

# Nucleoside H-Phosphonates XX. Efficient Method for the Preparation of Nucleoside H-Phosphonoselenoate Monoesters

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**Abstract:** The preparation of an H-phosphonoselenoyl group transferring reagent, 9-fluorenemethyl H-phosphonoselenoate, and its application to the synthesis of separate diastereomers of nucleoside 3'-H-phosphonoselenoate monoesters, are described.

**Key words:** nucleotides, phosphorylation, selenium, protecting groups, elimination

## Introduction

A close resemblance of selenium to biologically important element sulfur, constitutes a strong rationale for incorporation of selenium into potential medicinal agents. The toxic nature of most selenium compounds may pose a serious obstacle in drug development but does not *per se* rule out this type of modification for drug use.<sup>1</sup> Kindled by hopes of finding novel, useful properties, selenium has been incorporated into various biologically important compounds, e.g. carbohydrates,<sup>2</sup> lipids,<sup>3,4</sup> nucleosides,<sup>5</sup> oligonucleotides.<sup>6,7</sup>

Selenophosphates are usually prepared via selenization of suitable P(III) precursors, e.g. phosphite triesters,<sup>6,8</sup> H-phosphonate<sup>3,7</sup> and H-phosphonothioate<sup>7</sup> diesters, although derivatives accessible by these routes are usually limited to the corresponding selenophosphate di- and triesters, and selenothiophosphate diesters. Recently, we started to explore a new type of synthetic intermediates, H-phosphonoselenoate monoesters<sup>9,10</sup> that can provide access to new selenophosphate derivatives with double modifications at the phosphorus centre.

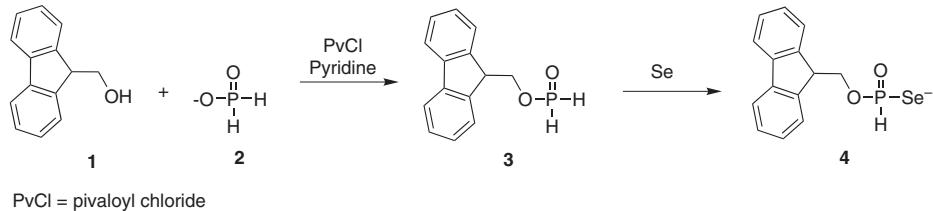
In this paper we report on the synthesis and application of a dedicated H-phosphonoselenoyl group-transferring reagent, namely, 9-fluorenemethyl H-phosphonoselenoate, that permits an easy preparation of nucleoside H-phosphonoselenoate monoesters either as diastereomeric mixtures or as separate P-diastereomers. This latter possibility can be potentially exploited in a stereospecific synthesis of P-chiral selenophosphate derivatives.

## Results and Discussion

Since nucleoside H-phosphonoselenoate monoesters are chiral at the phosphorus centre, access to the separate  $R_P$  and  $S_P$  diastereomers is required to exploit in full their synthetic potential. Unfortunately, the presence of a negative charge in an H-phosphonoselenoate moiety usually overshadows possible chromatographic differences between isomeric compounds and effectively prevents separation of their P-diastereomers on silica gel.<sup>9</sup> To overcome this problem, we designed a dedicated H-phosphonoselenoyl group transferring reagent, 9-fluorenemethyl H-phosphonoselenoate **4** (Scheme 1), which upon reaction with a hydroxylic component (e.g. a nucleoside) can produce uncharged H-phosphonoselenoate diesters of type **6** (Scheme 2). These are much more easy to separate into diastereomers and after the removal of the fluorenemethyl group,  $R_P$  and  $S_P$  diastereomers of H-phosphonoselenoate monoesters **7** can be obtained. The fluorenemethyl group in H-phosphonoselenoate diesters **6** acts as a lipophilic handle that facilitates separation of these compounds by a silica gel column chromatography, and since it is removable via a  $\beta$ -elimination mechanism, its deprotection does not affect stereochemical integrity of the phosphorus center. The added value of this approach is that, if separate diastereomers of H-phosphonoselenoates **7** are not required the synthesis can be simplified by subjecting H-phosphonoselenoate diesters **6** to deprotection directly after condensation to afford diastereomeric mixture of the corresponding H-phosphonoselenoate monoesters.

### Synthesis of H-Phosphonoselenoyl Group Transferring Reagent **4**

9-Fluorenemethyl H-phosphonoselenoate **4**<sup>10</sup> was prepared using standard phosphinate approach previously developed for the synthesis of H-phosphonothioate monoesters.<sup>11</sup> Thus treatment of 9-fluorenemethanol (**1**) with triethylammonium phosphinate (**2**) (<sup>31</sup>P NMR:  $\delta = 2.85$ ,  $^1J_{P,H} = 517$  Hz, t) in pyridine in the presence of pivaloyl chloride, followed by selenization of the produced phosphinate intermediate **3** (<sup>31</sup>P NMR:  $\delta = 14.22$ ,  $^1J_{P,H} = 572$  Hz,  $^3J_{P,H} = 9.8$  Hz, tt) with elemental selenium for two hours, produced 9-fluorenemethyl H-phosphonoselenoate **4** as the major product (>90%, <sup>31</sup>P NMR:  $\delta = 50.41$ ,  $^1J_{P,H} = 570$  Hz,  $^3J_{P,H} = 9.1$  Hz, dt;  $^1J_{P,Se} = 685.2$  Hz) (Scheme 1). Since reagent **4** in the form of triethylam-



Scheme 1

monium salt was a sticky oil and difficult to handle, it was converted into a *S*-(*p*-chlorobenzyl)isothiuronium salt (88% overall yield). This solid salt was stable, non-hygroscopic and could be used directly for the condensations without necessity of changing the cationic part.

### Synthesis of Nucleoside 3'-H-Phosphonoselenoate Monoesters 7

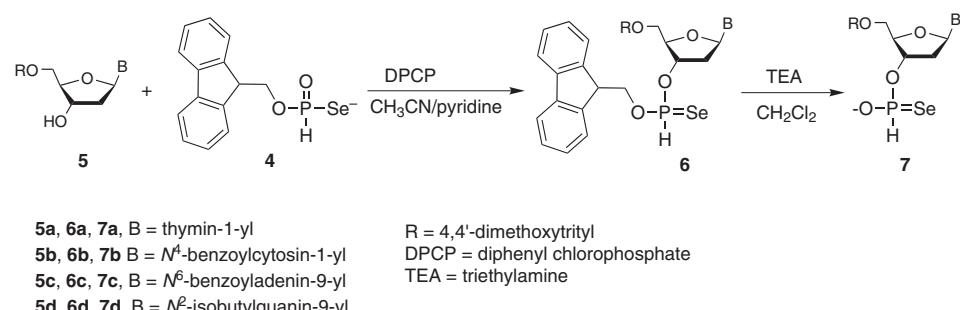
First, reaction conditions for condensation of H-phosphonoselenoate **4** with a hydroxyl component **5** to produce 9-fluorenemethyl derivatives **6** were investigated. Among the condensing agents tried [pivaloyl chloride, 2-chloro-2-oxo-5,5-dimethyl-1,3,2-dioxaphosphinane, diphenyl phosphorochloridate (DPCP)],<sup>12</sup> the best in terms of chemoselectivity, purity of the produced compounds and the shortest reaction time, turned out to be DPCP. With this reagent (2.5 equiv), a model reaction of nucleoside **5a** with H-phosphonoselenoate **4** (1.1 equiv) in acetonitrile containing pyridine (5 equiv) was complete within 10 minutes affording the expected H-phosphonoselenoate **6a** as the sole nucleotidic material [<sup>31</sup>P NMR:  $\delta$  = 77.29 ( $^1J_{P,Se}$  = 870 Hz) and 76.43 ( $^1J_{P,Se}$  = 871 Hz)]. These reaction conditions worked well also for other nucleosides **5** investigated (Scheme 2).

Due to some instability of nucleoside H-phosphonoselenoate diesters **6** in aqueous basic media, the work-up of the reaction mixtures after condensations was critical. Optimal conditions found consisted of quenching the reaction mixtures with water containing pyridine, followed by the dilution with toluene–ethyl acetate, and passing the mixture through silica gel pad. This procedure provided fluorenemethyl nucleoside H-phosphonate **6** of purity >98% (<sup>1</sup>H NMR spectroscopy) and in consistently high yields.

Removal of a fluorenemethyl group from H-phosphonoselenoates **6** to produce the corresponding H-phosphonoselenoate monoesters **7** was rather straightforward and could be effected quantitatively by treatment of **6** in dichloromethane with triethylamine (TEA) for 1 h. Progress of this reaction can be conveniently monitored by TLC analysis (decrease in mobility on a silica gel upon conversion to **7**) or by <sup>31</sup>P NMR spectroscopy. In the latter method, the removal of the fluorenemethyl group from **6** is accompanied by a huge change in the chemical shift from that of ca. 70 ppm for compounds of type **6** to ca. 45 ppm for compounds **7**. Other diagnostic parameters that vary significantly with structure are one-bond phosphorus–selenium ( $^1J_{P,Se}$ ) and phosphorus–hydrogen ( $^1J_{P,H}$ ) coupling constants (see experimental section).

If separated diastereomers of nucleoside H-phosphonoselenoate monoester **7** would be required, fluorenemethyl derivatives **6** could be separated into *R*<sub>P</sub> and *S*<sub>P</sub> diastereomers via silica gel chromatography prior to the deprotection step. As expected, the removal of the fluorenemethyl group from **6** was completely stereospecific and occurred most likely with retention of configuration.

Although the absolute configurations at the phosphorus centre in compounds **6** and **7** are yet remain to be determined, there seems to be a relationship between the configuration and the observed <sup>31</sup>P NMR chemical shift or the mobility on silica gel, as reported previously for other P-chiral nucleotide analogues.<sup>13</sup> For the compounds investigated, chromatographically faster moving diastereomers of **6** always resonated at higher field in the <sup>31</sup>P NMR spectra, while those of the lower chromatographic mobility, gave signals at lower field. For H-phosphonoselenoate monoesters **7**, the diastereomers showed almost identical mobility on silica gel, but they differed in the <sup>31</sup>P NMR shift values. Since P-diastereomers of **7** resonating at high



Scheme 2

field were formed ‘high field’ (or ‘fast’) diastereomers of **6**, while those at low field, were formed from ‘low field’ (or ‘slow’) **6**, these established a configurational relationship between diastereomeric compounds **6** and **7**.

## Conclusion

In conclusion, we have developed a dedicated reagent, 9-fluorenemethyl H-phosphonoselenoate **4**, for transferring an H-phosphonoselenoate moiety into hydroxylic components, and used it for a simple preparation in high yields of the separate *R*<sub>P</sub> and *S*<sub>P</sub> diastereomers of nucleoside H-phosphonoselenoate monoesters **7**. The reagent is stable, easy to prepare, and has a fluorenemethyl group as a lipophilic handle that facilitates separation into diastereomers of the intermediate produced, 9-fluorenemethyl nucleoside H-phosphonoselenoate diesters **6**. The fluorenemethyl group can be removed via  $\beta$ -elimination to keep the integrity of the configuration at the phosphorus intact. In addition, 9-fluorenemethyl nucleoside H-phosphonoselenoates **6**, which are accessible in diastereomerically pure forms, can be considered as potentially useful intermediates on their own for the preparation of various P-chiral phosphoroselenoate monoester analogues.

<sup>1</sup>H and <sup>31</sup>P NMR spectra were recorded on a Varian Unity 400 BB VT spectrometer. The <sup>31</sup>P NMR experiments were carried out at 25 °C in 5 mm tubes using 0.1 M concentrations of phosphorus-containing compounds in appropriate solvents (0.6 mL), and the spectra were referenced to 2% H<sub>3</sub>PO<sub>4</sub> in D<sub>2</sub>O (external standard). TLC analyses were carried out on Merck silica gel 60 F<sub>254</sub> precoated plates using the following solvent systems: (A) CHCl<sub>3</sub>–MeOH (9:1); (B) toluene–EtOAc (1:1). Pyridine (LabScan Ltd.) and anhyd MeCN (LabScan Ltd.) were stored over molecular sieves 4 Å. The starting materials for the synthesis, suitably protected nucleoside **5** were prepared by modifications of published procedures.<sup>14</sup> Anhyd triethylammonium phosphinate (**2**) was prepared by neutralization of 50% aq phosphinic acid with Et<sub>3</sub>N, followed by repeated evaporation of added pyridine.

The assignments of signals in the <sup>31</sup>P NMR spectra to particular products or intermediates were done on the basis of their chemical shifts, multiplicity of the signals in <sup>1</sup>H-coupled and <sup>1</sup>H-decoupled spectra, by spiking the reaction mixtures with appropriate species and, if possible, by isolation of a compound in question from reaction mixtures. The assignment of proton and carbon resonances of **4**, **6** and **7** was done on the basis of known or expected chemical shifts in conjunction with <sup>1</sup>H–<sup>1</sup>H, <sup>1</sup>H–<sup>13</sup>C, and DEPT correlated NMR spectroscopy.

### Fluorenemethyl H-Phosphonoselenoate (S-Chlorobenzyl)isothiuronium Salt (**4**)

Fluorenylmethanol (1.0 g, 5.1 mmol, 1.5 equiv) and triethylammonium phosphinate (568 mg, 3.4 mmol, 1 equiv) were rendered anhydrous by coevaporation of added pyridine and then dissolved in pyridine (15 mL). The solution was cooled to 0 °C, pivaloyl chloride (628 μL 5.1 mmol, 1.5 equiv), followed by elemental Se (805 mg, 10.2 mmol, 3 equiv) were added, and the reaction mixture was allowed to attain r.t. After stirring for 3 h, the solution was filtered through Celite onto H<sub>2</sub>O (200 μL), the filtrate was concentrated and then dissolved in CHCl<sub>3</sub> (100 mL). This solution was washed with H<sub>2</sub>O (3 × 50 mL; to the last extraction 200 μL of Et<sub>3</sub>N was added to improve the separation of layers), the organic phase was dried

(Na<sub>2</sub>SO<sub>4</sub>), filtered and then concentrated. The residue was co-evaporated with toluene (100 mL) to remove pyridine. The residue was added on top of a silica gel column that had been equilibrated with CH<sub>2</sub>Cl<sub>2</sub> containing 1% MeOH and 0.2% Et<sub>3</sub>N, and the product was eluted with CH<sub>2</sub>Cl<sub>2</sub> containing 1% MeOH. This afforded fluorenemethyl H-phosphonoselenoate **4** (1.32 g, triethylammonium salt) in 96% yield as a sticky, colorless oil. The residue was dissolved in Et<sub>2</sub>O (100 mL), and H<sub>2</sub>O (100 mL) together with S-(4-chlorobenzyl)isothiuronium chloride (1.1 equiv) was added. The H<sub>2</sub>O phase was extracted with Et<sub>2</sub>O (2 × 100 mL). The organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated to yield the fluorenemethyl H-phosphonoselenoate S-(4-chlorobenzyl)isothiuronium salt as a white foam in 88% overall yield (1.51 g). The foam can be dissolved in minimal amount of CHCl<sub>3</sub> and precipitated from pentane to give the product as a white powder (1.32 g) in 77% overall yield; *R*<sub>f</sub> 0.42 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 8.53 (br, 4 H, NH), 8.49 (d, *J* = 570 Hz, PH), 7.68 (d, *J* = 7.32 Hz, 2 H, H-4 and H-5), 7.56 (dd, *J* = 7.32, 2.54, 2 H, H-1, H-8), 7.33 (t, *J* = 7.32 Hz, 2 H, H-3 and H-6), 7.21 (m, 2 H, H-2 and H-7), 7.15–7.07 (q, 4 H, *J* = 8.42 Hz, ArH of chlorobenzyl), 4.29–4.24 (m, 1 H, H-9), 4.17–4.14 (m, 4 H, 2 CH<sub>2</sub>).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 169.94 (C of thiourea), 144.10, 144.02, 141.44 and 141.36 (4 C of fluorenyl), 134.63 (*ipso* C of chlorobenzyl), 131.31 (*para* C of chlorobenzyl), 130.50 and 129.38 (4 CH of chlorobenzyl), 127.85 (C-3 and C-6), 127.26 and 127.20 (C-1 and C-2), 125.36 and 125.31 (C-1 and C-8), 120.06 (C-4 and C-5), 67.41 (C-9), 48.20 (CH<sub>2</sub> at C-9), 35.40 (CH<sub>2</sub>).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 44.64 (<sup>1</sup>J<sub>P,H</sub> = 570 Hz, <sup>1</sup>J<sub>P,Se</sub> = 685.2 Hz, <sup>3</sup>J<sub>P,H</sub> = 9.1 Hz).

HRMS: *m/z* calcd for C<sub>22</sub>H<sub>23</sub>ClN<sub>2</sub>O<sub>2</sub>PSe [M + H]<sup>+</sup>: 525.0072; found: 525.0066.

Anal. Calcd for C<sub>22</sub>H<sub>22</sub>ClN<sub>2</sub>O<sub>2</sub>PSe: C, 50.44; H, 4.23; N, 5.35. Found: C, 50.14; H, 4.10; N, 5.42.

### 5'-O-Dimethoxytritylnucleoside Fluorenemethyl 3'-H-Phosphonoselenoate Diesters **6**; General Procedure

To a solution of 5'-O-dimethoxytritylnucleoside **5** (272 mg, 0.5 mmol) and fluorenemethyl H-phosphonoselenoate S-(4-chlorobenzyl)isothiuronium salt (**4**; 278 mg, 0.55 mmol, 1.1 equiv) in MeCN (5 mL) containing pyridine (201 μL, 2.5 mmol, 5 equiv), was added diphenyl chlorophosphate (DPCP, 259 μL, 1.25 mmol, 2.5 equiv). After 10 min, the reaction mixture was quenched with H<sub>2</sub>O (2 mL) and pyridine (806 μL, 20 equiv), stirred for 10 min and then diluted with EtOAc (50 mL) and toluene (50 mL). The insoluble material was removed and the organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>). After concentration, the residue was passed through a silica gel pad (3 × 3 cm) and washed with toluene–EtOAc (1:1, 200 mL). This gave the desired products **6** as a mixture of diastereoisomers of purity >98% (<sup>1</sup>H NMR spectroscopy). Chromatography of the diastereomeric mixture of **6** on a silica gel column using toluene–EtOAc (2:1) as an eluent, afforded separate diastereomers of **6**, designated as ‘fast’ and ‘slow’, according to their chromatographic mobility.

### 5'-O-Dimethoxytritylthymidine Fluorenemethyl 3'-H-Phosphonoselenoate Diester (**6a**)

White solid (1:1 mixture of diastereomers); yield: 357 mg (84%); *R*<sub>f</sub> 0.62 and 0.57 (System B).

HRMS: *m/z* calcd for C<sub>45</sub>H<sub>44</sub>N<sub>2</sub>O<sub>8</sub>PSe [M + H]<sup>+</sup>: 851.2000; found: 851.2010.

#### Fast Moving Diastereoisomer of **6a**

*R*<sub>f</sub> 0.62 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 9.19 (s, 1 H, NH), 8.20 (d, *J* = 650 Hz, 1 H, PH), 7.76 (dd, *J* = 7.32, 3.66 Hz, 2 H, H-4 and H-5 of fluorenyl), 7.63 (dd, *J* = 7.01, 2.13 Hz, 2 H, H-1 and H-8 of fluorenyl), 7.45–

7.20 (m, 13 H, ArH), 6.84 (d,  $J = 8.84$  Hz, 4 H, ArH *ortho* to OMe), 6.38 (t,  $J = 7.62$  Hz, 1 H, H-1'), 5.36 (m, 1 H, H-3'), 4.58–4.40 (m, 2 H, CH<sub>2</sub>), 4.25 (t,  $J = 6.40$  Hz, 1 H, H-9 of fluorenyl), 4.14 (m, 1 H, H-4'), 3.78 (s, 6 H, 2 CH<sub>3</sub>O), 3.41 (m, 2 H, H-5'), 2.3–2.2 (m, 2 H, H-2'), 1.47 (s, 3 H, CH<sub>3</sub> at C-5).

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 164.02$  (C-4), 159.02 [2 C of dimethoxytrityl (DMT)], 150.62 (C-2), 144.47 (2 C of DMT), 143.16 and 143.03 (2 C of fluorenyl), 135.49, 135.44 and 135.37 (2 C of DMT, C-6), 130.31, 128.39, 128.31, 127.55, 127.51 and 127.44 (13 ArCH), 125.26 (C-1 and C-8 of fluorenyl), 120.45 and 120.42 (C-4 and C-5 of fluorenyl), 113.62 (4 C of DMT), 111.85 (C-5), 87.50 (C of DMT), 85.12 (d,  $J = 5.44$  Hz, C-4'), 84.57 (C-1'), 77.79 (C-3'), 69.42 (d,  $J = 8.00$  Hz, CH<sub>2</sub> of fluorenyl), 63.29 (C-5'), 55.51 (2 CH<sub>3</sub>O), 48.03 (d,  $J = 7.73$  Hz, CH of fluorenyl), 39.25 (d,  $J = 4.00$  Hz, C-2'), 12.00 (CH<sub>3</sub> at C-5).

<sup>31</sup>P NMR (CDCl<sub>3</sub>):  $\delta = 70.89$  ( $^1J_{P,Se} = 870.8$  Hz,  $^1J_{P,H} = 649.5$  Hz).

### Slow Moving Diastereoisomer of 6a

$R_f$  0.57 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 9.49$  (s, 1 H, NH), 8.31 (d,  $J = 647$  Hz, 1 H, PH), 7.90–7.10 (m, 17 ArH and H-6), 6.85 (d,  $J = 8.84$  Hz, 4-H, ArH *ortho* to OMe), 6.47–6.42 (m, 1 H, H-1'), 5.53–5.47 (m, 1 H, H-3'), 4.53–4.43 (m, 2 H, CH<sub>2</sub> of fluorenyl), 4.27–4.15 (m, 1 H, CH of fluorenyl), 4.04 (br, 1 H, H-4'), 3.78 (s, 6 H, 2 CH<sub>3</sub>O), 3.40–3.30 (m, 2 H, H-5'), 2.56–2.27 (m, 2 H, H-2'), 1.50 (s, 3 H, CH<sub>3</sub> at C-5).

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 164.14$  (C-4), 159.03 (2 C of DMT), 150.80 (C-2), 144.40 (2 C of DMT), 143.14 and 142.98 (2 C of fluorenyl), 141.63 and 141.58 (2 C of fluorenyl), 135.50, 135.41 and 135.34 (2 C of DMT and C-6), 130.29, 128.32 and 127.50 (13 ArCH), 125.37 and 125.16 (C-1 and C-8 of fluorenyl), 120.39 (C-4 and C-5 of fluorenyl), 113.65 (4 C of DMT), 111.95 (C-5), 87.50 (C of DMT), 85.05 (C-4'), 84.69 (C-1'), 78.37 (C-3'), 69.27 (d,  $J = 6.87$  Hz, CH<sub>2</sub> of fluorenyl), 63.19 (C-5'), 55.51 (2 CH<sub>3</sub>O), 47.95 (d,  $J = 8.02$  Hz, CH of fluorenyl), 39.41 (d,  $J = 5.73$ , C-2'), 12.06 (CH<sub>3</sub> at C-5).

<sup>31</sup>P NMR (CDCl<sub>3</sub>):  $\delta = 71.66$  ( $^1J_{P,Se} = 871.2$  Hz,  $^1J_{P,H} = 647.0$  Hz).

### 5'-O-Dimethoxytrityl-N<sup>4</sup>-benzoylcytidine Fluorenemethyl 3'-H-Phosphonoselenoate Diester (6b)

White solid (1:1 mixture of diastereomers); yield: 436 mg (90%);  $R_f$  0.45 and 0.37 (System B).

HRMS: *m/z* calcd for C<sub>51</sub>H<sub>47</sub>N<sub>3</sub>O<sub>8</sub>PSe [M + H]<sup>+</sup>: 940.2266; found: 940.2276.

### Fast Moving Diastereoisomer of 6b

$R_f$  0.45 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 8.87$  (br, 1 H, NH), 8.16 (d,  $J = 649$  Hz, 1 H, PH), 8.14 (d,  $J = 7.42$  Hz, ArH), 7.89 (d,  $J = 7.42$  Hz, 1 H, ArH), 7.73 (t,  $J = 7.42$  Hz, 1 H, ArH), 7.61 (d,  $J = 7.42$  Hz, 1 H, ArH), 7.58–7.11 (m, 12 H, ArH, H-5 and H-6), 6.84 (d,  $J = 8.79$  Hz, 4 H, *ortho* to OMe), 6.28–6.21 (m, 1 H, H-1'), 5.27–5.21 (m, 1 H, H-3'), 4.57–4.39 (m, 2 H, H-4' and CH of fluorenyl), 3.76 (s, 6 H, 2 CH<sub>3</sub>O), 3.43–3.41 (m, 2 H, H-5'), 2.66–2.60 (m, 1 H, H-2'), 2.19–2.08 (m, 1 H, H-2').

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 162.56$  (C=O of Bz, C-4 and C-2), 158.99 (2 C of DMT), 144.57 (ArC), 144.21 and 144.19 (2 C of DMT), 143.15 and 143.00 (2 C of fluorenyl), 141.61 and 141.58 (2 C of fluorenyl), 135.39, 135.26 and 135.24 (2 C of DMT and ArC), 133.40 (ArCH), 130.36, 130.30, 129.29, 129.24, 128.33, 127.89 and 127.51 (ArCH, C-6 and C-5), 125.44 and 125.25 (C-1 and C-8 of fluorenyl), 120.42 (ArCH), 113.64 (4 C of DMT), 87.49 (C-1'), 85.78 (C-4'), 69.43 (CH<sub>2</sub> of fluorenyl), 62.64 (C-5'), 55.49 (2 CH<sub>3</sub>O), 48.05 (d,  $J = 8.43$  Hz, CH of fluorenyl), 40.73 (C-2').

<sup>31</sup>P NMR: (CDCl<sub>3</sub>):  $\delta = 74.57$  ( $^1J_{P,Se} = 872.44$  Hz,  $^1J_{P,H} = 648.9$  Hz).

### Slow Moving Diastereoisomer of 6b

$R_f$  0.37 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 8.69$  (br, 1 H, NH), 8.26 (d,  $J = 644$  Hz, 1 H, PH), 8.07 (d,  $J = 7.42$  Hz, 1 H, ArH), 7.89 (d,  $J = 7.23$  Hz, 2 H, ArH), 7.69 (d,  $J = 7.62$  Hz, 2 H, ArH), 7.55 (t, 2 H,  $J = 7.42$  Hz, 2 H, ArH), 7.51–7.10 (m, 12 H, ArH, H-5 and H-6), 6.80–6.77 (m, 4 H, *ortho* to OMe), 6.26 (t,  $J = 6.44$  Hz, 1 H, H-1'), 5.90–5.32 (m, 1 H, H-3'), 4.81–4.40 (m, 1 H, CH<sub>2</sub> of fluorenyl), 4.22–4.12 (m, 3 H, H-4', CH and CH<sub>2</sub> of fluorenyl), 3.73 (s, 3 H, CH<sub>3</sub>O), 3.72 (CH<sub>3</sub>O), 3.36–3.31 (m, 2 H, H-5'), 2.83–2.78 (m, 1 H, H-2'), 2.25–2.17 (m, 1 H, H-2').

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 162.39$  (C=O of Bz, C-4 and C-2), 158.94 (2 C of DMT), 144.42 (ArC), 144.07 (2 C of DMT), 143.10 and 142.93 (2 C of fluorenyl), 141.55 (2 C of DMT), 135.31 and 135.12 (2 C of DMT), 133.41 (ArCH), 130.36, 130.19, 129.26, 128.28, 127.75 and 127.46 (ArCH, C-5 and C-6), 125.35 and 125.14 (C-1 and C-8 of fluorenyl), 120.33 (ArCH), 113.59 (4 C of DMT), 87.43 (C-1'), 85.72 (C-4'), 69.21 (CH<sub>2</sub> of fluorenyl), 62.58 (C-5'), 55.44 (2 CH<sub>3</sub>O), 47.91 (d,  $J = 7.67$  Hz, CH of fluorenyl), 40.72 (C-2').

<sup>31</sup>P NMR (CDCl<sub>3</sub>):  $\delta = 75.137$  ( $^1J_{P,Se} = 872.4$  Hz,  $^1J_{P,H} = 644$  Hz).

### 5'-O-Dimethoxytrityl-N<sup>6</sup>-benzoyladenosine Fluorenemethyl 3'-H-Phosphonoselenoate Diester (6c)

White solid (1:1 mixture of diastereomers); yield: 433 mg (90%);  $R_f$  0.43 and 0.38 (System B).

HRMS: *m/z* calcd for C<sub>52</sub>H<sub>47</sub>N<sub>5</sub>O<sub>7</sub>PSe [M + H]<sup>+</sup>: 964.2378; found: 964.2389.

### Fast Moving Diastereoisomer of 6c

$R_f$  0.43 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 9.10$  (s, 1 H, NH), 8.71 (s, 1 H, H-2), 8.28 (d,  $J = 648$  Hz, 1 H, PH), 8.11 (s, 1 H, H-8), 8.03 (d,  $J = 7.32$  Hz, 2 H, ArH), 7.71 (d,  $J = 6.71$  Hz, 1 H, ArH), 7.67–7.48 (m, 5 H, ArH), 7.40–7.12 (m, 14 H, ArH), 6.79 (d,  $J = 8.84$  Hz, 4 H, *ortho* to OMe), 6.32 (dd,  $J = 8.38$ , 5.64 Hz, 1 H, H-1'), 5.40–5.33 (m, 1 H, H-3'), 4.62–4.47 (m, 2 H, H-4' and CH of fluorenyl), 3.76 (s, 6 H, 2 CH<sub>3</sub>O), 3.39–3.37 (m, 2 H, H-5'), 2.85–2.74 (m, 1 H, H-2'), 2.33–2.24 (m, 1 H, H-2').

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 164.83$  (C=O of Bz), 158.84 (2 C of DMT and C-6), 152.87 (C-2), 151.81 (C-6), 149.78 (C-4), 144.56 (2 C of DMT), 143.23 and 142.99 (2 C of fluorenyl), 141.72 (C-8), 141.57 (2 C of DMT), 135.69 and 135.64 (2 C of DMT), 133.87, 133.04, 130.28, 129.12, 128.34, 128.28, 128.18, 128.10, 127.51 and 127.23 (ArCH), 125.22 (C-1 and C-8 of fluorenyl), 120.41 and 120.34 (ArCH), 113.49 (4 C of DMT), 87.05 (C of DMT), 85.38 (d,  $J = 6.58$  Hz, C-4'), 84.66 (C-1'), 77.91 (d,  $J = 5.44$  Hz, C-3'), 69.20 (d,  $J = 6.30$  Hz, CH<sub>2</sub> of fluorenyl), 63.20 (C-5'), 55.47 (2 CH<sub>3</sub>O), 48.06 (d,  $J = 7.73$  Hz, CH of fluorenyl), 38.88 (C-2').

<sup>31</sup>P NMR (CDCl<sub>3</sub>):  $\delta = 71.07$  ( $^1J_{P,Se} = 872.5$  Hz,  $^1J_{P,H} = 648$  Hz).

### Slow Moving Diastereoisomer of 6c

$R_f$  0.38 (System B).

<sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta = 9.07$  (s, 1 H, NH), 8.70 (s, 1 H, H-2), 8.30 (d,  $J = 644$  Hz, 1 H, PH), 8.14 (s, 1 H, H-8), 8.02 (d,  $J = 7.01$  Hz, 2 H, ArH), 7.72 (dd,  $J = 7.32$ , 2.74 Hz, 2 H, ArH), 7.70–7.47 (m, 5 H, ArH), 7.41–7.16 (m, 14 H, ArH), 6.76 (d,  $J = 8.84$  Hz, 4 H, *ortho* to OMe), 6.44 (dd,  $J = 8.38$ , 5.64 Hz, 1 H, H-1'), 5.53–5.46 (m, 1 H, H-3'), 4.60–4.50 (m, 1 H, CH<sub>2</sub> of fluorenyl), 4.33–4.15 (m, 3 H, H-4', CH<sub>2</sub> and CH of fluorenyl), 3.74 (s, 6 H, 2 CH<sub>3</sub>O), 3.41–3.30 (m, 2 H, H-5'), 3.00–2.90 (m, 1 H, H-2'), 2.68–2.60 (m, 1 H, H-2').

<sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta = 164.81$  (C=O), 158.84 (2 C of DMT), 152.87 (C-2), 151.78 (C-6), 149.79 (C-4), 144.48 (2 C of DMT), 143.23 and 143.02 (2 C of fluorenyl), 141.59 (2 C of DMT), 135.62 (2 C of DMT), 133.86, 133.04, 130.23, 129.11, 128.33, 128.18, 128.09,

127.49 and 127.25 (ArCH), 125.34 and 125.13 (C-1 and C-8 of fluorenyl), 120.37 (ArCH), 113.49 (4 C of DMT), 87.09 (C of DMT), 85.42 (C-4'), 84.83 (C-1'), 78.50 (d,  $J = 6.01$  Hz, C-3'), 69.29 (d,  $J = 6.59$  Hz,  $\text{CH}_2$  of fluorenyl), 63.07 (C-5'), 55.46 (2  $\text{CH}_3\text{O}$ ), 48.00 (d,  $J = 7.73$  Hz, CH of fluorenyl), 38.96 (C-2').

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 71.05$  ( $^1J_{\text{P},\text{Se}} = 875.05$  Hz,  $^1J_{\text{P},\text{H}} = 644.9$  Hz).

#### 5'-O-Dimethoxytrityl-N<sup>2</sup>-isobutyrylguanosine Fluorenemethyl 3'-H-Phosphonoselenoate Diester (6d)

White solid (1:1 mixture of diastereomers); yield: 378 mg (80%);  $R_f$  0.09 and 0.08 (System B).

HRMS:  $m/z$  calcd for  $\text{C}_{49}\text{H}_{49}\text{N}_5\text{O}_8\text{PSe}$  [ $\text{M} + \text{H}$ ]<sup>+</sup>: 946.2484; found: 946.2475.

#### Fast Moving Diastereoisomer of 6d

$R_f$  0.09 (System B).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 11.94$  (s, 1 H, NH), 8.21 (d,  $J = 650$  Hz, 1 H, PH), 7.99 (s, 1 H, H-8), 7.74–7.57 (m, 4 H, ArH), 7.42–7.12 (m, 13 H, ArH), 6.79–6.73 (m, 4 H, *ortho* to OMe), 5.96 (t,  $J = 6.56$  Hz, 1 H, H-1'), 5.66–5.57 (m, 1 H, H-3'), 4.57–4.42 (m, 2 H,  $\text{CH}_2$  of fluorenyl), 4.14–4.10 (m, 1 H, H-4'), 3.75 (s, 6 H, 2  $\text{CH}_3\text{O}$ ), 3.35 (dd,  $J = 10.82$ , 3.20 Hz, 1 H, H-5'), 3.20 (dd,  $J = 10.67$ , 3.66 Hz, 1 H, H-5'), 2.86–2.76 (m, 1 H, H-2'), 2.18–2.10 (m, 1 H, H-2'), 2.08–1.97 [m, 1 H, CH of IBu (isobutylguanin)], 1.06 (d,  $J = 7.01$  Hz, 3 H,  $\text{CH}_3$  of IBu), 0.93 (d,  $J = 7.01$  Hz, 3 H,  $\text{CH}_3$  of IBu).

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 178.40$  (C=O), 158.90 (2 C of DMT), 155.61 (C-6), 147.86 (C-3), 147.28 (C-4), 144.81 (2 C of DMT), 143.11 and 142.95 (2 C of fluorenyl), 141.70 (2 C of DMT), 138.45 (C-8), 135.95 and 135.68 (2 C of DMT), 130.22, 129.26, 128.28, 128.19, 127.47, 127.28 and 127.17 (ArCH), 120.43 and 120.37 (ArCH), 113.44 (4 C of DMT), 86.70 (C of DMT), 84.30 (C-1' and C-4'), 69, 32 ( $\text{CH}_2$  of fluorenyl), 62.77 (C-5'), 55.47 (2  $\text{CH}_2\text{O}$ ), 48.00 (d,  $J = 8.00$  Hz, CH of fluorenyl), 38.04 (C-2'), 36.57 (CH of IBu), 19.98 and 18.89 (2  $\text{CH}_3$  of IBu).

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 71.59$  ( $^1J_{\text{P},\text{Se}} = 869.08$  Hz,  $^1J_{\text{P},\text{H}} = 649.6$  Hz).

#### Slow Moving Diastereoisomer of 6d

$R_f$  0.08 (System B).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 11.98$  (s, 1 H, NH), 8.24 (d,  $J = 645$  Hz, 1 H, PH), 8.23 (s, 1 H, H-8), 7.75–7.13 (m, 17 H, ArH), 6.78–6.71 (m, 4 H, *ortho* to OMe), 6.09 (dd,  $J = 7.62$ , 5.79 Hz, 1 H, H-1'), 5.61–5.53 (m, 1 H, H-3'), 4.53–4.43 (m, 1 H,  $\text{CH}_2$  of fluorenyl), 4.25–4.09 (m, 3 H, H-4',  $\text{CH}_2$  and CH of fluorenyl), 3.72 (s, 3 H,  $\text{OCH}_3$ ), 3.71 (s, 3 H,  $\text{OCH}_3$ ), 3.34 (dd,  $J = 10.67$ , 3.05 Hz, 1 H, H-5'), 3.18 (dd,  $J = 10.52$ , 3.81 Hz, 1 H, H-5'), 3.06–2.96 (m, 1 H, H-2'), 2.53–2.44 (m, 1 H, H-2'), 2.20–2.03 (m, 1 H, CH of IBu), 1.06 (d,  $J = 6.71$  Hz, 3 H,  $\text{CH}_3$  of IBu), 0.94 (d,  $J = 6.71$  Hz, 3 H,  $\text{CH}_3$  of IBu).

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 178.62$  (C=O), 158.91 (2 C of DMT), 155.67 (C-6), 148.07 (C-3), 147.45 (C-4), 144.75 (2 C of DMT), 143.05 and 142.96 (2 C of fluorenyl), 141.58 (2 C of DMT), 138.19 (C-8), 135.88 and 135.67 (2 C of DMT), 130.18, 129.27, 128.37, 128.32, 128.207, 127.46, 127.32, 127.28 and 125.04 (ArCH), 120.39 (ArCH), 113.48 (4 C of DMT), 86.74 (C of DMT), 84.77 (d,  $J = 4.58$  Hz, C-4'), 84.57 (C-1'), 69.25 (d,  $J = 6.30$  Hz,  $\text{CH}_2$  of fluorenyl), 62.93 (C-5'), 55.47 (2  $\text{CH}_3$ ), 47.94 (d,  $J = 7.73$  Hz, CH of fluorenyl), 38.67 (C-2'), 36.50 (CH of IBu), 18.99 (CH<sub>3</sub> of IBu), 18.90 (CH<sub>3</sub> of IBu).

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 71.43$  ( $^1J_{\text{P},\text{Se}} = 871.64$  Hz,  $^1J_{\text{P},\text{H}} = 645$  Hz).

#### 5'-O-Dimethoxytritylnucleoside 3'-H-Phosphonoselenoate, Triethylammonium Salts 7; General Procedure

Separated diastereomers of nucleoside fluorenemethyl 3'-H-phosphonoselenoate diesters **6** (425 mg, 0.5 mmol) were dissolved in  $\text{CH}_2\text{Cl}_2$  (15 mL) and treated with Et<sub>3</sub>N (695  $\mu\text{L}$ , 5 mmol, 10

equiv) for 1 h. Nucleoside 3'-H-phosphonoselenoate monoesters **7** were isolated by precipitation from pentane–Et<sub>2</sub>O (2:1) as white solids of purity >98% (<sup>1</sup>H NMR spectroscopy).

#### 5'-O-Dimethoxytritylthymidine 3'-H-Phosphonoselenoate, Triethylammonium Salt (7a)

##### 7a Fast

From faster moving diastereoisomer of **6a**; white solid; yield: 71 mg (74%);  $R_f$  0.30 (System A).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 11.56$  (br, 1 H, NH), 9.38 (s, 1 H, NH), 8.61 (d,  $J = 570.1$  Hz, 1 H, PH), 7.59 (d,  $J = 0.92$  Hz, 1 H, H-6), 7.42–7.16 (m, 10 ArH), 6.83–6.79 (m, 4 ArH *ortho* to OMe), 6.44 (dd,  $J = 8.84$ , 5.83 Hz, 1 H, H-1'), 5.48–5.41 (m, 1 H, H-3'), 4.25–4.24 (m, 1 H, H-4'), 3.76 (s, 6 H, 2  $\text{CH}_3\text{O}$ ), 3.46 (dd,  $J = 10.52$ , 2.59 Hz, 1 H, H-5'), 3.36 (dd,  $J = 10.52$ , 2.59 Hz, 1 H, H-5'), 3.16–3.06 (m, 6 H, 3  $\text{CH}_2$  of Et<sub>3</sub>N), 2.70–2.34 (m, 2 H, H-2'), 1.37–1.28 (m, 12 H, 3  $\text{CH}_3$  of Et<sub>3</sub>N and CH<sub>3</sub> at C-5).

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 164.24$  (C-4), 158.89 (2 C of DMT), 150.80 (C-2), 144.62 (2 C of DMT), 135.96, 135.75 and 135.57 (2 C of DMT and C-6), 130.37, 128.42, 128.23 and 127.26 (8 C of DMT), 113.54 (4 C of DMT), 111.46 (C-5), 87.27 (C of DMT), 85.11 (C-4'), 84.86 (C-1'), 76.23 (d,  $J = 4.87$  Hz, C-3'), 63.93 (C-5'), 55.48 (2  $\text{CH}_3\text{O}$ ), 45.95 (3  $\text{CH}_2$  of Et<sub>3</sub>N), 40.12 (C-2'), 11.86 (CH<sub>3</sub> at C-5), 8.87 (3  $\text{CH}_3$  of Et<sub>3</sub>N).

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 44.785$  (dd,  $^1J_{\text{P},\text{Se}} = 714.8$  Hz,  $^1J_{\text{P},\text{H}} = 570.3$  Hz,  $^3J_{\text{P},\text{H}} = 12.57$  Hz).

Anal. Calcd for  $\text{C}_{37}\text{H}_{48}\text{N}_3\text{O}_8\text{PSe}$ : C, 57.51; H, 6.26; N, 5.44. Found: C, 57.33; H, 6.08; N, 5.51.

##### 7a Slow

From slower moving diastereoisomer of **6a**; white solid; yield: 80 mg (98%);  $R_f$  0.30 (System A).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 11.57$  (br, 1 H, NH), 9.28 (br, 1 H, NH), 8.69 (d,  $J = 564.7$  Hz, 1 H, PH), 7.60 (br, 1 H, H-6), 7.42–7.14 (m, 10 ArH), 6.83–6.80 (m, 4 H, *ortho* to OMe), 6.47 (dd,  $J = 8.54$ , 5.49 Hz, 1 H, H-1'), 5.41–5.35 (m, 1 H, H-3'), 4.36 (br, 1 H, H-4'), 3.76 (s, 6 H, 2  $\text{CH}_3\text{O}$ ), 3.57–3.34 (m, 2 H, H-5'), 3.12–3.05 (m, 6 H, 3  $\text{CH}_2$  of Et<sub>3</sub>N), 2.60–2.31 (m, 2 H, H-2'), 1.38–1.27 (m, 12 H, 3  $\text{CH}_3$  of Et<sub>3</sub>N and CH<sub>3</sub> at C-5).

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 164.21$  (C-4), 158.88 (2 C of DMT), 150.76 (C-2), 144.67 (2 C of DMT), 135.97, 135.80 and 135.61 (2 C of DMT and C-6), 130.39, 128.55, 128.44, 128.23 and 127.26 (8 C of DMT), 113.55 (4 C of DMT), 111.42 (C-5), 87.28 (C of DMT), 85.93 (d,  $J = 5.44$  Hz, C-4'), 84.94 (C-1'), 63.76 (C-5'), 55.49 (2  $\text{CH}_3\text{O}$ ), 45.93 (3  $\text{CH}_2$  of Et<sub>3</sub>N), 39.58 (d,  $J = 3.48$  Hz, C-2'), 11.82 (CH<sub>3</sub> at C-5), 8.87 (3  $\text{CH}_3$  of Et<sub>3</sub>N).

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 45.69$  (dd,  $^1J_{\text{P},\text{Se}} = 714.8$  Hz,  $^1J_{\text{P},\text{H}} = 564.7$  Hz,  $^3J_{\text{P},\text{H}} = 12.6$  Hz).

Anal. Calcd for  $\text{C}_{37}\text{H}_{48}\text{N}_3\text{O}_8\text{PSe}$ : C, 57.51; H, 6.26; N, 5.44. Found: C, 57.18; H, 6.15; N, 5.60.

#### 5'-O-Dimethoxytrityl-N<sup>4</sup>-benzoylcytidine 3'-H-Phosphonoselenoate, Triethylammonium Salt (7b)

##### 7b Fast

From faster moving diastereoisomer of **6b**; white solid; yield: 65 mg (92%);  $R_f$  0.28 (System A).

$^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 8.60$  (d,  $J = 573$  Hz, 1 H, PH), 8.21 (d,  $J = 7.62$  Hz, 1 H, ArH), 7.88 (d,  $J = 7.62$  Hz, 2 H, ArH), 7.61–7.10 (m, 13 H, 11 ArH, H-5 and H-6), 6.87–6.83 (m, 4 H, *ortho* to OMe), 6.30 (t,  $J = 6.10$  Hz, 1 H, H-1'), 5.45–5.37 (m, 1 H, H-4'), 3.78 (s, 6 H, 2  $\text{CH}_3\text{O}$ ), 3.54–3.49 (m, 2 H, H-5'), 3.10–3.02 (m, 6 H, 3  $\text{CH}_2$  of Et<sub>3</sub>N), 2.97–2.91 (m, 1 H, H-2'), 2.41–2.29 (m, 1 H, H-2'), 1.33–1.25 (m, 9 H, 3  $\text{CH}_3$  of Et<sub>3</sub>N).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 162.27 (C=O of Bz, C-2 and C-4), 158.85 (2 C of DMT), 144.96 and 144.40 (2 C of DMT), 135.71 and 135.50 (2 C of DMT), 133.29 (ArCH), 130.41, 130.34, 129.22, 128.46, 128.25, 127.79 and 127.26 (ArCH), 113.56 (4 C of DMT), 87.54 (C-1'), 85.67 (C-4'), 74.83 (C-3'), 63.11 (C-5'), 55.46 (2 CH<sub>3</sub>O), 45.89 (3 CH<sub>2</sub> of Et<sub>3</sub>N), 41.58 (C-2'), 8.92 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 44.76 (d, <sup>1</sup>J<sub>P,Se</sub> = 713.5 Hz, <sup>1</sup>J<sub>P,H</sub> = 573 Hz, <sup>3</sup>J<sub>P,H</sub> = 11.93 Hz).

Anal. Calcd for C<sub>43</sub>H<sub>51</sub>N<sub>4</sub>O<sub>8</sub>PSe: C, 59.93; H, 5.96; N, 6.50. Found: C, 59.71; H, 5.76; N, 6.81.

### 7b Slow

From slower moving diastereoisomer of **6b**; white solid; yield: 44 mg (79%); R<sub>f</sub> 0.28 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 8.75 (d, J = 562 Hz, 1 H, PH), 8.21 (d, J = 7.32 Hz, 1 H, ArH), 7.88 (d, J = 7.01 Hz, 2 H, ArH), 7.62–7.17 (m, 13 H, 11 ArH, H-5 and H-6), 6.87–6.83 (m, 4 H, *ortho* to OMe), 6.31 (t, J = 6.10 Hz, 1 H, H-1'), 5.28–5.20 (m, 1 H, H-3'), 4.48–4.45 (m, 1 H, H-4'), 3.78 (s, 6 H, 2 CH<sub>3</sub>O), 3.56–3.42 (m, 2 H, H-5'), 3.11–3.02 (m, 6 H, 3 CH<sub>2</sub> of Et<sub>3</sub>N), 2.92–2.84 (m, 1 H, H-2'), 2.37–2.27 (m, 1 H, H-2'), 1.31–1.25 (m, 9 H, CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 162.28 (C=O of Bz, C-2 and C-4), 158.85 (2 C of DMT), 144.95 and 144.41 (2 C of DMT), 135.72 and 135.50 (2 C of DMT), 133.30 (ArCH), 130.42, 130.34, 129.22, 128.46, 128.25, 127.81 and 127.26 (ArCH), 113.57 (4 C of DMT), 87.62 (C-1'), 86.35 (d, J = 6.30 Hz, C-4'), 75.88 (C-3'), 62.92 (C-5'), 55.47 (2 CH<sub>3</sub>O), 45.88 (3 CH<sub>2</sub> of Et<sub>3</sub>N), 40.95 (C-2'), 8.95 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 45.99 (<sup>1</sup>J<sub>P,Se</sub> = 717.3 Hz, <sup>1</sup>J<sub>P,H</sub> = 562 Hz, <sup>3</sup>J<sub>P,H</sub> = 12.15 Hz).

Anal. Calcd for C<sub>43</sub>H<sub>51</sub>N<sub>4</sub>O<sub>8</sub>PSe: C, 59.93; H, 5.96; N, 6.50. Found: C, 59.63; H, 5.70; N, 6.72.

### 5'-O-Dimethoxytrityl-N<sup>6</sup>-benzoyladenosine H-Phosphonate, Triethylammonium Salt (7c)

#### 7c Fast

From faster moving diastereoisomer of **6c**; white solid; yield: 82 mg (88%); R<sub>f</sub> 0.37 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 9.04 (br, 1 H, NH), 8.67 (s, 1 H, H-2), 8.62 (d, J = 568 Hz, 1 H, PH), 8.14 (s, 1 H, H-8), 7.96 (d, J = 7.62 Hz, 2 H, ArH), 7.54 (t, J = 7.32 Hz, 1 H, ArH), 7.45 (t, J = 7.52 Hz, 2 H, ArH), 7.36 (d, J = 7.22 Hz, 2 H, ArH), 7.27–7.11 (m, 7 H, ArH), 6.73 (d, J = 8.79 Hz, 4 H, *ortho* to OMe), 6.56–6.51 (m, 1 H, H-1'), 5.41–5.36 (m, 1 H, H-3'), 4.39–4.38 (m, 1 H, H-4'), 3.71 (s, 6 H, 2 CH<sub>3</sub>O), 3.40 (dd, J = 10.25, 4.39 Hz, 1 H, H-5'), 3.34 (dd, J = 10.25, 3.81 Hz, 1 H, H-5'), 3.11–3.05 (m, 6 H, 3 CH<sub>2</sub> of Et<sub>3</sub>N), 2.95–2.83 (m, 1 H, H-2'), 1.30–1.26 (m, 9 H, 3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 164.79 (C=O), 158.69 (2 C of DMT), 152.76 (C-2), 151.76 (C-6), 149.55 (C-4), 144.73 (2 C of DMT), 141.65 (2 C of DMT), 135.88 (2 C of DMT), 132.92, 132.92, 130.30, 129.04, 128.39, 128.06 and 127.04 (ArCH), 113.37 (4 C of DMT), 86.78 (C of DMT), 85.64 (C-4'), 84.63 (C-1'), 75.99 (C-3'), 63.78 (C-5'), 55.42 (2 CH<sub>3</sub>O), 45.82 (3 CH<sub>2</sub> of Et<sub>3</sub>N), 40.00 (C-2'), 8.84 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 48.61 (d, <sup>1</sup>J<sub>P,Se</sub> = 715.9 Hz, <sup>1</sup>J<sub>P,H</sub> = 568 Hz, <sup>3</sup>J<sub>P,H</sub> = 12.48 Hz).

Anal. Calcd for C<sub>44</sub>H<sub>53</sub>N<sub>6</sub>O<sub>8</sub>PSe: C, 59.66; H, 5.80; N, 9.49. Found: C, 59.38; H, 5.48; N, 9.73.

#### 7c Slow

From slower moving diastereoisomer of **6c**; white solid; yield: 80 mg (96%); R<sub>f</sub> 0.37 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 9.04 (s, 1 H, NH), 8.66 (s, 1 H, H-2), 8.66 (d, J = 567 Hz, 1 H, PH), 8.14 (s, 1 H, H-8), 7.97–7.94 (m, 2 H, ArH), 7.53 (t, J = 7.32 Hz, 1 H, ArH), 7.45 (t, J = 7.41 Hz, 2 H, ArH), 7.36 (d, J = 7.81 Hz, 2 H, ArH), 7.27–7.10 (m, 7 H, ArH), 6.73 (d, J = 8.79 Hz, 4 H, *ortho* to OMe), 6.56–6.52 (m, 1 H, H-1'), 5.42–5.35 (m, 1 H, H-3'), 4.52–4.48 (m, 1 H, H-4'), 3.70 (s, 6 H, 2 CH<sub>3</sub>O), 3.45 (dd, J = 10.25, 4.2 Hz, 1 H, H-5'), 3.35 (dd, J = 10.25, 3.42 Hz, 1 H, H-5'), 3.09–3.02 (m, 6 H, 3 CH<sub>2</sub> of Et<sub>3</sub>N), 2.92–2.84 (m, 1 H, H-2'), 2.78–2.72 (m, 1 H, H-2'), 1.29–1.19 (m, 9 H, 3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 164.80 (C=O), 158.68 (2 C of DMT), 152.76 (C-2), 151.77 (C-6, 149.55 (C-4), 144.73 (2 C of DMT), 141.65 (2 C of DMT), 135.87 and 135.82 (2 C of DMT), 133.92, 132.92, 130.32, 129.64, 128.39, 128.05 and 127.04 (ArCH), 113.37 (4 C of DMT), 86.79 (C of DMT), 86.29 (C-4'), 84.98 (C-1'), 76.49 (C-3'), 63.71 (C-5'), 55.41 (2 CH<sub>3</sub>O), 45.79 (3 CH<sub>2</sub> of Et<sub>3</sub>N), 39.73 (C-2'), 8.81 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 48.66 (<sup>1</sup>J<sub>P,Se</sub> = 714.7 Hz, <sup>1</sup>J<sub>P,H</sub> = 567 Hz, <sup>3</sup>J<sub>P,H</sub> = 12.48 Hz).

Anal. Calcd for C<sub>44</sub>H<sub>53</sub>N<sub>6</sub>O<sub>7</sub>PSe: C, 59.66; H, 5.80; N, 9.49. Found: C, 59.47; H, 5.55; N, 9.65.

### 5'-O-Dimethoxytrityl-N<sup>2</sup>-isobutyrylguanosine H-Phosphonate, Triethylammonium Salt (7d)

#### 7d Fast

From faster moving diastereoisomer of **6d**; white solid; yield: 21 mg, (74%); R<sub>f</sub> 0.35 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 11.98 (s, 1 H, NH), 8.54 (d, J = 572 Hz, 1 H, PH), 7.77 (s, 1 H, H-8), 7.37–7.13 (m, 9 H, ArH), 6.75–6.71 (m, 4 H, *ortho* to OMe), 6.13 (dd, J = 7.01, 4.88 Hz, 1 H, H-1'), 5.87–5.75 (m, 1 H, H-3'), 4.23–4.18 (m, 1 H, H-4'), 3.74 (s, 6 H, OCH<sub>3</sub>), 3.36 (dd, J = 10.67, 3.05 Hz, 1 H, H-5'), 3.22 (dd, J = 10.67, 4.12 Hz, 1 H, H-5'), 3.14–3.06 (m, 7 H, H-2' and 3 CH<sub>2</sub> of Et<sub>3</sub>N), 2.69–2.59 (m, 1 H, H-2'), 2.55–2.45 (m, 1 H, CH of Ibu), 1.31–1.27 (m, 9 H, 3 CH<sub>3</sub> of Et<sub>3</sub>N), 1.14 (d, J = 7.01 Hz, 3 H, CH<sub>3</sub> of Ibu), 1.08 (d, J = 6.71, 3 H, CH<sub>3</sub> of Ibu).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 179.31 (C=O), 158.67 (2 C of DMT), 156.01 (C-6), 147.87 (C-3), 147.45 (C-4), 144.89 (2 C of DMT), 138.97 (C-8), 136.10 and 135.98 (2 C of DMT), 130.26, 128.39, 127.97 and 127.00 (8 C of DMT), 113.27 (4 C of DMT), 86.44 (C of DMT), 84.39 (C-4'), 74.31 (C-3'), 62.99 (C-5'), 55.44 (2 CH<sub>3</sub>O), 45.96 (3 CH<sub>2</sub> of Et<sub>3</sub>N), 38.50 (C-2'), 36.39 (CH of Ibu), 19.20 and 19.11 (CH<sub>3</sub> of Ibu), 8.89 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

<sup>31</sup>P NMR (CDCl<sub>3</sub>): δ = 45.50 (<sup>1</sup>J<sub>P,Se</sub> = 703.7 Hz, <sup>1</sup>J<sub>P,H</sub> = 572 Hz, <sup>3</sup>J<sub>P,H</sub> = 14.07 Hz).

Anal. Calcd for C<sub>41</sub>H<sub>53</sub>N<sub>6</sub>O<sub>8</sub>PSe: C, 56.74; H, 6.16; N, 9.68. Found: C, 56.39; H, 6.03; N, 9.88.

#### 7d Slow

From slower moving diastereoisomer of **6d**; white solid; yield: 41 mg (76%); R<sub>f</sub> 0.35 (System A).

<sup>1</sup>H NMR (CDCl<sub>3</sub>): δ = 8.50 (d, J = 578 Hz, 1 H, PH), 7.78 (s, 1 H, H-8), 7.40–7.13 (m, 9 H, ArH), 6.75–6.71 (m, 4 H, *ortho* to OMe), 6.18–6.13 (m, 1 H, H-1'), 5.84–5.74 (m, 1 H, H-3'), 4.36–4.31 (m, 1 H, H-4'), 3.74 (s, 6 H, 2 CH<sub>3</sub>O), 3.51–3.35 (m, 2 H, H-5'), 3.09–3.00 (m, 7 H, H-2' and 3 CH<sub>2</sub> of Et<sub>3</sub>N), 2.65–2.46 (m, 2 H, H-2' and CH of Ibu), 1.34–1.19 (m, 9 H, 3 CH<sub>3</sub> of Et<sub>3</sub>N), 0.90–0.84 (m, 6 H, 2 CH<sub>3</sub> of Ibu).

<sup>13</sup>C NMR (CDCl<sub>3</sub>): δ = 179.41 (C=O), 158.67 (2 C of DMT), 156.02 (C-6), 147.95 (C-3), 147.55 (C-4), 144.94 (2 C of DMT), 138.78 (C-8), 136.08 (2 C of DMT), 130.36, 128.48, 127.94 and 126.95 (8 C of DMT), 113.24 (4 C of DMT), 86.45 (C of DMT), 85.13 (C-4'), 84.64 (C-1'), 75.08 (C-3'), 63.61 (C-5'), 55.40 (2 CH<sub>3</sub>O), 45.95 (3 CH<sub>3</sub> of Et<sub>3</sub>N), 38.22 (C-2'), 36.34 (CH of Ibu), 19.21 and 19.07 (CH<sub>3</sub> of Ibu), 8.99 (3 CH<sub>3</sub> of Et<sub>3</sub>N).

$^{31}\text{P}$  NMR ( $\text{CDCl}_3$ ):  $\delta = 44.97$  ( $^1J_{\text{P},\text{Se}} = 707.1$  Hz,  $^1J_{\text{P},\text{H}} = 578.4$  Hz,  $^3J_{\text{P},\text{H}} = 12.57$  Hz).

Anal. Calcd for  $\text{C}_{41}\text{H}_{53}\text{N}_6\text{O}_8\text{PSe}$ : C, 56.74; H, 6.16; N, 9.68. Found: C, 56.48; H, 6.09; N, 9.77.

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