



Pergamon

SCIENCE @ DIRECT®

Tetrahedron Letters 44 (2003) 8887–8891

TETRAHEDRON  
LETTERS

# Synthesis of an external $\beta$ -turn based on the GLDV motif of cell adhesion proteins

David E. Davies,<sup>a</sup> Paul M. Doyle,<sup>b</sup> R. Duncan Farrant,<sup>a</sup> Richard D. Hill,<sup>c</sup> Peter B. Hitchcock,<sup>c</sup>  
Paul N. Sanderson<sup>a</sup> and Douglas W. Young<sup>c,\*</sup>

<sup>a</sup>GlaxoSmithKline, Medicines Research Centre, Gunnels Wood Road, Stevenage, Herts SG1 2NY, UK

<sup>b</sup>BioFocus Discovery PLC, Sittingbourne Research Centre, Sittingbourne, Kent ME9 8AZ, UK

<sup>c</sup>Sussex Centre for Biomolecular Design and Drug Development, Department of Chemistry, University of Sussex, Falmer, Brighton BN1 9QJ, UK

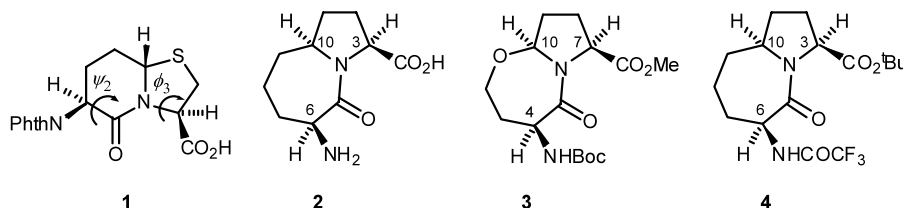
Received 18 August 2003; revised 8 September 2003; accepted 19 September 2003

**Abstract**—The (3*S*,6*S*,10*S*)-7/5 bicyclic lactam **4**, designed as an external turn constraint, was synthesised by a new stereoselective route involving Eschenmoser condensation. Calculated preferred conformations compare well with the preferred solid state conformation, obtained by X-ray crystallography. The lactam **4** was not a turn mimic in its own right but could be used as an external constraint to prepare the cyclic peptide **29** containing the integrin recognition motif GLDV. High-resolution NMR measurements were consistent with this compound having a single backbone conformation.

© 2003 Elsevier Ltd. All rights reserved.

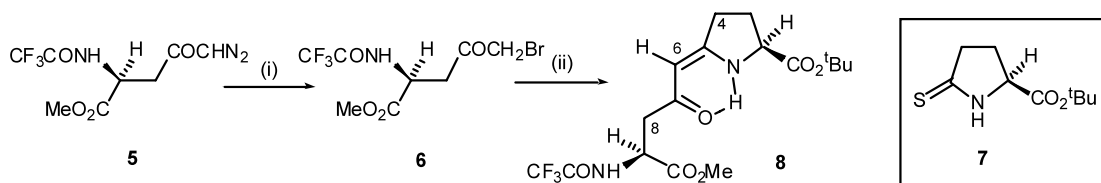
Reverse turns connect elements of protein secondary structure and usually occur on the surface of the protein. They can, therefore, be important in molecular recognition processes.<sup>1,2</sup> Synthetic molecules which mimic these turns, such as the classic 5/6 bicyclic lactam BTD, **1**, prepared by Nagai,<sup>3–5</sup> have been used to replace  $\beta$ -turns in various proteins without affecting their overall biological properties.<sup>4–6</sup> BTD, **1**, has been shown to be a type II'  $\beta$ -turn mimic from the angles  $\psi_2$  ( $-161^\circ$ ) and  $\phi_3$  ( $-69.4^\circ$ ) in the X-ray crystal structure.<sup>7</sup> One of us has used BTD<sup>8</sup> as a strong structural constraint on which to build a cyclic GLDV motif. This was shown to be a type I  $\beta$ -turn and inhibited the interaction of the integrin  $\alpha_4\beta_1$  with vascular cell adhesion molecule-1.<sup>8</sup>

Amino acid 7/5-bicyclic lactams of type **2** have found use as dipeptide surrogates in angiotensin-converting enzyme (ACE) inhibitors<sup>9</sup> and have more recently been used as external constraints for tripeptide RGD turns which were inhibitors of the  $\alpha_v\beta_3$  integrin receptor.<sup>10</sup> The 7/5 bicyclic lactam **3** was synthesised using an electrochemical approach and conformational analysis showed that the (4*S*,7*S*,10*S*)-stereoisomer had families of minimum conformations with torsion angles close to those of classical  $\beta$ -turns.<sup>11</sup> In continuation of our work on externally constrained GLDV turns,<sup>8</sup> we have now developed a new and stereoselective synthesis of the 7/5 bicyclic lactam, **4**, via a route involving Eschenmoser coupling. The crystal structure of our new constraint will indicate its promise as a turn mimic, and attach-

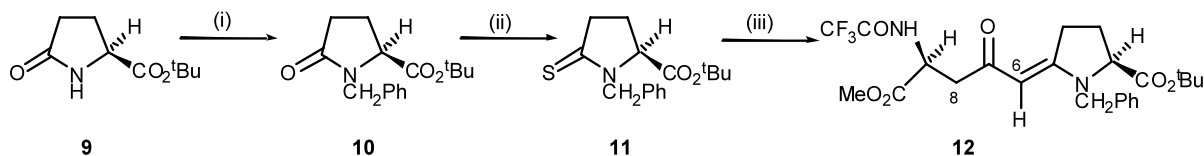


**Keywords:**  $\beta$ -turn; protein mimetic; amino acid; bicyclic lactam.

\* Corresponding author.



**Scheme 1.** Reagents and conditions: (i) HBr/CHCl<sub>3</sub>, rt, 89%; (ii) **7**/MeCN, reflux, then Ph<sub>3</sub>P/Et<sub>3</sub>N, 25%.



**Scheme 2.** Reagents and conditions: (i) NaHMDS/PhCH<sub>2</sub>Br/THF/−78°C, 64%; (ii) P<sub>4</sub>S<sub>10</sub>/THF/rt, 95%; (iii) **6**/Ph<sub>3</sub>P/Et<sub>3</sub>N/CH<sub>3</sub>CN, rt, 91%.

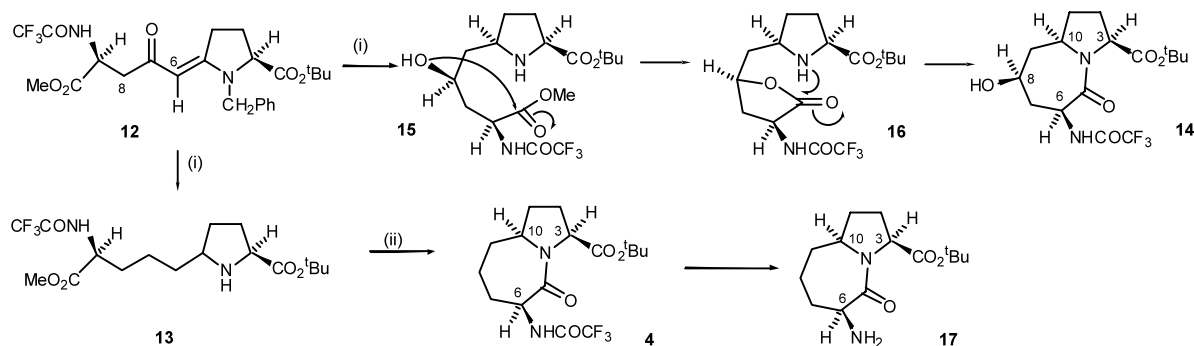
ment of the GLDV recognition motif to it will allow it to be assessed as an external turn constraint. GLDV is present in the integrin family of cell adhesion proteins.<sup>12</sup>

As a first step in our synthesis of the bicyclic lactam **4**, we attempted to prepare the bromide **6** by the method used by Weygand<sup>13</sup> to prepare the corresponding ethyl ester bromide but without success. Modification of the method by reacting the diazoketone **5**<sup>14</sup> with HBr in CHCl<sub>3</sub>, however, gave the bromide **6**,<sup>†</sup> mp 117–8°C, [ $\alpha$ ]<sub>D</sub><sup>30</sup> +3 (*c* 1.01, CH<sub>2</sub>Cl<sub>2</sub>), in 89% yield, as shown in Scheme 1. The bromide was heated at reflux with the thiolactam **7**<sup>15</sup> in CH<sub>3</sub>CN, followed by addition of Ph<sub>3</sub>P and triethylamine to give the desired product **8**<sup>†</sup> as an oil, [ $\alpha$ ]<sub>D</sub><sup>30</sup> +12.3 (*c* 1, CHCl<sub>3</sub>). This was shown to be the *Z*-isomer, since irradiation of the olefinic proton, H-6, caused enhancements to all four protons at C-4 and C-8 in the <sup>1</sup>H NMR spectrum. The hydrogen bond between the N–H of the vinylogous amide and its carbonyl group, as shown, presumably accounts for this specificity. The yield of the product **8** could not be optimised above 25%.

Since sulfur contraction reactions have been reported to be more effective when the thiolactam is trisubsti-

tuted,<sup>16</sup> we prepared the *N*-benzyl thiolactam **11** as shown in Scheme 2. This compound had previously been prepared by Rapoport<sup>17</sup> by a synthesis involving *N*-alkylation of glutamic acid prior to cyclisation to avoid the danger of racemisation. We were able to alkylate *tert*-butyl pyroglutamate **9**<sup>18</sup> directly to the *N*-benzyl derivative **10** in 64% yield and reaction with P<sub>4</sub>S<sub>10</sub> gave the thiolactam **11** in 95% yield with spectroscopic and optical properties similar to those reported by Rapoport.<sup>17</sup> The sulfur contraction reaction with the bromide **6**, using Ph<sub>3</sub>P/Et<sub>3</sub>N in CH<sub>3</sub>CN then gave the vinylogous amide **12**,<sup>†</sup> as an oil, [ $\alpha$ ]<sub>D</sub><sup>28</sup> +122.6 (*c* 1.025, CHCl<sub>3</sub>), in 91% yield.

When the olefinic proton, H-6, in the *N*-benzyl derivative **12** was irradiated, both of the protons H-8 and one of the benzylic CH<sub>2</sub> protons showed enhancement, implying that, unlike the *N*-H analogue **8**, the *N*-benzyl compound **12** had *E*-geometry. In order to reduce the vinylogous amide and hydrogenolyse the *N*-benzyl group in **12**, it was necessary to hydrogenate the compound at a pressure of 60–200 psi for 50 hours using 10% palladium on activated carbon (75% by weight) in methanol containing trifluoroacetic acid as shown in Scheme 3. The resultant single diastereoisomeric product **13**<sup>†</sup> was obtained as an oil, [ $\alpha$ ]<sub>D</sub><sup>28</sup> +8.5 (*c* 1,



**Scheme 3.** Reagents and conditions: (i) H<sub>2</sub>/Pd-C/60–200 psi, MeOH, TFA, 65%; (ii) *t*-BuMgCl/Et<sub>2</sub>O/0°C, 29%; (iii) K<sub>2</sub>CO<sub>3</sub>/MeOH/Δ, 86%.

<sup>†</sup> These compounds had the appropriate analytical and spectroscopic properties.

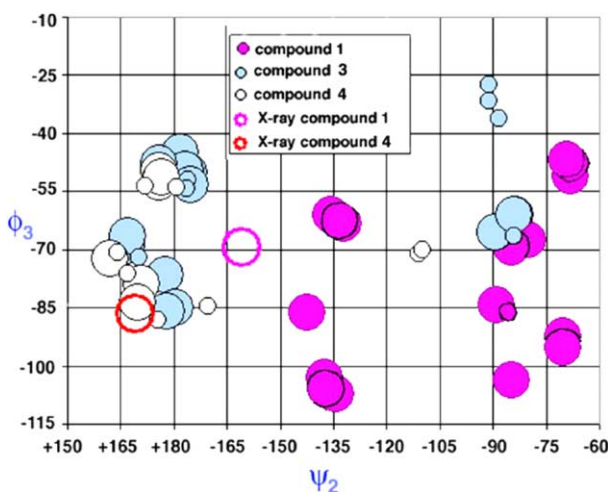
$\text{CHCl}_3$ ), in 65% yield but NOE experiments were inconclusive as to its stereochemistry. Interestingly, in some hydrogenation reactions, a small yield of a by-product was obtained. This proved to be the bicyclic lactam alcohol **14**,<sup>†</sup> mp 141–3°C,  $[\alpha]_{\text{D}}^{28} +14.6$  ( $c$  1,  $\text{CHCl}_3$ ). The (3*S*,6*S*,8*S*,10*S*)-stereochemistry of this compound was implied from NOE studies, since irradiation at H-3 caused enhancement of H-10 and irradiation at H-10 caused enhancements at both H-8 and H-6. It was not possible to optimise the yield of the bicyclic compound **14** but the mechanism suggested in Scheme 3 would account for its formation. Here, incomplete reduction to the alcohol **15** would be followed by lactonisation, giving the product **16** from which final cyclisation would give the product **14**.

The amino ester **13** was cyclised using  $t\text{BuMgCl}$  in ether at 0°C, giving the bicyclic lactam **4**,<sup>†</sup> mp 127–9°C,  $[\alpha]_{\text{D}}^{28} -20.6$  ( $c$  1,  $\text{CHCl}_3$ ), in 29% yield. Calculations of the conformational preferences<sup>19</sup> of the

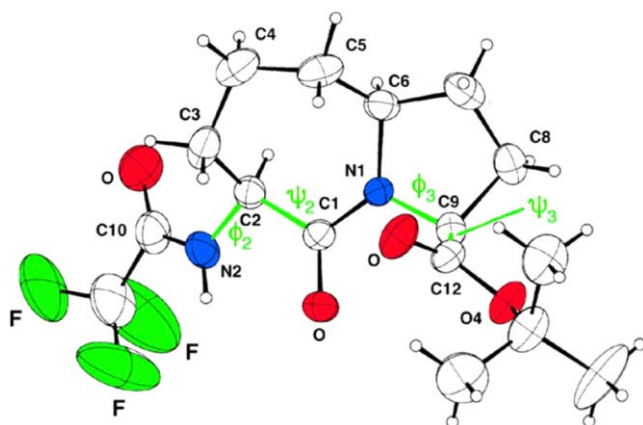
compound **4**, **BTD**, **1**, and compound, **3**, are shown in Figure 1. Except for minor conformers, those torsional minima found for compound **4** are not compatible with  $\beta$ -turns, and this agrees with the X-ray crystal structure analysis (Fig. 2)<sup>20</sup> which showed torsion angles  $\phi_3 -86.5^\circ$  and  $\psi_2 +169^\circ$  which are highlighted in Figure 1. The conformational minima found for **BTD**, **1**, are almost entirely  $\beta$ -turns, whilst the dispersion of minima indicate a more flexible scaffold than our compound **4**. The torsion angles found for the X-ray structure of **17** are somewhat shifted relative to those calculated but this is likely to be due to the combination of a flexible scaffold and strong lattice interactions, since the crystal structure was that of the free acid. The oxa compound **3** is seen to have more rigidity than compound **1** whilst maintaining three distinct local minima that enable the scaffold to mimic both the torsional regions displayed by compound **4** and the region associated with  $\beta$ -turns.

We now wished to examine the usefulness of our turn mimic as an external constraint by adding GLDV across the amino and carboxyl groups. Treatment of the lactam **4** with  $\text{K}_2\text{CO}_3$  in methanol at reflux, as shown in Scheme 3, gave the free amine **17**<sup>†</sup> in 86% yield and reaction of this with Fmoc-valine, TBTU and DIPEA gave the product **18**,<sup>†</sup> mp 129–131°C,  $[\alpha]_{\text{D}}^{28} -24.4$  ( $c$  1.1,  $\text{CHCl}_3$ ), in 80% yield, as shown in Scheme 4. Given the need to protect the  $\beta$ -carboxyl of the aspartate residue in GLDV as the *tert*-butyl ester to prevent intramolecular cyclisation reactions, we now needed to change the orthogonality of the protecting groups. The Fmoc-*tert*-butyl ester **18** was therefore deprotected using TFA to give the acid **19**,<sup>†</sup> mp 131–2°C,  $[\alpha]_{\text{D}}^{28} -69.6$  ( $c$  0.47,  $\text{CHCl}_3$ ), in quantitative yield. Reprotection using diphenyldiazomethane yielded the ester **20**<sup>†</sup> as an oil,  $[\alpha]_{\text{D}}^{30} -32.6$  ( $c$  1,  $\text{CHCl}_3$ ) in 64% yield. This was now sequentially deprotected with piperidine and reacted in turn with Fmoc-Asp(O*t*Bu)-OH, Fmoc-Leu-OH and Cbz-Gly-OH to give the protected tetrapeptide **26**,<sup>†</sup> as summarised in Scheme 4. Hydrogenolysis, cyclisation and final deprotection were then carried out, as in Scheme 4, and the final cyclic peptide **29** was purified by repeated reverse phase HPLC.

High-resolution NMR spectroscopic data were obtained to establish whether the constrained peptide **29** adopted a dominant conformation in solution. Inter-proton distance constraints derived from 2D-ROESY data were incorporated into molecular dynamics calculations, but were not sufficient to define a unique solution conformation of the backbone. However, additional NMR spectral parameters including the wide range of amide proton temperature coefficients, the non-'conformationally averaged' amide to  $\alpha$ -H coupling constants and the approximately 0.4 ppm difference in the glycine methylene proton chemical shifts together with their 'non-equal' vicinal coupling constants to the glycine amide proton (Table 1) were consistent with a single backbone conformation.

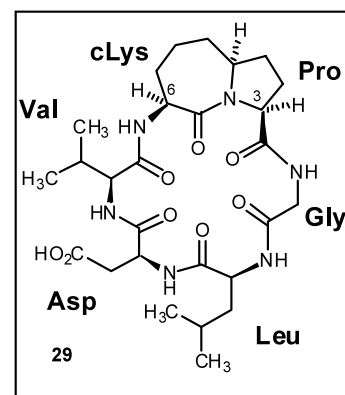


**Figure 1.** Calculated conformations based on energy minimisation for compounds **1**, **3**, and **4**. Large circles denote conformations within 5 kcal of minimum energy, smaller circles within 12 kcal. X-ray derived angles for **1** and **4** are also shown.



**Figure 2.** X-ray crystal structure of the bicyclic lactam **4**.

Gly	Leu	Asp	Val	7/5	
			H	OtBu	17
			Fmoc	(i) OtBu	18
			Fmoc	(ii) OH	19
			Fmoc	(iii) OCHPh <sub>2</sub>	20
			H	(iv) OCHPh <sub>2</sub>	21
		O <sup>t</sup> Bu		(v) OCHPh <sub>2</sub>	22
		Fmoc		(iv) OCHPh <sub>2</sub>	23
		H		(vi) OCHPh <sub>2</sub>	24
	Fmoc	O <sup>t</sup> Bu		(iv) OCHPh <sub>2</sub>	25
	H	O <sup>t</sup> Bu		(vii) OCHPh <sub>2</sub>	26
Cbz		O <sup>t</sup> Bu		(vii) OH	27
H		O <sup>t</sup> Bu		(ix) ]	28
cyclo		O <sup>t</sup> Bu		(x) ]	29
cyclo		OH			



**Scheme 4.** Reagents and conditions; (i) TBTU/DIPEA/DMF/Fmoc-Val-OH, 80%; (ii) CF<sub>3</sub>CO<sub>2</sub>H/CH<sub>2</sub>Cl<sub>2</sub>, quant; (iii) Ph<sub>2</sub>CHN<sub>2</sub>/CH<sub>2</sub>Cl<sub>2</sub>, 64%; (iv) piperidine/DMF, >82%; (v) TBTU/DIPEA/Fmoc-Asp(O<sup>t</sup>Bu)-OH, 88%; (vi) TBTU/DIPEA/DMF/Fmoc-Leu-OH, 82%; (vii) TBTU/DIPEA/Cbz-Gly-OH, 81%; (viii) H<sub>2</sub>/Pd-C, 92%; (ix) TBTU/DMAP/DMF; (x) CF<sub>3</sub>CO<sub>2</sub>H/ TES.

**Table 1.** <sup>1</sup>H NMR data for **29** in 90% H<sub>2</sub>O:10% D<sub>2</sub>O (ref δ 4.70) at 298 K

AA unit	NH	αH	βH	γH	δH	εH	<sup>3</sup> J <sub>NH,αH</sub> (Hz)	Δδ/ΔT (ppb/K)
Gly (G)	8.68	3.62 and 4.01	—	—	—	—	5.4 and 7.0	−7.8
Leu (L)	8.40	4.78	1.42 and 1.75	1.50	0.83 and 0.89	—	9.0	−2.8
Asp (D)	8.58	4.53	2.81 and 2.91	—	—	—	5.5	−4.3
Val (V)	7.24	4.35	2.36	2.40	0.77 and 0.84	—	9.5	−2.6
cLys (cK)	7.53	4.45	1.61	1.88	1.39	4.48	5.6	−1.7
Pro (P)	—	4.47	1.94 and 2.10	1.61 and 1.67	—	—	—	—

The concentration of monomeric peptide was approximately 0.6 mM; a substantial amount of aggregated peptide material was also present.

### Acknowledgements

We thank GlaxoSmithKline and the EPSRC for a CASE studentship (to R.D.H.) and Dr B. Sherborne for conformational calculations.

### References

- Rose, G. D.; Gierasch, L. M.; Smith, J. A. *Adv. Protein Chem.* **1985**, *37*, 1–109.
- Smith, J. A.; Pease, L. G. *CRC Crit. Rev. Biochem.* **1980**, *8*, 315–399.
- Nagai, U.; Sato, K. *Tetrahedron Lett.* **1985**, *26*, 647–650.
- Sato, K.; Nagai, U. *J. Chem. Soc., Perkin Trans. 1* **1986**, 1231–1234.
- Nagai, U.; Sato, K.; Nakamura, R.; Kato, R. *Tetrahedron* **1993**, *49*, 3577–3592.
- Wisskirchen, F. M.; Doyle, P. M.; Gough, S. L.; Harris, C. J.; Marshall, I. *Br. J. Pharm.* **1999**, *126*, 1163–1170.
- Osano, Y. T.; Nagai, U.; Matsuzaki, T. *Anal. Sci.* **1969**, *5*, 625–626 The data are available from the Cambridge Crystallographic Data Centre, Cambridge, code number SEYZUR.
- Doyle, P. M.; Harris, J. C.; Moody, C. M.; Sadler, P. J.; Sims, M.; Thornton, J. M.; Uppenbrink, J.; Viles, J. H. *Int. J. Peptide Protein Res.* **1996**, *47*, 427–436.
- Robl, J. A.; Cimarusti, M. P.; Simpkins, L. M.; Brown, B.; Ryono, D. E.; Bird, J. E.; Asaad, M. M.; Schaeffer, T. R.; Trippodo, N. C. *J. Med. Chem.* **1996**, *39*, 494–502.
- Belvisi, L.; Bernardi, A.; Checchia, A.; Manzoni, L.; Potenza, D.; Scolastico, C.; Castorina, M.; Cupelli, A.; Giannini, G.; Carminati, P.; Pisano, C. *Org. Lett.* **2001**, *3*, 1001–1004.
- Cornille, F.; Slomczynska, U.; Smythe, M. L.; Beusen, D. D.; Moeller, K. D.; Marshall, G. R. *J. Am. Chem. Soc.* **1995**, *117*, 909–917.
- Haubner, R.; Finsinger, D.; Kessler, H. *Angew. Chem., Int. Ed.* **1997**, *36*, 1374–1389.
- Weygand, F.; Klinke, P.; Eigen, I. *Chem. Ber.* **1957**, *90*, 1896–1905.
- Gani, D.; Young, D. W. *J. Chem. Soc., Perkin Trans. 1* **1983**, 2393–2398.
- Andersen, T. P.; Rasmussen, P. B.; Thomsen, I.; Lawesson, S.-O.; Jørgensen, P.; Lindhardt, P. *Annalen* **1986**, 269–279.
- Shiosaki, K. In *Comprehensive Organic Synthesis*; Trost, B., Fleming, I., series Eds., Vol. 2, Heathcock, C. H., Vol. Ed.; Pergamon Press: Oxford, 1991; pp. 865–892.
- Petersen, J. S.; Fels, G.; Rapoport, H. *J. Am. Chem. Soc.* **1984**, *106*, 4539–4547.
- Kolasa, T.; Miller, M. J. *J. Org. Chem.* **1990**, *55*, 1711–1721.
- Conformational analysis:** All calculations were performed with Sybyl 6.9, available from Tripos Inc., 1699 South Hanley Rd., St. Louis, MI, 63144, USA. Random searching was performed with 1000 cycles and an energy cut-off

of 20 kCal. Energy minimisations were performed with the Tripos forcefield with Gasteiger-Hückel charges and 1000 iterations.

20. **Crystal data:** compound **4**,  $C_{16}H_{23}F_3N_2O_4$ ,  $M=364.4$ , monoclinic, space group  $P2_1$  (No 4),  $a=5.485(2)$ ,  $b=11.970(4)$ ,  $c=14.437(5)$  Å,  $V=947.9(6)$  Å<sup>3</sup>,  $Z=2$ ,  $D_{\text{calc}}=1.28$  mg/m<sup>3</sup>,  $\mu$  (Mo-K $\alpha$ )  $0.11$  mm<sup>-1</sup>,  $T=293(2)$  K, 1757 independent reflections, 1295 with  $I>2\sigma(I)$ , final residuals

were  $R1=0.058$ ,  $wR2=0.152$ ; Data collection was carried out using a Nonius CAD4 diffractometer, structure analysis using program package WinGX, and refinement using SHELXL-97. The atomic coordinates are available on request from The Director, Cambridge Crystallography Data Centre, University Chemical Laboratory, Lensfield Road, Cambridge, CB2 1EW (deposition number CCDC 216301).