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New metal-free route towards imidazole substituted uridine

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Abstract: Nucleosides with a bi(hetero)aryl nucleobase have unique potential applications as antiviral drugs and molecular probes. The need for transition metal catalysis to synthetize these nucleosides from pre-functionalized building blocks and the use of nucleobase protection groups results in expensive and tedious syntheses. Herein we report that 5-imidazolyl-uracil can be obtained by scalable Van Leusen imidazole synthesis and regioselectively introduced on ribose to obtain the desired nucleoside in a 5 step synthesis (total yield 55%). The 5-imidazolyl moiety leads to improved fluorescence properties. The only side-product formed was characterized by 2D-NMR and X-ray crystallography and could be suppressed during synthesis in favor of the desired product.

Modified nucleobases, nucleosides and nucleotides have provided a wide range of biologically active compounds such as antibiotics, anti-cancer and antiviral drugs.[1] Furthermore, they provide new building blocks to construct and interfere with living systems, with potential application in synthetic biology. [2] A particular class of modified nucleosides are fleximers^[3], where the nucleobase has been modified to a bi(hetero)aryl inspired by disconnecting purine nucleobases into their respective imidazole and pyrimidine ring. Nucleosides forming the nucleosidic bond with the pyrimidine ring (also dubbed "reverse" fleximers[3b]) are extremely suitable for probing helix and base pair formation.^[4] Additionally, both reverse and conventional fleximers, nucleosidic bond with the imidazole moiety, have shown promising antiviral activity, including coronaviruses.^[5] However the classical way to obtain such biheteroaryl systems occurs via palladium-catalysed reactions such as Suzuki and Stille coupling. [3b, 4-6] Although these methods access a wide scope of compounds, they pose problems when scaling up requiring expensive reagents, pre-functionalised starting materials (mostly halogens) and safety concerns (as is the case for stannous derivatives).

Our research group took particularly interest in efficiently obtaining imidazole substituted uracil and uridine as they embody the broad applications of (reverse) fleximers^[6b] and the

introduction of imidazoles has led to the functionalisation of DNA in order to obtain a more stable version of a ribozyme, a DNAzyme.[7] Retrosynthetically, we aimed at preparing the substituted base before nucleoside formation, simultaneously avoiding the use of protection groups where possible; instead of the classical, transition metal catalysed route where the heterocycle was coupled to the nucleoside. Literature study only resulting in one paper by Ressner et al[8] where the desired imidazole substituted uracil was reported. However after the Biginelli-condensation to obtain 5-acetyluracil we did not obtain the α-brominated species using several methods (Br₂, NBS, HBr/AcOH) to conduct the subsequent imidazole ring formation with formamide. Therefore another route was investigated starting from commercial uracil; which was hydroxymethylated at the 5position and oxidized to the aldehyde 1 using potassium peroxodisulfate and a catalytic amount of silver nitrate, both using previously reported methods.[9] The aldehyde was then used to investigate the Van Leusen imidazole synthesis[10] using tosylmethylisocyanide (TOSMiC) (Table 1). To the best of our knowledge, there are no examples of Van Leusen imidazole synthesis performed on a nucleobase and mostly N-substituted derivatives are obtained by using primary amines. NH-analogues are either obtained by deprotection or using ammonium hydroxide as a base. In our system, using commercially available 7M NH₃ in methanol as the base in a pressurized reaction vessel results in the desired product precipitating out of the reaction mixture, which is isolated in good yield without further need for purification (Table 1, Entry 1-5). Scaling up increases the efficiency of the precipitation to yield the desired product (Table 1, Entry 1-5). Using ammonium hydroxide solution as amine source gives a similar precipitate although less easily filtrated, and much is lost during work-up (Table 1, Entry 6-7). The amine source was exchanged for various other bases in order to obtain the oxazole analogue, however this was unsuccessful (Table 1, Entry 8-11).

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Table 1. Van Leusen reaction on 5-formyluracil.

Entry	TOSMiC	Base	Solvent	Scale	Yield	^[a] (%)
	(eq)	(eq)		(g)	2a	2b
1	3	NH ₃ (6)	MeOH	0.125	40	-
2	1.6	NH ₃ (3)	MeOH	0.125	48	-
3	1.6	NH ₃ (3)	MeOH	0.25	59	-
4	1.6	NH ₃ (3)	MeOH	0.5	66	-
5	1.6	NH ₃ (3)	MeOH	1	85	-
6	1.6	NH ₄ OH (3)	MeOH	0.25	23	-
7	1.6	NH ₄ OH (3)	H ₂ O	0.25	Deg ^[b]	-
8	1.6	K ₂ CO ₃ (3)	MeOH	0.25	-	m.s. ^[c]
9	1.6	tBuOK (3)	MeOH	0.25	-	m.s. ^[c]
10	1.6	Et ₃ N (3)	MeOH	0.25	-	m.s. ^[c]
11	1.6	DBU (3)	MeOH	0.25	-	n.r. ^[d]

[a] Isolated yield, [b] degradation of starting material, [c] multiple spots, multiple products (monitored by TLC, MS), [d] no reaction

Thin layer chromatography (TLC) showed a myriad of products forming, an intermediate before elimination of the tosyl group mediated by base. Harsher conditions only resulted in the degradation of the uracil ring (Table 1, Entry 7-11).

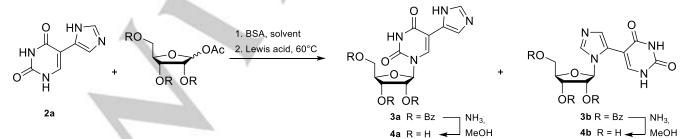
Nucleoside formation with the imidazole substituted uracil was done via the Vorbrüggen method^[11], by transiently protecting the nucleobase **2a** with trimethylsilyl-groups using bis(trimethylsilyl)acetamide (BSA). This was immediately reacted (one-pot) with 1-O-acetyl-2,3,5-tri-O-benzoyl-D-ribofuranose, promoted by the addition of a Lewis acid (Scheme 1). At room temperature, no reaction products were found on TLC nor isolated (Table 2, Entry 1), gentle heating at 60 °C (Table 2, Entry 2) showed the formation of two products, a major and a minor one.

Table 2. Optimization of Vorbrüggen conditions.

Entry	Lewis	Solvent	Т	t	Y	ield ^[a] (%))
	Acid		(°C)	(d)	3a	3b	Ratio
			- 4		9		(a:b)
1	TMSOTf ^[b]	ACN	rt	1	n.o.	[d,e]	ND ^[f]
2	TMSOTf[b]	ACN	60	1./	34	11	3:1
3	TMSOTf[b]	ACN	60	2	58	21	3:1
4	TMSOTf[b]	DCE	60	2	67	5	13:1
5	TMSOTf[c]	DCE	60	2	73	9	12:1
6	SnCl ₄ [b]	DCE	60	2	trace	s ^[e]	$ND^{[f]}$
7	TMSOTf[b]	THF	60	1	Insepa	rable mix	ture ^[g]
8	TMSOTf[b]	DMF	60	1	n.o.	[d,e]	$ND^{[f]}$

[a] Isolated yield, [b] 1 eq. [c] 1.5 eq [d] not observed, [e] monitored by TLC, [f] not determined, [g] more than 6 spots (TLC)

Longer reaction time led to increased product formation, with both products increasing similarly (Table 2, Entry 3). Changing the solvent to DCE shifted the obtained product ratio towards 3a (Table 2, Entry 4). However, it was now necessary to reflux the first silylation step in order to obtain a transparent solution, which was not necessary using ACN as a solvent. Slightly improved yields were obtained for both products when increasing the amount of TMSOTf (Table 2, Entry 5). Another Lewis acid used in nucleosidic bond formation (SnCl₄), did not show any significant product formation. THF and DMF were not suitable as solvents for this reaction (Table 2, Entry 7-8). The final products 4a and 4b were obtained by overnight deprotection of the benzoyl groups using 7M NH₃ in MeOH (Scheme 1). They were analyzed by 2D-NMR to ascertain which regioisomers were formed in the earlier mentioned Vorbrüggen reaction (Supporting Information, SI). As the designation of the regiochemistry of 4b relied on the possibly ambiguous absence of some 2D-NMR (HMBC) cross peaks, the structure was confirmed by x-ray crystallography (CCDC 1996102, Figure 1, SI). Compound 4b was crystallized as HCl salt with the imidazole ring being protonated at the N2 atom (SI, Figure 1). The puckering mode of the ribose ring is C3'-endo (pseudorotation phase angle $P = 9.8(\pm 0.4)^{\circ}$, puckering amplitude $\tau m = 38.1(\pm 0.3)^{\circ}$). The torsion angle O4-C1-N1-C8 (-139.5(\pm 0.4)°), comparable to the χ angle used to describe the orientation about the glycosyl bond, indicates an anti conformation with the bulk of the substituent pointing away from the sugar.



Scheme 1. Nucleoside synthesis via Vorbrüggen reaction with 5-imidazolyl uracil. Detailed reaction conditions for the formation of 3a/3b can be found in Table 2. BSA = Bis(trimethylsilyl)acetamide.

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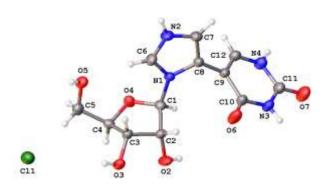


Figure 1. Single crystal X-ray structure of **4b**·HCl with thermal ellipsoids at 50% probability level.

The dihedral angle between the best planes through the imidazole and uracil ring is 67.4(±0.2)°. The crystal packing is consolidated through a series of O-H...O/Cl and N-H...O/Cl interactions between all possible hydrogen bond donors and acceptor, except sugar atom O4 (Figure S1, Table S2).

Similar DNA fleximers bearing furan and oxazole moieties have practical applications as fluorescent probe to measure the polarity at DNA abasic sites. [4a] Therefore the steady-state spectroscopy of both products (4a and 4b) was investigated in ACN, DCM, MeOH and phosphate buffered saline (PBS). The results are presented in Figures 2, S2, S3 and Tables 3, S3, S4. The S0→ S1 band of 4a is very similar in all four solvents, no vibrational structure is observed and the position of the maximum varies only slightly from 305 to 308 nm. The latter suggests an apolar ground state. At higher energy, the onset of a very intense transition is observed. Having the same regiochemistry as the previously mentioned fleximers, 4a exhibits similar albeit improved fluorescent properties. A broad and structureless band is observed in all solvents, shifting to longer wavelengths with increasing polarity with similar results for ACN and DCM, in agreement with literature. [4a] This suggests a slightly polar excited state. This is no surprise considering the chromophore resembles a merocyanine moiety. The quantum yield in water (PBS) is increased twofold with respect to a 5-furane substituted deoxyuridine, from 0.03 to 0.06.[4a] In order to further evaluate its suitability as a sensor, the excited state dynamics of 4a in PBS were evaluated. In Figure S3 the fluorescence decay trace and the corresponding fit are displayed.

Table 3. Summary of absorption and emission properties of ${\bf 4a}$ in different solvents.

		- 4			
Solvent	$\lambda_{\text{abs}}^{[a]}$	λ _{em} ^[b]	$\bar{\nu}_{abs} - \bar{\nu}_{em}^{[c]}$	FWHM _{em} ^[d]	QY ^[e]
	(nm)	(nm)	(cm ⁻¹)	(cm ⁻¹)	(%)
ACN	306	399	7610 ± 50	5200 ± 50	2.3 ± 0.2
DCM	305	400	7790 ± 50	5400 ± 50	3.7 ± 0.4
MeOH	307	417	8590 ± 50	5460 ± 50	5.7 ± 0.6
PBS	308	442	9840 ± 50	5610 ± 50	6.4 ± 0.6

[a] Absorption maxima, [b] emission maxima, [c] Stokes' shift, [d] full width at half maximum of the emission band, [e] fluorescence quantum yield. For all measurements, the concentration was 1×10^{-5} M

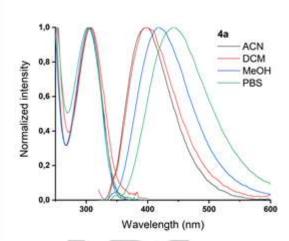


Figure 2. Normalized absorption and emission spectra of 4a $(1\times10^{-5} \text{ M})$ in acetonitrile (ACN), dichloromethane (DCM), methanol (MeOH) and phosphate buffered saline (PBS). The shoulder observed in the emission spectrum at higher energy is due to Raman scattering. The excitation wavelength for the emission spectra was 310 nm

The data could be fitted satisfactorily ($\chi^2=1.015$) with a monoexponential model, resulting in a lifetime (τ) of 1.88 ns. This uncomplicated behaviour is beneficial for a probe and unusual for a nucleoside. For **4b** on the other hand, the S0 \rightarrow S1 transition is observed at lower wavelengths, with maxima between 270 and 276 nm. No fluorescence could be observed.

Although recent developments have introduced the use of enzymatic techniques[6b, 6c, 13] in accessing the broad scope of (reverse) fleximers, they still rely on the use of transition metalcatalyzed reactions. Here we show that two isomers, 4a and 4b can be obtained without needing to resort to additional protecting groups and heavy metal catalysis. All this can be achieved in a simple, scalable reaction scheme of 5 steps which obtains 4a in a total yield of 55% starting from commercially available uracil and involving only a single use of chromatographic separation. The solvent dependency of the regioselective reaction is interesting. Attempts to increase the yield of 3b/4b by means of a different Lewis acid or solvent were unfortunately unsuccessful. Further investigation was beyond the scope of this endeavor but would be interesting as 4b, belongs to the family of distal fleximers whose synthesis still remains non trivial. [3a, 6b, 13] The introduction of an imidazole on the 5-position of uracil (4a), improves the spectroscopic characteristics when compared to furan, thiophene, oxazole, thiazole and benzimidazole. [4a, 14] Furthermore, it allows the possibility of wobble base pair formation[4b] and catalytic activity[7]. On the other hand, isomer 4b is a novel fleximer reminiscent of a disconnected xanthine, of which we provide the first detailed structural determination by 2D-NMR and XRD. Testing these newly accessed privileged structures against the current COVID19 pandemic is currently underway.

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- [1] a) L. P. Jordheim, D. Durantel, F. Zoulim, C. Dumontet, *Nat. Rev. Drug. Discov.* 2013, *12*, 447; b) M. Serpi, V. Ferrari, F. Pertusati, *J. Med. Chem.* 2016, *59*, 10343-10382.
- [2] S. A. Benner, A. M. Sismour, Nat. Rev. Genet. 2005, 6, 533-543.
- [3] a) K. Seley-Radtke, Antiviral Chem. Chemother. 2018, 26, 1-12; b) S. C. Zimmermann, K. L. Seley-Radtke, in Chemical Biology of Nucleic Acids: Fundamentals and Clinical Applications (Eds.: V. A. Erdmann, W. T. Markiewicz, J. Barciszewski), Springer Berlin Heidelberg, Berlin, Heidelberg, 2014, pp. 149-165.
- [4] a) N. J. Greco, Y. Tor, J. Am. Chem. Soc. 2005, 127, 10784-10785; b) E.
 Rozners, R. Smicius, C. Uchiyama, Chem. Commun. 2005, 5778-5780.
- [5] a) H. L. Peters, D. Jochmans, A. H. de Wilde, C. C. Posthuma, E. J. Snijder, J. Neyts, K. L. Seley-Radtke, *Bioorg. Med. Chem. Lett.* 2015, *25*, 2923-2926; b) P. Wigerinck, C. Pannecouque, R. Snoeck, P. Claes, E. De Clercq, P. Herdewijn, *J. Med. Chem.* 1991, *34*, 2383-2389; c) M. K. Yates, M. R. Raje, P. Chatterjee, C. F. Spiropoulou, S. Bavari, M. Flint, V. Soloveva, K. L. Seley-Radtke, *Bioorg. Med. Chem. Lett.* 2017, *27*, 2800-2802.
- [6] a) M. C. Jetter, A. B. Reitz, Synthesis 1998, 829-831; b) S. Vichier-Guerre, L. Dugué, F. Bonhomme, S. Pochet, Org. Biomol. Chem. 2017, 15, 8193-8203; c) S. Vichier-Guerre, L. Dugué, F. Bonhomme, S. Pochet, Org. Biomol. Chem. 2016, 14, 3638-3653.
- [7] S. K. Silverman, Acc. Chem. Res. 2015, 48, 1369-1379.
- [8] E. C. Ressner, P. Fraher, M. S. Edelman, M. P. Mertes, J. Med. Chem. 1976, 19, 194-196.
- [9] R. H. E. Hudson, F. Wojciechowski, Can. J. Chem. 2005, 83, 1731-1740.
- [10] D. V. Leusen, A. M. V. Leusen, in *Organic Reactions*, Vol. 57, John Wiley & Sons. Inc.. 2004. pp. 417-666.
- [11] a) H. Vorbrüggen, K. Krolikiewicz, B. Bennua, Chemische Berichte-Recueil 1981, 114, 1234-1255; b) H. Vorbrüggen, C. Ruh-Pohlenz, in Organic Reactions, John Wiley & Sons, Inc., 2004.
- [12] D. Onidas, D. Markovitsi, S. Marguet, A. Sharonov, T. Gustavsson, J. Phys. Chem. B 2002, 106, 11367-11374.
- [13] S. Vichier-Guerre, T. C. Ku, S. Pochet, K. L. Seley-Radtke, ChemBioChem 2020, 21, 1412-1417.
- [14] P. Guo, X. Xu, X. Qiu, Y. Zhou, S. Yan, C. Wang, C. Lu, W. Ma, X. Weng, X. Zhang, X. Zhou, Org. Biomol. Chem. 2013, 11, 1610-1613.



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Metal-free nucleosides: Bi(hetero)arylic nucleosides have unique potential as antiviral drugs and molecular probes. However their synthesis is tedious; requiring metal catalysis, pre-functionalized building blocks and nucleobase protection groups. Herein we report the scalable, metal-free synthesis of 5-imidazolyl-uracil and its regioselective nucleoside formation. Improved fluorescence and characterization of a new fleximer side-product are reported.

