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Effects of positional and geometrical isomerism on the biological activity of some novel oxazolidinones $^{\bigstar, \overleftrightarrow \overleftrightarrow}$

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Abstract—Some novel oxazolidinone derivatives with benzotriazole as pendant have been synthesized and tested for antibacterial activity. Linearly attached benzotriazole derivative showed more potency compared to angular one in vitro. Out of E/Z-isomers of angularly attached derivatives E-isomer was found to be more potent than Z-isomer. Either less active or inactive molecules were obtained, when benzotriazole was replaced with benzimidazole, benzthiazole, or benzoxazole. Finally, thioacetamide analogue of linear compound gave a lead having activity similar to linezolid in vitro.

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1. Introduction

Tuberculosis is one of the leading causes of death due to infection worldwide. Today, nearly one third of world population is infected with *Mycobacterium tuberculosis*; and according to WHO report this number is increasing every year.¹ Both the developed and developing countries are facing a challenge to combat this disease. The complexity is multifold in the case of immunocompromised patients acquiring *Mycobacterium avium* complex (MAC) infections. The emergence of multidrug resistant (MDR) *M. tuberculosis* poses a serious threat to the current therapies used for the treatment of tuberculosis. Therefore, there is an urgent need to develop novel drugs for tuberculosis.

Oxazolidinones, a completely synthetic antibacterial class, elicited lots of interest amongst medicinal chemists because of its activity against resistant Gram-positive

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organisms such as methicillin-resistant *Staphylococcus aureus* (MRSA), vancomycin-resistant *Enterococci* (VRE), and glycopeptide intermediate resistant *S. aureus* (GISA). Oxazolidinones bind selectively to 50S ribosomal subunit thereby inhibiting bacterial protein biosynthesis at an early stage.² Linezolid is the first drug from this class of compounds launched in 2000 by Pharmacia. Subsequently it was tested against 249 clinical isolates of rapidly growing mycobacteria and MIC of 4μ g/mL was reported against most of the strains studied.³ However, thiomorpholine analogue of linezolid, PNU 100480 (Fig. 1) was found to be more potent having MIC of 0.125μ g/mL against mycobacterial strains.⁴

We have a long-standing interest to develop drugs for tuberculosis and oxazolidinone class fits well into our objectives for its unique mechanism of action. Moreover oxazolidinones are completely synthetic class of molecules having less possibility of developing resistance.⁵

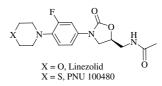


Figure 1.

Keywords: Oxazolidinones; *E–Z*-isomers; Positional isomers; Antibacterial; Benzotriazole.

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X,Y,Z = N, C; M = 2 H, CR₂, CHR, PhCH, p-NO₂C₆H₄CH, p-NR₂C₆H₄CH etc.

Figure 2.

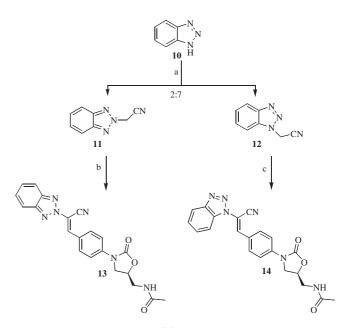
Literature search revealed the following common structures having antituberculosis activity (Fig. 2).^{6,7}

The combination of two antibacterial agents with different mechanisms of action into a single entity to achieve drugs with dual action has received considerable attention in the literature.⁸ We deliberated on the above chemical structures and decided to hybridize them with oxazolidinones to explore the antituberculosis activity of the resulting molecules (Fig. 3).

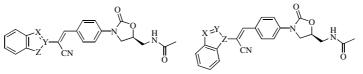
2. Chemistry

The key step involved in the synthesis of this series of compounds is the Knoevenagel condensation of aldehyde 1 with acetonitrile derivative 2 to give the hybrid molecule 3 (Scheme 1).

Synthesis of aldehyde **1** has been depicted in Scheme 2. Reduction of both nitro and aldehyde groups of 4-nitrobenzaldehyde was achieved with NiCl₂ and NaBH₄ to obtain the amino alcohol **5**. Protection of amine as benzylcarbamate and alcohol as TBDMS ether gave



Scheme 3. Reagents and conditions: (a) ClCH₂CN, TEA, DMF, 110 °C, 50%; (b) ald. 1, benzoic acid, piperidine, toluene, reflux, 40%; (c) same as (b), 50%.

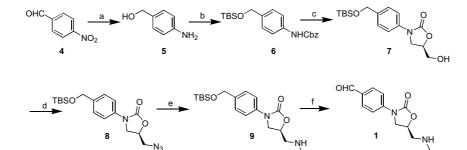


X,Y,Z = N, C, O, S

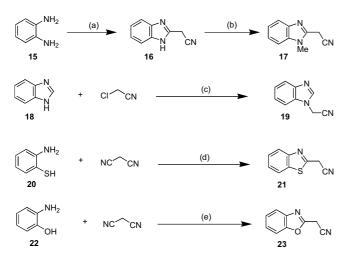


Scheme 1.

Figure 3.



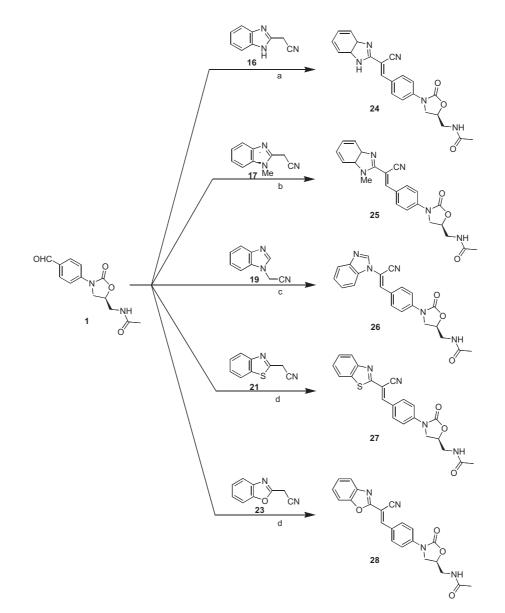
Scheme 2. Reagents and conditions: (a) NaBH₄, NiCl₂, 71%; (b) 1. Cbz–Cl, NaHCO₃, 2. TBDMS–Cl, Im, 43%; (c) *n*-BuLi, (*R*)-glycidyl butyrate, -78 °C to rt, 55%; (d) 1. MsCl, TEA, 2. NaN₃, 76%; (e) MeCOSH, 66%; (f) 1. TBAF, 2. PDC, 63%.



Scheme 4. Reagents and conditions: (a) CNCH₂COOH, 4N HCl, 50%; (b) K_2CO_3 , DMF, MeI, 75%; (c) TEA, DMF, 50%; (d) acetic acid/ethanol, 20%; (e) same as (d), 35%.

compound **6**. This compound was converted to **9** following well known methods in the literature.⁹ Deprotection of TBDMS ether and subsequent oxidation of the resulting alcohol with PDC produced the required aldehyde **1**.

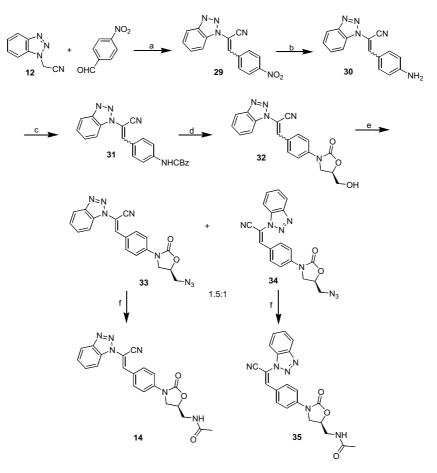
Initially we decided to synthesize benzotriazole derivatives 3 (Scheme 1, X = Y = Z = N) because both the regioisomers of benzotriazole 11 and 12 are readily available and a systematic study of *E*- and *Z*-isomers involving 11 and 12 with substituted benzaldehydes have been carried out by Sanna et al.⁷ Acetonitrile derivatives 11 and 12 were obtained from benzotriazole and chloroacetonitrile in 2:7 ratio.⁷ These two compounds were separated by column chromatography and spectral data were compared with the reported values. Compounds 11 and 12 were then condensed with aldehyde 1 under Knoevenagel conditions to give 13 and 14, respectively, and only *E*-isomers were isolated in both the examples (Scheme 3).



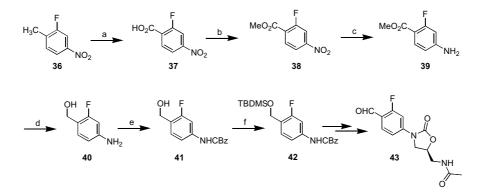
Scheme 5. Reagents and conditions: (a) KF, dry ethanol, 52%; (b) same as (a), 41%; (c) TEA, toluene, reflux, 50%; (d) CsF, dry ethanol, 15%.

Various acetonitrile derivatives were then prepared for condensation with aldehyde 1 (Scheme 4). 1,2-Phenylenediamine was converted to C-linked imidazol derivative 16 by heating with cyanoacetic acid in 4N HCl, which was further methylated with methyl iodide to give compound 17. N-Linked imidazol derivative 19 was prepared from imidazol and chloroacetonitrile in DMF with triethyl amine as base. Benzthiazole derivative 21 and benzoxazole derivative 23 were synthesized from *o*-aminothiophenol **20** and *o*-aminophenol **22**, respectively, with malononitrile in ethanol and acetic acid.

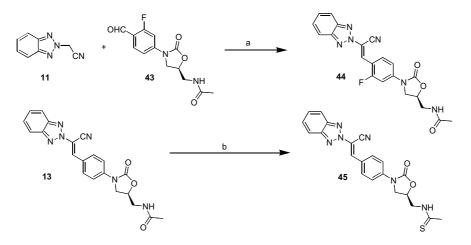
Condensation of acetonitrile derivatives with aldehyde **1** under Knoevenagel condition was tried with different base/solvent combinations. However, Knoevenagel condensation was base sensitive and a single base/solvent combination could not be utilized for all the conversions.¹⁰ Compound **16** was condensed with aldehyde **1**



Scheme 6. Reagents and conditions: (a) TEA, toluene, 50%; (b) NiCl₂/NaBH₄, 84%; (c) CbzCl, NaHCO₃, 57%; (d) *n*-BuLi, (*R*)-glycidyl butyrate, $-78 \degree$ C to rt, 50%; (e) 1. MsCl, TEA, 2. NaN₃, DMF, 51%; (f) CH₃COSH, 65%.



Scheme 7. Reagents and conditions: (a) Na₂Cr₂O₇, H₂SO₄, 90 °C, 70%; (b) MeOH, H₂SO₄, 93%; (c) PtO₂/H₂, 95%; (d) LAH, 60%; (e) CbzCl, NaHCO₃, 93%; (f) TBDMSCl, TEA, DMAP, 63%.



Scheme 8. Reagents and conditions: (a) KF/dry ethanol, 15%; (b) Lawesson's reagent, dioxane, 41%.

to produce 24 in dry ethanol and KF. Similarly compound 17 gave 25 under identical conditions whereas compound 26 was obtained from 19 and aldehyde 1 with triethylamine as base. Compounds 21 and 23 could not be condensed under both the conditions. However, compounds 27 and 28 could be obtained in modest yields on treatment with CsF in dry ethanol (Scheme 5).

Z-Isomer of 14 was prepared following the sequence described in Scheme 6. Knoevenagel condensation of 12 with 4-nitrobenzaldehyde gave a mixture of *E*- and *Z*-isomers 29. The mixture of isomers was reduced to the corresponding mixture of amines 30 with NiCl₂·6H₂O and NaBH₄. Isomeric amines 30 were converted to corresponding azidomethyl oxazolidinones 33 and 34 in 3:2 ratio following the known sequence of reactions (Scheme 2); and at this stage isomers could be separated by column chromatography. Finally, both *E*- and *Z*-isomers 14 and 35 were obtained from azides 33 and 34, respectively, on treatment with thiolacetic acid.

Synthesis of fluoro derivative of 13 is described in Scheme 7. 2-Fluoro-4-nitro toluene 36 was oxidized to 2-fluoro-4-nitro benzoic acid 37, which was then esterified with methanol to give 38. Reduction of both ester and nitro groups of 38 to amino alcohol 40 in one pot could not be accomplished. Therefore reduction of nitro group to amine 39 was carried out with PtO_2/H_2 and then to alcohol 40 by lithium aluminum hydride. Amino group of compound 40 was protected as benzylcarbamate (41) and hydroxy group as TBDMS ether to give compound 42. Aldehyde 43 was then obtained following the same procedure for aldehyde 1.

Condensation of acetonitrile derivative 11 with aldehyde 43 in presence of KF in dry ethanol yielded 44, the fluoro analogue of 13. In a separate reaction, 13 was converted to thioacetamide derivative 45 using Lawesson's reagent (Scheme 8).

It is our observation that Knoevenagel condensation of aromatic aldehydes with the acetonitrile derivatives discussed here produced in most of the cases the *E*-isomer as the only isolable product. *E*-Isomers have been characterized by higher δ values for vinylic protons⁷ in ¹H NMR and also by ${}^{3}J_{C-H}$ (13–14Hz) coupling for vinylic H and nitrile C by using the 'gate decoupling' technique.¹¹ The ¹H NMR values for vinylic protons of all the compounds have been compiled in Table 1. The vinylic proton peaks in ¹H NMR appeared at δ 8.21– 8.70 for all the *E*-isomers in accordance with the reported values except compound 14. In this case the vinylic proton appeared at δ 6.08. The upfield shift of vinylic proton for compound 14 was abnormal compared to other *E*-isomers. However, the vinylic proton for *Z*-isomer was found to appear at δ 6.22 as expected. The reason for the abnormal behavior of compound 14 is not clear to us at this moment. In order to prove that compound 14 is really E-isomer we have isolated both Eand Z-isomers from mixtures of 29 (Scheme 6) by careful column chromatography along with mixture of Eand Z-isomers and identified them on the basis of spectral data reported.⁷ Both the isomers were then converted to 14 and 35 following the same sequence of reactions as depicted in Scheme 6 independently.

3. Biology

All the compounds listed in Table 1 were tested against *Mycobacterium smegmatis* MTCC 006 in addition to the panel of organisms consisted of six reference Gram-positive bacteria including both sensitive and resistant strains: *S. aureus* ATCC 29213 (MSSA), *S. aureus*

Table 1. ¹H NMR (δ) values for vinylic protons

Entry	Compd no	δ values		
1	13	8.55		
2	14	6.08		
3	35	6.22		
4	24	8.31		
5	25	8.22		
6	26	8.45		
7	27	8.21		
8	28	8.32		
9	44	8.70		
10	45	8.52		

Entry	Compd		MIC (µg/mL)						
	no	structure	M.sm ^a 006	<i>S.a</i> 29213	<i>S.a</i> 49951	<i>S.a</i> 33591	<i>E.f</i> 29212	<i>E.f</i> 12201	E.fm 12202
1	13	$ \begin{array}{c} & & \\ & & $	>32	8	8	8	8	8	8
2	14	$ \underset{H}{\overset{N \in \mathbb{N}}{\longrightarrow}} \underset{H}{\overset{C N}{\longrightarrow}} \underset{N \to \mathbb{N}}{\overset{O}{\longrightarrow}} \underset{N \to \mathbb{N}}$	>32	16	16	16	32	32	32
3	24	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \\ \end{array} \\ \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \\ \end{array} \\ \\ \\ \\ $	>32	>32	>32	>32	>32	>32	>32
4	25	Me H O H	>32	>32	>32	>32	>32	>32	>32
5	26		>32	16	16	16	32	32	32
6	27	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \\ $	>32	>32	>32	>32	>32	>32	>32
7	28	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \\ \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} $	>32	>32	>32	>32	>32	>32	>32
8	35		>32	>32	>32	>32	>32	>32	>32
9	44	$ \begin{array}{c} H \\ H $	>32	8	16	16	32	32	32
10	45	$ \begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \\ \end{array} \\ \\ \\ \\ \\ \\ \\ \end{array} \\$	>32	2	2	2	4	4	4
11 12		Linezolid Streptomycin	2 0.5	2 4	2 2	2 >32	2 >32	2 >32	2 32

Table 2. In vitro potency of oxazolidinones

^a *M.sm* 006: *Mycobacterium smegmatis* MTCC 006; *S.a* 29213: *S. aureus* ATCC 29213 (MSSA); *S.a* 49951: *S. aureus* ATCC 49951 (Smith strain); *S.a* 33591: *S. aureus* ATCC 33591 (MRSA); *E.f* 29212: *E. faecalis* ATCC 29212; *E.f* 12201: *E. faecalis* NCTC 12201 (VREf); *E.fm* 12202: *E. faecium* NCTC 12202 (VREfm).

ATCC 49951 (Smith), S. aureus ATCC 33591 (MRSA), E. faecalis ATCC 29212, E. faecalis NCTC 12201 (VREf) E. faecium NCTC 12202 (VREfm). MIC was determined by brothmicro dilution as per the guidelines prescribed by NCCLS.¹² Doubling dilution of the test compound, in the range of $32-1\,\mu$ g/mL, was carried out in Mueller Hinton Medium (Difco). The organisms were inoculated to obtain a final concentration of $1-5 \times 10^5$ cfu/mL and the microtiter plates were incubated overnight, in ambient air, at $35\,^{\circ}$ C. Linezolid and streptomycin were included as controls.

4. Results

Our hypothesis to hybridize oxazolidinones with fragments having antituberculosis activity to achieve better molecules for the treatment of tuberculosis was found to be illusive. None of the compounds tested showed antibacterial activity against *M. smegmatis* (Table 2). However, some of these compounds were found to be active against the Gram-positive strains mentioned above. Benzotriazole derivative 13, having attachment through middle nitrogen (linear attachment), exhibited moderate activity (MIC, 8µg/mL). On the other hand compound 14, a positional isomer of 13 (having attachment through terminal nitrogen, i.e., angular attachment), was found to be less active than the linear one. C-Linked benzimidazole derivative 24 and its methyl derivative 25 did not show antibacterial activity against the strains tested whereas N-linked benzimidazole derivative 26 retained the activity of analogues compound 14. Derivatives of benzthiazole 27 and benzoxazole 28 were found to be inactive against the strains tested. Compound 35, Z-isomer of 14 was found to be inactive too. It is a general observation in oxazolidinone series that replacement of hydrogen of central aromatic ring with fluorine, having meta relationship with oxazolidinone group, is beneficial as far as MIC is concerned. However, in this series compound 44, fluoro analogue of 13, showed reduced antibacterial activity. On the other hand, thioacetamide analogue (45) of 13 was found to poses good antibacterial activity (MIC, 2- $4\mu g/mL$) and was the most potent amongst the molecules screened.

5. Conclusion

We have synthesized various oxazolidinone derivatives having positional and geometrical substitutions with benzotriazole, benzimidazole, benzthiazole, and benzoxazole. Contrary to our expectations none of the compounds showed antimycobacterial activity. However, some of them showed antibacterial activity against Gram-positive pathogens. Compound having linear attachment with benzotriazole was found to be the most potent among the compounds tested. Deviation from the position of attachment or the geometry of the molecule has dramatic effect on the activity. Heterocycle, other than benzotriazole, was also not tolerated. Finally, a lead molecule having activity comparable to linezolid has been identified.

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