Synthesis of Glycosyl Phosphonates by Michael-Type Addition to 2-Nitroglycals

Kandasamy Pachamuthu,^[a] Ignacio Figueroa-Perez,^[a] Ibrahim A. I. Ali,^[a] and Richard R. Schmidt^{*[a]}

Keywords: Carbohydrates / Nitroglycals / Michael addition / Phosphonates / Anomerisation

A new synthetic strategy, which allows to synthesize both α - and β -anomers of 2-acetylaminoglycosyl phosphonate or exclusively β -anomer is described. The present strategy is a successful Michael-type addition of dimethyl hydrogen phosphonate to 2-nitroglycals.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2004)

Introduction

During the last several years there has been considerable interest in the preparation and investigation of phosphonic acids and their derivatives that might be considered to be analogues of naturally occurring phosphates.^[1] It is understood that the carbon-phosphorus bond is incapable of being hydrolyzed by the enzymes involved in phosphate cleavage. Thus, the use of phosphonic acids as analogues of natural phosphates represents a systematic approach to metabolic regulation, enhancement or inhibition studies.



The outer surface of the outer membrane of *Escherichia coli* and other Gram-negative bacteria is made up primarily of lipid A, the hydrophobic anchor for lipopolysaccharides. Lipid A is essentially a β -(1'-6)-linked D-glucosamine disaccharide carrying phosphate residues at C-1 and C-4', and several *N*- and *O*-bound long-chain acyl groups.^[2] Lipid X (I), a monosaccharide isolated from *E. coli* mutants, is a biosynthetic precursor of Lipid A corresponding to the reducing end.^[3] Although it was identified in 1984, the function of the phosphate group at C-1 is not fully understood. Therefore, synthesis of phosphonate analogues of type II

E-mail: Richard.Schmidt@uni-konstanz.de

might help to study the function of the phosphate group.

Vasella et al.^[4-6] described the synthesis of such a type of nonisosteric glycosyl phosphonate analogues starting from 2-azido-2-deoxy sugars, employing the trichloroacetimidate method. In continuation of our work on Michaeltype addition of various nucleophiles to 3,4,5-tri-O-benzyl-2-nitroglycals,^[7-14] we now wish to report a convenient synthesis of such nonisosteric phosphonate analogues by Michael-type addition of dimethyl hydrogen phosphonate to 2-nitroglycals.

Results and Discussion

A straightforward way to synthesize the desired phosphonate analogues is Michael-type addition of HPO(OMe)₂ to 2-nitroglycals. The advantage of this method is mainly due to the easy preparation of the starting material in high yields and the convenient transformation of the nitro group into the corresponding amino group and hence, into an acylamino group. Treatment of 3,4,5-tri-O-benzyl-2-nitrogalactal with HPO(OMe)₂ in the presence of potassium tert-butoxide in toluene yielded the corresponding glycosyl phosphonates 2 and 3 in the ratio of 1:1.2 in 82% yield. Careful examination of the reaction revealed that the anomeric (α / β) ratio varies with time. A ratio of 1:1.2 for α/β was observed after 5 min at 0 °C, but after 2 h at the same reaction temperature only the β -product was observed in 60% yield, which indicates that the α -anomer was slowly transformed under basic reaction conditions into the thermodynamically more stable β -anomer presumably by ring opening to intermediate A followed by ring closure (Scheme 1).

Department of Chemistry, University of Konstanz, Fach M 725, 78457 Konstanz, Germany Fax: (internat.) + 49-7531-883135

Supporting information for this article is available on the WWW under http://www.eurjoc.org or from the author.

SHORT COMMUNICATION



Scheme 1. Michael-type addition of $HPO(OMe)_2$ to 2-nitrogalactal 1

The anomeric configuration is evident from the ¹H and ¹³C NMR spectroscopic data. In the ¹H NMR spectra, the signal of 1-H of **2** appears at $\delta = 4.7$ ppm (${}^{3}J_{1,2} = 7.1$ Hz) and for **3** at $\delta = 4.19$ ppm (${}^{3}J_{1,2} = 10.2$ Hz). In the ¹³C NMR spectra, the C-1 signals for **2** and **3** appear at $\delta = 70.6$ and 73.5 ppm, respectively; the signal for the α -anomer **2** occurs at a higher field than that for the corresponding β -anomer **3**.

By employing different weak bases and lowering the reaction temperature, the same result was obtained. Reduction of the nitro group in compounds 2 and 3 with the use of platinized Raney nickel in ethanol yielded the corresponding amines, which were acetylated to give the known compounds 4 and 5 in 90 and 92% yield, respectively,^[4] as shown in Scheme 2. The spectroscopic data are identical to literature values.^[4]



Scheme 2. Conversion of 2-nitrogalactosyl phosphonate to 2-(acetylamino)galactosyl phosphonate

Reaction of 2-nitrorhamnal **6** with HPO(OMe)₂ at 0 °C yielded the corresponding glycosyl phosphonates **7** and **8** in 5 min and with a ratio of 1:1.2 (α/β) in 80% yield. After



Scheme 3. Michael-type addition of $HPO(OMe)_2$ to 2-nitrorhamnal $\mathbf{6}$

2 h, only product 8 was isolated in 55% yield (Scheme 3 and Table 1).

On the other hand, reaction of 2-nitroglucal 9 with HPO(OMe)₂ at 0 °C for 5 min yielded mannose-configured α -anomer 10 and glucose-configured β -anomer 11 in a ratio of 1:1.2 in 81% yield (Scheme 4 and Table 1). A prolonged reaction time yielded only the β -anomer **11** in 80% yield, as was the case for the galactal derivative. The structures of 10 and 11 were deduced from their ¹H and ¹³C NMR spectroscopic data. In the ¹H NMR spectra of compound **10**, the signal for 1-H appears at $\delta = 4.63$ (${}^{3}J_{1,2} = 2.6$ Hz) and in the spectra of 11 at $\delta = 4.18$ ppm (${}^{3}J_{1,2} = 9.1$ Hz), whereas the signal for 3-H for 10 appears at $\delta = 4.31$ ppm (${}^{3}J_{3,4} =$ 8.3, ${}^{3}J_{3,2} = 5.1$ Hz) and that for 11 at $\delta = 4.22$ ppm (${}^{3}J_{3,4} =$ ${}^{3}J_{3,2} = 9.1$ Hz). Likewise, reaction of 2-nitromaltal 12 also yielded the same result. After 5 min at 0 °C, the products obtained were 13 and 14 in a ratio of 1:1.5, and after 2 h, only β -gluco-configured 14 was found. All compounds were characterized by spectral and analytical means.



Scheme 4. Michael-type addition of $HPO(OMe)_2$ to 2-nitroglucals 9 and 12

Conclusion

In conclusion, an efficient and very simple method for the synthesis of various glycosyl phosphonates by Michaeltype addition of $HPO(OMe)_2$ to 2-nitroglycals in high yields is described. Applications of this methodology to synthesize various phosphonate analogues of glycosyl con-

Table 1. Results obtained with 2-nitroglycals 6, 9, and 12

Starting compound	Reaction time	Products	Yield [%]	Reaction time	Yield [%]
6	5 min	7 + 8	81 (1:1.2)	2 h	55 (only 8)
9	5 min	10 + 11	80 (1:1.2)	2 h	80 (only 11)
12	5 min	13 + 14	80 (1:1.5)	2 h	80 (only 14)

jugates and glycopeptides are under progress in our laboratory.

Experimental Section

General Remarks: Solvents were purified according to standard procedures. NMR spectroscopic measurements were performed at 22 °C with a Bruker DRX600 instrument. TMS or the resonance of the deuterated solvent was used as internal standard. CDCl₃ ($\delta = 7.24$ ppm) was used as external standard. MALDI mass spectra were recorded with a Kratos Kompact Maldi 1 instrument, and 2,5-dihydroxybenzoic acid (DHB) or 6-aza-2-thiothymine (ATT) were used as matrices. Optical rotations were measured with a Perkin–Elmer polarimeter 241/MS in a 1-dm cell at 22 °C. Thinlayer chromatography (TLC) was performed on Merck silica gel 60 F_{254} plastic plates or Merck amino phase glass plates. Compounds were visualized by treatment with a solution of (NH₄)₆Mo₇O₂₄·4H₂O (20 g) and Ce(SO₄)₂ (0.4 g) in 10% sulfuric acid (400 mL). Flash chromatography was performed on J. T. Baker silica gel 60 (0.040–0.063 mm) at a pressure of 0.3 bar.

General Experimental Procedure for the Synthesis of 2, 3, 7, 8, 10, 11, 13 and 14: To a stirred solution of the 2-nitroglycal (1 mmol) and HPO(OMe)₂ (1.1 mmol) in dry toluene at 0 °C was added potassium *tert*-butoxide (1 mmol), and the resulting solution was stirred for 5 min. The solvent was then evaporated under reduced pressure, followed by extraction of the residue with ethyl acetate, washing of the combined extracts with dild. HCl, water, brine, and drying with anhydrous MgSO₄. After filtration, evaporation of the solvent gave the corresponding glycosyl phosphonates; provided are isolated yields.

General Experimental Procedure for the Synthesis of 4 and 5: To a stirred solution of freshly prepared Pt/Raney nickel (1 g) in ethanol was added the phosphonate 2 or 3, and the mixture stirred under hydrogen overnight. Filtration of the catalyst and evaporation of

the solvent yielded the corresponding amines, which were dissolved in acetic anhydride (5 mL) and stirred overnight at room temperature. Evaporation of the acetic anhydride, followed by flash column purification afforded the 2-acetylamino-2-deoxygalactosyl phosphonates **4** and **5**.

Supporting Information: Data for 2, 3, 7, 8, 10, 11, 13 and 14.

Acknowledgments

This work was supported by the Deutsche Forschungsgemeinschaft and the Fonds der Chemischen Industrie. K. P. is grateful for an Alexander von Humboldt Fellowship.

- ^[1] R. Engel, Chem. Rev. 1977, 77, 349-367.
- [2] E. Th. Rietschel, H. Brade, L. Brade, W. Kaca, K. Kawahara, B. Lindner, T. Lüderitz, T. Tomita, U. Schade, U. Seydel, U. Zähringer, in *Progress in Clinical and Biological Research* (Eds.: J. W. ten Cate, H. R. Büller, A. Sturk, J. Levin, Alan R. Liss), Alan Liss, Inc., New York, **1985**, vol. 189, pp. 31–45.
- [3] E. T. Rietschel, H.-W. Wollenweber, H. Brade, in *Handbook of Endotoxin, Chemistry of Endotoxin* (Ed.: E. Th. Rietschel), Elsevier, Amsterdam, **1984**, vol. 1, pp. 187–220.
- ^[4] K. Briner, A. Vasella, Helv. Chim. Acta 1987, 70, 1341-1356.
- ^[5] R. Meuwly, A. Vasella, Helv. Chim. Acta 1986, 69, 25-34.
- ^[6] A. Vasella, R. Wyler, Helv. Chim. Acta 1991, 74, 451-463.
- ^[7] J. Das, R. R. Schmidt, Eur. J. Org. Chem. 1998, 1609-1612.
- [8] G. A. Winterfeld, Y. Ito, T. Ogawa, R. R. Schmidt, *Eur. J. Org. Chem.* 1999, 1167–1171.
- [9] G. A. Winterfeld, R. R. Schmidt, Angew. Chem. 2001, 113, 2718–2721; Angew. Chem. Int. Ed. 2001, 40, 2654–2657.
- ^[10] K. Pachamuthu, A. Gupta, J. Das, R. R. Schmidt, Y. D. Vankar, *Eur. J. Org. Chem.* **2002**, 1479–1483.
- ^[11] K. Pachamuthu, R. R. Schmidt, Synlett 2003, 1355-1357.
- ^[12] G. A. Winterfeld, A. I. Khodair, R. R. Schmidt, *Eur. J. Org. Chem.* 2003, 1009–1021.
- ^[13] A. I. Khodair, K. Pachamuthu, R. R. Schmidt, *Synthesis* **2004**, 53–58.
- ^[14] J. Geiger, N. Barroca, R. R. Schmidt, Synlett 2004, 836–840. Received April 19, 2004