

Enantioselective Synthesis of Aurilide, a Cytotoxic 26-Membered Cyclodepsipeptide of Marine Origin

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Abstract: Aurilide (**1**), a novel cyclodepsipeptide isolated from the Japanese sea hare *Dolabella auricularia*, was enantioselectively synthesized, and the present result unambiguously confirmed its stereostructure. In addition, the cytotoxicity of **1** was evaluated by employing synthetic **1**.

Recently, we isolated aurilide (**1**) as a very minute constituent from the Japanese sea hare *Dolabella auricularia* and elucidated the absolute stereostructure of **1** on the basis of its spectral analysis and organic synthetic methods.¹ Although aurilide (**1**) was isolated from a cytotoxic fraction of the sea hare, the scarcity of a natural supply has prevented the evaluation of its cytotoxicity. This fact and the novel 26-membered cyclodepsipeptide structure prompted us to synthesize aurilide (**1**). We describe herein the enantioselective synthesis of aurilide (**1**) and its biological evaluation.

A key step in the synthesis of aurilide (**1**) is the 26-membered ring closure (Scheme 1). After careful inspection of the aurilide structure (**1**), we planned to construct the cyclic structure of **1** by the macrolactamization of amino acid **2**, which is synthesized from pentapeptide **3** and the protected dihydroxy acid **4**.

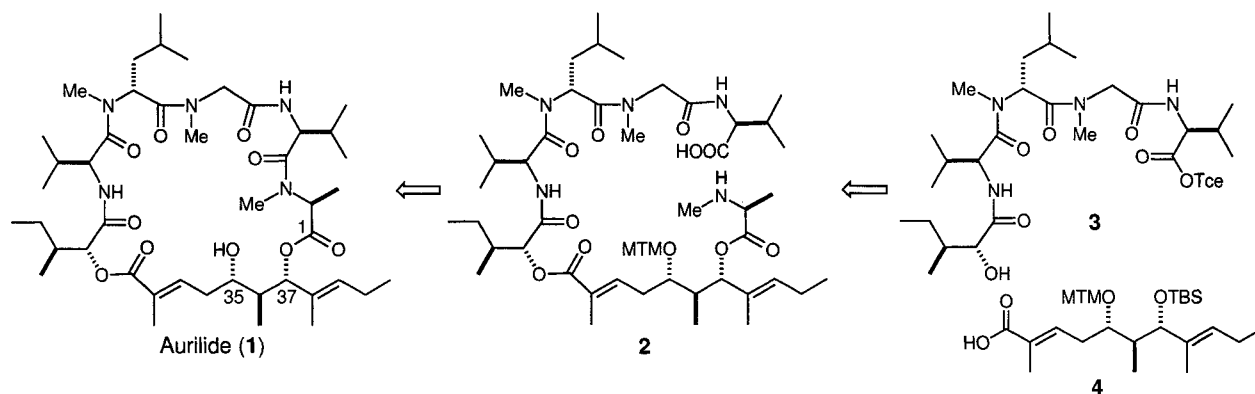
The synthesis of pentapeptide **3** was carried out starting from *N*-methylglycine *tert*-butyl ester hydrochloride in a stepwise manner in 84% overall yield (Scheme 2).

The protected dihydroxy acid **4** was synthesized from (4*S*,5*R*)-4-methyl-5-phenyl-3-propionyl-2-oxazolidinone (**5**), as shown in Scheme 3. The *anti* selective aldol reaction² between **5** and *trans*-2-methyl-2-pentenal afforded aldol **6**^{3,4} (67%), which was converted into aldehyde **7** by the standard synthetic reactions (78%, 3 steps). The vinylogous

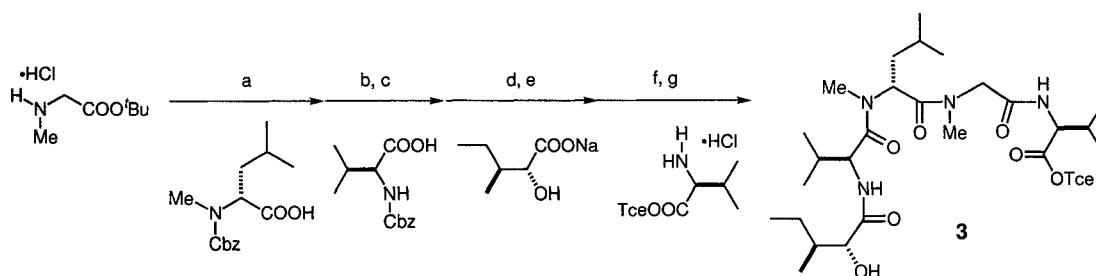
Mukaiyama aldol reaction⁵ between **7** and 2-methyl-1-trimethylsiloxy-1,3-butadiene provided a conjugated aldehyde, oxidation of which and subsequent treatment with diazomethane gave methyl ester **8** as a single diastereomer (59%, 3 steps).⁶ Configuration inversion of the C35 hydroxyl group in **8** was effected as follows: Dess-Martin oxidation⁷ of **8** afforded keto ester **9**, and reduction of the resulting keto group in **9** stereoselectively proceeded to give alcohol **10** (81% from **8**),⁸ which has the desired stereochemistry concerning the C35 hydroxyl group.⁶ Protection of the hydroxyl group in **10** followed by hydrolysis provided the protected dihydroxy acid **4** (68%, 2 steps).

The coupling reaction of pentapeptide **3** and the protected dihydroxy acid **4** was effected with EDCI·HCl⁹ and DMAP to provide ester **11**, which was converted into alcohol **12** (91% from **4**). Esterification of **12** with *N*-Fmoc-*N*-methyl-L-alanine gave the *N*-methylalanine ester **13** (92%). The trichloroethyl group and the Fmoc group in **13** were removed in two steps to give amino acid **2**, which was subjected to macrolactamization with Bop-Cl¹⁰ to afford cyclodepsipeptide **15** (29% from **13**).¹¹ Finally, the MTM group in **15** was removed to give aurilide (**1**) in 93% yield. Synthetic aurilide (**1**) was found to be identical with the natural product **1** by comparison of their spectral (UV, IR, ¹H NMR, MS, and α_D) and chromatographic properties.

In conclusion, the enantioselective synthesis of aurilide (**1**) was carried out in 3.9% overall yield based on the longest linear sequence (18 steps),¹⁵ and the stereostructure of aurilide was unambiguously confirmed to be **1**. Aurilide (**1**) was found to exhibit a strong cytotoxicity against HeLa S₃ cells with an IC₅₀ of 0.011 μg/mL by employing the synthetic specimen.

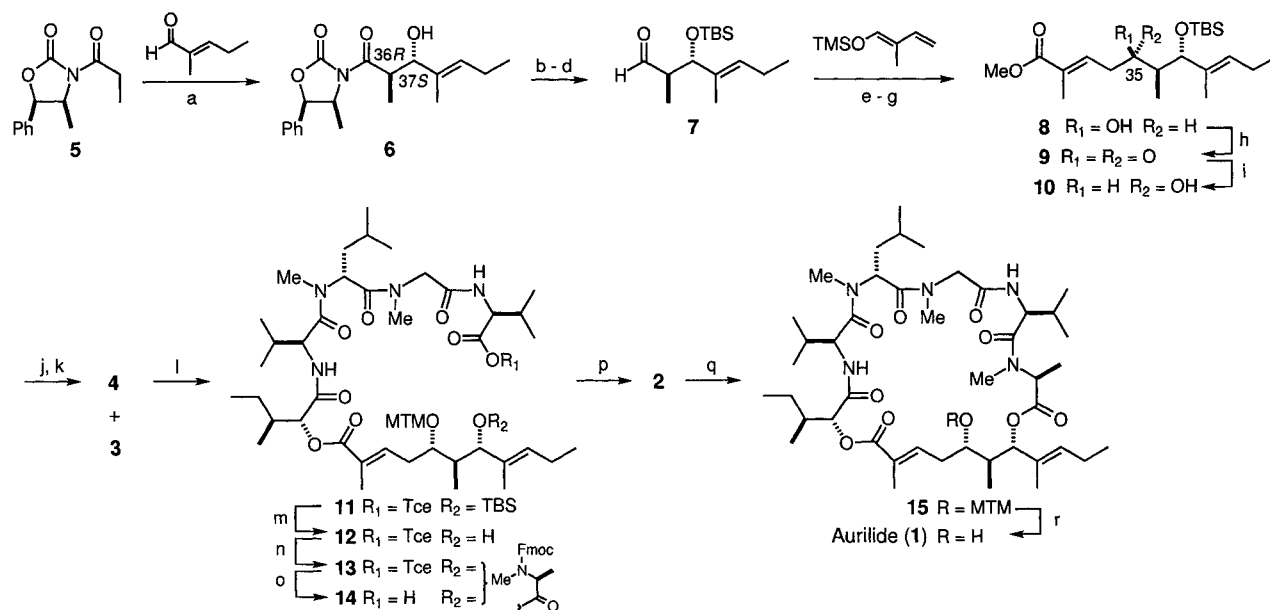


Scheme 1



Reagents and conditions: (a) DEPC,¹³ Et₃N, DMF, 23 °C (98%); (b) H₂, Pd-C, EtOH, 23 °C; (c) PyBOP,¹⁴ *i*-Pr₂NEt, CH₂Cl₂, 23 °C (92%, 2 steps); (d) H₂, Pd-C, EtOH, 23 °C; (e) EDCI·HCl,⁹ HOBT, DMF, 23 °C (95%, 2 steps); (f) TMSOTf, 2,6-lutidine, CH₂Cl₂, 0 °C (100%); (g) EDCI·HCl, HOBT, Et₃N, DMF, CH₂Cl₂, 23 °C (98%).

Scheme 2



Reagents and conditions: (a) *trans*-2-methyl-2-pentenal (1.25 equiv.), Bu₂BOTf (2.0 equiv.), *i*-Pr₂NEt (1.15 equiv.), Et₂O, -100 °C → -78 °C (67%); (b) MeNH(OMe)·HCl, Me₃Al, THF, 50 °C (84%); (c) TBSCl, imidazole, DMF, 23 °C (100%); (d) DIBAL, THF, -78 °C (93%); (e) 2-methyl-1-trimethylsiloxy-1,3-butadiene, BF₃·OEt₂, CH₂Cl₂, Et₂O, -78 °C (59%); (f) NaClO₂, NaH₂PO₄, 2-methyl-2-butene, *t*-BuOH, H₂O, 23 °C; (g) CH₂N₂, Et₂O, 0 °C (100%, 2 steps); (h) Dess–Martin periodinane, CH₂Cl₂, 23 °C (99%); (i) NaBH₄, MeOH, -23 °C (82%); (j) DMSO, Ac₂O, AcOH, 40 °C (74%); (k) LiOH, MeOH, H₂O, 30 °C (92%); (l) EDCI·HCl, DMAP, CH₂Cl₂, 23 °C (91%); (m) HF·pyridine, pyridine, THF, 40 °C (100%); (n) *N*-Fmoc-*N*-methyl-L-alanine, EDCI·HCl, DMAP, CH₂Cl₂, 23 °C (92%); (o) Zn, NH₄OAc, THF, H₂O, 23 °C (91%); (p) Et₂NH, MeCN, 23 °C; (q) Bop-Cl, Et₃N, CH₂Cl₂, 23 °C (32%, 2 steps); (r) AgNO₃, 2,6-lutidine, THF, H₂O, 70 °C (93%).

Scheme 3

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References and Notes

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- (3) Raimundo, B. C.; Heathcock, C. H. *Synlett*, **1995**, 1213.
- (4) The (36*R*,37*R*)-isomer of **6** was obtained as a minor product (14%).
- (5) The stereochemistry of aldol **6** was determined as follows. Transamidation of **6** and its *syn*-diastereomer **16** that was previously¹ prepared by Evans aldol reaction gave diastereomeric amides **17a** and **17b**, respectively. This finding indicated that the relative stereochemistry between C36 and C37 in **6** was *anti*. On the other hand, oxidation of **6** and **16** afforded diastereomeric ketones **18a** and **18b**, respectively. This fact indicated that the absolute configuration of C36 in **6** was *R*. From these results, the stereochemistry of **6** was determined to be 36*R*,37*S* (*anti*), as expected according to Heathcock and coworkers.²
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- (10) The stereoselectivity of reduction of **9** was 1:19 (**8**:**10**). The mixture of the diastereomeric alcohols **8** and **10** was separated by column chromatography (silica gel, 1:1 hexane/CH₂ClCH₂Cl).
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- (19) All new compounds gave spectroscopic data in agreement with the assigned structures. Data for the selected key compounds follow.
3: a colorless amorphous powder; [α]_D²⁷ +46.3 (c 1.41, CHCl₃); IR (CHCl₃) 3420, 3360 (br), 1755, 1675, 1630, 1515, 1465, 1390, 1140 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.78 (d, *J* = 6.8 Hz,

2.55 H), 0.81 (d, $J = 6.8$ Hz, 0.45 H), 0.85–1.07 (m, 21 H), 1.23–1.90 (m, 6 H), 2.05 (m, 1 H), 2.32 (m, 1 H), 2.89 (d, $J = 4.9$ Hz, 0.15 H), 2.93 (d, $J = 4.9$ Hz, 0.85 H), 2.96 (s, 0.45 H), 3.10 (s, 3 H), 3.12 (s, 2.55 H), 3.93 (d, $J = 17.1$ Hz, 0.15 H), 3.99 (d, $J = 15.6$ Hz, 0.85 H), 4.06 (d, $J = 15.6$ Hz, 0.85 H), 4.13 (dd, $J = 2.0$, 4.9 Hz, 1 H), 4.27 (d, $J = 17.1$ Hz, 0.15 H), 4.62 (d, $J = 11.7$ Hz, 1 H), 4.64 (dd, $J = 4.9$, 8.8 Hz, 0.85 H), 4.69 (dd, $J = 4.9$, 8.3 Hz, 0.15 H), 4.82 (m, 0.15 H), 4.85 (dd, $J = 6.8$, 8.8 Hz, 0.85 H), 4.90 (d, $J = 11.7$ Hz, 0.85 H), 4.92 (d, $J = 11.7$ Hz, 0.15 H), 5.29 (dd, $J = 5.4$, 9.3 Hz, 0.15 H), 5.47 (dd, $J = 6.3$, 8.3 Hz, 0.85 H), 6.74 (d, $J = 8.8$ Hz, 0.85 H), 6.82–6.89 (m, 0.3 H), 6.93 (d, $J = 8.8$ Hz, 0.85 H); ^{13}C NMR (100 MHz, CDCl_3) δ 11.8 (q), 12.6 (q), 17.39 [17.25] (q), 17.41 [17.7] (q), 19.0 (q), 19.5 (q), 22.0 [21.8] (q), 22.9 [23.1] (q), 24.7 [24.6] (d), 26.2 (t), 30.59 [30.65] (d), 30.74 (q), 31.1 [30.9] (d), 36.5 (q), 37.7 [37.8] (t), 38.6 (d), 51.2 [50.7] (d), 52.6 [52.4] (t), 53.7 [54.0] (d), 57.0 [57.3] (d), 73.95 [74.00] (d), 74.35 [74.40] (t), 94.4 [94.3] (s), 168.6 [168.3] (s), 170.2 [170.7] (s), 171.8 [171.6] (s), 172.3 [172.4] (s), 173.6 [173.7] (s) (the minor counter parts of doubled signals in the ratio of 5.6:1 are

in brackets); MS (FAB) m/z 681 ($\text{M} + \text{Na}$) $^+$; HRMS (FAB) calcd for $\text{C}_{28}\text{H}_{49}^{35}\text{Cl}_3\text{N}_4\text{NaO}_7$ [$(\text{M} + \text{Na})^+$] 681.2564, found 681.2579.

4: a colorless amorphous powder; $[\alpha]_{\text{D}}^{28} -90.3$ (c 1.09, CHCl_3); IR (CHCl_3) 3100 (br), 1685, 1645, 1460, 1290, 1250, 1105, 1055, 840 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ -0.05 (s, 3 H), 0.02 (s, 3 H), 0.72 (d, $J = 6.8$ Hz, 3 H), 0.89 (s, 9 H), 0.96 (t, $J = 7.3$ Hz, 3 H), 1.54 (br s, 3 H), 1.86 (br s, 3 H), 1.95–2.11 (m, 3 H), 2.15 (s, 3 H), 2.23–2.38 (m, 2 H), 3.67 (d, $J = 9.3$ Hz, 1 H), 4.16 (ddd, $J = 3.4$, 3.4, 8.8 Hz, 1 H), 4.53 (d, $J = 11.7$ Hz, 1 H), 4.63 (d, $J = 11.7$ Hz, 1 H), 5.30 (br t, $J = 6.8$ Hz, 1 H), 7.06 (m, 1 H) (signals of one proton (COOH) were not observed); ^{13}C NMR (67.8 MHz, CDCl_3) δ -5.3 (q), -4.4 (q), 10.4 (q), 10.7 (q), 12.3 (q), 13.8 (q), 14.0 (q), 18.1 (s), 20.8 (t), 25.8 (q, 3 C), 28.8 (t), 38.2 (d), 73.1 (t), 75.5 (d), 80.9 (d), 127.8 (s), 130.0 (d), 134.9 (s), 143.3 (d), 173.1 (s); MS (FAB) m/z 453 ($\text{M} + \text{Na}$) $^+$; HRMS (FAB) calcd for $\text{C}_{22}\text{H}_{42}\text{NaO}_4\text{SSi}$ [$(\text{M} + \text{Na})^+$] 453.2471, found 453.2495.