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Synthesis and structure–activity relationship (SAR) study of 4-azabenzoxazole analogues as H₃ antagonists

Ning Shao^{a,*}, Robert Aslanian^a, Robert E. West Jr.^b, Shirley M. Williams^b, Ren-Long Wu^b, Joyce Hwa^b, Christopher Sondey^b, Jean Lachowicz^b, Anandan Palani^a

^a Chemical Research Department, Department of Chemical Research, Merck Research Laboratories, 2015 Galloping Hill Road, Kenilworth, NJ 07033, USA ^b Cardiovascular/Metabolic Disease, Department of Chemical Research, Merck Research Laboratories, 2015 Galloping Hill Road, Kenilworth, NJ 07033, USA

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ABSTRACT

The synthesis and SAR of a novel series of 4-azabenzoxazole histamine H_3 antagonists is described. Introduction of substituted phenyl, pyridyl and fused heterocyclic groups to the 6-position of the 4-azabenzoxazole core gave a series of compounds with good H_3 antagonist activity in both ex vivo and in vivo assays.

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Histamine plays very important roles in physiological processes. H₁ and H₂ receptors have been successfully targeted, and therapeutic agents have been developed to treat conditions such as allergies (H₁ receptor antagonist) and gastric ulcers (H₂ receptor antagonist). The newly discovered H₄ receptor is found in immune cells, suggesting its role in regulating inflammatory responses.¹ The H₃ receptor, widely distributed in the central nervous system, modulates the release of histamine and other neurotransmitters such as acetylcholine, dopamine, norepinephrine and serotonin.² Central administration of a histamine H₃ antagonist leads to increased central histamine levels and hence may be useful for the treatment of a variety of CNS disorders,³ such as attention deficit and hyperactivity disorder (ADHD),⁴ sleep disorders,⁵ schizophrenia⁶ or obesity.⁷ Early generations of H₃ antagonists, based on structures containing the imidazole moiety found in histamine,⁸ suffered from CYP450 inhibition and drug-drug interactions.⁹ For this reason, recent efforts have been directed to the non-imidazole H₃ antagonists, such as ABT-239,¹⁰ GSK-189254,¹¹ UCL-2190,¹² A-331440 and JNJ-5207852.^{13,14}

Walczyński et al. identified a structurally novel non-imidazole histamine H₃ receptor antagonist I (Fig. 1), which has 5-azabenzo-thiazole moiety as the core structure.¹⁴ Compound I showed $pA_2 = 7.25 \pm 0.07$ (electric field stimulation assay on guinea-pig jejunum). In our own medicinal chemistry effort, we became interested in structurally similar cores II and III. The development of a

* Corresponding author. E-mail address: Ning.shao@merck.com (N. Shao). series of derivatives of **II** as potent H_3 antagonists has been published recently.¹⁵ Herein we describe our effort in developing 4-azabenzoxazole derivatives **III** as non-imidazole antagonists.

Synthesis of the key intermediate began with the commercially available 2-aminopyridin-3-ol **1** (Scheme 1). Condensation of **1** with carbon disulfide gave compound **2**. Methylation and substitution with piperidinopiperidine, which was identified in the previous study as a privileged scaffold, afforded compound **4**. Regioselective bromination at the 6-position gave compound **5**, which served as the key intermediate for the subsequent analogue synthesis. Suzuki coupling or Buchwald–Hartwig couplings gave a series of derivatives, and their activity was evaluated.¹⁶

The binding affinities of the analogues were determined as K_i values in the human H₃ recombinant assay.¹⁷ Compound **4** (human K_i = 1200 nM) and **4a** (human K_i = 4000 nM) showed weak binding affinity, but putting substitutions groups onto the 6-position of benzoxazole core increased the binding activity (Table 1). We first added smaller groups such as vinyl (compound 5), acetamido (compound 6), and small cyclic amino (compound 7) groups, and the binding affinities improved to as potent as 150 nM (compound 7). Further extension of the substituent either by fusing a phenyl ring (compound **8**) or attaching a heterocycle (compound **9**) reduced binding affinities. Aromatic rings with different substitution groups were then directly attached to the core. Polar substituents (compounds 10, 11, 12, 14) resulted in compounds which have binding affinities at around 300 nM, with an exception (compound **13**) that the ortho acetamido substitution has a fivefold drop in binding affinity. Presumably the appended phenyl group is twisted out of



Figure 1. Structures of target molecules of study.



Scheme 1. Synthesis of the 4-azabenzoxazole derivatives.

Table	1
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Compound	R	K_{i} (H ₃ , nM)	Compound	R	<i>K</i> _i (H ₃ , nM)
5	1	370	12	AcHN	233
6	O N ³ ² H	657	13	NHAc	1526
7	N. S.	150	14	H ₂ N H ₂ s ²	288
8	NyE	260	15	F ₃ C	238
9		357	16	O H	58
10	HO	249	17	NC 35	36
11	HO	379	18	NC St.	37

plane with the azabenzoxazole ring due to the presence of the *ortho* acetamido group, thus reducing binding affinity for the H3 receptor. Smaller substituents on the phenyl ring resulted in compounds (**16–18**) with 10-fold increased binding activity.

The active compounds were further tested in the ex vivo occupancy assay in the ICR mouse model.¹⁸ Four hours following the oral administration of compounds at 10 mg/kg, ex vivo receptor displacement of control in mouse brain slices was determined. Compound **18** exhibited good receptor occupancy (59%) in mouse at the 4 h time point. It also has favourable rat p*K*, with an AUC_{0-6h} of 706 ng·h/mL (10 mg/kg) and a brain/plasma ratio of 11.3 at the 6 h time point. This is very important since H₃ activity is centrally mediated. However, compounds **17** and **18** showed significant inhibition of the hERG channel (96% and 92% at 10 μ M), a classic problem associated with historic H₃ antagonists.^{19a} The less active compound **13** displayed no hERG inhibition at 10 μ M (-2.0%), probably due to its polar substituent group.^{19b}

In order to retain activity and reduce the hERG inhibition (Table 2) we next evaluated replacing the phenyl group with heteroclyes at the 6-position. This may also be beneficial to the binding activity since it is well documented that introducing a second amine into the molecule normally improves the binding activity as potent H₃ antagonist.²⁰ However, one drawback of those dibasic amine compounds is that it may result in phospholipidolysis, a type of toxicity that occurs when molecules partition into lipid bilayer and impair normal phospholipid turn over.²¹ We envisioned that heterocycles containing a less basic nitrogen would retain the similar activity-boosting effect, and in addition, should also have a better hERG profile due to the its reduced lipophilicity.

Pyridine was first evaluated as the phenyl replacement with 2-, 3- and 4-pyridyl substituents attached to the 6 position of the core structure (**19**, **20**, **21**). All three compounds retained good binding affinity (95, 37, 47 nM), with the 3-pyridyl slightly less active than the 2- and 4-pyridyl substituents. Then we appended

Compound	R	K_i (H ₃ , nM)	Compound	R	K_i (H ₃ , nM)
19	N St.	95	26	N St.	43
20	N	37	27	O N st	23
21	N 35	47	28		37
22	NC N 35	253	29	N	15
23	H ₂ N N	278	30	N St.	15
24	F N S	70	31	F ₃ C N 3 ⁵	143
25	N J J J J J J J J J J J J J J J J J J J	196	32	N 35	79

Table 3

N N	≻n∖ >
·N· N	

	R	K_{i} (H ₃ , nM)	hERG (10 µM) (%)		R	K_i (H ₃ , nM)	hERG (10 µM) (%)
33		117	95	38	N N N S ⁵	33	5
34	O States	109	N.D.	39		119	23
35		153	15	40	O N N N	25	20
36		79	96	41	N 3t	4	62
37	S ^r	35	92	42	N 35t	18	17

different substituents onto the pyridines. Substitution of 3-pyridine with cyano and amino groups (**22**, **23**) were detrimental to activity while the F/H replacement (**24**) did not offer much benefit either. For the 4-substituted pyridine, adding methyl or methoxyl group (compounds **25**, **26**) gave compounds with inferior/comparable binding activity. Gratifyingly, substitution of 2-pyridyl with methyl, chloro, or methoxyl groups gave compounds with improved potency (**27–30**, compared to **21**). Changing methyl to ethyl or a trifluoromethyl group reduced the potency (**31**, **32** compared to **29**), suggesting there may be a size limit at the ortho position.

Two of the most active compounds **29** and **30** were evaluated in the hERG binding assay and they demonstrated a better profile (49% and 32% inhibition at 10 μ M). Unfortunately, these compounds showed very low receptor occupancy in the ex vivo mouse study, with compound **29** showing 17% occupancy and compound **30** 18% at 4 h time point, probably due to poor pK and/or low brain penetration. To improve ex vivo activity and further improve hERG profile of these H₃ antagonists, we then turned to fused heterocycles to combine the beneficial effect of pyridine basic nitrogen and the small non-polar substitution groups (Table 3). Phenyl fused heterocycles such as compounds **33–35** showed no dramatic improvement in binding activity, but compounds **36** and **37** showed improved binding activity probably due to the basic nitrogen at the ortho position. Pyridine-fused heterocycles (**38–41**) generally exhibited better binding activity, such as compound **41** with binding affinity of 4 nM and compound **42** with binding of 18 nM. The hERG activity of these compounds was also tested, and they showed better hERG profile compared to the previous compounds (Table 3). Compound **41** had a moderate hERG inhibition of 62% while **42** showed only 17% inhibition.

Compound **41** was further evaluated in the ex vivo study. Fortunately, it exhibited very good receptor occupancy (65%) at 4 h time point with a dose of 10 mg/kg. We then studied it in the in vivo STZ-DIO mouse assay.²² After a two-day feed with 30 mg/kg dose, **41** demonstrated both antidiabetic (63 mg/L, glucose level reduction) and anti-obesity effect (3.85% food inhibition, 0.64% body weight loss).

In summary, a new series of 4-azabenzoxazole derivatives was evaluated as H_3 antagonists. Appending the phenyl, pyridyl or fused heterocyclic moieties to the 6-position of the core gave a series of H_3 antagonists with good in vitro binding activity. The hERG issues within this series were also addressed by incorporation of fused pyridine derived heterocycles. Compound **41** was identified as an active H_3 antagonist, with single digit in vitro binding activity and good ex vivo binding activity. It also demonstrated good in vivo activity in the rodent biological assay for obesity.

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- 17. For binding assays, membranes (P2 pellet) from rHu H3-HEK cells (3 µg protein) were incubated in 200 µl 50 mM Tris–HCl, pH 7.4, with 1 nM [3H]N- α -methylhistamine (82 Ci/mmol) and compounds at concentrations equivalent to half orders of magnitude over a five-order of magnitude range. Nonspecific binding was determined in the presence of 10⁻⁵ M thioperamide. After 30 min at 30 °C, assay mixtures were filtered through 0.3% polyethylenimine-soaked GF/B glass fiber filters, which were rinsed thrice with buffer, dried, impregnated with Meltilex wax scintillant, and counted. *K*_i values were determined from curves fit to the data using GraphPad Prism nonlinear, least squares, curve-fitting program.
- 18. ICR, Imprinting Control Region mice (from Taconics, nomenclature IcrTacICR). Individual mouse cortexes were dissected and homogenized in ice cold assay buffer, 50 mM Na₂HPO₄–KH₂PO₄ buffer (pH 6.8). Samples were then frozen at -80 °C for at least 12 h. Protein concentration was determined by BCA Protein Assay. A 30 min room temperature incubation was performed, containing 140 µg/assay homogenized cortex sample, 0.1% BSA, 1 nM 3*H*-*N*-∞ methylhistamine and 10 µM thioperamide for non-specific binding or assay buffer for total binding. Incubations were performed in quadruplet and stopped by rapid filtration on a Brandel Harvester using Unifilter-96, GF/B plates presoaked in 0.3% PEI for 30 min. The remaining radioactivity was measured on a Packard TopCount-NTX. Specific binding was calculated by subtracting the non-specific binding from the total.
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- 22. Five week old male ICR mice (Taconic, Tac:Icr:Ha (ICR)fBR) were fed high fat/ cholesterol diet (45% kcal fat and 0.12% cholesterol, Research Diets D04012801) for 3 weeks, and were given streptozotocin (STZ) intraperitoneally at 80 mg/kg to induce type 2 diabetes. T2D mice were chosen for the study two weeks after STZ injection (*n* = 12 per group, with non-fasting glucose between 250 and 500 mg/dl) and were balance into groups. Compounds were administered by PO dosing with 0.4% MC daily before the dark onset. Non-fasting glucose was monitored daily at 4 h after the lights on. Individual body weight and food intake were monitored daily.

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