

R(–)-*N*-alkyl-11-hydroxy-10-hydroxymethyl- and 10-methyl-aporphines as 5-HT_{1A} receptor ligands

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Abstract—Several *N*-substituted-11-hydroxy-10-hydroxymethyl- and 11-hydroxy-10-methylaporphines were synthesized and their binding affinities at dopamine D₁ and D₂ receptors and serotonin 5-HT_{1A} and 5-HT_{2A} receptors in rat forebrain tissue were evaluated. Tested compounds displayed moderate to high affinity to 5-HT_{1A} receptors but low affinity to D₁ and D₂ receptors. The most potent novel 5-HT_{1A} agent was *R*(–)-*N*-methyl-10-hydroxymethyl-11-hydroxyaporphine.
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R-Apomorphine, first synthesized in 1869, is an agonist on central dopamine (DA) D₁ and D₂ receptors.¹ Small changes in the structure of apomorphine can lead to major changes in pharmacological profiles.² For example, elimination of the 10-hydroxy group of apomorphine produced a dopamine DA D₁ receptor antagonist.³ Cannon reported that replacement of the C10-hydroxy moiety with a methyl group resulted in potent binding affinity at the serotonin (5-hydroxytryptamine) 5-HT_{1A} receptor but low affinity at DA receptors.⁴ A previously prepared series of 10-substituted-11-oxygenated *R*(–)-aporphines also lacked DA receptor affinity but showed potent and selective affinity toward the 5-HT_{1A} receptor.⁵ Hedberg proposed that the selective serotonin receptor affinity of these aporphines appears to be due to a C10-methyl group, and a binding-site model suggested the presence of a ‘methyl pocket’ in the 5-HT_{1A} receptor binding site.^{5c} In contrast, the C10-methyl group of these aporphines was not accommodated by a binding-site model for DA receptors.⁵

To develop additional insight into the importance of C10 as well as *N*-substituents in aporphines for 5-HT and DA receptor affinity, we synthesized several *N*-alkyl-11-hydroxy-10-hydroxymethyl- and 11-hydroxy-10-methylaporphines and evaluated their affinity at DA

(D₁ and D₂) and 5-HT (5-HT_{1A} and 5-HT_{2A}) receptors (Fig. 1).

R(–)-10-methyl-11-hydroxyaporphine **2** was synthesized starting from morphine by a procedure reported by Hedberg.^{5b} Triflation of the 3-hydroxy moiety of morphine followed by a palladium-catalyzed coupling reaction led to **8**. Acid-catalyzed rearrangement of **8** yielded the desired aporphine **2** (Scheme 1). Scheme 2 shows the synthesis of 2-methoxy-10-methyl-11-hydroxy-aporphines **3** and **4**. Thebaine **9** and *N*-*n*-propylnorthebane **10** were *O*-demethylated to **11** and **12**, respectively, using the procedure reported by Coop,⁶ and then *O*-triflated to produce compounds **13** and **14**. A palladium-catalyzed coupling reaction of **13** and **14** with Sn(Me)₄ gave **15** and **16**, followed by their acid-catalyzed rearrangement produced target compounds **3** and **4**.^{5a} The synthesis of 11-hydroxy-10-hydroxymethyl-

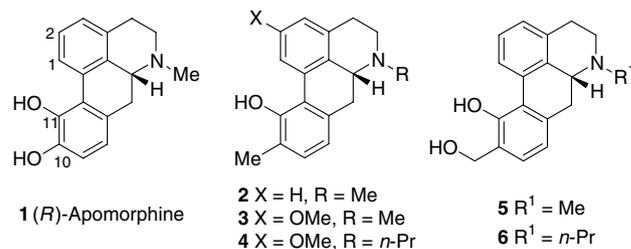
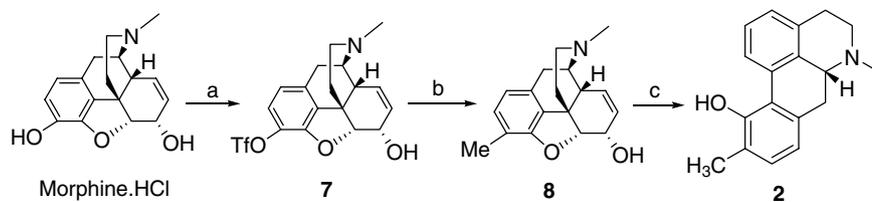


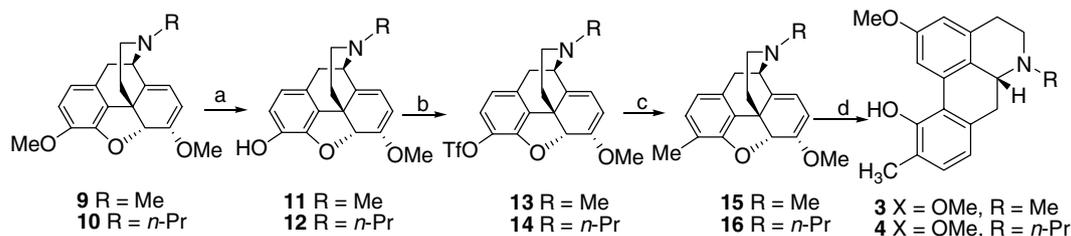
Figure 1. Structures of aporphine analogs.

Keywords: Aporphines, Binding affinities; 5-HT_{1A} receptor ligands.

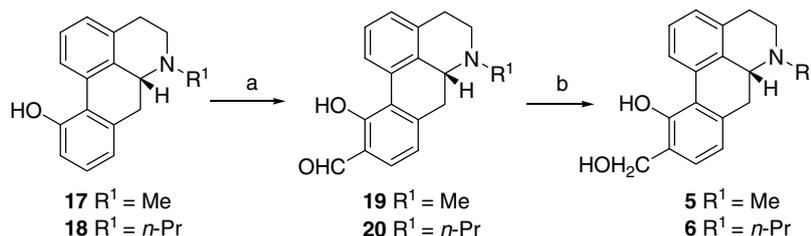
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Scheme 1. Reagents: (a) PhNTf₂, Et₃N, CH₂Cl₂, 90%; (b) Me₄Sn, (Ph₃P)PdCl₂, PPh₃, LiCl, DMF, 75%; (c) MeSO₃H, 65%.



Scheme 2. Reagents: (a) L-selectride, THF, 23–28%; (b) PhNTf₂, Et₃N, CH₂Cl₂, 90–93%; (c) Me₄Sn, (Ph₃P)₂PdCl₂, PPh₃, LiCl, DMF, 41–50%; (d) MeSO₃H, 45–50%.



Scheme 3. Reagents: (a) MeMgBr, HMPA, (HCHO)_n, benzene, 85–91%; (b) NaBH₄, MeOH, 90–92%.

porphines **5** and **6** is shown in **Scheme 3**. We synthesized the *R*-(–)-enantiomer of 11-hydroxyaporphines **17** and **18** using a previously reported procedure.⁷ *Ortho*-formylation of the 11-hydroxy aporphines **17** and **18** employed a modification of Cannon's procedure^{4b} and afforded the desired products **19** (85%) and **20** (91%) in high yields, followed by reduction of **19** and **20** with NaBH₄ to produce the target compounds **5** and **6**. Spectral (¹H NMR and ¹³C NMR) data and combustion analysis for the target compounds were consistent with their proposed structures.⁸

The affinities of compounds **2–6** for DA (D₁ and D₂) and 5-HT (5-HT_{1A} and 5-HT_{2A}) receptors were assessed using competitive binding assays with membrane homogenates of whole rat brain tissue (5-HT_{1A} and 5-HT_{2A}) or rat striatal tissue (D₁ and D₂). The following tritiated radioligands were used: [³H]SCH23390 (D₁), [³H]nemonapride (D₂), [³H]8-OH-DPAT (5-HT_{1A}), and [³H]ketanserin (5-HT_{2A}).⁹ The results are summarized in **Table 1**.

The *N*-substituted 11-hydroxy-10-hydroxymethyl aporphine **5** and 11-hydroxy-2-methoxy-10-methyl congener **3** displayed selective and potent affinity for the serotonin 5-HT_{1A} receptor but low affinity at DA receptors D₁ and D₂ (**Table 1**). These findings support the proposal that *ortho*-dihydroxy substitution in the aporphine D

ring enhances affinity to DA receptors, whereas analogous methyl or hydroxymethyl substitution enhances interactions with the 5-HT_{1A} receptor. Of note, the 10-hydroxymethyl-substituted compound **5** displayed 100-fold higher 5-HT_{1A} receptor affinity ($K_i = 2.4$ nM) than the 10-methyl substituted compound **2** ($K_i = 216$ nM), suggesting that a 10-methyl group is not required for affinity to the 5-HT_{1A} receptor. The 2-methoxy group in compound **3** seems to increase the affinity to 5-HT_{1A} receptors ($K_i = 21.5$ nM) 10-fold higher than compound **2** ($K_i = 216$ nM). That the 10-hydroxymethyl compounds **5** and **6** were inactive at DA receptors supports the impression that a 10-hydroxymethyl group is not required for high DA-receptor activity and that the interaction of 10-hydroxymethylaporphines with DA receptors does not involve hydrogen bonding. We also evaluated compounds **3** and **5** for affinity to 5-HT_{2A} receptors and found 108-fold and 57-fold lower potency than at 5-HT_{1A} receptors, respectively (for compound **5**: $K_i = 137$ vs 2.4 nM; for **3**: $K_i = 271$ vs 2.5 nM). It is worthy to note that the *N*-propyl substitution is preferred over *N*-methyl substitution in the interaction with DA receptors.¹⁰ In contrast *N*-methyl substitution ($K_i = 2.4$ for compound **5** at 5-HT_{1A}; $K_i = 21.5$ for compound **3** at 5-HT_{1A}) is preferred in the interaction with 5-HT receptor than the *N*-*n*-propyl substitution ($K_i = 375$ for compound **6** at 5-HT_{1A}; $K_i = 480$ for compound **4** at 5-HT_{1A}).

Table 1. Affinities (K_i) for rat brain D₁, D₂, 5-HT_{1A}, and 5-HT_{2A} receptors

Compound	K_i (nM)			
	[³ H]SCH23390 (D ₁)	[³ H]nemonapride (D ₂)	[³ H]8-OH-DPAT (5-HT _{1A})	[³ H]Ketanserin (5-HT _{2A})
1	214 ± 18 ^a	13 ± 2 ^a	296 ± 15 ^b	—
2	9650 ± 1250	11500 ± 1900	216 ± 40	—
3	1780 ± 320	3760 ± 760	21.5 ± 2.7	271 ± 19
4	2790 ± 640	1350 ± 250	480 ± 62	—
5	1390 ± 160	7000 ± 850	2.4 ± 0.4	137 ± 12
6	1980 ± 380	6060 ± 1110	375 ± 84	—

^a From Ref. 7.^b From Ref. 5c.

In addition, both 10-methyl and 10-hydroxymethyl substituted 11-hydroxyaporphines displayed high affinity toward serotonin 5-HT_{1A} receptors but very low affinity at DA (D₁ and D₂) receptors. Finally, the *N*-methyl-substituted 11-hydroxy-10-methyl- and 10-hydroxymethyl-aporphines were more potent than the *N*-*n*-propyl analogs at 5-HT_{1A} receptors.

Acknowledgments

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References and notes

- Baldessarini, R. J.; Kula, N. S.; Zong, R.; Neumeyer, J. L. *Eur. J. Pharmacol.* **1994**, *254*, 199.
- Zhang, A.; Zhang, Y.; Branfman, A. R.; Baldessarini, R. J.; Neumeyer, J. L. *J. Med. Chem.* **2007**, *50*, 171.
- Schaus, J. M.; Titus, R. D.; Foreman, M. M.; Mason, N. R.; Truex, L. L. *J. Med. Chem.* **1990**, *33*, 600.
- (a) Cannon, J. G.; Mohan, P.; Bojarski, J.; Long, J. P.; Bhatnagar, R. K.; Leonard, P. A.; Flynn, J. R.; Chatterjee, T. K. *J. Med. Chem.* **1988**, *31*, 313; (b) Cannon, J. G.; Moe, S. T.; Long, J. P. *Chirality* **1991**, *3*, 19.
- (a) Hedberg, M. H.; Johansson, A. M.; Hacksell, U. *J. Chem. Soc., Chem. Commun.* **1992**, 845; (b) Hedberg, M. H.; Johansson, A. M.; Nordvall, G.; Yliniemela, A.; Li, H.-B.; Martin, A. R.; Hjorth, S.; Unelius, L.; Sundell, S.; Hacksell, U. *J. Med. Chem.* **1995**, *38*, 647; (c) Hedberg, M. H.; Jansen, J. M.; Nordvall, G.; Hjorth, S.; Unelius, L.; Johansson, A. M. *J. Med. Chem.* **1996**, *39*, 3491.
- Coop, A.; Lewis, J. W.; Rice, K. C. *J. Org. Chem.* **1996**, *61*, 6774.
- Csutoras, C.; Zhang, A.; Zhang, K.; Kula, N. S.; Baldessarini, R. J.; Neumeyer, J. L. *Bioorg. Med. Chem.* **2004**, *12*, 3553.
- Compound 2: mp (free base) 270–271 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.88 (d, *J* = 8.1 Hz, 1H), 7.25 (dd, *J* = 7.5 and 7.5 Hz, 1H), 7.07 (d, *J* = 7.5 Hz, 1H), 7.00 (d, *J* = 7.8 Hz, 1H), 6.77 (d, *J* = 7.5 Hz, 1H), 3.23–3.14 (m, 1H), 3.08–3.00 (m, 3H), 2.75 (dd, *J* = 3.3 and 16.5 Hz, 1H), 2.55–2.45 (m, 2H), 2.52 (s, 3H), 2.28 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 150.8, 135.6, 135.4, 133.8, 131.5, 129.6, 127.6, 126.5, 123.6, 123.5, 120.9, 119.9, 62.39, 52.9, 44.0, 34.9, 29.1, 16.1; Anal. Calcd for C₁₈H₁₉NO: C, 71.63; H, 6.68; N, 4.64. Found: C, 71.34; H, 6.65; N, 4.60.
- Compound 3: mp (HCl salt) 183–185 °C; ¹H NMR (base, 300 MHz, CDCl₃) δ 7.50 (d, *J* = 2.7 Hz, 1H), 7.00 (d, *J* = 7.5 Hz, 1H), 6.78 (d, *J* = 7.2 Hz, 1H), 6.61 (d, *J* = 2.4 Hz, 1H), 3.82 (s, 3H), 3.24–3.00 (m, 4H), 2.72 (dd, *J* = 3.0 and 16.2 Hz, 1H), 2.56–2.47 (m, 2H), 2.52 (s, 3H), 2.29 (s, 3H); ¹³C NMR (base, 75 MHz, CDCl₃) δ 158.1, 150.8, 135.9, 135.0, 132.6, 129.7, 123.4, 120.1, 111.9, 110.3, 109.8, 62.1, 55.2, 53.0, 44.0, 35.3, 29.5, 16.1; Anal. Calcd for C₁₉H₂₂ClNO₂·H₂O (salt): C, 65.23; H, 6.91; N, 4.00. Found: C, 64.88; H, 6.56; N, 3.85.
- Compound 4: mp (HCl salt) 159–160 °C; ¹H NMR (base, 300 MHz, CDCl₃) δ 7.48 (d, *J* = 2.1 Hz, 1H), 7.01 (d, *J* = 7.8 Hz, 1H), 6.78 (d, *J* = 7.5 Hz, 1H), 6.61 (d, *J* = 2.1 Hz, 1H), 3.82 (s, 3H), 3.31–2.39 (m, 9H), 2.30 (s, 3H), 1.67–1.54 (m, 2H), 0.96 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (base, 75 MHz, CDCl₃) δ 158.1, 150.8, 136.0, 135.4, 132.8, 129.7, 123.4, 120.7, 120.0, 111.9, 110.2, 59.5, 56.4, 55.2, 48.9, 35.2, 29.6, 19.5, 16.1, 12.1; Compound 5: mp (HCl salt) >250 °C; (free base) 203–205 °C; ¹H NMR (base, 300 MHz, DMSO-*d*₆) δ 8.97 (br, 1H), 8.12 (d, *J* = 7.8 Hz, 1H), 7.19 (t, *J* = 7.8 Hz, 1H), 7.07 (d, *J* = 7.5 Hz, 1H), 7.02 (d, *J* = 7.5 Hz, 1H), 6.82 (d, *J* = 7.2 Hz, 1H), 5.74 (br, 1H), 4.66 (s, 2H), 3.15–2.90 (m, 4H), 2.7 (m, 1H), 2.44 (s, 3H), 2.42–2.25 (m, 2H); ¹³C NMR (base, 75 MHz, DMSO-*d*₆) δ 152.2, 136.3, 134.7, 132.5, 131.5, 127.4, 126.8, 126.3, 125.7, 125.6, 121.5, 119.4, 61.9, 60.9, 52.4, 43.8, 34.5, 28.8; Anal. Calcd for C₁₈H₁₉NO₂·HCl·H₂O (salt): C, 64.38; H, 6.50; N, 4.17. Found: C, 63.83; H, 6.01; N, 4.09.
- Compound 6: (free base) 160–161 °C; ¹H NMR (base, 300 MHz, CDCl₃) δ 8.16 (d, *J* = 7.8 Hz, 1H), 7.25 (t, *J* = 7.5 Hz, 1H), 7.05 (d, *J* = 7.5 Hz, 1H), 6.83 (d, *J* = 7.5 Hz, 1H), 6.72 (d, *J* = 7.8 Hz, 1H), 4.85 (ab, *J* = 12.9 and 36.0 Hz, 2H), 3.35–2.46 (m, 9H), 1.60 (m, 2H), 0.96 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (base, 75 MHz, CDCl₃) δ 153.8, 138.4, 133.1, 131.8, 127.7, 126.6, 126.3, 126.1, 124.5, 122.3, 119.6, 119.2, 65.0, 59.7, 56.5, 49.1, 35.0, 29.2, 19.5, 12.3; Anal. Calcd for C₂₀H₂₃NO₂: C, 77.64; H, 7.49; N, 4.53. Found: C, 77.03; H, 7.52; N, 4.50.
- Kula, N. S.; Baldessarini, R. J.; Kebabian, J. W.; Bakthavachalam, V.; Xu, L. *Eur. J. Pharmacol.* **1997**, *331*, 333.
- Menon, M. K.; Clark, W. G.; Neumeyer, J. L. *Eur. J. Pharm.* **1978**, *52*, 1.