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N-{4-[4-(2,3-Dichlorophenyl)piperazin-1-yl]butyl, Butenyl and Butynyl}arylcaboxamides as Novel Dopamine D₃ Receptor Antagonists

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Abstract—The dopamine D₃ receptor subtype has been targeted as a potential neurochemical modulator of the behavioral actions of psychomotor stimulants, such as cocaine. Previous synthetic studies provided structural requirements for high affinity binding to D₃ receptors which included a 2,3-dichloro-phenylpiperazine linked to an arylamido function via a butyl chain. To reduce lipophilicity of these agents and further investigate optimal conformation, a second series of 15 novel ligands was designed that included heteroaromatic substitution and unsaturated alkyl linkers. These compounds were synthesized and evaluated for binding at rat D₃ and D₂ receptors stably expressed in Sf9 cells. D₃ binding affinities ranged from $K_i = 0.6$ –1080 nM, with a broad range of D₃/D₂ selectivities (2–97). The discovery of potent, selective and bioavailable D₃ receptor ligands will provide essential molecular probes to elucidate the role D₃ receptors play in the psychomotor stimulant and reinforcing effects of cocaine.

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The dopamine D₃ receptor, a member of the D₂ receptor family, resides in brain regions associated with emotional and cognitive function, such as the nucleus accumbens (for review see refs 1 and 2). Efforts to further elucidate the function and potential therapeutic advantages of targeting D₃ receptors have resulted in compounds that may be useful in treating Parkinson's disease, schizophrenia and drug abuse.^{2–4} The discovery of D₃ receptor selective antagonists and partial agonists has received particular attention for potential treatment of cocaine abuse since the compound BP 897 was first reported to block cue-controlled cocaine-seeking in rats.⁵ Additional studies with BP 897⁶ and the potent and D₃ selective antagonist SB-277011^{7,8} further support the development of D₃ selective ligands as potential cocaine abuse medications.

As we have described⁹ and others have more recently confirmed^{10–12} optimal D₃ receptor binding affinities are obtained when a 2,3-dichlorophenylpiperazine is linked

to an aryl amide via a butyl chain (Fig. 1). However, the 2- or 4-substituted fluorenyl analogues, which gave the highest D₃ binding affinities and selectivities rendered these molecules highly lipophilic and possibly limited their bioavailability.⁹ In vivo investigation of these ligands will be required to elucidate mechanisms associated with reduction in cocaine-seeking behavior. Thus, improving physical properties of the molecules by reducing lipophilicity and further identifying structural modifications that would yield highly potent and selective D₃ ligands, was prioritized. In this pursuit, heteroaromatic replacement of the fluorenyl ring system and conformational optimization by adding unsaturation to the butyl linker was explored. The target molecules are shown in Figures 2 and 3.

Synthesis of novel amides **21–35** was achieved as depicted in Scheme 1. 2,3-Dichlorophenylpiperazine was linked to the *N*-phthalimido-protected butyl (**11**), butenyl (*cis*; **16a** or *trans*; **16b**) or butynyl (**19**) amines using standard *N*-alkylation conditions followed by deprotection with hydrazine.⁹ Amidation via the acid chloride using Schotten–Baumann conditions (Method A) or directly using CDI (Method B) gave the desired

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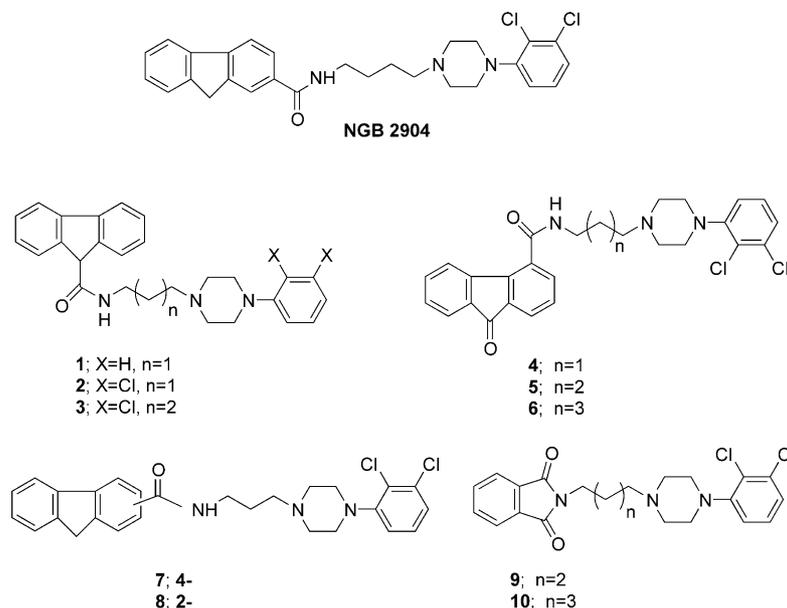


Figure 1. Previously synthesized D₃ ligands.

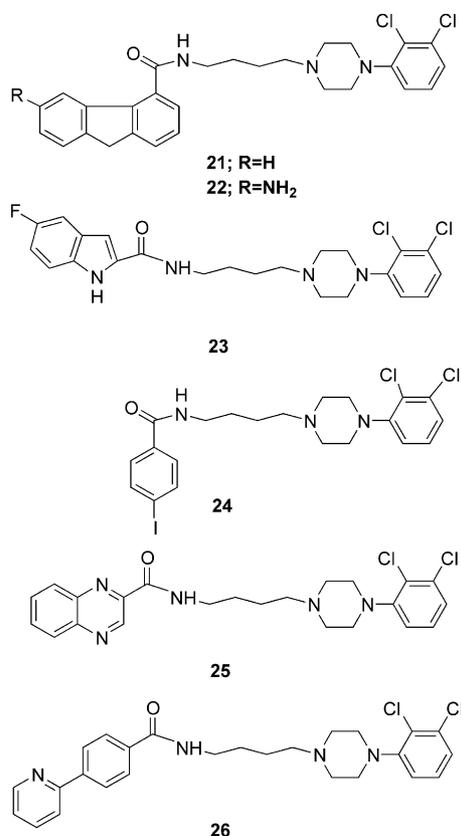


Figure 2. Novel D₃ ligands with saturated butyl chain link.

products (**21** and **23–35**). For compound **22**, the 5-nitrofluorenyl carboxylic acid was first reduced by catalytic hydrogenation (10% Pd/C) to the intermediate aniline that was then coupled to amine **13** using DCC and HOBt (Method C). All carboxylic acids were either commercially available or prepared by literature methods as indicated in Table 1 where physical properties of all final products are provided.

D₃ and D₂ receptor binding affinities for the 15 novel compounds were determined by competitive radioligand binding experiments using (a) the D₂-like receptor selective radioligand ¹²⁵I-IABN¹³ and (b) either rat D₂ or D₃ receptors expressed in Sf9 cells using a recombinant baculovirus. The binding methods used are as previously reported¹⁴ and briefly described in the Table 1 legend. Ten previously reported compounds **1–10** and NGB 2904, shown in Figure 1, were also evaluated for comparison purposes. cLogD values¹⁵ are also shown in Tables 2 and 3.

The binding data in Table 2 revealed that when the aromatic ring system contains heteroatoms, high affinity binding at D₃ is retained, while lipophilicity is decreased, for example compounds **5** and **21** compared to **23**, **25**, and **26**. Compound **26** demonstrated the highest D₃ binding affinity ($K_i=0.6$ nM) in this series and was 97-fold selective for D₃ receptors over D₂. Recently, similar findings have been reported with other heteroaromatic substitutions.^{10,11} However, smaller aryl ring systems on the saturated butyl linker also demonstrate moderately high affinity binding at D₂ receptors, limiting their utility as selective D₃ compounds (e.g., **9** and **10** as compared to **5**). Substitution of a phenyl ring with the sterically bulky 4'-iodo atom in compound **24**, somewhat improved D₃-selectivity, nevertheless, this compound was still quite lipophilic (cLogD=6.20). Alternatively, the investigation into structurally more rigid analogues obtained by unsaturation of the butyl linker led to a promising lead.

Whereas the alkyne linker (**33–35**) served to significantly reduce binding affinities at both D₃ and D₂, introduction of a *cis*, and particularly a *trans*, olefin resulted in retention of high affinity binding at D₃ receptors (Table 3). This was particularly true when the lipophilic fluorenyl ring system was replaced with a simple phenyl ring, where compound **31** showed high affinity ($K_i=1.3$ nM), as well as 39-fold selectivity for D₃ over D₂ receptors.

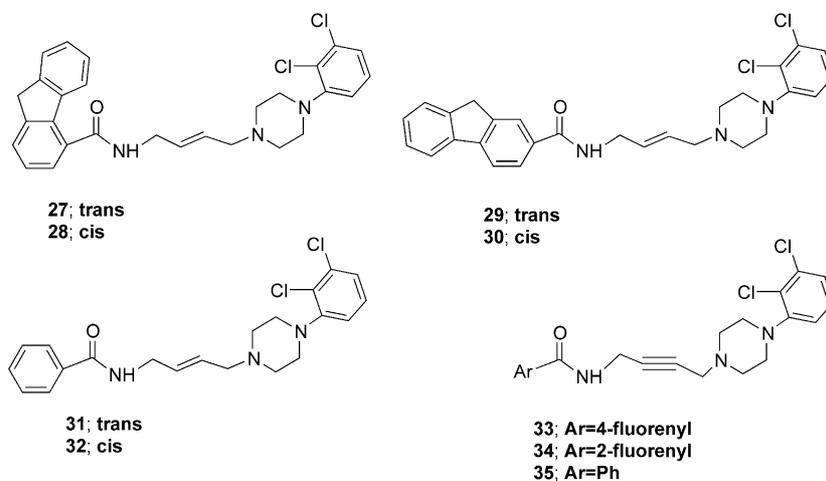
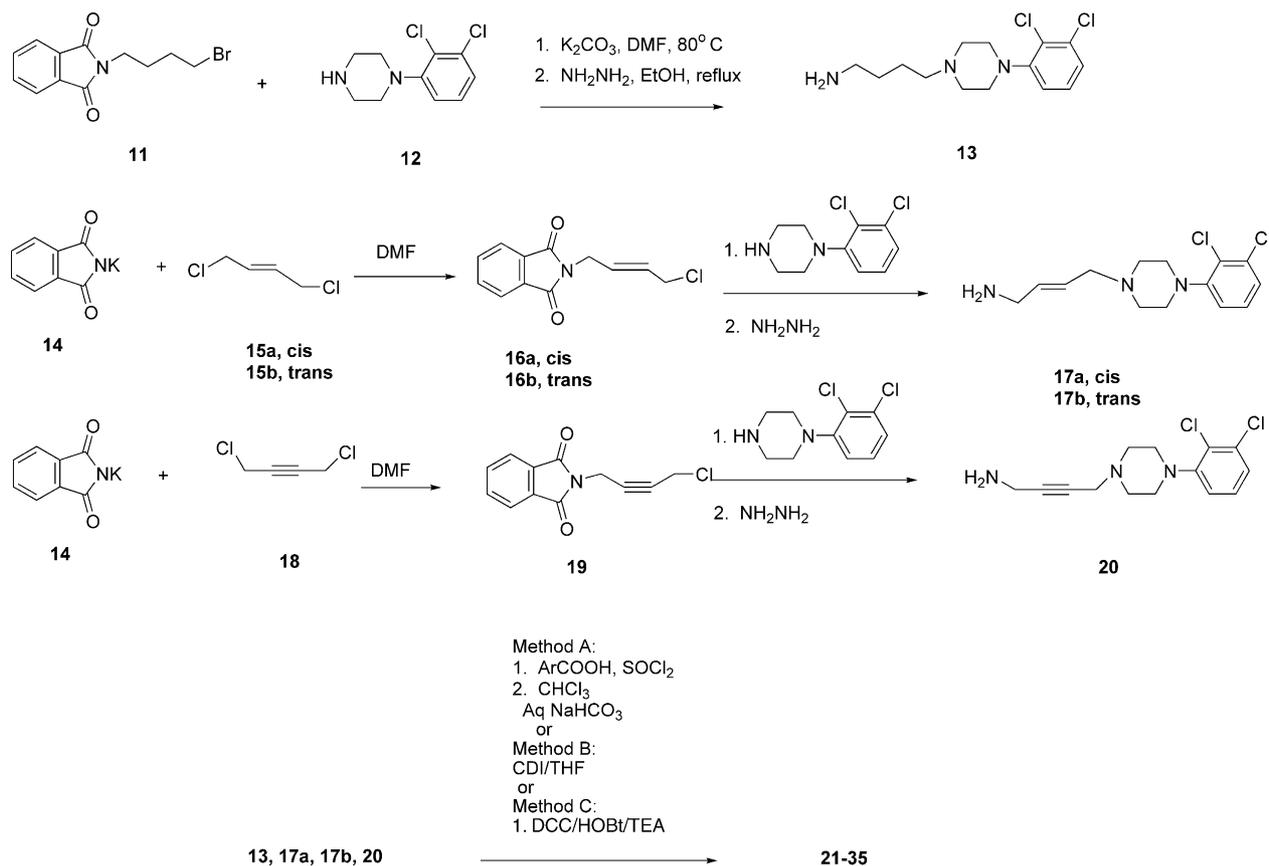


Figure 3. Structurally rigid D₃ receptor ligands with unsaturated chain link.



Scheme 1. Synthesis of novel D₃ ligands.

This modification reduced lipophilicity in this series of analogues by 2 orders of magnitude (e.g., compounds **27–30** compared to **31** and **32**). In fact, the smaller ring system improved binding affinity at D₃ significantly. Furthermore, for the phenyl-ring substituted compounds, **31** compared to **32**, the *trans* olefin is the preferred conformation for optimal D₃ binding affinity and selectivity over D₂. This conformational preference is in accord with *trans*-cyclohexamides previously reported as high affinity D₃ ligands.^{16,17} Investigation into optimal substitution of the phenyl ring as well as heteroaryl

substitutions in the *trans* butenyl series, is currently underway.

Evaluation of compounds **23–26**, **29**, **31**, and **32** in a functional assay using stimulation (agonist) or inhibition of quinpirole stimulation (antagonist) of mitogenesis in human D₃ transfected CHO cells is shown in Table 4. Data to support an antagonist profile for compounds **23**, **25**, **31** and **32**, which was predicted based on NGB 2904¹⁸ and other more recently reported D₃ antagonists having the 2,3-dichlorophenyl moiety¹⁰ is

Table 1. Synthetic methods and physical properties of novel D₃ ligands

Compd	Method ^c	Yield	Salt/rec solv.	Mp °C	Formula ^c
21	B	10 ^b	Fumarate/MeOH	187~190	C ₂₈ H ₂₉ N ₃ OCl ₂ ·C ₄ H ₄ O ₄ ·0.5H ₂ O
22	C	81	Oxalate/2-PrOH	140~142	C ₂₈ H ₂₈ Cl ₂ N ₄ O·2C ₂ H ₂ O ₄ ·2H ₂ O
23	B	46 ^b	HCl/MeOH	d 271~274	C ₂₃ H ₂₅ N ₄ OFCI ₂ ·HCl
24	B	25 ^b	HCl/2-PrOH	227~229	C ₂₁ H ₂₄ N ₃ OCl ₂ I·HCl
25	B	20 ^b	HCl/MeOH/ether	209~211	C ₂₃ H ₂₅ N ₃ OCl ₂ ·HCl
26	A ^a	40 ^b	diHCl/2-PrOH	234~237	C ₂₆ H ₂₈ N ₄ OCl ₂ ·2HCl
27	B	87	Oxalate/MeOH/ether	187~190	C ₂₈ H ₂₇ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·1.5H ₂ O
28	B	89	Oxalate/MeOH	170~172	C ₂₈ H ₂₇ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·0.5H ₂ O
29	B ^d	99	Oxalate/MeOH/ether	174~176	C ₂₈ H ₂₇ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·0.25H ₂ O
30	B ^d	94	Oxalate/MeOH	176~178	C ₂₈ H ₂₇ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·0.5H ₂ O
31	A	99	Oxalate/Acetone	118~122	C ₂₁ H ₂₃ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·0.5H ₂ O
32	A	99	HCl/2-PrOH	100~102	C ₂₁ H ₂₃ Cl ₂ N ₃ O·HCl
33	B	69	Oxalate/2-PrOH	180~181	C ₂₈ H ₂₅ Cl ₂ N ₃ O·C ₂ H ₂ O ₄ ·0.25H ₂ O
34	B ^d	96	Oxalate/MeOH	194~195	C ₂₈ H ₂₅ Cl ₂ N ₃ O·C ₂ H ₂ O ₄
35	A	98	Oxalate/2-PrOH	126~130	C ₂₁ H ₂₁ Cl ₂ N ₃ O·C ₂ H ₂ O ₄

^a4-Pyridin-2-yl benzoic acid.²¹

^b%Yield is based on the purified salt, all others based on crude free base.

^cAll compounds were purified through their respective salts and the free bases were characterized using ¹H and ¹³C NMR, FT-IR and GC-MS. Combustion analysis results agreed to ±0.4% of C, H and N with theoretical values.

^d2-Fluorenylcarboxylic acid.⁹

^eMethod A⁹; B¹⁸; C.²²

Table 2. D₃ and D₂ receptor binding data and clog D values for non-rigid molecules

Compd	D ₃ (nM)±SEM ^a	D ₂ (nM)±SEM ^a	D ₂ /D ₃	clog D ^b
1	84.5±19	1020±110	12	4.16
2	34.0±11	250±82	7	6.08
3	11.1±3.8	55.2±22	5	6.36
4	43.0±23	153±57	4	6.36
5	1.6±0.9	150±20	94	6.64
6	2.0±1.1	35.3±9.6	18	6.85
7	139±5.0	376±170	3	6.66
8	199±52	396±170	2	6.66
9	4.8±2.2	102±33	21	5.70
10	5.6±3.9	86.4±20	15	5.91
21	4.5±2.2	134±45	30	6.95
22	3.5±2.7	79.3±18	23	5.66
23	2.6±1.4	83.2±20	32	5.45
24	1.4±0.5	87.5±33	63	6.20
25	1.9±0.7	109.4±5.3	58	4.92
26	0.6±0.2	57.9±5.1	97	5.30
NGB 2904	1.1±0.2	911±190	830	6.94

^aThe methods for the binding assays have been previously described.¹⁴ K_i values are the mean of at least three independent determinations. The IC₅₀ values obtained from competition experiments were converted to K_i values using the Cheng and Prusoff correction.²³ The radioligand used for the competitive radioligand binding studies was ¹²⁵I-IABN. A recombinant baculovirus (Bv) expression system was used to express either rat D₂ (BvD₂) or rat D₃ (BvD₃) receptors in Sf9 cells. Competition curves were modeled as a one site fit using the TABLECURVE program. Human D₃ and D₂ receptor binding in CHO cells has previously been reported for compounds **1–10** and NGB 2904.⁹

^bSee ref 15.

shown. The *trans* olefins are particularly potent antagonists in this assay with IC₅₀ values in the low nanomolar range. Nevertheless, compounds **24**, **26**, and **29** show a partial agonist profile, which is consistent with the 2-methoxy phenylpiperazine compound BP 897 that was initially tested in a similar in vitro model of D₃ receptor function.⁵ However, subsequently BP 897 was shown to have an antagonist profile in other in vitro models of D₃ function.^{19,20} Although the initial report on BP 897 suggested that D₃ partial agonists would be predicted to

Table 3. D₃ and D₂ receptor binding data and ClogD values for rigid analogues

Compd	D ₃ (nM)±SEM ^a	D ₂ (nM)±SEM ^a	D ₂ /D ₃	clog D
27	19±7.8	106±22	6	7.07
28	6.5±1.8	10.7±5	1.6	7.07
29	5.9±1.8	198±39	34	7.07
30	6.0±1.6	87.3±19	15	7.07
31	1.3±0.4	50.1±6	39	5.13
32	4.9±1.9	20.0±1.8	4	5.13
33	1080±380	891±250	1	7.04
34	214±63	1000±180	5	7.04
35	394±140	1700±450	4	5.10

^aK_i values were obtained as described in Table 1.

Table 4. D₃ functional assay using stimulation or inhibition of quinpirole stimulation of mitogenesis in CHO cells (hD₃)

Compd	Agonist EC ₅₀ (nM)±SEM ^a	%Max Stim.±SEM ^a	Antagonist IC ₅₀ (nM)±SEM ^a
23	> 10,000	—	52.4±1.2
24	31.7±10	30.0±0.1	—
25	> 10,000	—	26.9±7.7
26	6.31±1.7	29.7±0.2	—
29	173±9.6	44.1±0.9	—
31	> 10,000	—	7.72±1.6
32	> 10,000	—	6.00±0.59

^aThese data were obtained through the service of CTDTP, Division of Treatment Research and Development, NIDA, using a contract (N01DA-1-8816) protocol.

block cocaine-seeking, recent reports have shown that the potent and selective D₃ antagonist SB-277011 showed remarkable inhibition of cocaine seeking and cocaine-enhanced brain reward in rat.⁸ These studies point to potential inconsistencies between in vitro and in vivo models of D₃ receptor function and support further investigation of novel D₃ compounds in vivo. Hence, the development of potent and selective D₃ receptor ligands, such as compounds **26** and **31**, will provide the pharmacological tools to clarify these

mechanistic questions and relate chemical structure, D₃ receptor function and behavior in models of cocaine abuse.

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References and Notes

1. Levant, B. *Pharm. Rev.* **1997**, *49*, 231.
2. Joyce, J. N. *Pharmacol. Ther.* **2001**, *90*, 231.
3. Le Foll, B.; Schwartz, J. C.; Sokoloff, P. *Eur. J. Psychiatry* **2000**, *15*, 140.
4. Richtand, N. M.; Goldsmith, R. J.; Nolan, J. E.; Berger, S. P. *J. Addictive Dis.* **2001**, *20*, 19.
5. Pilla, M.; Perachon, S.; Sautel, F.; Garrido, F.; Mann, A.; Wermuth, C. G.; Schwartz, J. C.; Everitt, B. J.; Sokoloff, P. *Nature* **1999**, *400*, 371.
6. Beardsley, P. M.; Sokoloff, P.; Balster, R. L.; Schwarz, J.-C. *Behav. Pharmacol.* **2001**, *12*, 1.
7. Stemp, G.; Ashmeade, T.; Branch, C. L.; Hadley, M. S.; Hunter, A. J.; Johnson, C. N.; Nash, D. J.; Thewlis, K. M.; Vong, A. K. K.; Austin, N. E.; Jeffrey, P.; Avenell, K. Y.; Boyfield, I.; Hagan, J. J.; Middlemiss, D. N.; Reavill, C.; Riley, G. J.; Routledge, C.; Wood, M. *J. Med. Chem.* **2000**, *43*, 1878.
8. Vorel, S. R.; Ashby, C. R.; Paul, M.; Liu, X. H.; Hayes, R.; Hagan, J. J.; Middlemiss, D. N.; Stemp, G.; Gardner, E. L. *J. Neurosci.* **2002**, *22*, 9595.
9. Robarge, M. J.; Husbands, S. M.; Kieltyka, A.; Brodbeck, R.; Thurkauf, A.; Newman, A. H. *J. Med. Chem.* **2001**, *44*, 3175.
10. Bettinetti, L.; Schlotter, K.; Hubner, H.; Gmeiner, P. *J. Med. Chem.* **2002**, *45*, 4594.
11. Leopoldo, M.; Berardi, F.; Colabufo, N. A.; De Giorgio, P.; Lacivita, E.; Perrone, R.; Tortorella, V. *J. Med. Chem.* **2002**, *45*, 5727.
12. Hackling, A. E.; Stark, H. *ChemBioChem.* **2002**, *3*, 946.
13. Luedtke, R. R.; Freeman, R. A.; Boundy, V. A.; Martin, M. W.; Mach, R. H. *Synapse* **2000**, *38*, 438.
14. Huang, Y. S.; Luedtke, R. R.; Freeman, R. A.; Wu, L.; Mach, R. H. *J. Med. Chem.* **2001**, *44*, 1815.
15. Calculated partition coefficient at physiological pH 7.4; ACD/LogD Suite, Advanced Chemistry Development Inc.: Toronto, Canada.
16. Belliotti, T. R.; Kesten, S. R.; Rubin, J. R.; Wustrow, D. J.; Georgic, L. M.; Zoski, K. T.; Akunne, H. C.; Wise, L. D. *Bioorg. Med. Chem. Lett.* **1997**, *18*, 2403.
17. Austin, N. E.; Avenell, K. Y.; Boyfield, I.; Branch, C. L.; Hadley, M. S.; Jeffrey, P.; Johnson, C. N.; Macdonald, G. J.; Nash, D. J.; Riley, G. J.; Smith, A. B.; Stemp, G.; Thewlis, K. M.; Vong, A. K. K.; Wood, M. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 2553.
18. Yuan, J.; Chen, X.; Brodbeck, R.; Primus, R.; Braun, J.; Wasley, J. W. F.; Thurkauf, A. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2715.
19. Wood, M. D.; Boyfield, I.; Nash, D. J.; Jewitt, F. R.; Avenell, K. Y.; Riley, G. J. *Eur. J. Pharmacol.* **2000**, *407*, 47.
20. Wicke, K.; Garcia-Ladona, J. *Eur. J. Pharmacol.* **2001**, *424*, 85.
21. Gong, Y.; Pauls, H. W. *Synlett* **2000**, 829.
22. Agoston, G. E.; Wu, J. H.; Izenwasser, S.; George, C.; Katz, J. L.; Kline, R. H.; Newman, A. H. *J. Med. Chem.* **1997**, *40*, 4329.
23. Cheng, Y. C.; Prusoff, W. H. *Biochem. Pharmacol.* **1973**, *22*, 3099.