

View Article Online View Journal

ChemComm

Accepted Manuscript

This article can be cited before page numbers have been issued, to do this please use: A. M. Olaru, A. Burt, P. Rayner, S. Hart, A. C. Whitwood, G. G. Green and S. B. Duckett, *Chem. Commun.*, 2016, DOI: 10.1039/C6CC07109K.



This is an Accepted Manuscript, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about Accepted Manuscripts in the **author guidelines**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the ethical guidelines, outlined in our <u>author and reviewer resource centre</u>, still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this Accepted Manuscript or any consequences arising from the use of any information it contains.



rsc.li/chemcomm

Published on 23 November 2016. Downloaded by Athabasca University on 24/11/2016 01:16:03.



Journal Name

COMMUNICATION

Using Signal Amplification by Reversible Exchange (SABRE) to hyperpolarise ¹¹⁹Sn and ²⁹Si NMR nuclei

Received 00th January 20xx, Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/

Alexandra M. Olaru, ^aAlister Burt, ^a Peter J. Rayner, ^a Sam J. Hart, ^a Adrian C. Whitwood, ^a Gary G. R. Green^a and Simon B. Duckett^{*a}

achieve this aim.

and 8.6% amplifies this effect even further.

Organotin reagents find wide use in synthetic chemistry in

carbon-carbon sigma bond forming reactions that are often

mediated by Pd(0) catalysts.² This approach reflects a major

advantage over traditional boronic acid methods due to the

mild conditions that can be employed and the reaction's high

functional group tolerance. Organosilanes react in a similar

way with organohalides or triflates via the Hyama coupling.³ They find use as protecting groups, reducing agents and

isosteres in drug discovery.⁴ Hence as these nuclei feature in

many significant synthetic procedures their fast NMR

detection would enable their analytical use as reaction probes

and potentially the development of a role in pharmaceutical

analysis. We exploit here the fact that molecular hydrogen exists in quantum states that span two manifolds, triplet or

orthohydrogen $(o-H_2)$ and singlet or parahydrogen $(p-H_2)$, to

the spin order of $p-H_2$ could be transformed into accessible

nuclear spin polarisation through its chemical addition to a

suitable molecular acceptor.^{5, 6} The resulting non-Boltzmann

spin state distribution creates what is now referred to as a

hyperpolarised state which, when examined, gives rise to

larger than normal NMR signals.⁷ In this work we use the signal

amplification by reversible exchange (SABRE) variant of $p-H_2$

induced polarisation, which achieves catalytic magnetisation

transfer into a substrate.⁸⁻¹⁰ This effect occurs while the

substrate is bound to a complex that also contains spin

polarisation derived from $p-H_2$ in the form of a pair of hydride

ligands and the magnetisation is transferred through the J-

coupling network into the nuclei of the ligand. The enhanced spin polarisation of the bound substrate is retained after dissociation and, when interrogated by NMR spectroscopy

methods, it yields signals of substantially larger intensity than

those which would normally be observed. In the case of

pyridine, a ¹H-signal enhancement factor of over 5500-fold has

been reported at 9.4 $\ensuremath{\mathsf{T}^{10}}$, and several reports on the

performance of SABRE in biocompatible solvents such as

ethanol and water have since been published.^{11, 12} While a

It was originally theorized by Bowers and Weitekamp that

The hyperpolarisation of the ¹¹⁹Sn and ²⁹Si nuclei in 5-(tributylstannyl)pyrimidine (A_{Sn}) and 5-(trimethylsilyl)pyrimidine (B_{Si}) is achieved through their reaction with [IrCl(COD)(IMes)] (1a) or [IrCl(COD)(SIMes)] (1b) and parahydrogen via the SABRE process. 1a exhibits superior activity in both cases. The two inequivalent pyrimidine proton environments of Asn readily yielded signal enhancements totalling ~2300-fold in its ¹H NMR spectrum at a field strength of 9.4 T, with the corresponding ¹¹⁹Sn signal being 700 times stronger than normal. In contrast, B_{si} produced analogous ¹H signal gains of ~2400-fold and a ²⁹Si signal that could be detected with a signal to noise ratio of 200 in a single scan. These sensitivity improvements allow NMR detection within seconds using micromole amounts of substrate and illustrate the analytical potential of this approach for high-sensitivity screening. Furthermore, after extended reaction times, a series of novel iridium trimers of general form $[Ir(H)_2 CI(NHC)(\mu-pyrimidine-\kappa N:\kappa N')]_3$ precipitate from these solutions whose identity was confirmed crystallographically for Bsi-

Nuclear magnetic resonance (NMR) spectroscopy is an incredibly powerful technique which yields chemically diagnostic information that is highly useful for structure elucidation.¹ It achieves this result despite suffering from inherently low sensitivity as a consequence of the fact that when atoms with a nuclear spin are placed in a magnetic field, their energy levels are only weakly split. While this Zeeman splitting increases with magnetic field strength, for standard NMR spectrometers the population difference, governed by the Boltzmann distribution, is very small. In fact, for protons at 298 K in a field of 9.4 T, the acquired signal is effectively derived from only 1 in every 32000 of these nuclei, while the corresponding ²⁹Si and ¹¹⁹Sn proportions are just 1 in ~156000 and ~83000 respectively. Their natural abundance of just 4.7

^a Department of Chemistry, University of York, Heslington, York, YO10 5DD, UK. Tel: +44 1904 322564, E-mail: simon.duckett@york.ac.uk

⁺Electronic Supplementary Information (ESI) available: general procedures and experimental conditions, sample preparation, performing SABRE experiments, ligand exchange rates, field dependent polarisation transfer studies, synthetic methods. See DOI: 10.1039/x0xx00000x

⁺⁺ Data created during this research are available by request from the University of York Data Catalogue http://dx.doi.org/10.15124/12345

⁺ These authors contributed equally to the work of this manuscript.

Published on 23 November 2016. Downloaded by Athabasca University on 24/11/2016 01:16:03.

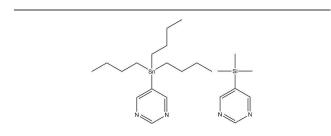
DOI: 10.1039/C6CC07109K

significant amount of attention has been dedicated to 1 H SABRE NMR, ${}^{13-17}$ this technique has also been shown to enable the hyperpolarisation of the heteronuclei ${}^{13}C^{13, 18-20}$, ${}^{15}N$ ${}^{21-27}$ or ${}^{31}p^{28, 29}$ with the ${}^{15}N$ reported signal gains being particularly noteworthy. 24

In this report we use SABRE to hyperpolarise the $^{\rm 119}{\rm Sn}$ and 29 Si nuclei of 5-(tributylstannyl)pyrimidine (A_{Sn}) and (trimethylsilyl)pyrimidine (\mathbf{B}_{s_i}) respectively (Scheme 1). In order to achieve this result they are reacted with H_2 and [IrCl(COD)(IMes)] (1a) or [IrCl(COD)(SIMes)] (1b) (where COD is 1,5-cyclooctadiene, IMes is 1.3-bis(2.4.6trimethylphenyl)imidazol-2-ylidene and SIMes is 1,3-bis(2,4,6trimethylphenyl)-4,5-dihydroimidazol-2-ylidene). This reaction $[Ir(H)_2(IMes)(A_{sn})_3]Cl$ forms the complexes (2a), $[Ir(H)_2(SIMes)(A_{sn})_3]CI$ (2b), $[Ir(H)_2(IMes)(B_{si})_3]CI$ (3a) and $[Ir(H)_2(SIMes)(B_{si})_3]CI$ (3b) according to the pathway shown in Scheme 2 and they are SABRE active.

In the case of **2a**, the original orange solution turns pale yellow over a 5 minute period at 295 K and a hydride signal at $\delta_{\rm H}$ –22.06 for [Ir(H)₂(IMes)(**A**_{sn})₃]Cl (see ESI, Table S27) appears in the corresponding ¹H NMR spectrum. When such a sample is shaken with *p*-H₂, SABRE is visible in the ¹H resonances of free **A**_{sn}. By examining the role that the iridium catalysts concentration and the ligand excess play on the level of signal enhancement (see ESI) we found that a 2.5 mM concentration and 7-fold excess led to an 803 ± 73 fold increase in size of the H-2 signal, and a 1486 ± 156 fold increase for the combined H-4, H-6 signals of **A**_{sn}, respectively, after transfer at 70 G.

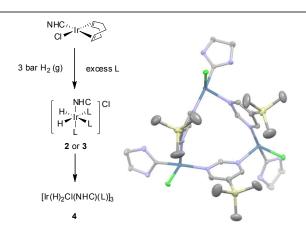
When SABRE transfer was repeated in a magnetic field of 25 G a ¹¹⁹Sn signal could be readily detected (Figure 1). In this case, the ¹¹⁹Sn response receives its polarisation indirectly via the ¹H polarisation of A_{Sn} rather than through a direct route as exemplified recently by Theis et *al.* for ¹⁵N²² and Koptyug for ³¹P.²⁹ Figure 2 shows how the resulting ¹¹⁹Sn signal enhancements and SNR values change with free ligand concentration when the concentration of **1a** is 5 mM. The 100 mM substrate loading, which corresponds to an excess of **A**_{Sn} to **2a** of 17:1, yielded the most intense ¹¹⁹Sn signal, with a SNR of 375:1 in the fully coupled spectrum, and 1099:1 in the decoupled spectrum. The signal enhancement obtained with this 100 mM loading was 772-fold (see ESI) and illustrates a time saving of in excess of 400 hours when set against the corresponding measurement under Boltzmann conditions.



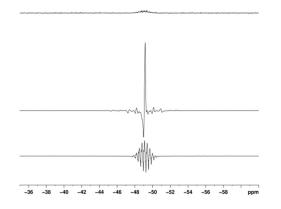
Scheme 1. 5-tributylstannylpyrimidine (A_{Sn}) and 5-trimethylsilylpyrimidine (B_{Si}) .

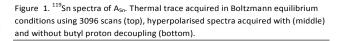
While the SIMes derived catalyst 1b system forms the analogous SABRE active complex 2b upon reaction with A_{Sn} it

yields consistently lower enhancements than those seen with 2a at 348 \pm 47 and 301 \pm 48 fold in the corresponding fully coupled and decoupled ¹¹⁹Sn NMR spectra. In order to rationalise this behaviour, the rates of H₂ elimination from 2a and 2b were determined by exchange spectroscopy (EXSY) in methanol- d_4 solution. At 295 K, H₂ loss from **2a** proceeds at a rate of 0.56 s⁻¹ whilst the analogous value for **2b** is 2.29 s⁻¹. Furthermore, for **2a**, $\Delta H^{\dagger} = 99.5 \pm 3.4$ kJ mol⁻¹ and $\Delta S^{\dagger} = 93.2 \pm$ 11.6 J K⁻¹ mol⁻¹ while for **2b**, ΔH^{\dagger} = 82.9 ± 2.9 kJ mol⁻¹ and ΔS^{\dagger} = 48.9 ± 10.1 J K⁻¹ mol⁻¹. These result in $\Delta G^{\dagger}_{(300)}$ values of 71.5 ± 0.1 kJ mol⁻¹ and 68.3 \pm 0.2 kJ mol⁻¹ respectively for these transformations and confirm that 2b undergoes more rapid ligand loss than 2a, which for this substrate is detrimental to SABRE activity. It should be noted that these exchange rates are far lower than those that are optimal for transfer to pyridine^{10,30, 31} which suggests the associated scalar coupling to a hydride ligand in 2 must be smaller in size.³²



Scheme 2. Reaction of [IrCl(COD)(NHC)] where NHC = IMes (1a) or SIMes (1b) with L $(A_{Sn} \text{ or } B_{Si})$ and H₂ yields [Ir(H)₂(IMes)(A_{Sn})₃]Cl (2a), [Ir(H)₂(IMes)(B_{Si})₃]Cl (3a), [Ir(H)₂(SIMes)(A_{Sn})₃]Cl (2b) or [Ir(H)₂(SIMes)(B)₃]Cl (3b). In each case, a trinuclear complex of general formula [[Ir(H)₂Cl(NHC)(μ -pyrimidine- κ N: κ N')]₃ eventually precipitates from these samples; the ORTEP for the product formed on reaction of 1b with B_{Si} is shown to the right.





This journal is C The Royal Society of Chemistry 20xx

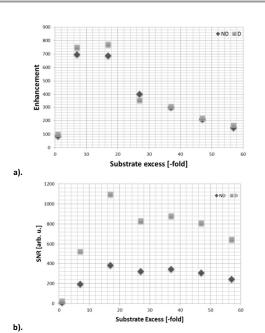


Figure 2. ¹¹⁹Sn NMR data for the hyperpolarization of 5-(tributylstannyl)pyrimidine (**2**) in methanol- d_4 through SABRE by **2a**; (a) Signal enhancement as a function of ligand excess, relative to a 5 mM concentration of catalyst, (b) SNR as a function of ligand excess for the 5mM loading. Data have been acquired with (D) and without (ND) butyl proton decoupling.

When the corresponding reaction between [IrCl(COD)(IMes)] (1a), 5-(trimethylsilyl)pyrimidine (B_{si}) and H₂ in methanol- d_4 is examined, the formation of **3a** is revealed. This complex yields a hydride signal at δ_{H} –22.44, which is almost identical in chemical shift to that of 2a. A series of SABRE experiments were then performed which showed that the resulting ¹H NMR signals of free **B**_{si} were also strongly enhanced. When 1a was used, transfer at 70 G ultimately yielded -761 ± 41 and -1524 ± 79 fold gains for the H-2 and the combined H-4, H-6 signals respectively. The corresponding values with **1b** were -450 ± 41 and -897 ± 82 . Additionally, transfer to ²⁹Si was observed and a non-decoupled SNR of 200 was obtained for a single shot measurement at a 25 mM concentration of substrate (Figure 3).

The rates of H₂ elimination from **3a** and **3b** were determined at 295 K and found to proceed at rates of 1.44 s⁻¹ and 5.11 s⁻¹ respectively. The corresponding activation parameters are $\Delta H^{\pm} = 104.7 \pm 10.3$ kJ mol⁻¹ and $\Delta S^{\pm} = 119.9 \pm 36.2$ J K⁻¹ mol⁻¹ and $\Delta H^{\pm} = 90.4 \pm 6.0$ kJ mol⁻¹ and $\Delta S^{\pm} = 82.3 \pm 21.8$ J K⁻¹ mol⁻¹ respectively. These result in $\Delta G^{\pm}_{(300)}$ values of 68.7 \pm 0.1 kJ mol⁻¹ and 65.7 \pm 0.1 kJ mol⁻¹ respectively and demonstrate again that the SIMes form undergoes more rapid ligand loss but this is detrimental to SABRE activity.

Interestingly, when these samples are left at 298 K the slow precipitation of a bright yellow powder is observed. X-ray diffraction studies on the analogous product formed with B_{Si} confirmed its identify as the iridium trimer $[\rm Ir(H)_2(CI)(\rm NHC)(\mu-pyrimidine-\kappa N:\kappa N')]_3$ (see ESI, sections 2.7 and 2.8). The formation of a related trinuclear iridium complex has been

reported, $^{\rm 33}$ alongside a series of related systems which feature bridging hydride ligands. $^{\rm 34,\,35}$

DOI: 10.1039/C6CC07109K COMMUNICATION

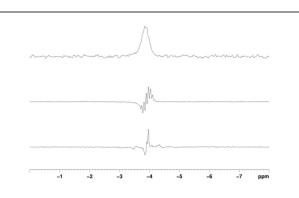


Figure 3. ²⁹Si spectra of a solution containing B_{Si} . Thermal trace (a) acquired using 128 averages, hyperpolarised trace acquired using 1 average with (middle) and without (bottom) methyl protons decoupling.

In this paper we have described how the addition of H_2 and the N-heterocycles 5-(tributylstannyl)pyrimidine (A_{Sn}) and 5-(trimethylsilyl)pyrimidine (B_{Si}) to [IrCl(COD)(IMes)] (1a) and [IrCl(COD)(SIMes)] (1b) result in the formation of a series of cationic dihydride complexes of the type [Ir(H)₂(NHC)(L)₃]Cl where L is A_{Sn} of B_{Si} . These complexes prove to be able to catalyse the transfer of polarisation from *parahydrogen* into their NMR active groups thereby making them readily detectable. For example, the ¹H NMR signal for the H-2 resonance of A_{Sn} shows an 827-fold intensity improvement when compared to that normally attained in Boltzmann equilibrium conditions. In the case of B_{Sn} the corresponding value is 761-fold.

When polarisation transfer to ¹¹⁹Sn is observed, a 700-fold intensity gain in its signal strength was measured which would require over 400 hours of measurement for comparable results. In contrast, the ²⁹Si resonance of B_{Si} provided a 200-fold SNR gain in the analogous experiment. We believe such remarkable improvements could be exploited in the future for reaction monitoring in synthetic procedures such as those outline in the introduction.

The values of $\Delta G^{\dagger}_{(300)}$ for H₂ loss in these complexes follow the order 2a>3a>2b>3b and confirm that the remote silicon and tin centres of these pyrimidine substrates play a role in determining the ligand exchange rates even though they are isolated from the metal centre by four bonds. Furthermore, SABRE is a catalytic process, in which the product and starting materials chemical identity remains the same, but their magnetic properties are changed. Hence both catalyst deactivation and selective reflect important points when analysing such processes, just like they would in a normal reaction. Here, the slow formation of a series of novel trimeric complexes with general formula [Ir(H)₂(Cl)(NHC)(µ-pyrimidine- $\kappa N:\kappa N'$]₃ that have been characterised by X-ray crystallography for \mathbf{B}_{si} results in the need to employ fresh samples during the SABRE analysis. Hence while high levels of signal gain are readily achieved through SABRE, there is a need

DOI: 10.1039/C6CC07109K

Journal Name

to refresh the catalyst periodically if activity is to be maintained over several hours.

The EPSRC (grant no. EP/G009546/1) and the Wellcome Trust (092506 and 098335) are thanked for funding. We also acknowledge Bruker Biospin for support and help from James Hayes.

References

- 1. W. P. Aue, E. Bartholdi and R. R. Ernst, *J. Chem. Phys.*, 1976, **64**, 2229-2246.
- 2. J. K. Stille, Angew. Chem., Int. Ed. Engl., 1986, **25**, 508-524.
- 3. Y. Hatanaka and T. Hiyama, *J. Org. Chem.*, 1988, **53**, 918-920.
- 4. G. A. Showell and J. S. Mills, *Drug Disc. Today*, 2003, **8**, 551-556.
- C. R. Bowers and D. P. Weitekamp, *Phys. Rev. Lett.*, 1986, 57, 2645-2648.
- C. R. Bowers and D. P. Weitekamp, J. Am. Chem. Soc., 1987, 109, 5541-5542.
- R. A. Green, R. W. Adams, S. B. Duckett, R. E. Mewis, D. C. Williamson and G. G. R. Green, *Prog. Nucl. Magn. Reson.* Spectrosc., 2012, 67, 1-48.
- R. W. Adams, J. A. Aguilar, K. D. Atkinson, M. J. Cowley, P. I. P. Elliott, S. B. Duckett, G. G. R. Green, I. G. Khazal, J. López-Serrano and D. C. Williamson, *Science*, 2009, **323**, 1708-1711.
- B. J. A. van Weerdenburg, S. Gloggler, N. Eshuis, A. H. J. Engwerda, J. M. M. Smits, R. de Gelder, S. Appelt, S. S. Wymenga, M. Tessari, M. C. Feiters, B. Blumich and F. Rutjes, *Chem. Commun. (Cambridge, U. K.)*, 2013, 49, 7388-7390.
- L. S. Lloyd, A. Asghar, M. J. Burns, A. Charlton, S. Coombes, M. J. Cowley, G. J. Dear, S. B. Duckett, G. R. Genov, G. G. R. Green, L. A. R. Highton, A. J. J. Hooper, M. Khan, I. G. Khazal, R. J. Lewis, R. E. Mewis, A. D. Roberts and A. J. Ruddlesden, *Catal. Sci. Technol.*, 2014, 4, 3544-3554.
- P. Spannring, I. Reile, M. Emondts, P. P. M. Schleker, N. K. J. Hermkens, N. G. J. van der Zwaluw, B. J. A. van Weerdenburg, P. Tinnemans, M. Tessari, B. Blümich, F. P. J. T. Rutjes and M. C. Feiters, *Chem. Eur. J.*, 2016, **22**, 9277-9282.
- M. L. Truong, F. Shi, P. He, B. Yuan, K. N. Plunkett, A. M. Coffey, R. V. Shchepin, D. A. Barskiy, K. V. Kovtunov, I. V. Koptyug, K. W. Waddell, B. M. Goodson and E. Y. Chekmenev, J. Phys. Chem. B., 2014, **118**, 13882-13889.
- R. E. Mewis, K. D. Atkinson, M. J. Cowley, S. B. Duckett, G. G. R. Green, R. A. Green, L. A. R. Highton, D. Kilgour, L. S. Lloyd, J. A. B. Lohman and D. C. Williamson, *Magn. Reson. Chem.*, 2014, **52**, 358-369.
- A. M. Olaru, S. S. Roy, L. S. Lloyd, S. Coombes, G. G. R. Green and S. B. Duckett, *Chem. Commun. (Cambridge, U. K.)*, 2016, **52**, 7842-7845.
- N. Eshuis, N. Hermkens, B. J. A. van Weerdenburg, M. C. Feiters, F. P. J. T. Rutjes, S. S. Wijmenga and M. Tessari, J. Am. Chem. Soc., 2014, 136, 2695-2698.
- 16. H. F. Zeng, J. D. Xu, J. Gillen, M. T. McMahon, D. Artemov, J. M. Tyburn, J. A. B. Lohman, R. E. Mewis, K. D. Atkinson,

G. G. R. Green, S. B. Duckett and P. C. M. van Zijl, *J. Magn. Reson.*, 2013, **237**, 73-78.

- 17. E. B. Ducker, L. T. Kuhn, K. Munnemann and C. Griesinger, *J. Magn. Reson.*, 2012, **214**, 159-165.
- J.-B. Hövener, N. Schwaderlapp, R. Borowiak, T. Lickert, S. B. Duckett, R. E. Mewis, R. W. Adams, M. J. Burns, L. A. R. Highton, G. G. R. Green, A. Olaru, J. Hennig and D. von Elverfeldt, *Anal. Chem.*, 2014, **86**, 1767-1774.
 - R. E. Mewis, R. A. Green, M. C. R. Cockett, M. J. Cowley, S.
 B. Duckett, G. G. R. Green, R. O. John, P. J. Rayner and D.
 C. Williamson, *J. Phys. Chem. B.*, 2015, **119**, 1416-1424.
- L. S. Lloyd, R. W. Adams, M. Bernstein, S. Coombes, S. B. Duckett, G. G. R. Green, R. J. Lewis, R. E. Mewis and C. J. Sleigh, J. Am. Chem. Soc., 2012, 134, 12904-12907.
- M. L. Truong, T. Theis, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, J. Phys. Chem. C., 2015, 119, 8786-8797.
- T. Theis, M. L. Truong, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, J. Am. Chem. Soc., 2015, 137, 1404-1407.
- 23. A. W. J. Logan, T. Theis, J. F. P. Colell, W. S. Warren and S. J. Malcolmson, *Chem. Eu. J.*, 2016, **22**, 10777-10781.
- D. A. Barskiy, R. V. Shchepin, A. M. Coffey, T. Theis, W. S. Warren, B. M. Goodson and E. Y. Chekmenev, J. Am. Chem. Soc., 2016, 138, 8080-8083.
- 25. R. V. Shchepin, D. A. Barskiy, D. M. Mikhaylov and E. Y. Chekmenev, *Bioconjugate Chem.*, 2016, **27**, 878-882.
- 26. R. V. Shchepin and E. Y. Chekmenev, J. Labelled Compd. Radiopharm., 2014, **57**, 621-624.
- F. Reineri, A. Viale, S. Ellena, D. Alberti, T. Boi, G. B. Giovenzana, R. Gobetto, S. S. D. Premkumar and S. Aime, J. Am. Chem. Soc., 2012, 134, 11146-11152.
- M. J. Burns, P. J. Rayner, G. G. R. Green, L. A. R. Highton, R. E. Mewis and S. B. Duckett, *J. Phys. Chem. B.*, 2015, 119, 5020-5027.
- 29. V. V. Zhivonitko, I. V. Skovpin and I. V. Koptyug, *Chem. Commun. (Cambridge, U. K.)*, 2015, **51**, 2506-2509.
- R. W. Adams, S. B. Duckett, R. A. Green, D. C. Williamson and G. G. R. Green, *The Journal of Chemical Physics*, 2009, 131, 194505.
- D. A. Barskiy, A. N. Pravdivtsev, K. L. Ivanov, K. V. Kovtunov and I. V. Koptyug, *Phys. Chem. Chem. Phys.*, 2016, 18, 89-93.
- N. Eshuis, R. Aspers, B. J. A. van Weerdenburg, M. C. Feiters, F. Rutjes, S. S. Wijmenga and M. Tessari, *J. Magn. Reson.*, 2016, 265, 59-66.
- 33. H. A. Mayer and W. C. Kaska, *Chem. Ber.*, 1995, **128**, 95-98.
- 34. V. Albano, P. Bellon and V. Scatturin, *Chem. Commun.* (*Cambridge, U. K.*), 1967, 730-731.
- S. P. Smidt, A. Pfaltz, E. Martinez-Viviente, P. S. Pregosin and A. Albinati, *Organometallics*, 2003, 22, 1000-1009.

COMMUNICATION

19.