 spectrometer. Elemental analysis was performed on FLASH EA 112 (Thermoelectron Corporation) analyzer at Department of Chemistry, Guru Nanak Dev University, Amritsar and the results are quoted in %. IR recorded on FTIR Shimadzu 8400 Fourier-transform spectrophotometer in the range 400–4000 cm⁻¹ using KBr. Melting points were determined in open capillaries and are uncorrected. For monitoring the progress of a reaction and for comparison purpose, thin layer chromatography (TLC) was performed on pre-coated aluminium sheets Merck (60F₂₅₄, 0.2 mm) using an appropriate solvent system. The chromatograms were visualized under UV light. For column chromatography silica gel (60-120 mesh) was employed and eluents were ethyl acetate/ hexane mixtures.

5.2. General procedure for the synthesis of N3-dichlorophosphinyl DHPM derivatives **7**

Appropriate DHPM 6 (0.01 mol) was suspended in phosphorous oxychloride (10 ml) and heated at 105 $^{\circ}$ C for 30 min. Excess phosphorous oxychloride (POCl₃) was removed under reduced

pressure and last traces of $POCl_3$ were removed through azeotropic distillation with dry benzene to furnish the N3-dichlorophosphinyl DHPM derivatives 7 in 90–95% yield, and were used for subsequent reactions without further purification.

5.3. General procedure for the synthesis of N3-substituted DHPM derivatives ${m 8}$

N3-dichlorophosphinyl DHPM derivative 7 (0.5 g) was treated with ammonia gas (evolved by warming 30% aqueous ammonia solution using standard assembly and dried through KOH trap) dissolved in THF at room temperature for 45 min. White precipitates formed in the reaction mixture were filtered and recrystallized from methanol to obtain pure compound. The characteristic data of the compounds is given below.

5.3.1. 5-Ethoxycarbonyl-6-methyl-3-(diaminophosphinyl)-4-phenyl-3,4-dihydropyrimidin-2(1H)-one (**8a**)

White crystalline solid. Rf: 0.4 (methanol:ethyl acetate/10:90). Yield: 95%. M.p. 258–260 °C (methanol). IR (KBr): ν_{max} 1090, 1220, 1615, 1680, 3320 cm⁻¹. ¹H NMR (300 MHz, DMSO- d_6 , 25 °C): δ 1.17 (t, J = 7.2 Hz, 3H, ester-CH₃), 2.23 (s, 3H, C6-CH₃), 4.02 (br, 2H, D₂O exchangeable, NH₂), 4.04–4.15 (m, 2H, ester-CH₂), 4.33 (br, 2H, D₂O exchangeable, NH₂), 6.09 (d, J = 7.2 Hz, 1H, C4-H), 7.17–7.32 (m, 5H, ArH), 9.62 (br, 1H, D₂O exchangeable, N1-H). ¹³C NMR (75 MHz, DMSO- d_6 , 25 °C) δ 14.2, 17.5, 54.5, 59.6, 102.6, 126.1, 127.0, 128.2, 142.7, 148.1, 153.3 and 165.4. ³¹P NMR (121 MHz, DMSO- d_6 , 25 °C): δ 11.95. Anal. Calcd. for C₁₄H₁₉N₄O₄P: C, 49.70; H, 5.62; N, 16.57; Found: C, 49.77; H, 5.93; N, 16.20. MS: *m*/*z* 339 (M⁺+1).

5.3.2. 5-Ethoxycarbonyl-6-methyl-3-(diaminophosphinyl)-4-(4-methoxyphenyl)-3,4-dihydropyrimidin-2(1H)-one (**8b**)

Wite crystalline solid. Rf: 0.2 (methanol:ethyl acetate/10:90). Yield: 95%. M.p. 260–262 °C (methanol). IR (KBr): ν_{max} 1091, 1235, 1631, 1690, 3399 cm⁻¹ ¹H NMR (300 MHz, DMSO- d_6 , 25 °C): δ 1.16 (t, *J* = 7.2 Hz, 3H, ester-CH₃), 2.23 (s, 3H, C6-CH₃), 3.36 (s, 3H, OCH₃), 3.95 (br, 2H, D₂O exchangeable, NH₂), 4.02–4.10 (m, 2H, ester-CH₂), 4.30 (br, 2H, D₂O exchangeable, NH₂), 6.00 (d, *J* = 6.9 Hz, 1H, C4-H), 6.80–6.83 (m, 2H, ArH), 7.22 (d, *J* = 8.7 Hz, 2H, ArH), 9.59 (br, 1H, D₂O exchangeable, N1-H). ¹³C NMR (75 MHz, DMSO- d_6 , 25 °C): δ 14.3, 17.4, 54.2, 55.0, 59.5, 102.7, 113.5, 127.5, 134.7, 147.8, 153.2, 158.3 and 165.3. ³¹P NMR (121 MHz, DMSO- d_6 , 25 °C): δ 12.04. Anal. Calcd. for C₁₅H₂₁N₄O₅P: C, 48.91; H, 5.71; N, 15.22; Found: C, 49.19; H, 6.01; N, 15.25. MS: *m/z* 369 (M⁺+1).

5.3.3. 5-Isopropoxycarbonyl-6-methyl-3-(diaminophosphinyl)-4-(3-nitrophenyl)-3,4-dihydropyrimidin-2(1H)-one (**8c**)

White crystalline solid. Rf: 0.5 (methanol:ethyl acetate/10:90). Yield: 95%. M.p. 268–270 °C (methanol). IR (KBr): ν_{max} 1090, 1225, 1525, 1635, 1680, 3320 cm⁻¹ ¹H NMR (300 MHz, DMSO- d_6 , 25 °C): δ 1.12 (d, J = 6.3 Hz, 3H, ester-CH₃), 1.21 (d, J = 6.3 Hz, 3H, ester-CH₃), 2.24 (s, 3H, C6-CH₃), 4.08 (br, 2H, D₂O exchangeable, NH₂), 4.41 (br, 2H, D₂O exchangeable, NH₂), 4.87–4.95 (m, 1H, ester-CH), 6.12 (d, J = 6.6 Hz, 1H, C4-H), 7.59 (t, J = 7.8 Hz, 1H, ArH), 7.74 (d, J = 7.8 Hz, 1H, ArH), 8.08–8.14 (m, 2H, ArH), 9.86 (br, 1H, D₂O exchangeable, N1-H). ¹³C NMR (75 MHz, DMSO- d_6 , 25 °C): δ 17.5, 21.6, 54.8, 67.1, 101.8, 121.1, 122.2, 130.0, 132.9, 145.2, 147.7, 148.6, 152.7 and 164.5. ³¹P NMR (121 MHz, DMSO- d_6 , 25 °C): δ 11.90. Anal. Calcd. for C₁₅H₂₀N₅O₆P: C, 45.34; H, 5.04; N, 17.63; Found: C, 45.19; H, 5.22; N, 17.43. MS: m/z 398 (M⁺+1).

5.3.4. 5-Methoxycarbonyl-6-methyl-3-(diaminophosphinyl)-4-phenyl-3,4-dihydropyrimidin-2(1H)-one (**8d**)

White crystalline solid. Rf: 0.7 (methanol:ethyl acetate/20:80). Yield: 93%. M.p. 253–255 °C (methanol). IR (KBr): ν_{max} 1024, 1237,

1551, 1631, 1698, 3317 cm⁻¹ ¹H NMR (300 MHz, DMSO- d_{6} , 25 °C): δ 2.23 (s, 3H, C6-CH₃), 3.62 (s, 3H, OCH₃), 4.06 (br, 2H, D₂O exchangeable, NH₂), 4.36 (br, 2H, D₂O exchangeable, NH₂), 6.09 (d, J = 7.2 Hz, 1H, C4-H), 7.20–7.29 (m, 5H, ArH), 9.67 (br, 1H, D₂O exchangeable, NH). ¹³C NMR (75 MHz, DMSO- d_{6} , 25 °C): δ 17.6, 51.3, 54.7, 102.5, 126.3, 127.3, 128.2, 142.5, 148.7, 152.4 and 166.0. ³¹P NMR (121 MHz, DMSO- d_{6} , 25 °C): δ 11.93. Anal. Calcd. for C₁₃H₁₇N₄O₄P: C, 48.15; H, 5.25; N, 17.28; Found: C, 48.42; H, 5.46; N, 16.96. MS: *m*/*z* 325 (M⁺+1).

5.3.5. 5-Ethoxycarbonyl-6-methyl-3-(diaminophosphinyl)-3,4dihydropyrimidin-2(1H)-one (**8e**)

White crystalline solid. Rf: 0.3 (methanol:ethyl acetate/15:85). Yield: 85%. M.p. 260–262 °C (methanol). IR (KBr): ν_{max} 1096, 1291, 1658, 1710, 1741, 3398 cm⁻¹ ¹H NMR (300 MHz, DMSO- d_6 , 25 °C): δ 1.19 (t, J = 7.2 Hz, 3H, ester-CH₃), 2.16 (s, 3H, C6-CH₃), 4.07 (q, J = 6.9 Hz, 2H, ester-CH₂), 4.19 (br, 4H, D₂O exchangeable, 2 × NH₂), 5.73 (s, 2H, C4-CH₂), 9.89 (br, 1H, D₂O exchangeable, N1-H). ¹³C NMR (75 MHz, DMSO- d_6 , 25 °C): δ 14.2, 16.9, 42.1, 59.3, 97.0, 148.1, 153.4 and 165.0. ³¹P NMR (121 MHz, DMSO- d_6 , 25 °C): δ 12.87. Anal. Calcd. for C₈H₁₅N₄O₄P: C, 36.64; H, 5.72; N, 21.37; Found: C, 36.68; H, 6.00; N, 20.89. MS: *m/z* 263 (M⁺+1).

5.3.6. 5-Ethoxycarbonyl-4,6-dimethyl-3-(diaminophosphinyl)-3,4dihydropyrimidin-2(1H)-one (**8**f)

White crystalline solid. Rf: 0.3 (methanol:ethyl acetate/20:80). Yield: 92%. M.p. 265–267 °C (methanol). IR (KBr): ν_{max} 1029, 1245, 1411, 1635, 1696, 3401 cm⁻¹ ¹H NMR (400 MHz, DMSO- d_6 , 25 °C): δ 0.91 (d, J = 6.4 Hz, 3H, C4-CH₃), 0.98 (t, J = 7.0 Hz, 3H, ester-CH₃), 1.96 (s, 3H, C6-CH₃), 3.82–3.92 (m, 2H, ester-CH₂), 3.95 (br, 2H, exchanged with D₂O, NH₂), 3.99 (br, 2H, exchanged with D₂O, NH₂), 3.99 (br, 2H, exchanged with D₂O, NH₂), 4.71 (t, J = 6.2 Hz, 1H, C4-H), 9.28 (br, 1H, exchanged with D₂O, NH). ¹³C NMR (100 MHz, DMSO- d_6 , 25 °C): δ 14.3, 17.3, 21.1, 48.6, 59.3, 103.2, 147.2, 153.0 and 165.0. ³¹P NMR (162 MHz, DMSO- d_6 , 25 °C): δ 12.02. Anal. Calcd. for C₉H₁₇N₄O₄P: C, 39.13; H, 6.16; N, 20.29; Found: C, 38.92; H, 6.44; N, 19.94. MS: m/z 277 (M⁺+1).

5.3.7. 5-Methoxycarbonyl-4,6-dimethyl-3-(diaminophosphinyl)-3,4-dihydropyrimidin-2(1H)-one (**8g**)

White crystalline solid. Rf: 0.2 (methanol:ethyl acetate/20:80). Yield: 83%. M.p. 272–274 °C (methanol). IR (KBr): ν_{max} 1080, 1212, 1487, 1642, 1703, 3431 cm⁻¹ ¹H NMR (300 MHz, DMSO- d_6 , 25 °C): δ 1.11 (d, J = 6.6 Hz, 3H, C4-CH₃), 2.16 (s, 3H, C6-CH₃), 3.61 (s, 3H, OCH₃), 4.16 (br, 4H, D₂O exchangeable, 2 × NH₂), 4.93 (t, J = 6.3 Hz, 1H, C4-H), 9.48 (br, 1H, D₂O exchangeable, NH). ¹³C NMR (75 MHz, DMSO- d_6 , 25 °C): δ 17.3, 21.1, 48.6, 51.0, 103.1, 147.5, 153.1 and 165.5. ³¹P NMR (121 MHz, DMSO- d_6 , 25 °C): δ 11.93. Anal. Calcd. for C₈H₁₅N₄O₄P: C, 36.64; H, 5.72; N, 21.37; Found: C, 36.52; H, 5.92; N, 21.13. MS: m/z 263 (M⁺+1).

5.4. General procedure for the synthesis of N3-substituted DHPM derivatives ${m 9}$

To a stirred solution of appropriate 7 (1 equiv.) in dry dioxane (10 ml) appropriate amine (1.5 equiv.) was added and resulting solution was stirred at room temperature for 2-4 h. After completion of reaction (TLC), dioxane was removed under reduced pressure and residue was dissolved in ethylacetate (20-30 ml) and washed with water (2×30 ml). Organic phase was dried over anhydrous sodium sulphate and compounds 9a-b were obtained through column chromatography using hexane and ethylacetate as eluents. The characteristic data of the compounds is given below.

5.4.1. 5-Ethoxycarbonyl-6-methyl-3-[bis(butylamino)phosphinyl]-3,4-dihydropyrimidin-2(1H)-one (**9a**)

White crystalline solid. Rf: 0.7 (methanol:ethyl acetate/10:90). Yield: 66%. M.p. 115–117 °C (dichloromethane/hexane). IR (KBr): ν_{max} 1023, 1235, 1407, 1634, 1697, 3356 cm⁻¹ ¹H NMR (400 MHz, CDCl₃, 25 °C): δ 0.76 (t, *J* = 7.2 Hz, 3H, CH₃), 0.88 (t, *J* = 7.2 Hz, 3H, CH₃), 1.03–1.16 (m, 4H, 2 × CH₂), 1.19 (t, *J* = 7.0 Hz, 3H, ester-CH₃), 1.29–1.36 (m, 2H, CH₂), 1.41–1.46 (m, 2H, CH₂), 2.38 (s, 3H, C6-CH₃), 2.48–2.58 (m, 2H, CH₂), 2.75 (br, 1H, D₂O exchangeable, NH), 2.90–2.98 (m, 2H, CH₂), 3.06 (br, 1H, D₂O exchangeable, NH), 4.05–4.14 (m, 2H, ester-CH₂), 6.12 (d, *J* = 6.0 Hz, 1H, C4-H), 7.23–7.32 (m, 3H, ArH), 7.46–7.48 (m, 2H, ArH), 8.44 (br, 1H, D₂O exchangeable, NH). ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ 13.6, 13.7, 14.1, 17.9, 19.6, 19.8, 33.7, 33.8, 40.0, 40.6, 56.9, 60.2, 104.2, 127.5, 127.9, 128.4, 142.5, 145.6, 154.2 and 165.2. ³¹P NMR (162 MHz, CDCl₃, 25 °C): δ 12.21; Anal. Calcd. for C₂₂H₃₅N₄O₄P: C, 58.67; H, 7.78; N, 12.44; Found: C, 58.30; H, 7.50; N, 12.22. MS: *m/z* 473 (M⁺+23).

5.4.2. 5-Ethoxycarbonyl-6-methyl-3-[bis(benzylamino) phosphinyl]-3,4-dihydropyrimidin-2(1H)-one (**9b**)

White crystalline solid. Rf: 0.7 (methanol:ethyl acetate/10:90). Yield: 60%. M.p. 183–185 °C (dichloromethane/hexane). IR (KBr): ν_{max} 1086, 1233, 1634, 1700, 3374 cm⁻¹ ¹H NMR (300 MHz, CDCl₃, 25 °C): δ 1.22 (t, J = 6.9 Hz, 3H, ester-CH₃), 2.29 (s, 3H, C6-CH₃), 3.11 (br, 1H, D₂O exchangeable, NH), 3.56 (t, J = 7.2 Hz, 1H, D₂O exchangeable, NH), 3.68–3.85 (m, 2H, CH₂), 4.08–4.19 (m, 4H, ester-CH₂ and CH₂), 6.13 (d, J = 6.0 Hz, 1H, C4-H), 6.71 (br, 1H, D₂O exchangeable, NH), 7.01–7.04 (m, 2H, ArH), 7.16–7.31 (m, 11H, ArH), 7.43–7.46 (m, 2H, ArH). ¹³C NMR (75 MHz, CDCl₃, 25 °C) δ 14.2, 18.8, 29.7, 55.8, 60.1, 101.5, 126.7, 128.0, 128.8, 143.7, 146.2, 153.3 and 165.7. ³¹P NMR (162 MHz, CDCl₃, 25 °C): δ 11.85. Anal. Calcd. for C₂₈H₃₁N₄O₄P: C, 64.86; H, 5.98; N, 10.81; Found: C, 64.66; H, 6.32; N, 10.52. MS: m/z 517 (M⁻-1).

5.4.3. 5-Ethoxycarbonyl-6-methyl-3-(diethylphosphonate)-4-phenyl-3,4-dihydropyrimidin-2(1H)-one (**9**c)

Compound 7 (R^1 = Ph, R^2 = Et, R^3 = H)was treated with ethanol at 80 °C for 3 h and after completion of reaction (TLC) solvent was removed under reduced pressure. Purification of the residue by column chromatography using hexane and ethylacetate as eluents resulted in the isolation of compound 9c as a white crystalline solid. Rf: 0.8 (methanol:ethyl acetate/20:80). Yield: 70%. M.p. 112–114 °C (dichloromethane/hexane). IR (KBr): $v_{\rm max}$ 1024, 1243, 1390, 1636, 1704, 3236 cm⁻¹ ¹H NMR (300 MHz, CDCl₃, 25 °C): δ 1.07 (t, J = 6.9 Hz, 3H, ester-CH₃), 1.22 (t, J = 7.2 Hz, 3H, CH₃), 1.32 (t, J = 7.2 Hz, 3H, CH₃), 2.38 (s, 3H, C6-CH₃), 3.52-3.67 (m, 1H, CHH), 3.72-3.82 (m, 1H, CHH), 4.07-4.26 $(m, 4H, 2 \times CH_2), 6.08 (d, I = 6.9 Hz, 1H, C4-H), 7.26-7.31 (m, 3H, C4-H), 7.26-7.31 (m, 3H, C4-H), 7.26-7.31 (m, 2H, C4-H), 7.26-7.31 (m, 2H,$ ArH), 7.40–7.46 (m, 2H, ArH), 7.69 (br, 1H, D₂O exchangeable, NH), ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ 14.1, 15.9, 17.9, 58.2, 60.3, 63.7, 64.4, 104.1, 127.2, 128.1, 128.5, 141.9, 145.8, 153.0 and 165.0. ³¹P NMR (162 MHz, CDCl₃, 25 °C): δ –0.53. Anal. Calcd. for C₁₈H₂₅N₂O₆P: C, 54.54; H, 6.31; N, 7.07.; Found: C, 54.20; H, 6.52; N, 6.80. MS: *m*/*z* 419 (M⁺+23).

5.5. General procedure for the synthesis of N3-substituted DHPM derivatives bearing phosphorous containing heterocycles **10**

To a stirred solution of appropriate 7 (1 equiv.) in dry dioxane (10 ml) an appropriate diamine or amino alcohols (1.5 equiv.) was added and the resulting solution was stirred at room temperature for 2-3 h. After completion of reaction (TLC) dioxane was removed under reduced pressure and residue was dissolved in ethylacetate (20–30 ml) and washed with water

 $(2\times30$ ml). Organic phase was dried over anhydrous sodium sulphate and compound 10 was obtained through crystallization with methanol. The characteristic data of the compounds is given below.

5.5.1. 2-[5-ethoxycarbonyl-6-methyl-4-phenyl-3,4dihydropyrimidin-2(1H)-one]-3-yl-1,3,4,5,6,7-hexahydro [1–3] diazaphosphepine-2-oxide (**10a**)

White crystalline solid. Rf: 0.3 (methanol:ethyl acetate/10:90). Yield: 72%. M.p. 218–220 °C (methanol). IR (KBr): ν_{max} 1029, 1228, 1390, 1641, 1690, 3369 cm⁻¹¹H NMR (300 MHz, CDCl₃, 25 °C): δ 1.23 (t, *J* = 7.2 Hz, 3H, ester-CH₃), 1.60–1.62 (m, 4H, 2 × CH₂), 2.39 (s, 3H, C6-CH₃), 3.03–3.12 (m, 3H, CH₂ and D₂O exchangeable, NH), 3.24–3.32 (m, 2H, CH₂), 3.71–3.73 (m, 1H, D₂O exchangeable, NH), 4.09–4.19 (m, 2H, ester-CH₂), 6.25 (d, *J* = 7.5 Hz, 1H, C4-H), 7.22–7.31 (m, 3H, ArH), 7.40–7.43 (m, 2H, ArH), 7.87 (br, 1H, D₂O exchangeable, NH). ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ 14.2, 16.0, 30.1, 34.6, 39.0, 111.0, 126.1, 126.9, 128.0, 144.0, 148.3, 158.0 and 166.9. ³¹P NMR (121 MHz, DMSO-*d*₆, 25 °C): δ 16.83; Anal. Calcd. for C₁₈H₂₅N₄O₄P: C, 55.10; H, 6.38; N, 14.28. Found: C, 54.82; H, 6.20; N, 14.40. MS: *m*/*z* 393 (M⁺+1).

5.5.2. 5-Ethoxycarbonyl-4,6-dimethyl-3-(1,3,2-

diazaphosphorinane-2-oxide)-3,4-dihydropyrimidin-2(1H)-one (**10b**)

White crystalline solid. Rf: 0.5 (methanol:ethyl acetate/20:80). Yield: 65%. M.p. 233–235 °C (methanol). IR (KBr): ν_{max} 1010, 1242, 1313, 1635, 1699, 3353 cm⁻¹ ¹H NMR (400 MHz, DMSO- d_6 , 25 °C): δ 0.87 (d, J = 6.5 Hz, 3H, C4-CH₃), 0.94 (t, J = 7.0 Hz, 3H, ester-CH₃), 1.65–1.72 (m, 2H, CH₂), 1.94 (s, 3H, C6-CH₃), 2.64 (t, J = 7.5 Hz, 4H, 2 × CH₂), 3.81–3.88 (m, 2H, ester-CH₂), 4.28 (br, 1H, D₂O exchangeable, NH), 4.56 (br, 1H, D₂O exchangeable, NH), 4.61 (q, J = 6.3 Hz, 1H, C4-H), 9.24 (br, 1H, D₂O exchangeable, N1-H). ¹³C NMR (100 MHz, DMSO- d_6 , 25 °C): δ 14.3, 17.2, 21.0, 25.0, 41.6, 49.0, 59.3, 103.2, 147.2, 153.5 and 165.0. ³¹P NMR (162 MHz, DMSO- d_6 , 25 °C): δ 4.81. Anal. Calcd. for C₁₂H₂₁N₄O₄P: C, 45.57; H, 6.64; N, 17.72; Found: C, 45.54; H, 6.51; N, 17.40. MS: *m/z* 317 (M⁺+1).

5.5.3. 5-Ethoxycarbonyl-6-methyl-3-(1,3,2-oxazaphosphino-2-yl)-4-phenyl-3,4-dihydropyrimidin-2(1H)-one (**10c**)

White crystalline solid. Rf: 0.2 (methanol:ethyl acetate/10:90). Yield: 65%. M.p. 208–210 °C (methanol). IR (KBr): ν_{max} 1019, 1247, 1365, 1635, 1705, 3333 cm⁻¹ ¹H NMR (400 MHz, DMSO- d_6 , 25 °C): δ 1.16 (t, J = 7.0 Hz, 3H, ester-CH₃), 1.72–1.84 (m, 2H, CH₂), 2.28 (s, 3H, C6-CH₃), 3.04–3.15 (m, 1H, CHH), 3.29–3.34 (m, 1H, CHH), 4.05–4.14 (m, 2H, ester-CH₂), 4.16–4.28 (m, 1H, CHH), 4.54–4.56 (m, 1H, CHH), 5.45–5.48 (m, 1H, D₂O exchangeable, NH), 6.07 (d, J = 7.5 Hz, 1H, C4-H), 7.20–7.32 (m, 5H, ArH). ¹³C NMR (100 MHz, DMSO- d_6 , 25 °C): δ 14.2, 17.4, 25.4, 41.2, 55.9, 59.8, 69.4, 102.7, 126.1, 127.3, 128.4, 142.1, 148.1, 153.3 and 165.2. ³¹P NMR (162 MHz, DMSO- d_6 , 25 °C): δ –0.94. Anal. Calcd. for C₁₇H₂₂N₃O₅P: C, 53.83; H, 5.80; N, 11.08. Found: C, 53.52; H, 5.63; N, 10.82. MS: *m/z* 402 (M⁺+23).

5.5.4. 5-Ethoxycarbonyl-1,6-dimethyl-3-(1,3,2-oxazaphosphino-2-yl)-4-phenyl-3,4-dihydropyrimidin-2(1H)-one (**10d**)

White crystalline solid. Rf: 0.4 (methanol:ethyl acetate/20:80). Yield: 65%. M.p. 213–215 °C (methanol). IR (KBr): ν_{max} 1015, 1225, 1345, 1665, 1710, 3343 cm⁻¹ ¹H NMR (400 MHz, DMSO- d_6 , 25 °C): δ 1.18 (t, J = 7.0 Hz, 3H, ester-CH₃), 1.74–1.87 (m, 2H, CH₂), 2.50 (s, 3H, C6-CH₃), 3.07–3.13 (m, 4H, N1-CH₃ and CH<u>H</u>), 3.28–3.34 (m, 1H, C<u>H</u>H), 4.13–4.18 (m, 2H, ester-CH₂), 4.21–4.29 (m, 1H, CH<u>H</u>), 4.58–4.64 (m, 1H, C<u>H</u>H), 5.45–5.48 (m, 1H, D₂O exchangeable, NH), 6.13 (d, J = 8.0 Hz, 1H, C4-H), 7.22–7.33 (m, 5H, ArH). ¹³C NMR (100 MHz, DMSO- d_6 , 25 °C): δ 14.1, 15.7, 25.3, 30.5, 41.1, 54.4, 59.9, 69.4, 106.2, 125.8, 127.3, 128.3, 141.2, 150.2, 153.8 and 165.2. Anal. Calcd. for C₁₈H₂₄N₃O₅P: C, 54.96; H, 6.10; N, 10.68; Found: C, 54.66; H, 6.21; N, 10.40. MS: *m/z* 416 (M⁺+23).

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References

- (a) R.A. Janis, D.J. Trigger, New developments in calcium ion channel antagonists, J. Med. Chem. 26 (1983) 775–785.
- [2] B. Loev, M.M. Goodman, K.M. Snader, R. Tedeschi, E. Macko, Hantzsch-type dihydropyridine hypotensive agents, J. Med. Chem. 17 (1974) 956–965.
- [3] P. Biginelli, Aldehyde-uridic derivative of ether acetyl and Diossalacetic, Gazz. Chim. Ital. 23 (1893) 360-416.
- [4] K.S. Atwal, B.N. Swanson, S.E. Unger, D.M. Floyd, S. Moreland, A. Hedberg, B.C.O'Reilly, Dihydropyrimidine calcium channel blockers. 3. 3-Carbamoyl-4aryl-1,2,3,4-tetrahydro-6-methyl-5-pyrimidinecarboxylic acid esters as orally effective antihypertensive agents, J. Med. Chem. 34 (1991) 806–811.
- [5] K.S. Atwal, G.C. Rovnyak, S.D. Kimball, D.M. Floyd, S. Moreland, B.N. Swanson, J.Z. Gougoutas, J. Schwartz, K.M. Smillie, M.F. Malley, Dihydropyrimidine calcium channel blockers. II. 3-Substituted-4-aryl-1,4-dihydro-6-methyl-5pyrimidinecarboxylic acid esters as potent mimics of dihydropyridines, J. Med. Chem. 33 (1990) 2629–2635.
- [6] C.O. Kappe, Biologically active dihydropyrimidones of the Biginelli-typea literature survey, Eur. J. Med. Chem. 35 (2000) 1043–1052.
- [7] K. Singh, D. Arora, K. Singh, S. Singh, Genesis of dihydropyrimidinone calcium channel blockers: recent progress in structure-activity relationships and other effects, Mini Rev. Med. Chem. 9 (2009) 95–106.
- [8] K. Singh, D. Arora, E. Poremsky, J. Lowery, R.S. Moreland, N1-Alkylated 3,4dihydropyrimidine-2(1*H*)-ones: Convenient one-pot selective synthesis and evaluation of their calcium channel blocking activity, Eur. J. Med. Chem. 44 (2009) 1997–2001.
- [9] K. Singh, D. Arora, D. Falkowski, Q. Liu, R.S. Moreland, An efficacious protocol for 4-Substituted 3,4-Dihydropyrimidinones: synthesis and calcium channel binding studies, Eur. J. Org. Chem. (2009) 3258–3264.
- [10] K. Singh, S. Singh, Chemical resolution of inherently racemic dihydropyrimidinones via a site selective functionalization of Biginelli compounds with chiral electrophiles: a case study, Tetrahedron 65 (2009) 4106–4112.
- [11] K. Singh, S. Singh, A mild and practical method for the regioselective synthesis of N-acylated 3,4-dihydropyrimidin-2-ones. New acyl transfer reagents, Tetrahedron Lett. 47 (2006) 8143–8146.
- [12] K. Singh, K. Singh, N1, N3-Diacyl-3,4-dihydropyrimidin-2(1H)-ones neutral acyl group transfer reagents, Tetrahedron 65 (2009) 10395–10399.
- [13] I. Morita, S.I. Tada, K. Kunimoto, M. Tsuda, M. Kise, K. Kimura, Synthesis and antihypertensive activities of 1,4-Dihydropyridine-5-phosphonate derivatives, Chem. Pharm. Bull. 35 (1987) 3898–3904.
- [14] F.H. Westheimer, Why nature chose phosphates, Science 235 (1987) 1173–1178.
- [15] H. Seto, T. Kuzuyama, Bioactive natural products with carbon phosphorus bonds and their biosynthesis, Nat. Prod. Rep. 16 (1999) 589–596.
- [16] B. Lejczak, D. Dus, P. Kafarski, Phosphonic and phosphinic acid analogues of tyrosine and 3,4-dihydroxyphenylalanine (dopa) as potential antimelanotic agents, Anti-Cancer Drug Des. 5 (1990) 351–358.
- [17] O.M. Colvin, An overview of cyclophosphamide development and clinical applications, Curr. Pharm. Des. 5 (1999) 555–560.
- [18] M.M. Mader, P.A. Bartlett, Binding energy and catalysis: the implications for transition-state analogs and catalytic antibodies, Chem. Rev. 97 (1997) 1281–1302.
- [19] P. Kafarski, B. Lejczak, Anticancer agents. Aminophosphonic acids of potential medical importance, Curr. Med. Chem. 1 (2001) 301–312.
- [20] Crystallographic data for 8b have been deposited with the Cambridge Crystallographic Data Centre (CCDC). The coordinates can be obtained on request from the Director, CCDC, 12 Union Road, Cambridge CB2 1 EZ, UK. The CCDC Number is 677322.
- [21] G.C. Rovnyak, S.D. Kimball, B. Beyer, G. Cucinotta, J.D. DiMarco, J. Gougoutas, A. Hedberg, M. Malley, J.P. McCarthy, R. Zhang, S. Moreland, Calcium entry blockers and activators: conformational and structural determinants of dihydropyrimidine calcium channel modulators, J. Med. Chem. 38 (1995) 119–129.
- [22] S. Moreland, R.S. Moreland, Effects of dihydropyridines on stress, myosin phosphorylation, and V0 in smooth muscle, Am. J. Physiol. 252 (1987) H1049-H1058.