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Synthesis and Antiallergic Activity of Pyridothienopyrimidines

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Abstract—The synthesis of a series of pyridothienopyrimidines and their evaluation as inhibitors or inducers of the release of histamine from rat mast cells is reported. The activity was measured after immunological stimulation with ovoalbumin and chemical stimulation with polymer 48/80 and the drugs adryamicin and vinorelbine. The experiments were carried out with and without preincubation of the stimulus with the cells before addition of the drug. Several pyridothienopyrimidines show inhibitory IC₅₀ values in the range $2-25\,\mu$ M, indicating they are up to 100 times more potent than cromoglycate (DSCG) and 10 times greater than Ketotifen. Compound 91 is a potent inhibitor in all the conditions tested and shows $IC_{50}=9-25\,\mu$ M. Pyridothienopyrimidines **4I** and **9e** are very strong inducers of histamine release in the immunological (41, 170-230%) and chemical (9e, 100-150%) assays, respectively. Compounds 41 and 9i are cytotoxic in vitro (IC₅₀=0.1-0.2 µg/mL) against P-388, A-549, HT-29, and MEL-28 tumor cell lines. © 1998 Elsevier Science Ltd. All rights reserved.

Introduction

Asthma is an inflammatory disease that affects a significant portion of the population, with recent data suggesting that the numbers of asthma sufferers is increasing.¹ One of the most useful drugs for asthma treatment is disodium cromoglycate^{2a,b} (DSCG, Chart 1). This compound inhibits IgE-mediated secretion of vasoactive mediators from mast cells both in vivo and in vitro, but unfortunately is not effective by oral administration and must be used as an insufflated powder. Several orally active drugs have been described and many of them are nitrogen or sulfur-containing heterocycles including tetrazoles,³ naphthyridines,⁴ pyridothienotriazines⁵ such as RHC 2963, and thiophenes as Ketotifen^{2c} (Chart 1).

Compounds containing a fused pyrimidine ring have attracted attention in the past few years owing to their

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wide range of biological activity, particularly in cancer and virus research.⁶ Among these heterocycles, the thienopyrimidine class is also of interest because some derivatives, such as Tiprinast⁷ (Chart 1), have been shown to be clinically effective antiallergics. In addition, antianaphilactic,8 anti-inflammatory9 analgesic and antipyretic,¹⁰ and antineoplastic¹¹ activities have been described for these compounds.

Our previous work^{12a} on the search for novel heterocyclic compounds of pharmacological interest has shown that certain guanadinocyanopyridines and pyridopyrimidines are good inhibitors, and other strong stimulants, of the release of histamine from mast rat cells. These findings, along with our interest in the development of structurally novel compounds acting as histamine release inhibitors, and the known effectiveness of pyridothienotriazine RHC29635 and thienopyrimidine Tiprinast⁷ as immunological inhibitors of the release of histamine, prompted us to report here our results on the preparation and study of the inhibition of the release of histamine from rat mast cells and the antitumor activity of a series of new pyridothienopyrimidines shown in Table 1.

Key words: Pyridothienopyrimidine; synthesis; histamine; rat mast cells ; antiallergic; cytotoxicity.



Chart 1.

Chemistry

For the preparation of the heterocycles we devised suitable procedures for the introduction of a variety of substituents with a minimum change in the nature of the chemistry used. Thus, substituents R, R₁, and R₂ are present in the pyridine or thienopyridine used as the starting material. R can be either an aryl ring or hydrogen, R₁ any alkoxy or amino group, and R₂ a cyano group or hydrogen. Substituent R₃ is introduced during the formation of the pyrimidine ring. Most of the compounds described in this paper have R₃=H or NMe₂, depending on the method employed for the cyclization. Finally, R_4 is introduced in the last step by means of an aromatic nucleophilic substitution of a chloro-substituent. Several nucleophiles of nitrogen, oxygen, and sulfur have been shown to be effective in this reaction.

The structure of the required starting compounds is mainly determined by the nature of the substituents on the pyrimidine ring and, as a rule, systems with an amino group adjacent to another functional group are the most widely used.

In this paper we have chosen bicyclic thienopyridines with either an ortho amino nitrile (1a-b) or an ortho amino carboxamide (5a-b) functionality as the starting materials for the preparation of the tricyclic pyridothienopyrimidines shown in Table 1. In accordance with these two functionalities, two different types of cyclization reaction were needed.

In the case of pyridothienopyrimidines,⁹ cyclization was carried out by the introduction of the required carbon atom with orthoformate, while in the case of pyridothienopyrimidines 4 and 21, carrying a dimethylamino group at position 2, phosgene iminium salts were employed.

These salts are very useful synthetic reagents that can function as Vilsmeier or Mannich acceptors and lead to the insertion of a one-carbon unit bearing a dialkylamino group.¹³ This makes them especially useful in one-step heterocyclization reactions such as those considered here.

Thus, treatment of 3-aminothieno[2,3-*b*]pyridine-2-carbonitriles $1a^{14}$ and $1b^{15}$ (Scheme 1) with phosgene iminium chloride in refluxing 1,2-dichloroethane gave the amide halide intermediates 2a-b, which were reacted with dry



hydrogen chloride to undergo cyclization to chloropyridothienopyrimidines **3a–b**. These compounds were easily converted into a variety of pyridothienopyrimidines **4a–1** by aromatic substitution with nucleophiles (Table 1).

The structures of **2** and **3** were consistent with their elemental analyses and spectral data. The mass spectra showed the expected molecular ion peaks and the IR spectrum of **2** exhibited an absorption band at $v = 1640 \text{ cm}^{-1}$ due to the imino group. The ¹³C NMR spectrum of **2a** showed two signals at $\delta = 113.8$ and 113.9 due to the cyano group, while compound **3a** showed only one signal for cyano groups at $\delta = 114.4$.

 R_2

CN

CN

CN

CN

CN

CN

Compounds **9a–1** were prepared in two ways by the reactions shown in Scheme 2. Refluxing heterocyclic carboxamides **5a**¹⁶ and **5b**¹⁵ with ethyl orthoformate gave the triheterocyclic compounds **7a–b**, which were converted to the corresponding chloro derivatives **8a–b** by treatment with phosphorous oxychloride. Aromatic nucleophilic substitution with a variety of nucleophiles afforded the new substituted pyridothienopyrimidine derivatives **9a–1**.

An alternative method for the preparation of 7a consisted of the reaction of 5a with dimethylformamide dimethylacetal to give 6, which could readily be converted into 7a by refluxing with sodium ethoxide.

 $mp(^{\circ}C)$

220-222

192-194

204-206

184 dec

225 dec

170 dec

yield (%)

86^f 72^b

71^b

 64^{f}

77

65^d

Table 1. Physicochemical data for pyridothienopyrimidines 3a, 4a-l, 9a-l, 13, and 21a-b

 R_3

NMe₂

NMe₂

NMe₂

NMe₂

NMe₂

NMe₂

Cl

OEt

piperidino

piperazino

4-methylpiperidino

4-methylpiperazino

 R_4

4f	Ph	EtO	CN	NMe ₂	4-ethoxycarbonylpiperazino	217-219	77 ^b	$C_{27}H_{29}N_7O_3S$
4g	Ph	EtO	CN	NMe ₂	4-(4-acetylphenyl) piperazino	212-214	65 ^b	$C_{32}H_{31}N_7O_2S$
4h	Ph	EtO	CN	NMe ₂	4-(trifluoro-m-tolyl)-piperazino	249-251	72 ^b	C31H28N7OF3S
4i	Ph	Ph	Η	NMe ₂	4-methylpiperidino	138-140	59e	$C_{29}H_{29}N_5S$
4j	Ph	Ph	Η	NMe ₂	4-methylpiperazino	200-202	64 ^b	$C_{28}H_{28}N_6S$
4k	Ph	Ph	Н	NMe ₂	4-(4-acetylphenyl) piperazino	214-216	58 ^b	C35H32N6OS
41	Ph	Ph	Н	NMe ₂	piperazino	> 300	72 ^d	$C_{27}H_{26}N_6S$
9a	Ph	EtO	CN	Н	piperidino	189–191	75 ^b	C23H21N5OS
9b	Ph	EtO	CN	Н	4-methylpiperidino	210-212	74 ^c	C24H23N5OS
9c	Ph	EtO	CN	Н	morpholino	220-222	80 ^b	$C_{22}H_{19}N_5O_2S$
9d	Ph	EtO	CN	Н	piperazino	212-214	74 ^d	$C_{22}H_{20}N_6OS$
9e	Ph	EtO	CN	Н	4-methylpiperazino	212-214	65 ^d	C23H22N6OS
9f	Ph	EtO	CN	Н	4-benzylpiperazino	229-231	81 ^d	$C_{29}H_{26}N_6OS$
9g	Ph	EtO	CN	Н	4-(4-acetylphenyl) piperazino	226-228	85 ^d	$C_{30}H_{26}N_6O_2S$
9h	Ph	EtO	CN	Н	4-(trifluoro-m-tolyl)-piperazino	213-215	83 ^d	$C_{29}H_{23}N_6OF_3S$
9i	Ph	Ph	Н	Н	piperazino	191–193	68°	$C_{25}H_{21}N_5S$
9j	Ph	Ph	Н	Н	4-methylpiperidino	145-148	65 ^e	$C_{27}H_{24}N_4S$
9k	Ph	Ph	Η	Н	4-methylpiperazino	190-192	84 ^b	$C_{26}H_{23}N_5S$
91	Ph	Ph	Н	Н	4-(4-acetylphenyl) piperazino	230-232	62 ^d	C33H27N5OS
13	Ph	EtO	CN	$2NO_2C_6H_4$	4-benzylpiperazino	209-211	71 ^d	C35H29N7O3S
21a	Н	morpholino	CN	NMe ₂	morpholino	235-237	77 ^b	$C_{20}H_{23}N_7O_2S$
21b	Н	morpholino	CN	NMe ₂	thiomorpholino	218-220	75 ^b	$C_{20}H_{23}N_7OS_2$

^aAll compounds analyzed for C, H, N; analytical results are within $\pm 0.4\%$ of theoretical values.

^bRecrystallized from ethanol/acetone.

^cRecrystallized from ethanol.

Compd

3a

4a

4h

4c

4d

4e

R

Ph

Ph

Ph

Ph

Ph

Ph

 R_1

EtO

EtO

EtO

EtO

EtO

EtO

^dRecrystallized from ethanol/dichloromethane.

 $^{e}\mbox{Purified}$ by flash chromatography with dichloromethane/hexane (3/1, v/v) as eluent.

^fPurified by flash chromatography with dichloromethane/hexane (1/2, v/v) as eluent.

formula^a

C20H16N5OCIS

C22H21N5O2S

C25H26N6O2S

 $C_{26}H_{28}N_6OS$

 $C_{24}H_{25}N_7O_2S$

C25H27N7OS

 Table 2.
 Selected spectroscopic data for pyridothienopyrimidines

Compd	¹ H NMR (CDCl ₃) δ ^{a,b}	$^{13}C \ NMR \ (CDCl_3) \ \delta^{a,b}$	MS (FAB) ^c m/z (%)
3a	2.82 (br s, 6H, NMe ₂)	36.9 (NMe ₂)	411 (M ⁺ +2, 9), 410 (40), 409 (M ⁺ , 26), 408 (100), 380 (54), 352 (33)
4 a	1.52 (t, 3H, $J=7.1$ Hz, CH ₃), 2.80 (s, 6H, NMe ₂), 4.52 (a) 2H, $J=7.1$ Hz, OCH ₂)	14.4 (CH ₃), 36.7 (NMe ₂), 62.3 (CH ₂ O)	419 (M ⁺ , 6), 391 (100), 376 (23) 362 (30)
4b	$4.52 (q, 2H, J = 7.1 Hz, 0CH_2)$	$24.8 (CH_2O)$	370(23), 302(30) $458(M^+, 100), 443(33)$
-10	$3.78 \text{ (br s, 4H, N(CH_2)_2)}$	47.5 (NCH ₂)	429 (26), 415 (29), 403 (12)
4c	0.96 (d, 2H, $J = 6.0$ Hz, CHCH ₃),	21.8 (CHCH ₃), 31.4 (CH ₃ CH),	473 [(MH) ⁺ , 100],
	$1.23-1.34$ (m, 2H, CHC H_2),	36.5 (NMe ₂), 46.8 (NCH ₂)	445 (23), 429 (8)
	1.59–1.76 (m, 3H, CHCH ₂ , CHCH ₂), 2.74 (s, 6H, NMe ₂),		
	3.00 (t, 2H, J=12.7 Hz, NCH ₂), 4.56–4.66 (m, 2H, NCH ₂)		
4d	2.66 (s, 6H, NMe ₂), 2.90 (br s, 4H, NCH ₂),		460 [(MH) ⁺ , 100],
	3.75 (br s, 4H, NCH ₂)		432 (19), 403 (16)
4e	2.33 (s, 3H, NCH ₃), 2.51 (t, 4H, <i>J</i> =4.9 Hz, NCH ₂),	36.5 (NMe ₂), 46.1,	474 [(MH) ⁺ , 100],
	2.74 (s, 6H, NMe ₂), 3.84 (t, 4H, <i>J</i> =4.9 Hz, NCH ₂)	54.7 (NCH ₂)	472 (12), 446 (10), 403 (12)
4f	1.28 (t, 3H, J =7.1 Hz, CH ₃), 2.74 (s, 6H, NMe ₂), 3.58–3.83 (m, 8H, NCH ₂), 4.17 (c, 2H, J =7.1 Hz, OCH ₂) 14.6 (CH ₃), 36.5 (NMe ₂), 43.4, 46.0 (NCH ₂), 61.6 (CH ₂ O), 155.4 (CO) 531 (M ⁺ 58) 403 (100) 375 (16)		
4g	2.52 (s. 3H, CH ₂ CO), 2.75 (s. 6H, NMe ₂).	26.1 (CH ₂ CO), 36.5 (NMe ₂).	578 [(MH) ⁺ , 39].
-8	3.49 (t, 4H, $J = 5.1$ Hz, NCH ₂).	45.5, 46.8 (NCH ₂), 113.2, 128.1.	403 (12), 217 (100)
	$4.00 (t, 4H, J=5.1 Hz, NCH_2).$	130.3. 153.6 (C ₄ H ₄). 194.6 (CO)	()
	6.87, 7.89 (AB system, 4H, $J=9.0$ Hz, C ₆ H ₄)		
4h	2.75 (s, 6H, NMe ₂), 3.34 (t, 4H, $J = 5.0$ Hz, NCH ₂),	36.5 (NMe ₂), 45.8, 48.4 (NCH ₂),	604 [(MH) ⁺ , 10],
	3.98 (t, 4H, $J = 4.9$ Hz, NCH ₂), 7.05–7.12 (m, 4H, C ₆ H ₄ CF ₃)	112.1, 112.2, 129.6, 131.7 (C ₆ H ₄)	403 (4), 217 (100)
4i	$1.00 (d, 2H, J = 6.1 Hz, CHCH_3), 1.33-1.40 (m, 2H, CHCH_2),$	21.9 (CHCH ₃), 31.4 (CH ₃ CH),	480 [(MH) ⁺ , 100],
	1.61–1.81 (m, 3H, CHCH ₂ , CHCH ₂), 2.88 (s, 6H, NMe ₂),	34.1 (CH ₂ CH), 36.6 (NMe ₂),	437 (13), 410 (4)
	$3.06 (dt, 2H, J=1.3 Hz, J=12.7 Hz, NCH_2),$	46.9 (NCH ₂)	
	4.71 (br s, 2H, NCH ₂)	·	
4j	2.37 (s, 3H, NCH ₃), 2.57 (t, 4H, J=4.8 Hz, NCH ₂),	36.7 (NMe ₂), 46.1 (NCH3),	481 [(MH) ⁺ , 100],
	2.86 (s, 6H, NMe ₂), 3.94 (t, 4H, <i>J</i> =4.8 Hz, NCH ₂)	46.2 (NCH ₂), 54.9 (NCH2)	467 (3), 424 (16), 410 (21)
4k	2.53 (s, 3H, COCH ₃), 2.86 (s, 6H, NMe ₂),	26.1 (CH ₃ CO), 36.7 (NMe ₂),	585 [(MH) ⁺ , 40],
	3.54 (t, $3H$, $J = 5.1$ Hz, NCH ₂),	45.7, 47.0 (NCH ₂), 113.3,	451 (69), 410 (46)
	4.07 (t, 3H, $J = 5.2$ Hz, NCH ₂),	127.5, 130.4, 153.8 (C ₆ H ₄),	
	6.90, 7.91 (AB system, 4H, $J = 9.0$ Hz, C_6H_4)	196.5 (CO)	
41	2.85 (s, 6H, NMe ₂), 3.00 (t, 3H, $J = 5.0$ Hz, NCH ₂),	36.6 (NMe ₂), 45.9, 47.5 (NCH ₂)	467 (M ⁺ , 25), 410 (100),
	3.87 (t, 3H, $J = 5.0$ Hz, NCH ₂)		397 (60), 382 (28), 368 (29)
9a	1.74 (br s, 6H, CH ₂ -CH ₂ -CH ₂), 3.89 (br s, 4H, NCH ₂),	24.6 (CH ₂), 26.0 (CH ₂),	415 (M ⁺ , 100), 386 (43),
	8.35 (s, 1H, H-2)	47.6 (NCH ₂)	359 (17), 332 (22)
9b	0.99 (d, 3H, <i>J</i> =6.2 Hz, CHCH ₃), 1.20–1.38 (m, 2H, CHCH ₂),	21.7 (CHCH ₃), 31.2 (CH ₃ CH),	430 (M ⁺ , 95), 417 (87),
	1.69-1.84 (m, 3H, CHCH ₂ + CHCH ₂),	34.2 (CH ₂ CH), 46.9 (NCH ₂)	232 (16)
	$3.10 (t, 2H, J = 12.3 Hz, NCH_2),$		
	4.63–4.70 (m, 2H, NCH ₂), 8.35 (s, 1H, H-2)		
9c	3.83–3.94 (m, 8H, CH ₂ -CH ₂), 8.39 (s, 1H, H-2)	46.6 (NCH ₂), 66.7 (OCH ₂)	417 (M ⁺ , 100), 388 (18), 386 (23), 360 (49)
9d	3.03 (t, 4H, <i>J</i> =4.9 Hz, NCH ₂), 3.91 (t, 4H, <i>J</i> =4.9 Hz, NCH ₂), 8.38 (s, 1H, H-2)	47.4 (NCH ₂), 52.9 (NCH ₂)	417 [(MH) ⁺ , 100], 389 (9)

(continued)

Table 2—contd

$ \begin{array}{c} & 346 (20) \\ & 343 (t, 4H, J=4.8 Hz, NCH_2), 3.56 (s, 2H, CH_2Ph), \\ & 3.53 (t, 1H, J=2) \\ & 9g \\ & 2.54 (s, 3H, CH_3CO), 3.55 (t, 4H, J=5.2 Hz, NCH_2), \\ & 4.12 (t, 4H, J=5.2 Hz, NCH_2), 6.90, \\ & 7.91 (AB system, 4H, J=9.0 Hz, C_6H_4), \\ & 840 (s, 1H, H=2) \\ & 9h \\ & 3.40 (t, 4H, J=5.1 Hz, NCH_2), \\ & 4.10 (t, 4H, J=5.1 Hz, NCH_2), \\ & 4.11 (t, 4H, J=4.2 Hz, NCH_2), \\ & 4.11 (t, 4H, J=4.2 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.2 Hz, NCH_2), \\ & 5.0 (s, 1H, H=2) \\ & 9i \\ & 3.11 (t, 4H, J=4.2 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.2 Hz, NCH_2), \\ & 4.03 (t, 2H, J=0.2 Hz, J=12.4 Hz, NCH_2), \\ & 4.03 (t, 2H, J=0.2 Hz, J=12.4 Hz, NCH_2), \\ & 4.03 (t, 2H, J=0.2 Hz, J=12.4 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.2 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.2 Hz, NCH_2), \\ & 5.1 (s, 3H, NCH_3), 2.50 (t, 4H, J=4.8 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.8 Hz, NCH_2), \\ & 5.1 (s, 3H, NCH_3), 3.55 (t, 3H, J=4.8 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.8 Hz, NCH_2), \\ & 5.1 (s, 3H, NCH_3), 3.55 (t, 3H, J=4.8 Hz, NCH_2), \\ & 4.01 (t, 4H, J=4.8 Hz, NCH_2), \\ & 5.4 (NCH_2), \\ & 13.8 (COH_3), \\ & 4.01 (t, 4H, J=5.9 Hz, C_6 H_4), 8.50 (s, 1H, H=2) \\ & 13.9 (C-2) \\ & 13. 2.61 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.90 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), \\ & 3.81 (4), 368 (10), 376 (10) \\ & 10100 \\ \\ & 14. 3.81, 3.58 (m, 16H, CH_2$	9e	2.36 (s, 3H, NCH ₃), 2.56 (t, 4H, <i>J</i> = 4.9 Hz, NCH ₂), 3.95 (t, 4H, <i>J</i> = 4.9 Hz, NCH ₂), 8.37 (s, 1H, H-2)	46.0 (NCH ₃), 46.2 (NCH ₂), 54.8 (NCH ₂)	431 [(MH) ⁺ , 100], 429 (10), 374 (13),
$ \begin{array}{llllllllllllllllllllllllllllllllllll$				346 (20)
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	9f	2.59 (t, 4H, $J = 4.8$ Hz, NCH ₂), 3.56 (s, 2H, CH ₂ Ph),	46.3 (CH ₂), 52.9 (NCH ₂),	506 [(MH) ⁺ , 20],
		$3.93 (t, 4H, J = 4.9 Hz, NCH_2), 7.24-7.35 (m, 5H, C_6H_5),$	62.9 (NCH ₂)	217 (100)
$ \begin{array}{llllllllllllllllllllllllllllllllllll$		8.35 (s, 1H, H-2)		
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	9g	2.54 (s, 3H, CH ₃ CO), 3.55 (t, 4H, $J = 5.2$ Hz, NCH ₂),	26.0 (CH ₃ CO), 45.5,	535 [(MH) ⁺ , 12],
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		4.12 (t, 4H, $J = 5.2$ Hz, NCH ₂), 6.90,	47.0 (NCH ₂), 112.3, 113.4,	360 (2)
$ \begin{array}{llllllllllllllllllllllllllllllllllll$		7.91 (AB system, 4H, $J = 9.0$ Hz, C_6H_4),	128.3, 130.4 (C ₆ H ₄), 196.5 (CO)	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$		8.40 (s, 1H, H-2)		
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	9h	$3.40 (t, 4H, J = 5.1 Hz, NCH_2),$	45.6, 48.6 (NCH ₂), 112.4 (C-6'),	561 [(MH) ⁺ , 20],
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		4.11 (t, 4H, $J = 5.1$ Hz, NCH ₂),	116.5, 116.6 (C-2', C-4'),	217 (100)
$ \begin{array}{c} 131.9 \ (C-3'), \ 151.0 \ (C-5') \\ 1424 \ [(MH)^+, \ 100], \\ 381 \ (25), \ 367 \ (14), \\ 381 \ (25), \ 367 \ (14), \\ 355 \ (11) \\ 391 \ 0.89 \ (d, 2H, J=6.2 \ Hz, \ CHCH_3), \\ 1.16-1.31 \ (m, 2H, \ CHCH_2), \\ 3.01 \ (dt, 2H, J=2.3 \ Hz, \ J=12.4 \ Hz, \ NCH_2), \\ 4.63 \ (br s, 2H, \ NCH_2), \ 8.37 \ (s, 1H, \ H-2) \\ 153.9 \ (C-2) \\ 9k \ 2.37 \ (s, 3H, \ NCH_3), \ 2.60 \ (t, 4H, J=4.8 \ Hz, \ NCH_2), \\ 4.01 \ (t, 4H, J=4.8 \ Hz, \ NCH_2), \ 8.51 \ (s, 1H, \ H-2) \\ 153.8 \ (C-2) \\ 9l \ 2.53 \ (s, 3H, \ COCH_3), \ 3.55 \ (t, 3H, J=4.8 \ Hz, \ NCH_2), \\ 4.13 \ (t, 3H, J=5.3 \ Hz, \ NCH_2), \ 6.89, \\ 7.90 \ (AB \ system, \ 4H, \ J=8.9 \ Hz, \ C_{6}H_4), \ 8.50 \ (s, 1H, \ H-2) \\ 153.8 \ (C-2) \\ 9l \ 2.53 \ (s, 3H, \ COCH_3), \ 3.55 \ (t, 3H, J=4.8 \ Hz, \ NCH_2), \\ 4.13 \ (t, 3H, J=5.3 \ Hz, \ NCH_2), \ 6.89, \\ 7.90 \ (AB \ system, \ 4H, \ J=8.9 \ Hz, \ C_{6}H_4), \ 8.50 \ (s, 1H, \ H-2) \\ 153.8 \ (C-2) \\ 9l \ 3.81 \ (2.2), \ 130.4 \ (C_{6}H_4), \\ 153.9 \ (C-2) \\ 13 \ 2.61 \ (s, 4H, \ NCH_2), \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.90 \ (s, 4H, \ NCH_2) \ 3.58 \ (s, 2H, \ CH_2Ph), \\ 3.81 \ (4), \ 368 \ (24), \ 281 \ (4), \ 368 \ (24), \ 281 \ (4), \ 381 \ (4), \ 368 \ (24), \ 281 \ (15) \ 381 \ (4), \ 368 \ (24), \ 281 \ (15) \ 339 \ (7$		7.09-7.43 (m, 4H, C ₆ H ₄ CF ₃), 8.42 (s, 1H, H-2)	122.1 (CF ₃), 131.4 (C-1'),	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$			131.9 (C-3'), 151.0 (C-5')	
$ \begin{array}{c} 4.01 \ (t, 4H, J=4.5 Hz, NCH_2), \\ 8.50 \ (s, 1H, H-2) \\ 9i \\ 0.89 \ (d, 2H, J=6.2 Hz, CHCH_3), \\ 1.16-1.31 \ (m, 2H, CHCH_2), \\ 1.57-1.75 \ (m, 3H, CHCH_2 + CHCH_2), \\ 3.01 \ (dt, 2H, J=2.3 Hz, J=12.4 Hz, NCH_2), \\ 4.63 \ (br \ s, 2H, NCH_2), \\ 8.51 \ (s, 3H, NCH_3), 2.60 \ (t, 4H, J=4.8 Hz, NCH_2), \\ 4.01 \ (t, 4H, J=4.8 Hz, NCH_2), \\ 9i \\ 2.53 \ (s, 3H, COCH_3), 3.55 \ (t, 3H, J=4.8 Hz, NCH_2), \\ 4.13 \ (t, 3H, J=5.3 Hz, NCH_2), 6.89, \\ 7.90 \ (AB \ system, 4H, J=8.9 Hz, C_{6H_4}), 8.50 \ (s, 1H, H-2) \\ 7.90 \ (AB \ system, 4H, J=8.9 Hz, C_{6H_4}), 8.50 \ (s, 1H, H-2) \\ 132 \ 2.61 \ (s, 4H, NCH_2), 3.58 \ (s, 2H, CH_2Ph), \\ 3.90 \ (s, 4H, NCH_2), 3.58 \ (s, 2H, CH_2Ph), \\ 3.90 \ (s, 4H, NCH_2) \\ 2.1a \ 3.81-3.88 \ (m, 16H, CH_2) \\ 2.74 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 4.77 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 4.77 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 4.77 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.90 \ (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 \ (m, 8H, CH), \\ 3.91 \ (t, 3H) \ (t$	9i	3.11 (t, 4H, $J = 4.2$ Hz, NCH ₂),		424 [(MH) ⁺ , 100],
		$4.01 (t, 4H, J = 4.5 Hz, NCH_2),$		381 (25), 367 (14),
$ \begin{array}{llllllllllllllllllllllllllllllllllll$		8.50 (s, 1H, H-2)		355 (11)
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	9j	$0.89 (d, 2H, J = 6.2 Hz, CHCH_3),$	21.7 (CHCH ₃),	437 [(MH) ⁺ , 100),
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		1.16–1.31 (m, 2H, CHCH ₂),	31.3 (CH ₃ CH),	435 (10), 367 (9)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		1.57–1.75 (m, 3H, CHCH ₂ +CHCH ₂),	34.2 (CH ₂ CH),	
$\begin{array}{c} 4.63 \ (br\ s,\ 2H,\ NCH_2),\ 8.37 \ (s,\ 1H,\ H-2) \\ 9k \\ 2.37 \ (s,\ 3H,\ NCH_3),\ 2.60 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2), \\ 4.01 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=4.8\ Hz,\ NCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=5.0\ Hz,\ SCH_2),\ 8.51 \ (s,\ 1H,\ H-2) \\ 4.01 \ (t,\ 4H,\ J=5.0\ Hz,\ SCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ SCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ SCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ NCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ NCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ NCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ NCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 4.77 \ (t,\ 4H,\ J=5.0\ Hz,\ NCH_2),\ 3.78-3.90 \ (m,\ 8H,\ CH), \\ 3.80 \ (s) \ 3.81 \ (s) \ 3.72 \ (30), \\ 3.89 \ (7) \ 3.81 \ (s) \ 3.81 \ (s) \ 3.72 \ (30), \\ 3.89 \ (7) \ 3.81 \ (s) \ 3.81 \ (s) \ 3.72 \ (30), \\ 3.89 \ (7) \ 3.81 \ (s) \ 3.72 \ (30), \\ 3.89 \ (7) \ 3.81 \ (s) \ 3.72 \ (30), \\ 3.89 \ (7) \ 3.81 \ (s) \ 3.81 \ (s$		$3.01 (dt, 2H, J=2.3 Hz, J=12.4 Hz, NCH_2),$	46.9 (NCH ₂),	
9k $2.37 (s, 3H, NCH_3), 2.60 (t, 4H, J=4.8 Hz, NCH_2), 4.01 (t, 4H, J=4.8 Hz, NCH_2), 8.51 (s, 1H, H-2)46.0 (NCH_3), 438 [(MH)^+, 100], 438 [(MH)^+, 100], 413 (t, 4H, J=4.8 Hz, NCH_2), 46.2 (NCH_2), 153.8 (C-2)9l2.53 (s, 3H, COCH_3), 3.55 (t, 3H, J=4.8 Hz, NCH_2), 4.13 (t, 3H, J=5.3 Hz, NCH_2), 6.89, 7.90 (AB system, 4H, J=8.9 Hz, C_6H_4), 8.50 (s, 1H, H-2)26.1 (COCH_3), 45.6 (47.0 (NCH_2)), 486 (19), 367 (19), 486 (19), 367 (19), 7.90 (AB system, 4H, J=8.9 Hz, C_6H_4), 8.50 (s, 1H, H-2)132.61 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), 3.90 (s, 4H, NCH_2)46.2 (NCH_2), 52.9 (NCH_2), 609 (18), 518 (4), 62.8 (C_6H_5CH_2)143.81-3.88 (m, 16H, CH_2)46.4 (NCH_2), 66.7 (CH_2O)425 (M^+, 41), 381 (4), 368 (24), 281 (15)21b2.74 (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 (m, 8H, CH), 4.77 (t, 4H, J=5.0 Hz, NCH)26.8 (SCH_2), 49.1 (NCH_2)441 (M^+, 57), 368 (100), 372 (30), 339 (7)$		4.63 (br s, 2H, NCH ₂), 8.37 (s, 1H, H-2)	153.9 (C-2)	
4.01 (t, 4H, $J = 4.8$ Hz, NCH ₂), 8.51 (s, 1H, H-2) 46.2 (NCH ₂), 481 (59), 367 (23) 54.9 (NCH ₂), 153.8 (C-2) 91 2.53 (s, 3H, COCH ₃), 3.55 (t, 3H, $J = 4.8$ Hz, NCH ₂), 4.13 (t, 3H, $J = 5.3$ Hz, NCH ₂), 6.89, 45.6, 47.0 (NCH ₂), 486 (19), 367 (19) 7.90 (AB system, 4H, $J = 8.9$ Hz, C ₆ H ₄), 8.50 (s, 1H, H-2) 13 2.61 (s, 4H, NCH ₂), 3.58 (s, 2H, CH ₂ Ph), 46.2 (NCH ₂), 52.9 (NCH ₂), 609 (18), 518 (4), 3.90 (s, 4H, NCH ₂) 21a 3.81–3.88 (m, 16H, CH ₂) 46.4 (NCH ₂), 66.7 (CH ₂ O) 21b 2.74 (t, 4H, $J = 5.0$ Hz, SCH ₂), 3.78–3.90 (m, 8H, CH), 4.77 (t, 4H, $J = 5.0$ Hz, NCH) 46.8 (SCH ₂), 49.1 (NCH ₂) 39 (S)	9k	2.37 (s, 3H, NCH ₃), 2.60 (t, 4H, J=4.8 Hz, NCH ₂),	46.0 (NCH ₃),	438 [(MH) ⁺ , 100],
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		4.01 (t, 4H, J=4.8 Hz, NCH ₂), 8.51 (s, 1H, H-2)	46.2 (NCH ₂),	481 (59), 367 (23)
91 $2.53 (s, 3H, COCH_3), 3.55 (t, 3H, J=4.8 Hz, NCH_2), 4.13 (t, 3H, J=5.3 Hz, NCH_2), 6.89, 7.90 (AB system, 4H, J=8.9 Hz, C_6H_4), 8.50 (s, 1H, H-2)26.1 (COCH_3), 45.6, 47.0 (NCH_2), 486 (19), 367 (19)132.61 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph), 3.90 (s, 4H, NCH_2)45.2 (NCH_2), 52.9 (NCH_2), 609 (18), 518 (4), 62.8 (C_6H_5CH_2)143.81-3.88 (m, 16H, CH_2)46.4 (NCH_2), 66.7 (CH_2O)425 (M^+, 41), 381 (4), 368 (24), 281 (15)21b2.74 (t, 4H, J=5.0 Hz, SCH_2), 3.78-3.90 (m, 8H, CH), 4.77 (t, 4H, J=5.0 Hz, NCH)26.8 (SCH_2), 49.1 (NCH_2)441 (M^+, 57), 368 (100), 372 (30), 339 (7)$			54.9 (NCH ₂),	
912.53 (s, 3H, COCH ₃), 3.55 (t, 3H, $J=4.8$ Hz, NCH ₂), 4.13 (t, 3H, $J=5.3$ Hz, NCH ₂), 6.89, 7.90 (AB system, 4H, $J=8.9$ Hz, C ₆ H ₄), 8.50 (s, 1H, H-2)26.1 (COCH ₃), 45.6, 47.0 (NCH ₂), 153.9 (C-2), 113.3, 114.2, 129.9, 130.4 (C ₆ H ₄), 196.4 (CO)542 [(MH) ⁺ , 40], 486 (19), 367 (19)132.61 (s, 4H, NCH ₂), 3.58 (s, 2H, CH ₂ Ph), 3.90 (s, 4H, NCH ₂)45.6 (C ₆ H ₂), 52.9 (NCH ₂), 62.8 (C ₆ H ₅ CH ₂)609 (18), 518 (4), 481 (9), 146 (11), 91 (100)21a3.81–3.88 (m, 16H, CH ₂)46.4 (NCH ₂), 66.7 (CH ₂ O)425 (M ⁺ , 41), 381 (4), 368 (24), 281 (15)21b2.74 (t, 4H, $J=5.0$ Hz, SCH ₂), 3.78–3.90 (m, 8H, CH), 4.77 (t, 4H, $J=5.0$ Hz, NCH)26.8 (SCH ₂), 49.1 (NCH ₂)441 (M ⁺ , 57), 368 (100), 372 (30), 339 (7)			153.8 (C-2)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	91	2.53 (s, 3H, COCH ₃), 3.55 (t, 3H, $J = 4.8$ Hz, NCH ₂),	26.1 (COCH ₃),	542 [(MH) ⁺ , 40],
7.90 (AB system, 4H, $J = 8.9$ Hz, C_6H_4), 8.50 (s, 1H, H-2) 153.9 (C-2), 113.3, 114.2, 129.9, 130.4 (C_6H_4), 196.4 (CO) 13 2.61 (s, 4H, NCH ₂), 3.58 (s, 2H, CH ₂ Ph), 3.90 (s, 4H, NCH ₂) 46.2 (NCH ₂), 52.9 (NCH ₂), 609 (18), 518 (4), 62.8 ($C_6H_5CH_2$) 21a 3.81–3.88 (m, 16H, CH ₂) 46.4 (NCH ₂), 66.7 (CH ₂ O) 425 (M ⁺ , 41), 381 (4), 368 (24), 281 (15) 21b 2.74 (t, 4H, $J = 5.0$ Hz, SCH ₂), 3.78–3.90 (m, 8H, CH), 4.77 (t, 4H, $J = 5.0$ Hz, NCH) 26.8 (SCH ₂), 49.1 (NCH ₂) 441 (M ⁺ , 57), 368 (100), 372 (30), 339 (7)		4.13 (t, 3H, $J = 5.3$ Hz, NCH ₂), 6.89,	45.6, 47.0 (NCH ₂),	486 (19), 367 (19)
$\begin{array}{c} 114.2, 129.9, 130.4 (C_{6}H_{4}), \\ 196.4 (CO) \\ 13 & 2.61 (s, 4H, NCH_{2}), 3.58 (s, 2H, CH_{2}Ph), \\ 3.90 (s, 4H, NCH_{2}) \\ 21a & 3.81-3.88 (m, 16H, CH_{2}) \\ 21b & 2.74 (t, 4H, J = 5.0 \text{Hz}, SCH_{2}), 3.78-3.90 (m, 8H, CH), \\ 4.77 (t, 4H, J = 5.0 \text{Hz}, NCH) \\ \end{array}$		7.90 (AB system, 4H, $J = 8.9$ Hz, C ₆ H ₄), 8.50 (s, 1H, H-2)	153.9 (C-2), 113.3,	
13 $2.61 (s, 4H, NCH_2), 3.58 (s, 2H, CH_2Ph),$ $3.90 (s, 4H, NCH_2)$ $196.4 (CO)$ $46.2 (NCH_2), 52.9 (NCH_2),$ $62.8 (C6H5CH2)609 (18), 518 (4),481 (9), 146 (11),91 (100)21a3.81-3.88 (m, 16H, CH_2)46.4 (NCH_2), 66.7 (CH_2O)425 (M^+, 41),381 (4), 368 (24),281 (15)21b2.74 (t, 4H, J = 5.0 Hz, SCH_2), 3.78-3.90 (m, 8H, CH),4.77 (t, 4H, J = 5.0 Hz, NCH)26.8 (SCH_2), 49.1 (NCH_2)441 (M^+, 57),368 (100), 372 (30),339 (7)$			114.2, 129.9, 130.4 (C ₆ H ₄),	
13 2.61 (s, 4H, NCH ₂), 3.58 (s, 2H, CH ₂ Ph), 3.90 (s, 4H, NCH ₂) 46.2 (NCH ₂), 52.9 (NCH ₂), 62.8 (C ₆ H ₃ CH ₂) 609 (18), 518 (4), 481 (9), 146 (11), 91 (100) 21a 3.81–3.88 (m, 16H, CH ₂) 46.4 (NCH ₂), 66.7 (CH ₂ O) 425 (M ⁺ , 41), 381 (4), 368 (24), 281 (15) 21b 2.74 (t, 4H, J = 5.0 Hz, SCH ₂), 3.78–3.90 (m, 8H, CH), 4.77 (t, 4H, J = 5.0 Hz, NCH) 26.8 (SCH ₂), 49.1 (NCH ₂) 441 (M ⁺ , 57), 368 (100), 372 (30), 339 (7)			196.4 (CO)	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	13	2.61 (s, 4H, NCH ₂), 3.58 (s, 2H, CH ₂ Ph),	46.2 (NCH ₂), 52.9 (NCH ₂),	609 (18), 518 (4),
21a $3.81-3.88 \text{ (m, 16H, CH}_2)$ 46.4 (NCH2), 66.7 (CH2O) 91 (100) 21b $2.74 \text{ (t, 4H, } J = 5.0 \text{ Hz, SCH}_2), 3.78-3.90 \text{ (m, 8H, CH)}, 4.77 \text{ (t, 4H, } J = 5.0 \text{ Hz, NCH}$ 26.8 (SCH2), 49.1 (NCH2) 91 (100) 411 (M ⁺ , 57), 368 (100), 372 (30), 339 (7)		3.90 (s, 4H, NCH ₂)	$62.8 (C_6H_5CH_2)$	481 (9), 146 (11),
21a $3.81-3.88 \text{ (m, 16H, CH}_2\text{)}$ $46.4 \text{ (NCH}_2\text{)}, 66.7 \text{ (CH}_2\text{O}\text{)}$ $425 \text{ (M}^+, 41),$ 21b $2.74 \text{ (t, 4H, } J = 5.0 \text{ Hz}, \text{ SCH}_2\text{)}, 3.78-3.90 \text{ (m, 8H, CH)},$ $26.8 \text{ (SCH}_2\text{)}, 49.1 \text{ (NCH}_2\text{)}$ $441 \text{ (M}^+, 57),$ $4.77 \text{ (t, 4H, } J = 5.0 \text{ Hz}, \text{ NCH}\text{)}$ $26.8 \text{ (SCH}_2\text{)}, 49.1 \text{ (NCH}_2\text{)}$ $368 \text{ (100)}, 372 \text{ (30)},$				91 (100)
21b $2.74 (t, 4H, J = 5.0 Hz, SCH_2), 3.78-3.90 (m, 8H, CH), 4.77 (t, 4H, J = 5.0 Hz, NCH)26.8 (SCH_2), 49.1 (NCH_2)381 (4), 368 (24), 281 (15)441 (M+, 57), 368 (100), 372 (30), 339 (7)$	21a	3.81–3.88 (m, 16H, CH ₂)	46.4 (NCH ₂), 66.7 (CH ₂ O)	425 (M ⁺ , 41),
21b $2.74 (t, 4H, J = 5.0 Hz, SCH_2), 3.78-3.90 (m, 8H, CH),$ $26.8 (SCH_2), 49.1 (NCH_2)$ $281 (15)$ $4.77 (t, 4H, J = 5.0 Hz, NCH)$ $368 (100), 372 (30),$ $339 (7)$				381 (4), 368 (24),
21b 2.74 (t, 4H, J=5.0 Hz, SCH ₂), 3.78–3.90 (m, 8H, CH), 26.8 (SCH ₂), 49.1 (NCH ₂) 441 (M ⁺ , 57), 4.77 (t, 4H, J=5.0 Hz, NCH) 368 (100), 372 (30), 339 (7)				281 (15)
4.77 (t, 4H, <i>J</i> =5.0 Hz, NCH) 368 (100), 372 (30), 339 (7)	21b	2.74 (t, 4H, J=5.0 Hz, SCH ₂), 3.78–3.90 (m, 8H, CH),	26.8 (SCH ₂), 49.1 (NCH ₂)	441 (M ⁺ , 57),
339 (7)		4.77 (t, 4H, J = 5.0 Hz, NCH)		368 (100), 372 (30),
				339 (7)

^aThe NMR spectra of 4d and 9c were obtained in DMSO- d_6 .

^dElectron impact MS data (EIMS) is shown for compounds **3a**, **4a**, **4b**, **4f**, **4l**, **9a–c**, **13**, **21a**, and **21b**. The IR spectra **21a–b** shows an absorption for the nitrile group at 2900 and 2220 cm⁻¹ (CN), while compounds **3a**, **4a–h**, **9a–h**, and **13** showed the CN band at 2210–2220 cm⁻¹.

^bThe ¹H NMR spectra of compounds **3a**, **4a–h**, **9a–h**, and **13** exhibited additional signals typical for C_6H_5 [7.40, 7.50–7.46, 7.58 (m, 5H)], and OCH₂CH₃ groups [1.41–1.54 (t, 3H, J=7.1 Hz), and 4.57–4.66 (q, 2H, J=7.1 Hz)]. Compounds **4i–l** and **9i–l** present also resonances for C_6H_5 [7.42, 7.49–7.59, 7.69 (m, 8H), and 8.02, 8.17–8.08, 8.22 (m, 2H)] and for H-8 [7.65–7.78 (s, 1H)]. Compounds **21a–b** exhibit signals for NMe₂ [3.22 (s, 6H)] and for H-9 [8.56–8.60 (s, 1H)].

^cThe ¹³C NMR spectra of **3a**, **4a–h**, **9a–h**, and **13** (compounds **4d** and **9i** are not sufficiently soluble in DMSO- d_6 or CDCl₃ to produce good ¹³C NMR data, showed signals at 14.2–14.4 (CH₃), 64.0–64.6 (CH₂O), 95.1–96.7 (C-8), 100.5–112.3 (C-4a), 114.5–115.0 (CN), 118.5–119.4 (C-9a), 127.7, 128.0–133.4, 134.4 (four peaks for C₆H₅), 154.9–155.7 (C-5a), 155.5–157.4 (C-9b), 157.3–158.6 (C-4), and 163.2–164.6 (two renonances for C-7 and C-9). Compounds **3a**, **4a–c**, **4e–h** show a resonance for C-2 at 160.2 ppm. In the **9a–h** series C-2 resonates in the range 154.1–154.5 ppm. Compounds **4i–l** and **9i–l** showed signals at 102.7 (C-4a) (**4i–l**) and at 114.0 (C-4a) (**9j–l**), 118.9–119.2 (C-8), 122.8–123.4 (C-9a), 127.3–138.6 (seven absorptions for two C₆H₅ groups), 149.0–149.7 (C-5a), 155.8–157.6 (C-9b), 157.4–158.2 (C-4), 158.1–158.8, and 162.7–163.4 (C-7 and C-9). Compounds **4i–l** showed one resonance at 160.1–160.4 for C-2. Compounds **21a–b** present resonances at 37.1 (NMe), 48.8 (NCH₂), 66.6 (CH₂O), 92.8 (C-8), 100.4 (C-4a), 118.2 (CN), 120.4 (C-9a), 139.5 (C-9), 156.6 (C-5a), 157.6–158.4 (C-9b), 160.4 (C-2), 160.8 (C-7), 165.0 (C-4).

Compound 13 was obtained from 5a by the sequence shown in Scheme 3. 5a was reacted with *o*-nitrobenzaldehyde in refluxing toluene containing a catalytic amount of *p*-toluensulfonic acid, with azeotropic removal of water, to give 10 in 81% yield. Refluxing compound 10 with DDQ in THF for a few hours afforded pyridothienopyrimidone 11, [EIMS m/z=427(M⁺-OCN) and 292 (M⁺-OCNRHC)]. Reaction of 10 with POCl₃ yielded the expected 4-chloropyridothienopyrimidine derivative 12, which gave 13 on reaction with 4-benzylpiperazine. The ¹H NMR spectrum of pyridothienopyrimidine 10 showed signals at $\delta=6.25$ for H-2, and two other signals at $\delta=5.13$ and $\delta=8.26$, which could be exchanged with deuterium oxide, assigned to the N-bonded protons at positions 1 and 3, respectively. The ¹³C NMR spectrum of **10** showed one signal at $\delta = 63.2$ corresponding to the carbon at position 2 of the newly formed pyrimidine ring. The EIMS of pyridothienopyrimidone **11** showed the expected molecular ion and other fragments at m/z = 427 (M⁺-OCN) and 292 (M⁺-OCNRHC).

For the preparation of pyridothienopyrimidines **21a–b** we started from the aminocyanopyridine **14**.¹⁷ This compound was first converted into the thienopyridine **19** and then cyclized to give the pyridothienopyrimidines using the phosgene iminium chloride method discussed above. The whole synthetic process is represented in Scheme 4. All compounds gave elemental analyses and spectra consistent with the assigned structures.



Scheme 2.



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Biological Results

Immunological stimulus

Initial information about the ability of the pyridothienopyrimidines to inhibit the release of histamine from rat mast cells was obtained by screening the activity using a single dose $(100 \,\mu\text{g/mL})$ of compound. In the experiments performed with preincubation (compound added to the cells 10 min before than the stimulus, Fig. 1), the inhibitory action of **4k**, **9i**, **9k**, **9l**, and **21a** is of around 40%, compound **21a** being the most active showing a 60% inhibitory activity.

When the experiments were carried out with simultaneous addition of the compound and the stimulus (i.e. without preincubation, Figure 2), the inhibitory pattern



Figure 1. Histamine release in mast cells stimulated with 3.5 mg/mL of antigen (ovoalbumin) in the presence of pyridothieno-pyrimidines. The compounds were added to the cells 10 min before the addition of the stimulus (10 min preincubation). The activity data are normalized to the response of the control experiments (29.63 ± 1.59% in the presence of 5 mg/mL of antigen).

is very similar to that in the previous figure, with a consistent inhibition of around 40% for several compounds. Nevertheless, **4h**, **4i**, **9a**, and **21a** show a higher inhibitory activity (60%) and values of 90% inhibition are obtained with **9b** and **9l**. From those experiments, pyridothienopyrimidines **9l** and **21a** emerge as especially interesting since they show consistent inhibition of the immunological stimulus and a low desensitization.

In contrast to the inhibitory activity found in those heterocycles, two compounds, 9d (40–60%) and 4l (170–220%) have been shown to be activators of the exocytosis in rat mast cells.

Non-immunological stimulus

The pyridothienopyrimidines were also tested when the release of histamine was induced by the chemical inducer compound 48/80.¹⁸

The results of single dose experiments (at $100 \mu g/mL$) are presented in Figures 3 and 4 and show that compounds **4c**, **4i**, **4k**, and **9l** produce a 50–60% inhibition when preincubated with the stimulus while in the experiments carried out with simultaneous addition only four compounds (**4c**, **4e**, **4k**, and **9l**) show some inhibitory action (20–30%). Overall, **4c**, **4k**, and **9l** are active in both conditions **9l** being the most effective inhibitor with inhibition values of 30% (simultaneous addition) and 60% (preincubated), respectively. Figure 3 indicates the presence of several histamine releasers too. The most

active one in the experiments with preincubation is 9e (100%). In the assays performed with simultaneous addition of the stimulus (Fig. 4) there is only one good inducer in the whole series and this is again 9e (170%), which therefore is a good releaser both with and without preincubation.

The activity of the pyridothienopyrimidines was also tested using the antineoplastic drugs adryamicin and vinorelbine, recently reported to induce histamine release,¹⁹ as chemical inducers. The results (not shown) obtained with a selection of pyridothienopyrimidines (4b, 9a, 9c, 9g, 9h, and 13) indicated that the amount of histamine released after simultaneous addition of adryamicin was practically the same as in the absence of the chemical. Experiments carried out with preincubation of the cells with the pyridothienopyrimidines before addition of the antineoplastic drug, showed a small increase in the inhibition values that reached up to 30% in some cases. Similar tests with vinorelbine as histamine releaser did not show any clear inhibitory action either with or without preincubation. In summary, no clear correlation was observed between the results of the experiments using adrvamicin, vinorelbine and compound 48/80 as chemical releasers of histamine.

In order to get a better picture of the ability of these heterocycles to inhibit the release of histamine, and to allow a direct comparison of their potency with that of known antiallergics as cromoglycate (DSCG) and ketotifen,²⁰ experiments using five concentrations of inhibitor were carried out. Table 3 shows the results



Figure 2. Histamine release in mast rat cells stimulated with 3.5 mg/mL of antigen (ovoalbumin) in the presence of pyridothienopyrimidines. The compounds were added to the cells simultaneously with the stimulus (no preincubation). The activity data are normalized to the response of the control experiments ($21.47 \pm 2.4\%$ in the presence of 5 mg/mL of antigen).

expressed as IC₅₀ values, for a selection of pyridothienopyrimidines. The figures are in the range $2-25 \,\mu$ M well below those of cromoglycate (DSCG) and similar to ketotifen in the immunologically stimulated assays. In the chemical stimulated experiments, the selected pyridothienopyrimidines are clearly much more active than both references. These results prove that these heterocycles are up to 10–100 times more active than the standards and confirm their potential interest.

Antitumor activity

The pyridothienopyrimidines shown in Table 1 have been tested in vitro for their antitumor activity against four standard tumor cell lines. The IC₅₀ figures presented in Table 4 show that four pyridothienopyrimidines can be considered to be active in these assays. These are **4d** and **9d** (IC₅₀: 2.5–0.5 µg/mL) and **9i** and **4l**, which show IC₅₀ values of 0.12–0.25 µg/mL. Structurally,



Figure 3. Histamine release in mast rat cells stimulated with $2 \mu g/mL$ of compound 48/80 in the presence of pyridothienopyrimidines. The compounds were added to the cells simultaneously with the stimulus (no preincubation). The activity data are normalized to the response of the control experiments ($32.0 \pm 4.7\%$ in the presence of $2 \mu g/mL$ of compound 48/80.



Figure 4. Histamine release in mast rat cells stimulated with $2 \mu g/mL$ of compound 48/80 in the presence of pyridothienopyrimidines. The compounds were added to the cells 10 min before the addition of the stimulus (10 min preincubation). The activity data are normalized to the response of the control experiments ($35.0 \pm 3.5\%$ in the presence of $2 \mu g/mL$ of compound 48/80).

Immunological s (5 mg/mL)	timulation	Chemical s (2 µg	stimulation /mL)	Immunologica (5 mg/	l stimulation mL)	Chemical st (2 µg/1	imulation mL)
Compd (Preinc.)	IC ₅₀ (µM)	Compd (Preinc.)	IC ₅₀ (µM)	Compd (No preinc.)	IC ₅₀ (µM)	Compd (No preinc.)	IC ₅₀ (µM)
DSCG ^a	400	DSCG ^a	200	4h	4 ± 0.73	4c	6 ± 1.1
Ketotifen ^a	10	Ketotifen ^a	100	4i	6 ± 1.2	4 e	2 ± 0.4
4k	10 ± 1.2	4c	10 ± 1.0	9a	20 ± 0.8	4k	5 ± 1.0
9i	25 ± 2.5	4i	20 ± 0.9	9b	25 ± 2.4	91	15 ± 1.7
9k	14 ± 1.3	4k	14 ± 1.3	91	9 ± 1.3		
91	20 ± 1.7	91	25 ± 1.7	21a	20 ± 3.2		
21 a	25 ± 3.5						

Table 3. Histamine release inhibition (IC₅₀ μ M) of selected pyridothienopyrimidines

^aSee ref. 20.

the last two compounds have significantly different combinations of R_1 , R_2 , and R_3 but have in common the piperazine substituent (R), suggesting that this unit plays an important role in their action. It is interesting to note that **41** and **9d** are the stronger stimulants for the liberation of histamine found in the series of pyridothienopyrimidines studied.

Conclusions

In conclusion, we have presented efficient synthetic procedures for the preparation of pyridothienopyrimidines of interest as antiallergics. Their activity has been evaluated with rat mast cells and the assays have been carried out under different conditions and with different types of stimulus. Under immunological stimulus with ovoalbumin (Figs 1 and 2, Table 3), several pyridothienopyrimidines are stronger inhibitors of the release of histamine than DSCG (up to 100 times more potent) and of the same order of potency as Ketotifen,²⁰ although only two compounds, 91 and 21a are active with and without preincubation. In the assays using compound 48/80 as chemical inducer (Figs 3 and 4, Table 3), there are also several inhibitors, 10-100 times more potent than DSCG and Ketotifen.²⁰ Three of them, 4c, 4k, and 9l, are active with and without preincubation.

Table 4. Antitumoral activity of selected pyridothienopirimidines expressed in IC_{50}

Compd	IC ₅₀ (µg/mL)					
	P-388	A-549	HT-29	MEL		
4d	2.5	2.5	2.5	2.5		
41	0.25	0.25	0.25	0.50		
9d	0.5	1	2	1		
9i	0.12	0.25	0.25	0.25		

Overall, the data shown in Table 3 demonstrates that pyridothienopyrimidines are worth considering for their potential antiallergic activity. Compound **91** is particularly interesting because it is the only one that conserves its strong inhibitory activity ($IC_{50}=9-25 \,\mu M$) in all the conditions tested (chemical and immunological stimulus, with/without preincubation).

From the structure–activity point of view, our results indicate that the modification of the substituents on the pyridothienopyrimidine skeleton is not as effective in improving the inhibitory action as similar changes in the cyanopyridines and pyridopyrimidine series, ^{12a} and this may explain why so many pyridothienopyrimidines show some inhibition. Nevertheless, some trends can be distinguished and for example, comparison of the inhibitors **9b** and **9l** with the inactive **9g** and **9j** may illustrate the influence of R_1 and R_2 . Similarly, the good inhibitory action of **9a** is lost in its *N*-dimethyl derivative **4b**.

The fact that some inhibitory action is observed in virtually all cases examined (Figs 1–4), the absence of any apparent link between the chemical and the immunologically stimulated experiments, prevents the formulation of any further conclusion. Similarly, comparison of the results obtained with compound $48/80^{18}$ and the drugs adryamicin and vinorelbine indicate that no apparent correspondence exists between those three chemical releasers.

A completely different picture is seen if our attention is focussed on the heterocycles acting as inducers of the release of histamine. This action is found to be very selective among the compounds studied. Indeed, only one pyridothienopyrimidine (**4**I) acts as a releaser of histamine under immunological conditions (170-220%), and another one (**9e**, 100-170%) in the chemically driven experiments. This effect is observed in both pre-

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incubated and non-preincubated experiments, because, as expected, is independent of the stimulus itself. Both structures share the same R substituent (R = piperidine) but comparison with other members of the series, and particularly with the inactive compounds 4d and 9i, suggests that the combination of substituents represented by 4l is necessary to obtain the maximum activity.

The discovery of strong histamine releasers among the compounds investigated is interesting because, although inducers have no clinical interest so far, knowledge of their structures may provide an aid to the prediction²¹ of unwanted side effects.

Some pyridothienopyrimidines¹¹ and pyridopyrimidines²² have been reported to be antineoplastic. In our tests, only four compounds were found to be active, so cytotoxicity would not pose a problem in the eventual use of these compounds as antiallergics. However, it is worth noting that two of the stronger stimulants, **41** and **9d**, are among the more cytotoxic pyridothienopyrimidines.

Experimental

Chemistry

All reagents used were commercial grade chemicals from freshly opened containers. The histamine releaser compound 48/80, is a condensation product of Nmethyl-p-methoxy-phenethylamine, was obtained from Sigma Chemical Co. Melting points were determined using a Büchi 510 apparatus and are uncorrected. IR spectra were recorded as potassium bromide disks using a Perkin-Elmer 783 spectrophotometer. ¹H and ¹³C NMR spectra were obtained using a Bruker AC 200F instrument at room temperature. Mass spectra were obtained using a VG QUATTRO spectrometer. The silica gel 60 HF254+366 used for analytical thin-layer chromatography and the silica gel 60 (230-400 mesh) employed for flash chromatography were purchased from Merck. Microanalyses for C, H, and N, were performed by the Servicio General de Analisis Elemental at the University of La Coruña.

Histamine release and neoplastic assays^{12b-d}

Rat mast cells and human and mouse tumoral cells were prepared and incubated as previously described.^{12b} Histamine release was measured spectrofluorometrically^{12c} both in the pellet and in the supernatants with and without preincubation (10 min) of each drug (100 μ g/mL in the single dose experiments) with the cells before addition of the stimulus. All the experiments were repeated at least three times in duplicate and the data was analyzed using the Student's *t*-test for umpaired data. A probability level of 0.05 or less was used for statistical significance and the results are expressed as mean \pm SEM. Five concentrations of inhibitor (0.1, 1, 10, 100, and 500 µg/mL) were used in order to obtain the IC₅₀ values.

The antineoplasic activity was measured^{12d} as IC_{50} against P-388 (ATCC CCL 46; suspension culture of a lymphoid neoplasm from a DBA/2 mouse), A-549 (ATCC CCL 185; monolayer culture of a human lung carcinoma), HT-29 (ATCC HTB-38; monolayer culture of a human colon carcinoma), and MEL-28 (ATCC HTB-72; monolayer culture of a human melanoma).

3-Chlorodimethylaminomethylenamino-2-cyanothieno[2,3*b*]-pyridine: General procedure (2a,b). A solution of 1a or 1b (1.3 mmol) and phosgene iminium salt (0.26 g, 1.6 mmol) in 1,2-dichloroethane (20 mL) was refluxed for 45 min. The solvent was removed under reduced pressure and the resulting solid was purified by flash chromatography using $CH_2Cl_2/hexane$ (2/1) as eluent.

3-Chlorodimethylaminomethylenamino-2,5-dicyano-6-ethoxy-4-phenylthieno[**2,3-***b*]pyridine (2a). Yield 78%; mp 220–222 °C; C₂₀H₁₆N₅OCIS calcd C, 58.61; H, 3.93; N, 17.09. found: C, 58.67; H, 4.07; N, 17.17; ¹H NMR (CDCl₃) δ 1.50 (t, 3H, *J*=7.1 Hz, CH₃), 2.70 (s, 6H, NMe₂), 4.61 (q, 2H, *J*=7.1 Hz, CH₂), 7.27–7.55 (m, 5H, C₆H₅); ¹³C NMR (CDCl₃) δ 14.4 (CH₃), 39.6 (NMe₂), 64.4 (CH₂), 90.9 (C-2), 96.6 (C-5), 113.8 (CN), 113.9 (CN), 119.2 (C-3a), 127.5, 128.4, 128.9, 133.6 (C₆H₅), 139.8, 151.1, 154.2, 162.1, 162.6; MS (EI, *m/z*, %): 411 (M⁺ + 2, 33), 409 (M⁺, 85), 374 (100), 346 (40), 289 (31); IR (KBr, cm⁻¹): 2220 (CN), 2200 (CN), 1640, 1550, 1430.

3-Chlorodimethylaminomethylenamino-2-cyano-4,6-phenylthieno[2,3-*b***]pyridine (2b). Yield 95%; mp 153–155°C; C_{23}H_{17}N_4ClS calcd C, 66.26; H, 4.11; N, 13.44. found: C, 66.07; H, 4.01; N, 13.58; ¹H NMR (CDCl₃) \delta 2.76 (s, 6H, NMe₂), 7.34–7.55 (m, 8H, C₆H₅), 7.65 (s, 1H, H-5), 8.09–8.15 (m, 2H, C₆H₅); ¹³C NMR (CDCl₃) \delta 39.7 (NMe₂), 93.3 (C-2), 114.5 (CN), 119.2 (C-5), 123.0 (C-3a), 127.3, 127.4, 127.9, 128.9, 129.1, 129.8, 137.8, 138.0 (C₆H₅), 140.0 (C–Cl), 148.3 (C-3), 150.9, 157.3, 161.6 (C-4, C-6, C-7a); MS (EI,** *m/z***, %): 418 (M⁺ + 2, 37), 416 (M⁺, 94), 381 (100), 365 (20), 336 (28), 324 (46); IR (KBr, cm⁻¹): 2200 (CN), 1640, 1560, 1320.**

4-Chloro-2-dimethylaminopyrido[3', 2':4,5]**thieno**[3,2-*d*]-**pyrimidine: General procedure (3a,b).** A stream of dry hydrogen chloride was passed through a solution of **2a** or **2b** (0.43 mmol) in 1,2-dichloroethane (10 mL) for 3 h. After that time, the reaction mixture was allowed to stand overnight at room temperature, the solvent was

removed under reduced pressure and the resulting solid was purified by flash chromatography eluting with $CH_2Cl_2/Hexane$ (2/1) to yield **3a** (Tables 1 and 2) or **3b**.

4- Chloro-7,9-diphenylpyrido[3',2':4,5]thieno[3,2-*d*]pyrimidine (3b). Yield 83%; mp 237–239 °C; $C_{23}H_{17}N_4ClS$ calcd C, 66.26; H, 4.11; N, 13.44. found: C, 66.34; H, 4.19; N, 13.51; ¹H NMR (CDCl₃) δ 2.94 (s, 6H, NMe₂), 7.48–7.65 (m, 8H, C₆H₅), 7.75 (s, 1H, H-8), 8.16–8.21 (m, 2H, C₆H₅); ¹³C NMR (CDCl₃) δ 37.1 (NMe₂), 119.2 (C-8), 122.9 (C-9a), 127.5, 127.7, 128.4, 128.9, 129.6, 130.0, 137.4, 138.1 (C₆H₅), 150.2, 154.3, 158.3, 158.7, 160.3, 165.2; MS (EI, *m/z*, %): 418 (M⁺ + 2, 40), 416 (M⁺, 100), 401 (37), 387 (86), 372 (32); IR (KBr, cm⁻¹): 1590, 1530, 1480, 1410.

8-Cyano-4,7-diethoxy-2-dimethylaminopyrido[3',2':4,5]thieno[3,2-*d*]pyrimidine (4a). A solution of 3a (0.15 g, 0.37 mmol) in ethanol (20 mL) containing NaOEt (0.08 g of Na, 3.7 mmol) was refluxed for 1 h. The solid was filtered off and recrystallized from ethanol/acetone to yield 4a (Tables 1 and 2).

2-Dimethylaminopyrido[3',2':4,5]thieno[3,2-d]pyrimidine-**4-substituted (4b–l): General procedure.** A solution of **3a** or **3b** (0.37 mmol) and the appropriate amine (0.44 mmol) in ethanol (10 mL) was refluxed until the starting material had disappeared (TLC). The solid was filtered off and recrystallized or was purified by flash chromatography (Tables 1 and 2).

5-Cyano-6-ethoxy-3-dimethylaminomethylenamino-4-phenylthieno[2,3-b]pyridine-2-carboxamide (6). A solution containing 5a (1.0 g, 3.0 mmol) and DMFDMA (0.54 g, 4.5 mmol) in DMF (5 mL) was stirred at 80 °C for 15 min. Ethanol (20 mL) was added and the solid was filtered off and recrystallized from ethanol/acetone to yield 6 (1.03 g, 89%): mp 242 °C; C₂₀H₁₉N₅O₂S calcd C, 61.05; H, 4.87; N, 17.80. found: C, 60.89; H, 4.99; N, 17.74; ¹H NMR (CDCl₃) δ 1.47 (t, 3H, J=7.1 Hz, CH₃), 3.12 (s, 3H, NMe₂), 3.12 (s, 3H, NMe₂), 4.59 (q, 2H, $J = 7.1 \text{ Hz}, \text{ CH}_2$, 7.45–7.59 (m, 5H, C₆H₅), 8.53 (s, 1H, CH = N; ¹³C NMR (CDCl₃) δ 14.3 (CH₃), 35.3 (NMe₂), 41.3 (NMe₂), 63.9 (CH₂), 94.4 (C-5), 105.6 (C-2), 114.7 (CN), 117.6 (C-3a), 128.1, 129.1, 130.1, 133.2 (C_6H_5) , 146.1, 153.1, 158.9, 162.2, 163.7, 174.5 (CO); MS (EI, *m*/*z*, %): 393 (M⁺, 28), 349 (100), 321 (59), 294 (9); IR (KBr, cm⁻¹): 3460 (NH), 3300 (NH), 2220 (CN), 1630 (CO), 1570, 1330.

Pyrido[3',2':4,5]thieno[3,2-d]pyrimidin-4(3H)-ones (7a,b): General procedure. Method A: A solution of 6 (0.58 g, 1.48 mmol) and an excess of NaOEt in ethanol (25 mL) was refluxed for 30 min. The solvent was removed under reduced pressure and water (40 mL) was added. The solution was neutralized with 2 N HCl and the resulting solid was filtered off to yield **7a** (0.42 g, 82%). **Method B**: A solution of **5a** or **5b** (3.00 mmol) and a catalytic amount of PTSA in HC(OEt)₃ (15 mL) was refluxed until the starting material had disappeared (TLC). The solid was filtered off and the crude product was used in the next step without further purification.

8-Cyano-7-ethoxy-9-phenylpyrido[3',2':4,5]thieno[3,2-*d*]pyrimidin-4(3*H*)-one (7a). Yield 98%; mp > 300 °C; C₁₈H₁₂N₄O₂S calcd C, 62.06; H, 3.47; N, 16.08; found: C, 62.20; H, 3.55; N, 16.25; ¹H NMR (DMSO-*d*₆) δ 1.43 (t, 3H, *J* = 7.1 Hz, CH₃), 4.61 (q, 2H, *J* = 7.1 Hz, CH₂), 7.49–7.67 (m, 5H, C₆H₅), 8.03 (s, 1H, H-2), 12.89 (br s, 1H, exchangeable with D₂O, NH). MS (EI, *m*/*z*, %): 348 (M⁺, 73), 319 (100), 306 (19), 292 (23), 264 (20), 236 (32), 165 (31); IR (KBr, cm⁻¹): 3240 (NH), 2220 (CN), 1680 (CO), 1330, 1230.

7,9-Diphenylpyrido[**3**',**2**':**4,5**]**thieno**[**3**,**2**-*d*]**pyrimidin-4**(**3***H*)**-one (7b).** Yield 93%; mp > 300 °C; C₂₁H₁₃N₃OS calcd C, 70.97; H, 3.69; N, 11.82; found: C, 71.07; H, 3.57; N, 11.75; ¹H NMR (DMSO-*d*₆) δ 7.47–7.70 (m, 8H, C₆H₅), 7.98 (s, 1H, H-8), 8.11 (s, 1H, H-2), 8.26–8.30 (m, 2H, C₆H₅), 12.85 (br s, 1H, exchangeable with D₂O, NH); ¹³C NMR (DMSO-*d*₆) δ 119.6 (C-8), 122.7 (C-9a), 123.5 (C-4a), 127.4, 127.6, 128.7, 129.0, 130.0, 130.1, 136.7, 137.2 (C₆H₅), 146.5 (C-2), 149.4, 157.4, 162.8 (C-5a, C-7, C-9), 151.4 (C-9b), 156.7 (CO); MS (EI, *m/z*, %): 355 (M⁺, 100), 299 (23), 255 (16), 227 (16); IR (KBr, cm⁻¹): 1660 (CO), 1540, 1400.

4-Chloropyrido[3',2':4,5]thieno[3,2-d]pyrimidine (8a,b). A stirred solution of 7a or 7b (2.37 mmol) and phosphorous pentachloride (3.55 mmol) in phosphorous oxychloride (6 mL) was refluxed until starting material had disappeared (TLC). The solvent was removed under reduced pressure and the resulting solid was purified by flash chromatography using CH_2Cl_2 /hexanes as eluent (3:2 for 8a and 2:1 for 8b).

4-Chloro-8-cyano-7-ethoxy-9-phenylpyrido]*3*',*2*':**4**,**5**]thieno-[**3**,**2**-*d*]**pyrimidine** (**8a**). Yield 85%; mp 202–204°C; C₁₈H₁₁N₄OCIS calcd C, 58.94; H, 3.02; N, 15.27; found: C, 59.14; H, 3.20; N, 15.09; ¹H NMR (CDCl₃) δ 1.56 (t, 3H, *J* = 7.1 Hz, CH₃), 4.72 (q, 2H, *J* = 7.1 Hz, CH₂), 7.48–7.61 (m, 5H, C₆H₅), 8.77 (s, 1H, H-2); ¹³C NMR (CDCl₃) δ 14.2 (CH₃), 65.1 (CH₂), 97.4 (C-8), 113.9 (CN), 118.7 (C-9a), 128.3, 129.1, 130.3, 132.6 (C₆H₅), 154.3 (C-5a), 154.7 (C-2), 156.6, 156.8, 164.8, 166.0. MS (EI, *m/z*, %): 368 (M⁺ + 2, 38), 366 (M⁺, 100), 337 (99), 324 (35); IR (KBr, cm⁻¹): 2220 (CN), 1540, 1320, 1030.

4-Chloro-7,9-diphenylpyrido[**3**',**2**':**4**,**5**]**thieno**[**3**,**2**-*d*]**pyrimidine** (**8b**). Yield 68%; mp 199–201°C; C₂₁H₁₂N₃CIS calcd C, 67.47; H, 3.24; N, 11.24; found: C, 67.22; H, 3.41; N, 11.06; ¹H NMR (CDCl₃) δ 7.45–7.66 (m, 8H, C₆H₅), 7.80 (s, 1H, H-8), 8.11–8.16 (m, 2H, C₆H₅), 8.81 (s, 1H, H-2); ¹³C NMR (CDCl₃) δ 120.0 (C-8), 122.4 (C-9a), 127.5, 128.0, 128.9, 129.1, 129.5, 130.3, 136.5, 137.4 (C₆H₅), 150.6, 153.9 (C-2), 154.4, 157.0, 159.4, 164.3; MS (EI, *m/z*, %): 375 (M⁺ + 2, 16), 373 (M⁺, 44), 372 (100), 337 (8), 309 (8); IR (KBr, cm⁻¹): 1560, 1540, 1490, 1420.

Pyrido[3',2':4,5]thieno[3,2-d]pyrimidine-4-substituted (9a–1): General procedure. A solution of 8a or 8b (0.27 mmol) and the appropriate amine (0.32 mmol) in ethanol (10 mL) was refluxed until the starting material had disappeared (TLC). The solvent was removed under reduced pressure and the resulting solid was purified by flash chromatography or, alternatively, filtered off and recrystallized to yield (9a–1) (Tables 1 and 2).

8-Cyano-7-ethoxy-9-phenyl-2-(2-nitrophenyl)-4-oxo-1,2,3,4 tetrahydropyrido[3',2':4,5]thieno[3,2-d] pyrimidine (10). A catalytic amount of PTSA was added to a solution of 5a (0.3g, 0.87 mmol) and 2-nitrobenzaldehyde (0.14 g, 0.95 mmol) in toluene (30 mL). The reaction mixture was refluxed for 2h and the water formed was continuously removed by means of a Dean-Stark trap. The desired product was filtered off and recrystallized from ethanol/acetone to yield 10 (0.33 g, 81%): mp 291-293 °C; C₂₄H₁₇N₅O₄S calcd C, 61.14; H, 3.63; N, 14.85; found: C, 61.31; H, 3.75; N, 14.64; ¹H NMR (DMSO d_6) δ 1.37 (t, 3H, J=7.1 Hz, CH₃), 4.52 (q, 2H, J = 7.1 Hz, CH₂), 5.13 (d, 1H, exchangeable with D₂O, J = 3.5 Hz, NH), 6.25 (t, 1H, J = 3.4 Hz, H-2), 7.28–7.79 (m, 9H, C_6H_5 and C_6H_4), 8.26 (d, 1H, exchangeable with D₂O, J = 3.5 Hz, NH); ¹³C NMR (DMSO- d_6) δ 14.1 (CH₃), 63.2 (C-2), 64.1 (CH₂O), 95.1, 103.3, 114.2 (CN), 116.8, 125.4, 127.9, 128.2, 128.4, 128.8, 129.1, 130.0, 130.3, 132.2, 134.0, 135.2, 143.1, 146.9, 153.5, 161.1, 162.0, 163.4; MS (EI, *m/z*, %): 471 (M⁺, 80), 349 (100), 322 (69), 294 (49); IR (KBr, cm⁻¹): 3410 (NH), 3170 (NH), 2220 (CN), 1650 (CO).

8-Cyano-7-ethoxy-9-phenyl-2-(2-nitrophenyl)-4-oxo-3,4dihydropyrido[3',2':4,5]thieno[3,2-*d*]pyrimidine (11). A solution of 10 (0.20 g, 0.45 mmol) and 2,3-dichloro-5,6dicyano-1,4-benzoquinone (DDQ) (0.15 g, 0.67 mmol) in tetrahydrofuran (10 mL) was refluxed for 3 h. After cooling, the precipitate obtained was filtered off and the crude solid 11 was used in the next step without further purification. Compound 11, (0.15 g, 70%): mp > 300 °C; $C_{24}H_{13}N_5O_4S$ calcd C, 61.40; H, 3.22; N, 14.92; found: C, 61.56; H, 3.14; N, 14.99; ¹H NMR (DMSO-*d*₆) δ 1.43 (t, 3H, *J*=7.1 Hz, CH₃), 4.60 (q, 2H, *J*=7.1 Hz, CH₂), 7.34–8.14 (m, 9H, C₆H₅ and C₆H₄), 13.21 (br s, 1H, exchangeable with D₂O, NH). MS (EI, *m/z*, %): 469 (M⁺, 61), 292 (69), 264 (42); IR (KBr, cm⁻¹): 2220 (CN), 1640 (CO).

4-Chloro-8-cyano-7-ethoxy-9-phenyl-2-(2-nitrophenyl)pyrido[3',2':4,5]thieno[3,2-d]pyrimidine (12). A stirred solution of 11 (0.14g, 0.29 mmol) and phosphorous pentachloride (0.06 g, 0.29 mmol) in phosphorous oxychloride (5 mL) was refluxed for 15 h. The more volatile components of the reaction mixture were evaporated under reduced pressure, and ice (10g) was added. The resulting solid was filtered off and recrystallized from ethanol/acetone to yield 12 (0.13 g, 90%): mp 241-243 °C; C₂₄H₁₃N₅O₃ClS calcd C, 59.08; H, 2.89; N, 14.35; found: C, 59.09; H, 2.75; N, 14.50; ¹H NMR $(DMSO-d_6) \delta 1.46 (t, 3H, J = 7.1 Hz, CH_3), 4.63 (q, 2H, d)$ $J = 7.1 \text{ Hz}, \text{ CH}_2$), 7.52–7.93 (m, 9H, C₆H₅ and C₆H₄); ¹³C NMR (CDCl₃) δ 14.3 (CH₃), 65.2 (CH₂), 97.3 (C-8), 114.0 (CN), 118.7 (C-9a), 127.5 (C-4a), 124.0, 128.3, 128.8, 129.9, 130.6, 131.2, 131.7, 131.8, 132.5, 149.5 (C₆H₄NO2 and C₆H₅), 154.1, 156.5, 156.8, 159.2, 164.8, 166.2 MS (EI, m/z, %): 489 (M⁺ + 2, 5), 487 (M⁺, 13), 470 (100), 440 (59), 411 (36), 221 (8); IR (KBr, cm⁻¹): 2220 (CN), 1540, 1340, 1310.

4-(4-Benzylpiperazino)-8-cyano-7-ethoxy-9-phenyl-2-(2nitrophenyl)-pyrido[3',2':4,5]thieno[3,2-*d***]pyrimidine (13). A solution of 12** (0.10 g, 0.22 mmol) and 4-benzylpiperazine (0.08 g, 0.44 mmol) in ethanol (7 mL) was refluxed for 3 h. The solid was filtered off and recrystallized from ethanol/dichloromethane (Tables 1 and 2).

2-Amino-3,5-dicyano-6-morpholinopyridine (15). A solution of 14^{17} (0.18 g, 1.12 mmol) and morpholine (0.12 g, 1.34 mmol) in ethanol/THF (1/3, 15 mL) was refluxed until the starting material had disappeared (TLC). The solid was filtered off and recrystallized from ethanol to yield **15** (0.14 g, 54%): mp 198–200 °C; C₁₁H₁₁N₅O calcd C, 57.63; H, 4.84; N, 30.55; found: C, 57.72; H, 4.68; N, 30.47; ¹H NMR (CDCl₃) δ 3.78 (m, 4H, NCH₂), 3.88 (m, 4H, CH₂O), 5.36 (br s, 2H, exchangeable with D₂O, NH₂), 7.77 (s, 1H, H-4); ¹³C NMR (CDCl₃) δ 47.3 (NCH₂), 66.5 (CH₂O), 81.4, 82.6 (C-3, C-5), 115.8 (CN), 117.8 (CN), 149.5 (C-4), 159.1, 159.4 (C-2, C-6); MS (EI, *m/z*, %): 229 (M⁺, 32), 172 (38), 144 (100), 117 (51), 89 (47); IR (KBr, cm⁻¹): 3400, 3315, 3210 (NH), 2210 (CN), 1650, 1600, 1540, 1480.

2-Chloro-3,5-dicyano-6-morpholinopyridine (16). To a stirred suspension of anhydrous $CuCl_2$ (4.4 g, 32.8 mmol) in dry CH₃CN (100 mL), containing isoamyl nitrite (3.8 g, 32.8 mmol), was added compound **15** (5.0 g, 21.8 mmol). The reaction mixture was heated at 65 °C for 2.5 h. After cooling, the reaction mixture was poured into 20% HCl (200 mL) and extracted with CH₂Cl₂ (3×50 mL). The organic layers were combined and evaporated under reduced pressure and the resulting solid was purified by flash chromatography using CH₂Cl₂/AcOEt (35/1) as eluent to yield **16** (2.9 g, 53%): mp 186–188 °C; C₁₁H₉N₄ClO calcd C, 53.13; H, 3.65;

N, 22.53; found: C, 53.35; H, 3.76; N, 22.57; ¹H NMR (CDCl₃) δ 3.82 (m, 4H, NCH₂), 4.02 (m, 4H, CH₂O), 7.96 (s, 1H, H-4), ¹³C NMR (CDCl₃) δ 47.5 (NCH₂), 66.4 (CH₂O), 89.9, 98.1 (C-3, C-5), 114.1 (CN), 115.9 (CN), 149.9 (C-4), 154.8, 157.2 (C-2, C-6); MS (EI, *m*/*z*, %): 250 (M⁺ + 2, 13), 248 (M⁺, 39), 217 (35), 205 (65), 191 (66), 163 (100), 128 (44); IR (KBr, cm⁻¹): 2220 (CN), 1600, 1535, 1410, 1275.

3,5-Dicyano-6-morpholinopyridine-2-(1*H***)-thione (17). A mixture of 16** (2.4 g, 9.8 mmol) and NaHS×H₂O (2.4 g) in ethanol (100 mL) was stirred at room temperature for 1.5 h. The solvent was removed under reduced pressure. After addition of water (50 mL) the solution was acidified with 2 N HCl and **17** deposited as a solid that was filtered off and used in the next step without further purification. Compound **17** (2.3 g, 9.5 mmol): mp 228–230 °C; C₁₁H₁₀N₄OS calcd C, 53.65; H, 4.09; N, 22.75; found: C, 53.47; H, 4.33; N, 22.59; ¹H NMR (acetone- d_6) δ 3.66 (br s, 1H, exchangeable with D₂O), 3.88 (m, 8H, NCH₂CH₂O), 8.05 (s, 1H, H-4); MS (EI, *m/z*, %): 246 (M⁺, 49), 215 (34), 189 (82), 161 (100), 134 (46), 128 (17); IR (KBr, cm⁻¹): 2210 (CN), 1590, 1495, 1280.

3,5-Dicyano-2-cyanomethylthio-6-morpholinopyridine (18). A mixture of 17 (0.1 g, 0.4 mmol), chloroacetonitrile (40 mg, 0.5 mmol), potassium carbonate (70 mg, 0.5 mmol), and a catalytic amount of KI in acetone (15 mL) was stirred at room temperature for 4h. The insoluble solid was removed by filtration and washed with acetone. The filtrate and the washings were combined and the solvent was evaporated. The resulting solid was recrystallized from acetone to yield 18 (75 mg, 0.3 mmol): mp 176-178 °C; C₁₃H₁₁N₅OS calcd C, 54.73; H, 3.89; N, 24.55; found: C, 54.62; H, 3.78; N, 24.42; ¹H NMR (CDCl₃) δ 3.87 (m, 6H, NCH₂ and SCH₂), 4.05 (m, 4H, CH₂O), 7.88 (s, 1H, H-4); ${}^{13}C$ NMR (CDCl₃) δ 16.1 (SCH₂), 47.7 (NCH₂), 66.5 (CH₂O), 88.5, 95.3 (C-3, C-5), 113.9 (CN), 115.4 (CN), 116.4 (CN), 148.4 (C-4), 157.5, 162.3 (C-2, C-6). MS (EI, *m*/*z*, %): 285 (M⁺, 46), 245 (100), 226 (61), 173 (25), 160 (20); IR (KBr, cm⁻¹): 2240 (CN), 2215 (CN), 1580, 1535, 1400, 1340.

5-Amino-3,6-dicyano-2-morpholinothieno[2,3-*b***]pyridine (19). Method A.** A mixture of **18** (2.0 g, 7.0 mmol) and potassium carbonate (1.2 g, 8.4 mmol) was stirred at room temperature for 1 h. The solid was filtered off and recrystallized from ethanol to yield **19** (1.7 g, 88%): mp 294–296 °C; $C_{13}H_{11}N_5OS$ calcd C, 54.73; H, 3.89; N, 24.55; found: C, 54.65; H, 3.76; N, 24.45; ¹H NMR (DMSO-*d*₆) δ 3.69 (m, 8H, NCH₂CH₂O), 7.28 (br s, 2H, exchangeable with D₂O, NH₂), 8.75 (s, 1H, H-4); ¹³C NMR (DMSO-*d*₆) δ 48.3 (NCH₂), 65.8 (CH₂O), 68.5 (C-6), 92.2 (C-3), 115.8 (CN), 116.5 (CN), 117.9 (C-4a), 140.1 (C-4), 150.1 (C-5), 158.8, 162.6 (C-2, C-7a); MS (EI, *m/z*, %): 285 (M⁺, 95), 270 (15), 254 (33), 240 (34),

228 (80), 200 (100), 173 (59); IR (KBr, cm⁻¹): 3400, 3340, 3220 (NH), 2210 (CN), 2190 (CN), 1655, 1595, 1565, 1555, 1510, 1435.

Method B (from compound 17). A mixture of 17 (2.3 g, 9.3 mmol), chloroacetonitrile (0.8 g, 11.2 mmol), potassium carbonate (1.5 g, 11.2 mmol), and a catalytic amount of KI in acetone (50 mL) was stirred at room temperature for 4 h. The solvent was removed under reduced pressure and the crude solid obtained was stirred at room temperature for 1 h with a solution of potassium carbonate (1.5 g, 11.2 mmol) in ethanol (50 mL). The solid was filtered off and recrystallized from ethanol to yield 19 (2.6 g, 99%).

4-Chloro-8-cyano-2-dimethylamino-7-morpholinopyrido-[3',2':4,5]thieno[3,2-d]pyrimidine (20). A solution of 19 (2.3 g, 8.1 mmol) and phosgene iminium salt (2.0 g, 12.2 mmol) in 1,2-dichloroethane (20 mL) was refluxed for 6 h. After cooling, a stream of dry hydrogen chloride was passed through the reaction mixture for 30 min, and the stirring continued at room temperature for 48 h. The resulting solid was filtered off and recrystallized from ethanol/acetone to yield 20 (1.6 g, 53%): mp 225-227 °C; C₁₆H₁₅N₆OClS calcd C, 51.26; H,4.03; N, 22.42; found: C, 51.37; H, 3.95; N, 22.49; ¹H NMR (CDCl₃) δ 3.25 (s, 6H, NMe₂), 3.89 (m, 8H, NCH₂CH₂O), 8.48 (s, 1H, H-9); ¹³C NMR (CDCl₃) δ 37.4 (NMe₂), 48.5 (NCH₂), 66.6 (CH₂O), 92.2 (C-8), 114.4 (C-4a), 117.9 (CN), 119.2 (C-9a), 140.0 (C-9), 153.7 (C-4), 157.4 (C-5a), 159.9 (C-9b), 160.8 (C-7), 167.0 (C-2); MS (EI, m/z, %): 374 (M⁺, 92), 359 (15), 344 (6), 330 (11), 317 (62), 287 (29), 238 (24); IR (KBr, cm⁻¹): 2970, 2210 (CN), 1600, 1580, 1550, 1490.

8-Cyano-2-dimethylamino-7-morpholinopyrido[3',2':4,5]-thieno[3,2-*d***]pyrimidine-4-substituted (21a,b).** A solution of **20** (0.15 g, 0.27 mmol) and the appropriate amine (0.32 mmol) in ethanol/THF (1/3, 10 mL) was refluxed until the starting material had disappeared (TLC). The solid was filtered off and recrystallized from ethanol/ acetone to yield **21a,b** (Tables 1 and 2).

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