

Available online at www.sciencedirect.com



Tetrahedron

Tetrahedron 62 (2006) 9628-9634

Synthesis of (S)-gizzerosine, a potent inducer of gizzard erosion in chicks

Yasuharu Shimasaki,^a Hiromasa Kiyota,^{a,*} Minoru Sato^b and Shigefumi Kuwahara^a

^aDivision of Bioscience and Biotechnology for Future Bioindustry, Graduate School of Agricultural Science, Tohoku University, 1-1 Tsutsumidori-Amamiya, Aoba-ku, Sendai 981-8555, Japan

1-1 Isutsumiaori-Amamiya, Aoba-ku, Senaal 981-8555, Japan

^bDivision of Biological Resource Science, Graduate School of Agricultural Science, Tohoku University, 1-1 Tsutsumidori-Amamiya, Aoba-ku, Sendai 981-8555, Japan

> Received 25 May 2006; revised 24 July 2006; accepted 26 July 2006 Available online 17 August 2006

Abstract—(S)-Gizzerosine, a potent inducer of gizzard erosion in chicks, was synthesized using successive zinc-mediated and palladiumcatalyzed coupling reactions as the key steps. © 2006 Elsevier Ltd. All rights reserved.

1. Introduction

Gizzerosine [(S)-2-amino-9-(4-imidazolyl)-7-azanonanoic acid (1) (Fig. 1)], generated during the heat treatment of brown fish meal, brings about gizzard erosion and ulceration in chicks, which cause a serious disease 'Black vomit'.^{1,2} The structure and absolute stereochemistry of 1 were confirmed by the chemical synthesis of racemic $^{3-5}$ and optically active samples.^{3,6} Biological studies of **1** showed it to be a potent agonist for the H₂-receptor of histamine (2),^{7,8} causing 10 times more gastric secretions than 2, and thereby, promoting acid indigestion and gizzard erosion. In addition, cAMP synthesis was enhanced by $1\,1000$ times more than 2.9 On the other hand, **1** did not show any visible effects on rat stomach, though the gastric secretions being promoted.^{2,10,11} This biological profile suggests 1 could be a possible drug candidate for gastric achlorhydria and osteoporosis. Since 1 can only be isolated in small amounts from fish meal (0.2 mg kg^{-1}) ,² synthetic supply is necessary for the standard of the quality control of fish meal and further biological studies. In a previous communication, we reported a facile and practical synthesis of (S)-1.^{12,13} Herein, we describe in detail this synthesis and report the development of another.



Figure 1.

2. Results and discussion

2.1. Synthesis of Mori's intermediate (formal synthesis)

Mori et al. used enzymatic optical resolution of α -acylaminoadipic acid 3 as the key reaction and determined the absolute configuration to be S (Scheme 1).⁶ We targeted the Mori's intermediates $4^{6,14}$ and 5 for the formal synthesis of **1** using (S)-serine as the starting material. (S)-Serine was first converted to the known iodide 6 in four steps.¹⁵ Conversion of 6 to the aldehyde 5 via 9 using a zinc-mediated coupling reaction^{16,17} with acrolein was attempted; however, 1.4-addition¹⁸ did not occur and only the corresponding alanine derivative 10 (formed by quenching of the intermediate zinc iodide 7) could be isolated. Next we aimed to prepare the alcohol 4 in a two-step procedure. The coupling reaction with allyl bromide¹⁹ afforded the desired olefin $\mathbf{8}$, but the use of propargyl bromide gave not alkyne 11 but allenyl compound 12.¹⁸ Hydroboration–oxidation of **8** under mildly basic conditions using sodium acetate²⁰ gave 4 without concomitant hydrolysis of the benzyl group, which was converted to (S)-gizzerosine 1 according to Mori's procedure.⁶ The overall yield of **4** from (S)-serine was 58% over six steps (Scheme 1).

2.2. New synthesis using successive metal-mediated coupling reactions

2.2.1. Initial zinc-mediated coupling reaction with enol acetate. Whilst preparing Mori's intermediate **5**, we found that the coupling of the organozinc iodide **7** with 3-bromo-1-propenyl acetate $(13)^{21}$ afforded allylic acetate **14** as a 2:1 diastereomeric mixture at the 4-position in 98% yield, instead of enol acetate **15** (Scheme 2). Allylic acetate **14** was

Keywords: Gizzerosine; Gizzerd erosion; Chicks; Total synthesis; Histamine.
 * Corresponding author. Fax: +81 22 7178783; e-mail: kiyota@biochem. tohoku.ac.jp



Scheme 1. Mori's synthesis and our formal synthesis of 1 using zinc-mediated coupling reaction: (a) i. Zn, $(CH_2Br)_2$, TMSCl, DMF, then 6, ii. CuCN, LiCl, allyl bromide (quant.); (b) i. BH₃·SMe₂, THF, ii. 30% H₂O₂, NaOAc (68%).

expected to be a good substrate for palladium-catalyzed coupling reactions with nitrogen nucleophiles,²² but the coupling reaction with histamine (2) did not afford the desired product 16 under various conditions.

2.2.2. Protection of histamine. It was thought that this unreactivity was due to the insolubility of histamine itself and/or the poor nucleophilicity of the terminal amino group. Therefore, we prepared several histamine derivatives bearing electron-donating or electron-withdrawing protecting groups. Table 1 shows the protection of histamine (2) by nucleophilic substitution reactions. Bis-Boc product **17a** was the main product in entry 1. The primary amino group was selectively masked for benzyl carbamate **17b**²³ and trityl (Tr) ether **17c** but in low yields. Formation of benzyl-oxymethyl (BOM) ether and *m*-nitrobenzenesulfonamide

Table 1. Protection of histamine by nucleophilic substitution reactions

$$H_{2N} \xrightarrow{N}_{H^{*}2HCI} \xrightarrow{HN}_{R'} H_{R'}$$

| Entry | Conditions | Products | R | R′ | Yields (%) |
|-------|---|----------|-----|-----|------------|
| 1 | (Boc) ₂ O, Et ₃ N, MeOH | 17a | Boc | Boc | 83 |
| 2 | ZCl, NaOMe, MeOH | 17b | Ζ | Н | 36 |
| 3 | TrCl, Et ₃ N, CHCl ₃ | 17c | Tr | Н | 36 |
| 4 | BOMCl, Et ₃ N, CHCl ₃ | _ | BOM | Н | Trace |
| 5 | NsCl, K ₂ CO ₃ , DMF | _ | Ns | Н | Trace |
| 6 | PMBCl, NaOMe, MeOH | _ | PMB | Н | _ |
| | | | | | |

(Ns) was observed only slightly by TLC. Since none of the product was formed in entry 6, benzyl derivatives were synthesized by reductive amination.



Scheme 2. Preparation of 14 and trials of palladium-catalyzed coupling reaction with histamine: (a) CuCN, LiCl, 13, DMF (98%).

Table 2. Protection of histamine by reductive amination reactions

| | $H_{2}N \xrightarrow{N} H \cdot 2HCI $ histamine (2) $H_{2}N \xrightarrow{N} H \cdot 2HCI $ | | IO (2.5 eq) H ₄ (2.0 eq) A, MeOH , 15 h R 18a-d | |
|-------|---|----------|--|---------------|
| Entry | ArCHO | Products | R | Yields (%) |
| 1 | Benzaldehyde | 18a | Benzyl | 93 |
| 2 | Anisaldehyde | 18b | p-Methoxybenzyl | 94 |
| 3 | Piperonal | 18c | Piperonyl | 89 |
| 4 | 3,4-Dimethoxy- benzaldehyde | 18d | 3,4-Dimethoxybenzyl | 10 |

As shown in Table 2, reductive amination of benzaldehyde, anisaldehyde, and piperonal with **2** afforded the desired compounds **18a**,²⁴ **18b**,²⁵ and **18c**,²⁵ respectively, in excellent yields.

2.2.3. Second palladium-catalyzed coupling reaction. The palladium-catalyzed coupling reactions of **14** with the histamine derivatives were examined under a variety of conditions (Table 3). The histamines with electron-withdrawing

Table 3. Palladium-catalyzed coupling reactions

groups 17a and 17b did not react even in the presence of strong bases. Trityl derivative 17c was also unreactive, probably due to its steric bulk. However, 18a bearing an electron-donating benzyl group afforded the desired product 19a in a yield of 50% when a mixture of PPh₃ (0.5 mol %) and Pd(II) (1 mol %) was used. Since increasing the amount of PPh₃ (1 mol %) gave multiple products, this mixture was subjected to further investigation. As expected, the more electron-donating derivatives 18b and 18c gave better yields of the coupling products 19b and 19c, respectively. The *E/Z* ratio of the products could not be ascertained due to overlappings in NMR spectra.

2.2.4. Total synthesis. We aimed to simultaneously carry out the deprotection of all three protecting groups and reduction of the chain double bond. Although the benzyl ester group and *N*-benzyloxycarbonyl group were easily deprotected and the double bond was reduced by hydrogenation, removal of the *N*-benzyl group of **19a** was unsuccessful (Table 4) and prolonged reaction times caused decomposition of gizzerosine framework. Additional experiments showed that gizzerosine itself was not stable under these reaction conditions. The *N*-PMB group (**19b**) was also largely inert to



| Entry | Histamines | Catalysts | Additive | Solvents | Products | R | Yields (%) | |
|-------|------------|----------------------------------|--|------------------|----------|-----------|------------|--|
| 1 | 17a | Pd ₂ dba ₃ | PPh ₃ , (base) ^a | THF | _ | _ | _ | |
| 2 | 17b | Pd ₂ dba ₃ | PPh_3 , $(base)^b$ | THF/DMF | _ | _ | _ | |
| 3 | 17c | Pd ₂ dba ₃ | PPh ₃ | THF | _ | _ | _ | |
| 4 | 18a | $Pd(OAc)_2$ | PPh ₃ | THF ^c | _ | _ | _ | |
| 5 | 18a | Pd ₂ dba ₃ | PPh ₃ | THF | 19a | Bn | 50 | |
| 6 | 18b | Pd ₂ dba ₃ | PPh ₃ | THF | 19b | PMB | 92 | |
| 7 | 18c | Pd ₂ dba ₃ | PPh ₃ | THF | 19c | Piperonyl | 71 | |
| 8 | 18d | Pd ₂ dba ₃ | PPh ₃ | THF | _ | _ | — | |

^a The following bases were used respectively: none, K₂CO₃, Et₃N, NaH, and KHMDS.

^b The following bases were used respectively: none, NaH, and KHMDS.

^c DMSO and MeCN were used respectively, besides THF.

Table 4. Hydrogenation and hydrolysis of 19a-c toward the total synthesis

| NHZ Bro I as a | $\int \bigvee$ | $ \begin{bmatrix} N\\N \end{bmatrix} $ |
|-------------------|----------------|--|
| 0 19a-c | N R R | (S)-1 N N H |

| Entry | Substrate | R | Conditions | Yields (%) |
|--------------------------|-----------|-----------|--|---|
| 1 2 3 | 19a | Bn | H ₂ , Pd/C, MeOH H ₂ , Pd(OH) ₂ /C, THF/EtOH/H ₂ O Raney Ni, MeOH/H ₂ O | Trace |
| 4 5 6 7 | 19b | РМВ | H ₂ , 5% Pd/C, THF/EtOH/H ₂ O H ₂ , 5% Pd/C, 2 M HCl aq/EtOH H ₂ , Pd(OH) ₂ /C, EtOH/H ₂ O H ₂ , Pd(OH) ₂ , 2 M HCl aq/THF/EtOH | Trace Trace Trace Trace |
| 8 9 10 11 12 | 19c | Piperonyl | H ₂ , 10% Pd/C, THF/EtOH/H ₂ O H ₂ , 10% Pd/C, Pd(OH) ₂ /C, THF/EtOH/H ₂ O H ₂ , Pd(OH) ₂ /C, THF/EtOH/H ₂ O H ₂ , Pd(OH) ₂ /C (excess), THF/EtOH/H ₂ O Raney Ni, THF/EtOH/H ₂ O | 22 Decomp. Trace 47 Decomp. |

hydrogenolysis. Selective removal under oxidative (CAN or DDQ), reductive (Li/NH₃), and acidic (2 M HCl aq) conditions also resulted in either decomposition or no reaction. On the other hand, the *N*-piperonyl group was readily removed, and finally, gizzerosine (S)-1 was obtained from **19c** in 47% yield using excess amount of $Pd(OH)_2/C$. The overall yield was 29% in seven steps from (S)-serine.

3. Conclusion

We have developed new and facile synthesis of (S)-gizzerosine (1), a potent inducer of gizzard erosion, using successive zinc-mediated and palladium-catalyzed coupling reactions as the key steps. *N*-Piperonyl group was successfully applied as a new *N*-protecting group.

4. Experimental

4.1. General

The melting points measured by Yanaco MP-J3 micromelting point apparatus were uncorrected. Optical rotation values were measured by a Horiba Sepa-300 polarimeter. IR spectra were recorded by a Jasco FT-IR 4100 spectrometer (ATR, Zn–Se). ¹H and ¹³C NMR spectra were recorded with Varian Inova 600 (150 MHz for ¹³C), Inova 500 (500 MHz for ¹H and125 MHz for ¹³C), and Gemini 2000 (300 MHz for ¹H and 75 MHz for ¹³C) spectrometers in CDCl₃ with tetramethylsilane ($\delta_{\rm H}$ 0 ppm) and CHCl₃ ($\delta_{\rm C}$ 77.00 ppm) or in D₂O with acetone ($\delta_{\rm H}$ 2.22 ppm and $\delta_{\rm C}$ 215.48 ppm) as internal standards. Mass spectra were recorded with a Jeol JMS-700 spectrometer using glycerol matrix. Merck silica gel 60 (63–212 µm) and Kanto silica gel 60N (spherical, neutral, 100–210 µm) were used for column chromatography.

4.1.1. Benzyl (S)-2-(benzyloxycarbonylamino)-5-hexenoate (8). A 500 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N2 inlet adapter, and a septum was placed under a nitrogen atmosphere. In the flask was placed zinc powder (15.2 g, 232 mmol) and 1,2-dibromoethane (1.00 ml, 11.6 mmol) in dry DMF (170 ml) was stirred at room temperature for 20 min. To the reaction mixture was added TMSCl (0.30 ml, 2.32 mol) and stirred at 60 °C for 30 min. Then to this mixture was added dropwise iodide 6 (MW: 439.24, 17.0 g, 38.7 mmol) in dry DMF (50 ml) and stirred at 60 °C for 20 min. A solution of CuCN (3.80 g, 42.5 mmol) and LiCl (3.60 g, 85.1 mmol) in dry DMF (40 ml) was added to the mixture at -55 °C. This was warmed to 0 °C and stirred for 10 min. Then the mixture was cooled to -55 °C again, allyl bromide (6.44 ml, 50.3 mmol) was added to this mixture. After the mixture was stirred at 0 °C for 2 h, unreacted zinc was filtrated through a Celite® pad and the filtrate was quenched with satd aq NH₄Cl soln. This mixture was extracted with EtOAc. The combined organic layer was washed with brine, dried over MgSO₄, and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with hexane/EtOAc gave 8 (MW: 353.41, 13.7 g, 38.7 mmol, quantitative yield from 6) as a colorless oil, which very slowly crystallized to form colorless needles; 9631

mp 37–38 °C, R_f 0.50 (hexane/EtOAc=3:1), $[\alpha]_{D}^{25}$ +1.80 (*c* 1.00, CHCl₃). ¹H NMR (300 MHz, CDCl₃): δ =7.35 (s, 10H), 5.82–5.64 (m, 1H, H-5), 5.33 (pseudo d, 1H, *J*=8.1 Hz, NH), 5.17 (pseudo d, 2H, *J*=5.4 Hz), 5.12 (s, 2H), 5.02–4.92 (m, 2H), 4.50–4.40 (m, 1H, H-2), 2.20–1.70 (m, 4H). ¹³C NMR (75 MHz, CDCl₃): δ =172.38, 155.94, 136.85, 136.29, 135.33, 128.69, 128.60, 128.56, 128.37, 128.26, 128.18, 115.87, 67.12, 66.97, 53.45, 31.76, 29.18. IR: ν =3341 (m, N–H), 3064 (w), 3034 (w), 2953 (w), 1718 (s, C=O), 1641 (w), 1521 (m), 1454 (m), 1387 (w), 1343 (m), 1254 (m), 695 (s), 584 (w) cm⁻¹. FABMS: *m*/*z*=354 (M+H)⁺, 310, 91 (Bn)⁺. FABHRMS: calcd for C₂₁H₂₄O₄N (M+H)⁺ *m*/*z*=354.1705; found, 354.1709.

4.1.2. Benzyl (S)-2-(benzyloxycarbonylamino)-5-hydroxy-5-hexenoate (4). A 100 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N₂ inlet adapter, and a septum was placed under a nitrogen atmosphere. To a solution of 8 (MW: 353.41, 1.70 g, 4.81 mmol) in dry THF (20 ml) was added dropwise BH3 · SMe2 (2.0 M in THF, 1.20 ml, 2.41 mmol) at 0 °C and the mixture was stirred while the reaction temperature gradually raised to 20 °C. To this was again added dropwise BH₃·SMe₂ (1.20 ml, 2.41 mmol) and the mixture was further stirred at 20 °C for 12 h. The reaction mixture was cooled to 0 °C before being quenched with H₂O. To the resulting solution were added successively aq H₂O₂ (30%, 8.82 M, 1.8 ml, 16 mmol) and NaOAc (0.99 g, 12 mmol) in H₂O (3 ml), and the mixture was stirred at 20 °C for 12 h. The reaction mixture was extracted with EtOAc. The extract was washed with brine, dried with MgSO₄ and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with hexane/ EtOAc gave 4 (MW: 371.43, 1.22 g, 3.28 mmol, 68.3%) as a colorless oil; $R_f 0.27$ (hexane/EtOAc=1:1), $[\alpha]_D^{26}$ -5.09 $(c \ 1.06, \text{CHCl}_3)$ {lit.⁶ $[\alpha]_D^{23} - 3.9 (c \ 1.06, \text{CHCl}_3)$ }. ¹H NMR spectral data were in agreement with the published data.¹⁴

4.1.3. Benzyl (2S,4RS)-4-acetoxy-2-(benzyloxycarbonylamino)-5-hexenoate (14). A 100 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N2 inlet adapter and a septum was placed under a nitrogen atmosphere. The flask was charged with zinc powder (5.36 g, 82.0 mmol) and 1,2-dibromoethane (0.353 ml, 4.10 mmol) in dry DMF (20 ml) and stirred at room temperature for 20 min. To the reaction mixture was added TMSCI (104 µml, 822 µmol) and stirred at 60 °C for 30 min. Then to this mixture was added dropwise iodide 6 (MW: 439.24, 6.00 g, 13.7 mmol) in dry DMF (10 ml) and stirred at 60 °C for 20 min. A solution of CuCN (1.23 g, 13.7 mmol) and LiCl (1.16 g, 27.4 mmol) in dry DMF (10 ml) was added to the mixture at -55 °C. After being warmed to 0 °C and stirred for 10 min, the mixture was cooled to $-55 \,^{\circ}\text{C}$ again, and then bromide 13 (MW: 179.01, 2.94 g, 16.4 mmol) in dry THF (7 ml) was added. After the mixture was stirred for 2 h, unreacted zinc was filtrated through Celite® pad and the filtrate was quenched with satd aq NH₄Cl soln. This mixture was extracted with EtOAc. The combined organic layer was washed with brine, dried over MgSO₄ and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with hexane/EtOAc gave 14 (a 2:1 diastereomeric mixture, MW: 411.45, 5.52 g, 13.4 mmol, 98% from 6) as a colorless oil; $R_f 0.28$ (hexane/EtOAc=4:1),

[α]_D²⁴ +16.9 (*c* 1.27, CHCl₃). ¹H NMR (500 MHz, CDCl₃): δ=7.35 (s, 10H), 5.81–5.64 (m, 1H, H-5), 5.50 (d, 1H, J=8.0 Hz, NH), 5.38–5.06 (m, 7H), 4.59–4.48 (m, 1H, H-2), 2.20 (t, 2H, J=6.0 Hz, H-3), 1.93 (s, 3H, Me). ¹³C NMR (125 MHz, CDCl₃): δ =171.71, 170.12, 155.72 (NCO), 136.09, 135.35, 135.15, 128.66, 128.64, 128.57, 128.53, 128.51, 128.40, 128.23, 128.20, 128.13, 128.10, 117.71, 117.55, 70.80, 70.78, 67.43, 67.29, 67.13, 67.09, 51.05, 50.95, 36.67, 36.12, 21.08, 20.89. IR: ν =3450–3200 (br, m), 3033 (w), 2947 (w), 1719 (s), 1523 (m), 1455 (m), 1372 (m), 1341 (m), 1229 (s), 1047 (m), 738 (m), 696 (m) cm⁻¹.FABMS: *m*/*z*=412 (M+H)⁺, 352 (M+H–AcOH)⁺, 308, 91 (Bn)⁺. FABHRMS: calcd for C₂₃H₂₆O₆N (M+H)⁺ *m*/*z*=412.7601; found, 412.1767.

4.1.4. N^{α} , N^{τ} -Bis(*tert*-butoxycarbonyl)histamine (17a). A 100 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N2 inlet adapter, and a septum was placed under a nitrogen atmosphere. The flask was charged with histamine \cdot 2HCl ($2 \cdot$ 2HCl, MW: 184.07, 1.00 g, 5.43 mmol) and Et₃N (1.65 g, 16.3 mmol) in dry MeOH (20 ml) at room temperature and stirred for 30 min. After to this being added di-tert-butyl dicarbonate (Boc₂O, 2.37 g, 10.9 mmol), the mixture was stirred for 30 min. The reaction mixture was guenched with water and concentrated in vacuo. The residue was diluted with water and extracted with EtOAc. The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with CHCl₃/EtOH (30:1) gave 17a (MW: 311.38, 1.41 g, 4.53 mmol, 83%) as a white powder; mp 129.5-130.5 °C, R_{f} 0.36 (CHCl₃/EtOH=30:1). ¹H NMR (300 MHz, CDCl₃): $\delta = 8.00$ (s, 1H), 6.82 (s, 1H), 4.99 (br s, 1H), 3.42 (t, 2H, J=6.3 Hz), 2.74 (t, 2H, J=6.6 Hz), 1.61 (s, 9H), 1.44 (s, 9H). IR: v=3258 (m), 3130 (w), 2980 (m), 2933 (m), 1736 (s), 1700 (s), 1536 (m), 1391 (s), 1365 (s), 1274 (s), 1255 (s), 1155 (s), 1009 (s), 846 (m), 777 (m), 750 (m) cm^{-1} . FABMS: m/z=312 (M+H)⁺, 256 (M+H-t-Bu)⁺, 200 $(M+H-t-Bu_2)^+$, 156 $(M+H-Boc)^+$, 112 $(M+H-Boc_2)^+$, 57. FABHRMS: calcd for $C_{15}H_{26}O_4N_3$ (M+H)⁺ m/z=312.1923; found, 312.1927.

4.1.5. N^α-(Benzyloxycarbonyl)histamine (17b). A 10 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, a Dimroth condenser, and a septum was charged with $2 \cdot 2$ HCl (30.0 mg, 0.163 mmol) and NaOMe (17.6 mg, 0.326 mmol) in MeOH (800 µl) and stirred at 50 °C for 2 h. Then the mixture was cooled to room temperature and then were added Na₂CO₃ (25.9 mg, 0.245 mmol), benzyl chloroformate (27.9 µl, 0.196 mmol), and H₂O (100 µl). This was stirred for 3 h. To the reaction mixture was added H₂O and extracted with EtOAc. The combined organic layer was washed with brine, dried over MgSO₄, and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with CHCl₃/MeOH (1:1) gave 17b (MW: 245.28, 14.2 mg, 59.1 μmol, 36%); R_f 0.63 (CHCl₃/MeOH=1:1). ¹H NMR (300 MHz, CDCl₃): δ =7.54 (s, 1H, NCH=N), 7.40-7.25 (m, 6H), 6.80 (s, 1H), 5.38 (s, 2H, ArCH₂), 3.37-3.45 (m, 2H), 2.72-2.82 (m, 2H). FABMS: m/z=246 (M+H)⁺, 136, 91 (Bn)⁺.

4.1.6. N^{α} -(Triphenylmethyl)histamine (17c). A 10 ml, round bottomed flask equipped with a magnetic stirrer bar,

was charged with $2 \cdot 2\text{HCl}$ (15.0 mg, 81.5 µmol), Et₃N (24.9 µl, 179 µmol), and triphenylmethyl chloride (TrCl, 37.0 mg, 133 µmol) in CHCl₃ (600 µl). The mixture was stirred for 20 h at room temperature. Then this mixture was quenched with cold H₂O and extracted with CHCl₃. The combined organic layer was washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with CHCl₃/EtOH (10:1) gave **17c** (MW: 353.49, 10.0 mg, 28.3 µmol, 35%); R_f 0.61 (CHCl₃/EtOH=10:1). ¹H NMR (300 MHz, CDCl₃): δ =7.40–7.10 (m, 16H), 6.50 (d, 1H, *J*=1.5 Hz), 2.72 (t, 2H, *J*=6.9 Hz), 2.39 (t, 2H, *J*=6.0 Hz).

4.1.7. N^{α} -Benzylhistamine (18a). A 100 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, a Dimroth condenser, and a septum was charged with $2 \cdot 2$ HCl (500 mg, 2.72 mmol) and NaOMe (293 mg, 5.43 mmol) in dry MeOH (20 ml) at room temperature. Then the mixture was stirred at 50 °C for 1 h. To the solution were added benzaldehyde (0.69 ml, 6.8 mmol) and MS3A (25 mg) at 0 °C and the mixture was stirred at 50 °C for 30 min. Then to the solution was added NaBH₄ (308 mg, 8.15 mmol) at -78 °C and the mixture was gradually warmed to room temperature. The reaction mixture was filtrated through a Celite[®] pad and concentrated in vacuo. The residue was diluted with water and extracted with CHCl₃. The combined extract was washed with brine, dried over Na₂SO₄, and concentrated in vacuo. The residue was chromatographed on neutral silica gel. Elution with CHCl₃/ EtOH (1:1) gave 18a (MW: 202.13, 513 mg, 2.54 mmol, 93%) as a pale yellow oil; $R_f 0.23$ (CHCl₃/MeOH=30:1, 1% Et₃N). ¹H NMR (300 MHz, CDCl₃): δ =7.51 (d. 1H. J=0.9 Hz, NCH=N), 7.38-7.20 (m, 5H), 6.79 (d, 1H, J=0.6 Hz), 3.82 (s, 2H), 2.95 (pseudo t, 2H, J=6.0 Hz), 2.80 (pseudo t, 2H, J=6.0 Hz). ¹³C NMR (150 MHz, CDCl₃): δ=139.88, 134.32, 128.49, 128.13, 127.10, 53.81, 48.63, 26.11. IR: v=3650-2200 (br s), 1567 (w), 1494 (m), 1453 (s), 1261 (w), 1104 (m), 938 (w), 820 (m), 733 (s), 697 (s), 662 (m), 624 (m) cm⁻¹. FABMS: m/z=202 $(M+H)^+$, 91 $(Bn)^+$. FABHRMS: calcd for $C_{12}H_{16}N_3$ (M+H)⁺ *m*/*z*=202.1344; found, 202.1342.

4.1.8. N^{α} -(*p*-Methoxybenzyl)histamine (18b). In the same manner as described for 18a, 18b (MW: 231.29, 2.95 g, 12.8 mmol) was obtained from $2 \cdot 2$ HCl (2.50 g, 13.6 mmol) and p-methoxybenzylaldehyde (4.14 ml, 34.0 mmol) in a yield of 94% based on $2 \cdot 2$ HCl, as a pale yellow oil; R_f 0.29 (CHCl₃/MeOH=5:1). ¹H NMR (300 MHz, CDCl₃): $\delta = 7.49$ (s, 1H, NCH=N), 7.20 (pseudo d, 2H, J=8.7 Hz), 6.85 (pseudo d, 2H, J=8.7 Hz), 6.77 (s, 1H), 3.79 (s, 3H, OMe), 3.74 (s, 2H), 2.92 (pseudo t, 2H, J=7.2 Hz), 2.76 (pseudo t, 2H, J=7.2 Hz). ¹³C NMR (75 MHz, CDCl₃): $\delta = 158.80, 134.51, 132.15, 129.38, 113.89, 55.18, 53.10,$ 48.50, 26.26. IR: ν =3500–2200 (br s), 1611 (m), 1584 (w), 1509 (s), 1460 (m), 1441 (m), 1301 (w), 1244 (s), 1175 (m), 1104 (m), 1032 (m), 938 (w), 813 (s), 773 (w), 662 (w), 626 (w) cm⁻¹. FABMS: m/z=232 (M+H)⁺, 121 (PMB)⁺. FABHRMS: calcd for $C_{13}H_{18}O_3N_3 (M+H)^+ m/z = 232.1450$; found, 232.1453.

4.1.9. N^{α} -**Piperonylhistamine** (18c). In the same manner as described for 18a, 18c (MW: 2456.28, 238 mg, 0.970 mmol) was obtained from 2.2HCl (200 mg, 1.09 mmol) and

piperonal (408 mg, 2.72 mmol) in a yield of 89% based on **2**·2HCl, as a pale yellow oil; R_f 0.79 (CHCl₃/EtOH/28% NH₃ aq=3:3:1). ¹H NMR (300 MHz, CDCl₃): δ =7.54 (d, 1H, *J*=1.2 Hz, NCH=N), 6.82–6.79 (m, 2H), 6.78–6.72 (m, 2H), 5.95 (s, 2H, OCH₂O), 3.73 (s, 2H, ArCH₂), 2.92 (pseudo t, 2H, *J*=5.7 Hz), 2.79 (pseudo t, 2H, *J*=5.7 Hz), 2.79 (pseudo t, 2H, *J*=5.7 Hz), 2.13C NMR (125 MHz, CDCl₃): δ =147.69, 146.58, 133.62, 121.30, 108.63, 108.11, 100.89, 53.46, 48.41, 26.29. IR: ν =3500–2400 (br s), 1503 (m), 1489 (s), 1442 (m), 1246 (s), 1215 (m), 1103 (w), 1039 (m), 931 (w), 810 (w), 744 (s), 665 (w), 626 (w) cm⁻¹. FABMS: m/z=246 (M+H)⁺, 135 (piperonyl)⁺. FABHRMS: calcd for C₁₃H₁₆O₂N₃ (M+H)⁺ m/z=246.1242; found, 246.1248.

4.1.10. N^{α} -(**3,4-Methoxybenzyl**)histamine (**18d**). In the same manner as described for **18a**, **18d** (MW: 245.28, 71.0 mg, 0.289 mmol, 11%) was obtained from **2**·2HCl (500 mg, 2.72 mmol) and 3,4-dimethoxybenzaldehyde (1.13 g, 6.79 mmol) in a yield of 11% based on **2**·2HCl, as a pale yellow oil; ¹H NMR (300 MHz, CDCl₃): δ =7.44 (s, 1H, NCH=N), 6.78 (s, 1H), 6.75 (s, 2H), 6.71 (s, 1H), 3.79 [s, 6H, (OMe)₂], 3.69 (s, 2H, ArCH₂), 2.86 (t, 2H, *J*=6.5 Hz), 2.73 (t, 2H, *J*=6.5 Hz).

4.1.11. Benzyl (2*S*,4*EZ*)-2-(benzyloxycarbonylamino)-6-(benzyl)[2-(4-imidazolyl)ethyl]amino-4-hexenoate (19a). In the similar manner as described for 19c, 19a (MW: 552.66, 671 mg, 1.22 mmol) was obtained from 14 (MW: 411.45, 1.00 g, 2.43 mmol) and N^{α} -benzylhistamine 18a (587 mg, 2.91 mmol) in a yield of 50% based on 14, as a yellow oil; R_f 0.29 (CHCl₃/MeOH=10:1). ¹H NMR (300 MHz, CDCl₃): δ =7.45 (s, 1H, NCH=N), 7.40–7.15 (m, 15H), 6.73 (s, 1H, NCH=C), 5.56–5.35 (m, 2H), 5.20–5.00 (m, 4H), 4.55–4.45 (m, 1H, H-2), 3.58 (s, 2H, NCH₂Ph), 3.00 (d, 2H, *J*=5.7 Hz, H-6), 2.80–2.40 (m, 6H). FABMS: m/z=553 (M+H)⁺, 471, 279, 91 (Bn)⁺. FABHRMS: calcd for C₃₃H₃₇O₄N₄ (M+H)⁺ m/z=553.2815; found, 553.2819.

4.1.12. Benzyl (2S,4EZ)-2-(benzyloxycarbonylamino)-6-[2-(4-imidazolyl)ethyl](p-methoxybenzyl)amino-4-hexenoate (19b). In the similar manner as described for 19c, 19b (MW: 582.69, 262 mg, 449 µmol) was obtained from 14 (MW: 411.45, 200 mg, 0.486 mmol) and N^{\alpha}-(p-methoxybenzyl)histamine 18b (135 mg, 533 µmol) in a yield of 92% based on 14, as a yellow oil; R_f 0.50 (CHCl₃/ EtOH=10:1). ¹H NMR (300 MHz, CDCl₃): δ =7.46 (s, 1H, NCH=N), 7.36-7.28 (m, 10H), 7.16 (d, 2H, J=8.4 Hz), 6.83 (d, 2H, J=8.7 Hz), 6.74 (s, 1H, NCH=C), 5.58-5.36 (m, 3H), 5.20-5.00 (m, 4H), 4.54-4.45 (m, 1H), 3.78 (s, 3H, OMe), 3.52 (s, 2H), 2.99 (d, 2H, J=5.4 Hz), 2.68 (dd, 2H, J=16.5, 5.4 Hz), 2.56 (t, 2H, J=5.7 Hz), 2.49 (t, 2H, J=6.6 Hz). FABMS: m/z=583 (M+H)⁺, 501, 230, 121 (PMB)⁺, 91 (Bn)⁺. FABHRMS: calcd for C₃₄H₃₉O₅N₄ (M+H)⁺ *m*/*z*=583.2921; found, 583.2922.

4.1.13. Benzyl (2*S***,4***EZ***)-2-(benzyloxycarbonylamino)-6-[2-(4-imidazolyl)ethyl](piperonyl)amino-4-hexenoate (19c).** A 20 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N₂ inlet adapter, and a septum was placed under a nitrogen atmosphere. The flask was charged with 14 (MW: 411.45, 540 mg, 1.31 mmol), Pd₂(dba)₃ (60.0 mg, 65.6 µmol), and PPh₃ (18.2 mg, 69.4 µmol) in dry THF, and stirred for 15 min at room temperature. To the suspension was added N^{α} -piperonylhistamine **19c** (555 mg, 1.58 mmol), warmed up to 50 °C, and the mixture was stirred for 1 h. The reaction mixture was filtrated through a Celite[®] pad, added water, and extracted with EtOAc. The combined organic layer was washed with brine, dried over MgSO₄, and concentrated in vacuo. The residue was chromatographed on silica gel. Elution with CHCl₃/EtOH (20:1) gave 19c (MW: 507.27, 550 mg, 0.922 mmol, 71% based on 14) as a yellow oil; $R_f = 0.57$ (CHCl₃/EtOH=30:1), $[\alpha]_D^{24} = -0.703$ (c 1.28, CHCl₃). ¹H NMR (300 MHz, CDCl₃): δ =7.47 (s, 1H, NCH=N), 7.40-7.25 (m, 10H), 6.75-6.65 (m, 4H), 5.91 (s, 2H, OCH₂O), 5.64–5.36 (m, 3H), 5.22–5.00 (m, 4H), 4.54– 4.45 (m, 1H, H-2), 3.47 (d, 2H, NCH₂Ar), 2.98 (d, 2H, J=5.4 Hz, NCH₂C=), 2.78–2.40 (m, 6H). ¹³C NMR (75 MHz, CDCl₃): δ =171.79, 155.91, 147.76, 146.66, 136.19, 135.19, 134.30, 132.84, 131.73, 128.67, 128.56, 128.36, 128.23, 128.05, 127.07, 121.97, 109.19, 107.93, 100.86, 67.20, 66.93, 57.85, 54.78, 53.66, 52.56, 35.35, 23.18. IR: v=3493-2100 (br s), 1717 (s), 1500 (s), 1487 (s), 1454 (m), 1440 (s), 1342 (w), 1244 (s), 1214 (s), 1187 (m), 1093 (w), 1038 (s), 975 (w), 930 (w), 810 (w), 749 (s), 696 (s), 665 (w) cm⁻¹. FABMS: m/z=597 (M+H)⁺, 154, 136, 91 (Bn)⁺, 73. FABHRMS: calcd for $C_{34}H_{37}O_6N_4$ (M+H)⁺ m/z=597.2713; found, 597.2718.

4.1.14. (S)-2-Amino-9-(4-imidazolyl)-7-azanonanoic acid (gizzerosine, 1). A 30 ml, two-necked round bottomed flask equipped with a magnetic stirrer bar, an N₂ inlet adapter, and a septum was placed under a hydrogen atmosphere. The flask was charged with 19c (MW: 596.67, 54.4 mg, 91.1 µmol) and Pd(OH)₂/C (110 mg) in 5 ml of EtOH/ THF/H₂O (3:1:1). The mixture was stirred for 5 h at room temperature. Then this was filtrated through a Celite[®] pad and the filtrate was concentrated in vacuo. The resulting solid was purified through Sephadex[®] LH-20 column. Elution with MeOH/H₂O (1:1) gave gizzerosine 1 (MW: 240.30, 10.4 mg, 0.433 mmol, 47%) as a white solid. This solid was dissolved in 1 M HCl aq and stirred for 5 min. Then this solution was concentrated in vacuo. This resulting white amorphous solid was used for analysis; R_f 0.09 (CHCl₃/MeOH/28% NH₃ aq=3:3:1), mp 250–251 °C (dec) [lit.⁶ 251–252 °C (dec)]; $[\alpha]_D^{22}$ +9.45 (*c* 0.555, H₂O) {lit.⁶ $[\alpha]_D^{22}$ +10.3 (*c* 1.28, H₂O)}. ¹H NMR (300 MHz, D₂O): δ =8.30 (br s, 1H), 7.26 (br s, 1H), 3.75 (t, 1H, J=6.0 Hz), 3.37 (pseudo t, 2H, J=7.2 Hz), 3.11 (pseudo t, 4H, J=6.6 Hz), 1.89 (m, pseudo q, 2H, J=7.4 Hz), 1.74 (quint, 2H, J=7.5 Hz), 1.60–1.35 (m, 2H). ¹³C NMR (75 Hz, D_2O): $\delta = 175.08$, 135.69, 131.45, 116.93, 54.99, 47.72, 46.86, 30.38, 25.65, 22.91, 22.04. IR: $\nu = 3300 - 2300$ (br s), 3116 (s), 2786 (s), 2450 (m), 1634 (s), 1601 (s), 1522 (s), 1462 (s), 1395 (s), 1348 (m), 1329 (m), 1235 (w), 1052 (w), 957 (w), 839 (w), 794 (w), 719 (w), 623 (m) cm⁻¹. FABMS: *m*/*z*=241 (M+H)⁺, 207, 185, 93, 75, 57, 45. FABHRMS: calcd for $C_{11}H_{21}N_4O_2$ (M+H)⁺ m/z=241.1665; found, 241.167. These spectral data are in good agreement with those reported.6

Acknowledgements

We thank Prof. Takeshi Sugai (Keio Univ., Japan) for useful advices and Prof. Hidenori Watanabe (Tokyo Univ., Japan) for kindly giving us the authentic sample.

References and notes

- 1. Johnson, D. C.; Pinedo, D. C. Avian Dis. 1971, 15, 835.
- Okazaki, T.; Noguchi, T.; Igarashi, K.; Sakagami, Y.; Seto, H.; Mori, K.; Naito, H.; Masumura, T.; Sugahara, M. Agric. Biol. Chem. 1983, 47, 2949.
- Mori, K.; Okazaki, T.; Noguchi, T.; Naito, H. Agric. Biol. Chem. 1983, 47, 2131.
- 4. Bazureaum, J. P.; Person, D.; Le Corre, M. *Tetrahedron Lett.* **1989**, *30*, 3065.
- Herrera, C.; Tello, M.; Ibanez, I.; Valenzuela, O.; Olivares, L. Bol. Soc. Chil. Quím. 1999, 44, 117.
- Mori, K.; Sugai, T.; Maeda, Y.; Okazaki, T.; Noguchi, T.; Naito, H. *Tetrahedron* 1985, *41*, 5307.
- Masumura, T.; Sugahara, M.; Noguchi, T.; Mori, K.; Naito, H. Poult. Sci. 1985, 64, 356.
- Sugahara, M.; Hattori, T.; Nakajima, T. Agric. Biol. Chem. 1987, 51, 3423.
- Ito, Y.; Terao, H.; Noguchi, T.; Naito, H. Poult. Sci. 1988, 67, 1290.
- Masumura, T.; Sugahara, M. Jpn. J. Zootech. Sci. 1982, 53, 743 (in Japanese).
- 11. Noguchi, T. Seikagaku 1988, 60, 1168 (in Japanese).
- Shimasaki, Y.; Kiyota, H.; Sato, M.; Kuwahara, S. Synthesis 2005, 3191.

- 13. Kiyota, H. Biosci. Biotechnol. Biochem. 2006, 70, 317.
- 14. Tice, T. M.; Ganem, B. J. Org. Chem. 1983, 48, 5043.
- Adlington, R. M.; Baldwin, J. E.; Basak, A.; Kozyrod, R. P. J. Chem. Soc., Chem. Commun. 1983, 944.
- 16. Knochel, P.; Singer, R. D. Chem. Rev. 1993, 93, 2117.
- 17. Gair, S.; Jackson, R. F. W. Curr. Org. Chem. 1998, 2, 527.
- 18. Dexter, C. S.; Jackson, R. F. W. J. Org. Chem. 1999, 64, 7579.
- Dunn, M. J.; Jackson, R. F. W.; Pietruszka, J.; Turner, D. J. Org. Chem. 1995, 60, 2210.
- Xue, C.-B.; Voss, M. E.; Nelson, D. J.; Duan, J. J.-W.; Cherney, R. J.; Jacobson, I. C.; He, X.; Roderick, J.; Chen, L.; Corbett, R. L.; Wang, L.; Meyer, D. T.; Kennedy, K.; DeGrado, W. F.; Hardman, K. D.; Teleha, C. A.; Jaffee, B. D.; Liu, R.-Q.; Copeland, R. A.; Covington, M. B.; Christ, D. D.; Trzaskos, J. M.; Newton, R. C.; Magolda, R. L.; Wexler, R. R.; Decicco, C. P. J. Med. Chem. 2001, 44, 2636.
- Lombard, M.; Girotti, R.; Morganti, S.; Trombini, C. Org. Lett. 2001, 3, 2981.
- Heck, R. F. Palladium Reagents in Organic Syntheses; Katritzky, A. R., Meth-Cohn, O., Rees, C. W., Eds.; Academic: London, 1985; pp 122–130.
- Nagao, Y.; Seno, K.; Kawabata, K.; Miyasaka, T.; Takao, S.; Fujita, E. *Tetrahedron Lett.* **1980**, *21*, 841.
- 24. Plimls, J.; Protiva, M. Chem. Listy 1953, 47, 1874.
- 25. van der Merwe, P. Z. Physiol. Chem. 1928, 177, 301.