

8-Piperazinyl-2,3-dihydropyrrolo[3,2-g]isoquinolines: Potent, selective, orally bioavailable 5-HT₁ receptor ligands

Tom D. Heightman,^{a,*} Laramie M. Gaster,^a Sarah L. Pardoe,^a Jean-Pierre Pilleux,^b Michael S. Hadley,^b Derek N. Middlemiss,^b Gary W. Price,^b Claire Roberts,^b Claire M. Scott,^b Jeannette M. Watson,^b Laurie J. Gordon,^c Vicky A. Holland,^c Jenifer Powles,^c Graham J. Riley,^c Tania O. Stean,^d Brenda K. Trail,^d Neil Upton,^d Nigel E. Austin,^b Andrew D. Ayrton,^e Tanya Coleman^d and Leanne Cutler^d

^aHigh Throughput Chemistry, Discovery Research, GlaxoSmithKline, New Frontiers Science Park, Third Avenue, Harlow Essex CM19 5AW, UK

^bPsychiatry Centre of Excellence for Drug Discovery, GlaxoSmithKline, New Frontiers Science Park, Third Avenue, Harlow Essex CM19 5AW, UK

^cDiscovery Research Biology, GlaxoSmithKline, New Frontiers Science Park, Third Avenue, Harlow Essex CM19 5AW, UK

^dNeurology and GI Centre of Excellence for Drug Discovery, GlaxoSmithKline, New Frontiers Science Park, Third Avenue, Harlow Essex CM19 5AW, UK

^eGlaxoSmithKline Drug Metabolism and Pharmacokinetics, Ware SG12 0DP, UK

Received 11 May 2005; revised 9 June 2005; accepted 9 June 2005

Available online 21 July 2005

Abstract—The novel 8-piperazinyl-2,3-dihydropyrroloisoquinoline template was synthesized in nine steps. The template was *N*-substituted to give a series of compounds showing binding to human cloned 5-HT_{1A}, 5-HT_{1B} and 5-HT_{1D} receptors with pK_i's greater than 9 and selectivities up to 1000-fold against other serotonin, dopamine and adrenergic receptors. Several compounds were shown to possess weak partial agonist activity in cloned receptors, which translated to antagonism in *in vitro* studies.

© 2005 Elsevier Ltd. All rights reserved.

Although the precise mechanism of antidepressant action remains poorly understood, many currently prescribed treatments for depression are believed to enhance neurotransmission by increasing extracellular levels of serotonin (5-hydroxytryptamine, 5-HT) in the brain.¹ Neuronal 5-HT release is modulated by presynaptic inhibitory 5-HT₁ autoreceptors. 5-HT_{1A} receptors are mainly somatodendritic, and cause a reduction in the rate of cell firing and 5-HT release when stimulated.² 5-HT_{1B} receptors are mainly located on cell terminals, where they mediate a reduction in the amount of 5-HT released on each firing event.³ The role of terminal 5-HT_{1D} receptors is less clear, although there is recent evidence to suggest that they play a role in modulating extracellular brain 5-HT levels in guinea pig.⁴ The com-

bined activation of all three receptor subtypes by extracellular 5-HT thus has the potential to cause a decrease in the amount and frequency of 5-HT released into the synaptic cleft. Stimulation of these autoreceptors may be responsible for the delayed onset of action of most antidepressants which work wholly or partly by inhibiting 5-HT reuptake.⁵ The two to four week onset time for these antidepressants is consistent with the time taken for these autoreceptors to desensitize in rats upon treatment with fluoxetine or citalopram.⁶ Therefore, a drug which antagonizes all 5-HT₁ inhibitory autoreceptors simultaneously should acutely mimic their chronic desensitization, and thereby lead to an immediate and sustained increase in levels of synaptic 5-HT for each transmission event.⁵ Such a drug could well provide a much needed rapidly acting antidepressant.

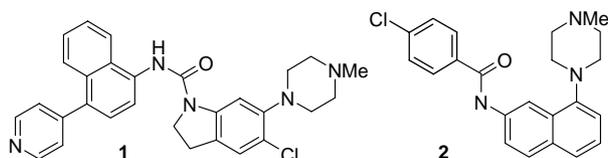
We have previously reported the *in vitro* pharmacology of SB-272183, the first selective 5-HT_{1A/B/D} receptor antagonist.⁷ In this paper we describe the synthesis

Keywords: 5-HT_{1A}; 5-HT_{1B}; 5-HT_{1D}; Receptor antagonist.

* Corresponding author. Tel.: +44 1279 622031; fax: +44 1279 627779; e-mail: Tom.D.Heightman@gsk.com

and pharmacology of a new series of highly potent and selective 5-HT_{1A/B/D} receptor partial agonists/antagonists.

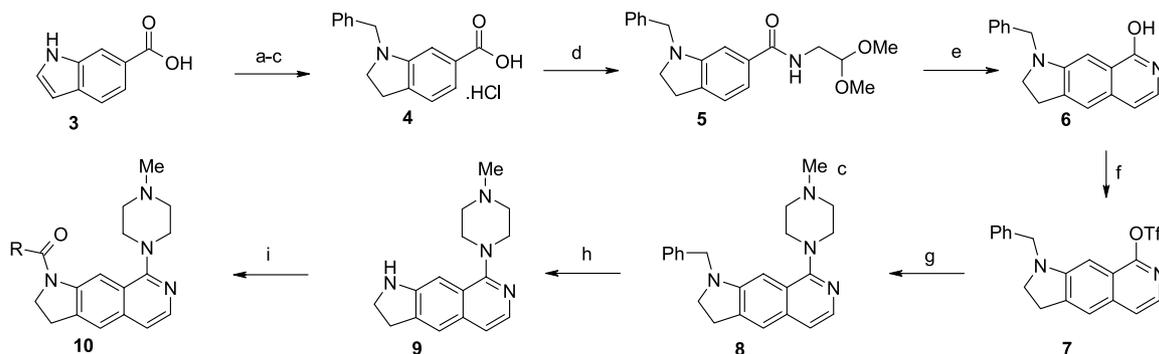
The novel dihydropyrrolo[3,2-*g*]isoquinoline template **10** was proposed from a pharmacophore alignment of the indoline SB-272183 (**1**) and a series of naphthylpiperazines,⁸ e.g., **2**, with reported 5-HT₁ p*K*_i's > 6.2.



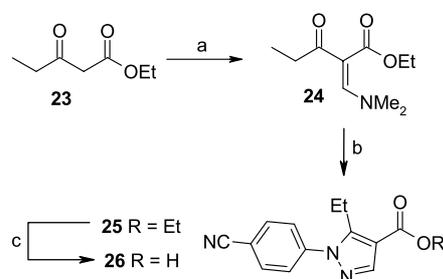
The synthesis is illustrated in Scheme 1. Indole-6-carboxylic acid (**3**) was converted to the *N*-monobenzylated indoline **4** by dibenylation; reduction of the five-membered ring; and saponification of the ester, all under standard conditions.

To construct the pyridyl ring, a procedure related to the Pomeranz–Fritsch method was used via the intermediate 2,2-dimethoxyethyl amide **5**. Treatment of **5** with Brønsted acids (H₂SO₄, TFA or HCl) gave rise to a mixture of cyclized regioisomers, presumably by partially favouring *N*-protonation of the indoline leading to intramolecular delivery of the proton to the acetal, so that the reactive oxonium species is formed in proximity to the 7-position of the indoline. The use of boron trifluoride-diethyl etherate in refluxing THF gave a single regioisomer **6**, easily distinguished from the alternative regioisomer by the presence of two new singlets in the aromatic region of the ¹H NMR. This is consistent with selective boron complexation of the acetal oxygen atoms, leading to formation of the reactive oxonium intermediate in the less hindered region of the molecule, and favouring reaction at the 5-position of the indoline. Attempts to cyclize analogues of **5** with *N*-acyl protecting groups were not successful.

The hydroxy group of isoquinoline **6** was readily activated by triflation, and displaced by *N*-methyl piperazine. Debenylation was effected by catalytic hydrogenation under strongly acidic conditions, giving the key intermediate **9**, which was coupled to form amides and ureas under standard conditions.



Scheme 1. Reagents: (a) NaH, BnBr, NMP, 90%; (b) NaBH₃CN, AcOH, 96%; (c) aq NaOH, dioxane, quant.; (d) EDC, HOBT, DMF, 98%; (e) BF₃·OEt₂, THF, 87%; (f) Tf₂O, pyridine, 63%; (g) *N*-methyl piperazine, 93%; (h) H₂/Pd/C, MeOH, HCl, 81%; (i) RCO₂H, DIC, HOBT, DMF.



Scheme 2. Reagents and conditions: (a) DMFDMA, DMF, 120 °C, quant.; (b) 4-cyanophenyl hydrazine, EtOH, 97%; (c) NaOH aq, dioxane.

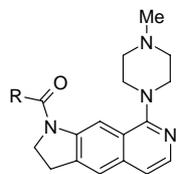
Analogues of the *N*-methyl piperazine moiety were introduced into intermediate triflate **7** and further derivatized using analogous procedures to those outlined in Scheme 1. The 4-cyanophenyl pyrazole carboxylic acid intermediate **26** used in the synthesis of **22** was prepared by combination of 4-cyanophenyl hydrazine with the dimethylaminomethylene oxopentanoate ester **24**⁹ followed by saponification, as per Scheme 2.

Compounds were tested for their activity at 5-HT₁ receptors using standard binding assays with [³H]5-HT or [³H]8-OH-DPAT as the radioligand (Table 1). Substituted benzamides showed clear SAR with 4-substituents showing higher affinity than two or three isomers. Heterocycles were also tolerated in this position with some loss of potency, but this was restored with biaryl groups.

The *N*-methyl piperazinyl group proved relatively insensitive to steric modifications: removal of the methyl group (**31**) resulted in only a slight loss of potency, but a dramatic loss of selectivity against the β₂ adrenoreceptor (Table 2).

Increasing the size of the *N*-alkyl substituent caused the affinity to diminish incrementally (**32**, **33**). The 3,5-dimethylpiperazine **34** showed somewhat reduced activity particularly at the 5-HT_{1A} receptor, but replacement of the *N*-methyl piperazine with either enantiomer of diazabicyclooctane was well tolerated (**35**, **36**).

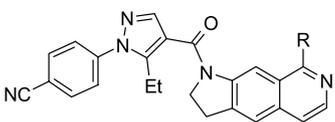
Compounds in this series showed a wide range of efficacies vs 5-HT for 5-HT_{1A} and 5-HT_{1B} receptor subtypes

Table 1. Receptor binding affinities of reference compound **1** and amide derivatives **11–22** in the 1-(4-methyl piperazinyl) series

Compound	R	pK_i^a			Selectivity ^b
		5-HT _{1A}	5-HT _{1B}	5-HT _{1D}	
1	—	8.0 ± 0.1	8.1 ± 0.1	8.7 ± 0.1	30
11		9.1 ± 0.1	9.5 ± 0.1	9.3 ± 0.1	320
12		9.0 ± 0.1	9.3 ± 0.1	9.1 ± 0.1	>1000
13		8.0 ± 0.1	7.8 ± 0.1	7.8 ± 0.1	30
14		8.3 ± 0.2	8.9 ± 0.1	8.6 ± 0.1	60
15		8.5 ± 0.1	9.6 ± 0.2	9.3 ± 0.1	400
16		8.4 ± 0.2	9.1 ± 0.2	8.4 ± 0.2	130
17		8.3 ± 0.1	8.7 ± 0.2	8.1 ± 0.2	250
18		8.2 ± 0.3	7.8 ± 0.2	8.6 ± 0.2	30
19		8.2 ± 0.2	8.3 ± 0.1	8.0 ± 0.1	160
20		9.5 ± 0.1	9.0 ± 0.1	8.9 ± 0.2	250
21		9.2 ± 0.4	9.2 ± 0.3	9.2 ± 0.4	500
22		8.9 ± 0.3	9.0 ± 0.4	9.2 ± 0.3	160

^a Values are means of at least three experiments.

^b Selectivity versus 5-HT_{2A,2B,2C}, 6 and 7; dopamine D₂, D₃; adrenergic α1b and β2.

Table 2. Receptor binding affinities of amino group analogues **31–36** from the cyanophenyl pyrazole series


Compound	R	p <i>K</i> _i ^a			Selectivity ^b
		5-HT _{1A}	5-HT _{1B}	5-HT _{1D}	
31		8.9 ± 0.3	8.6 ± 0.3	8.6 ± 0.2	4
32		8.5 ± 0.1	8.8 ± 0.1	8.6 ± 0.1	320
33		8.2 ± 0.1	8.4 ± 0.1	8.1 ± 0.1	100
34		7.5 ± 0.1	8.1 ± 0.1	8.1 ± 0.1	320
35		8.8 ± 0.1	9.0 ± 0.2	9.0 ± 0.1	200
36		8.9 ± 0.1	8.7 ± 0.1	8.5 ± 0.2	100

^a Values are means of at least three experiments.

^b Selectivity versus 5-HT_{2A,2B,2C}, 6 and 7; dopamine D₂, D₃; adrenergic α1b

in the GTPγS functional assay⁷ (Table 3). For both aryl and heteroaryl compounds, a *para*-substituent tended to engender higher intrinsic activity, whilst an *ortho*-substituent adjacent to the linking carboxamide showed lower intrinsic activity approaching silent antagonism, suggesting a role for the torsional angle of the ring–C=O bond. Despite this intrinsic activity in the recombinant GTPγS assay, compounds with intrinsic activities below ~0.5 behaved as antagonists in 5-HT_{1A} and 5-HT_{1B} receptor-mediated native tissue assays. Hence, the phenyl pyrazole **22** reversed the inhibition of rat dorsal raphe nucleus cell firing by the 5-HT_{1A} agonist 8-OH-DPAT (pA₂ ~ 7); and it potentiated the electrically stimulated release of 5-HT from guinea pig cortex slices, which has been previously demonstrated to be mediated by the 5-HT_{1B} receptor.⁷

After intravenous infusion, compounds in this series showed a wide range of clearance rates in rat, from less than 25% of liver blood flow to approximately 100%, and an equally broad range of brain penetration (Table 4).¹⁰ The 4-cyanobenzamide **12** showed a lower rate of clearance but also lower brain penetration than the isosteric 4-chlorobenzamide **11**, consistent with its lower lipophilicity. The quinolinyl amide **16** showed rapid clearance

Table 3. Partial agonist efficacy of analogues **11–14**, **21** and **22** in the 5-HT_{1A} and 5-HT_{1B} GTPγS functional assays

Compound	GTPγS	
	5-HT _{1A}	5-HT _{1B}
11	0.69 ± 0.03	0.75 ± 0.06
12	0.84 ± 0.05	0.86 ± 0.04
13	Inactive	0.23 ± 0.02
14	0.42 ± 0.04	0.42 ± 0.06
21	0.54 ± 0.05	0.59 ± 0.04
22	0.33 ± 0.03	0.47 ± 0.03

Values expressed as fraction of 5-HT efficacy.

Table 4. In vivo pharmacokinetics (rat, *n* = 1)¹⁰

Compound	CLb (ml/min/kg)	Brain: blood	Fpo (%)
11	29	1.3	ND ^a
12	19	0.7	ND
16	75	0.2	ND
18	39	0.5	55
19	48	0.1	ND
21	32	1.1	40
22	28	0.5	50

^a Not determined.

and limited brain penetration. In contrast, the smaller heterocycles showed more acceptable clearance; however whilst the dimethyloxazole **18** showed good brain penetration, the closely related methyl-trifluoromethyl pyrazole **19** did not. The addition of the phenyl ring restored the brain penetration and provided compounds with low to moderate clearance and promising oral bioavailability. In particular, compound **22** showed moderate to good bioavailability across species: rat 50%, guinea pig 20%, dog 89% and cynomolgous monkey 58%.

Compound **22** was further studied in in vivo pharmacodynamic models based on 5-HT_{1B} receptor antagonism. In rats, **22** reversed the effects of the selective 5-HT_{1B} receptor agonist SKF-99101 on electroshock seizure threshold, with an oral ID₅₀ of 3.5 ± 1.3 mg/kg.¹¹ In guinea pigs, **22** reversed SKF-99101-induced hypothermia with an oral ID₅₀ of 0.2 mg/kg.¹² Compound **22** and related compounds are under further investigation in preclinical models of depression.

References and notes

- Cowen, P. J. In *Selective Serotonin Re-uptake Inhibitors*; Feighner, J. P., Boyer, W. F., Eds.; John Wiley & Sons: Chichester, 1996, pp 63–86.

2. (a) Craven, R.; Grahame-Smith, D.; Newberry, N. *Eur. J. Pharmacol.* **1994**, *271*, R1; (b) Haj-Dahmane, S.; Hamon, M.; Lanfumey, L. *Neuroscience* **1991**, *41*, 495; (c) Sprouse, J. S.; Aghajanian, G. K. *Synapse* **1987**, *1*, 3.
3. (a) Roberts, C.; Watson, J.; Burton, M.; Price, G.; Jones, B. *Br. J. Pharmacol.* **1996**, *117*, 384; (b) Engel, G.; Gothert, M.; Hoyer, D.; Schlicker, E.; Hillenbrand, K. *Naunyn-Schmiedberg's Arch. Pharmacol.* **1986**, *332*, 1.
4. (a) Bonaventure, P.; Voorn, P.; Luyten, W. H. M. L.; Jurzak, M.; Schotte, A.; Leysen, J. E. *Neuroscience* **1998**, *9*, 641; (b) Bonaventure, P.; Langlois, X.; Leysen, J. E. *Neurosci. Lett.* **1998**, *254*, 113.
5. (a) Cryan, J. F.; Leonard, B. E. *Human Psychopharmacology* **2000**, *15*, 113; (b) Artigas, F. *TIPS* **1993**, *14*, 262.
6. Czachura, J. F.; Rasmussen, K. *Naunyn-Schmiedberg's Arch. Pharmacol.* **2000**, *362*, 266; Moret, C.; Briley, M. *Eur. J. Pharmacol.* **1990**, *180*, 351.
7. Procedures for cell firing and electrically stimulated release described in: Watson, J.; Roberts, C.; Scott, C.; Kendall, I.; Collin, L.; Day, N.; Harries, M. H.; Soffin, E.; Davies, C. H.; Randall, A. D.; Heightman, T. D.; Gaster, L.; Wyman, P.; Parker, C.; Price, G. W.; Middlemiss, D. N. *Br. J. Pharmacol.* **2001**, *133*, 797.
8. Chenard, B.L.; Macor, J.E.; Segelstein, B.E.; *PCT Int. Appl.* WO9421619; Chem. Abstr. **1994**, *122*, 314570.
9. Menozzi, G.; Mosti, L.; Schenone, P. *J. Heterocycl. Chem.* **1987**, *24*, 1669.
10. *DMPK procedures*. Clearance and brain penetration determination, iv dosing in male Sprague–Dawley rats, constant infusion over 1 h to achieve target dose of 1 mg free base/kg; oral dosing, solution or suspension administered at target dose of 3 mg free base/kg. Blood and brain homogenate samples analysed by HPLC/MS/MS.
11. Procedures for electroshock seizure threshold experiments described in: Stean, T. O.; Atkins, A. R.; Heidbreder, C. A.; Quinn, L. P.; Trail, B. K.; Upton, N. *Br. J. Pharmacol.* **2005**, *144*, 628.
12. Procedure for SKF-99101-induced hypothermia described in: Hagan, J. J.; Slade, P. D.; Gaster, L.; Jeffrey, P.; Hatcher, J. P.; Middlemiss, D. N. *Eur. J. Pharmacol.* **1997**, *331*, 169.