This article was downloaded by: [Umeå University Library] On: 18 November 2014, At: 20:18 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: <u>http://www.tandfonline.com/loi/lcar20</u>

# SYNTHESIS OF A MANNOTETRAOSE—THE REPEATING UNIT OF THE CELL-WALL MANNANS OF MICROSPORUM GYPSEUM AND RELATED SPECIES OF TRYCHOPHYTON

Linsen Heng<sup>a</sup>, Jun Ning<sup>a</sup> & Fanzuo Kong<sup>a</sup>

<sup>a</sup> Research Center for Eco-Environmental Science, Academia Sinica, P.O. Box 2871, Beijing, 100085, China Published online: 16 Aug 2006.

To cite this article: Linsen Heng, Jun Ning & Fanzuo Kong (2001) SYNTHESIS OF A MANNOTETRAOSE—THE REPEATING UNIT OF THE CELL-WALL MANNANS OF MICROSPORUM GYPSEUM AND RELATED SPECIES OF TRYCHOPHYTON, Journal of Carbohydrate Chemistry, 20:3-4, 285-296, DOI: <u>10.1081/CAR-100104864</u>

To link to this article: http://dx.doi.org/10.1081/CAR-100104864

## PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <a href="http://www.tandfonline.com/page/terms-and-conditions">http://www.tandfonline.com/page/terms-and-conditions</a>

#### J. CARBOHYDRATE CHEMISTRY, 20(3&4), 285–296 (2001)

## SYNTHESIS OF A MANNOTETRAOSE—THE REPEATING UNIT OF THE CELL-WALL MANNANS OF *MICROSPORUM GYPSEUM* AND RELATED SPECIES OF *TRYCHOPHYTON*

### Linsen Heng, Jun Ning, and Fanzuo Kong

Research Center for Eco-Environmental Science, Academia Sinica. P.O. Box 2871, Beijing 100085, China

## ABSTRACT

A tetrasaccharide,  $\alpha$ -D-mannopyranosyl- $(1\rightarrow 2)$ - $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ - $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -D-mannopyranose (1), the repeating unit of the cell-wall mannans of *Microsporum gypseum* and related species of *Trychophyton*, was synthesized using 6-*O*-acetyl-2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (5) and 2-*O*-acetyl-3,4,6-tri-*O*-benzoyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (13) as the glycosyl donors in "the inverse Schmidt" procedure.

## **INTRODUCTION**

Dermatophytes are a group of fungi which parasitise man and animals, and cause superficial cutaneous infections involving primarily the keratinised tissues of the epidermis, nails, and pilosebaceous follicles. The cell wall of dermatophytes is made up of chitin, a water-insoluble  $\beta$ -(1 $\rightarrow$ 3)-glucan, and water-soluble polysaccharides which are antigenically relevant. In many ascoycetous yeasts,<sup>1</sup> the antigenically relevant outer polysaccharides have a skeleton composed of  $\alpha$ -(1 $\rightarrow$ 6)linked mannopyranosyl residues, to most of which are attached one branching moiety, each composed of chains of various lengths containing mainly  $\alpha$ -,  $\beta$ (or both)-(1 $\rightarrow$ 2) and/or  $-(1\rightarrow3)$  links (the so-called comb-like structure, as for instance, in *Candida maltosa* and *C. Tropicalis*<sup>2</sup>). A more complex structure (a treelike structure) has been reported for *C. Albicans*,<sup>3,4</sup> which shows an analogous backbone of  $\alpha$ -(1 $\rightarrow$ 6)-linked mannopyranosyl residues with (1 $\rightarrow$ 2) and/or (1 $\rightarrow$ 3) side

ORDER		REPRINTS
-------	--	----------



chains, some of which are branched. A third type of sequence has been proposed for the *C*. *Krusei* mannan,<sup>5</sup> which contains a main chain of mannopyranose with  $(1\rightarrow 2)$  and  $(1\rightarrow 6)$  linkages in a 3:1 ratio but is lightly branched, either at the 2- or 6-positions.

In 1995, Jiménez-Barbero et al.<sup>6</sup> investigated the structures of cell-wall mannans isolated from *Microsporum gypseum* and related species of *Trychophyton* and found that all of them consist of an  $\alpha$ -D-mannopyranosyl- $(1\rightarrow 2)$ - $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ - $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -D-mannopyranose repeating tetrasaccharide. Syntheses of the fragments of polysaccharides are important for elucidation of biological functions of polysaccharides. In this paper, as a part of our continuous synthetic approach directed toward epitopes of fungal cell-wall mannans, we report the synthesis of a tetrasaccharide 1, the repeating unit of mannans isolated from *Microsporum gypseum* and related species of *Trychophyton*.

## **RESULTS AND DISCUSSION**

Tritylation of mannose (2) followed by benzoylation in a one-pot manner gave the 1,2,3,4-tetra-O-benzoyl-6-O-trityl-D-mannopyranose, selective acetolysis of which using CH<sub>2</sub>Cl<sub>2</sub>-AcOH-Ac<sub>2</sub>O-H<sub>2</sub>SO<sub>4</sub> in a ratio of 1:1:0.6:0.175 afforded the corresponding 1,6-diacetate **3** in 71.3% yield (for three steps). The diacetate **3** was selectively deacetylated at the anomeric position with benzylamine in THF in high yield to give the corresponding 6-O-acetyl-2,3,4-tri-O-benzoyl-D-mannopyranose (**4**). Subsequent reaction of **4** with CCl<sub>3</sub>CN/DBU in dichloromethane afforded the glycosyl donor **5**. A similar procedure gave another glycosyl donor **7** (Scheme 1).



Scheme 1. Reagents and conditions: (a) i.trityl chloride (1.2 equiv), pyridine, 50°C, 32 h; ii. Ph-COCl (4.8 equiv),  $<40^{\circ}$ C, 24 h; iii.CH<sub>2</sub>Cl<sub>2</sub>/HOAc/Ac<sub>2</sub>O/H<sub>2</sub>SO<sub>4</sub> = 1/1/0.6/0.175 (v/v), rt, 20 h, 71.3% (for three steps); (b) benzylamine (3.2 equiv), THF, rt, 24 h, 86.2%.(c) CCl<sub>3</sub>CN (2.3–3.2 equiv), DBU (0.18–0.23 equiv), rt, 5 h, 88.1%–85.6%.

286



	REPRINTS
--	----------



*Scheme 2.* Reagents and conditions: (a) PhCOCl (3.3 equiv), pyridine,  $<40^{\circ}$ C, 24 h, 94.8%; (b) F<sub>3</sub>CCOOH (90%), rt, 4 h, 89.0%; (c) (Ac)<sub>2</sub>O, pyridine, rt, 5 h, 100%; (d) benzylamine (4.0 equiv), THF, rt, 24 h, 88.4%.(e) CCl<sub>3</sub>CN (3.3 equiv), DBU (0.3 equiv), rt, 5 h, 86.3%.

Benzoylation of 1,2-*O*-ethylidene- $\beta$ -D-mannopyranose<sup>7</sup> (8) followed by hydrolysis with 90% CF<sub>3</sub>COOH afforded the 3,4,6-tri-*O*-benzoyl-D-mannopyranose (10), subsequent acetylation with acetic anhydride in pyridine furnished the diacetate 11. Selective removal of the 1-*O*-acetyl group with benzylamine in THF ( $\rightarrow$ 12), and then treatment with CCl<sub>3</sub>CN/DBU in dichloromethane afforded the glycosyl donor 13. (Scheme 2).

As shown in Scheme 3, allyl 6-*O*-acetyl-2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranoside (14) was prepared by the Helferich reaction using 3 as the glycosyl donor and allyl alcohol as the acceptor.<sup>8</sup> Selective removal of the acetyl group of 14 in



*Scheme 3.* Reagents and conditions: (a) Allyl alcohol (2 equiv.), TMSOTf (0.26 equiv), CH<sub>2</sub>Cl<sub>2</sub>, rt, 3 h, 88%. (b) methanol/0.5% HCl, rt, 12–14 h, 93%–96%. (c) **5** (1.4 equiv), CH<sub>2</sub>Cl<sub>2</sub>, TMSOTf (0.08 equiv), rt, 3 h, 86.5%. (d) **13** (1.4 equiv), CH<sub>2</sub>Cl<sub>2</sub>, TMSOTf (0.16 equiv), rt, 3 h, 85.7%. (e) **7** (5 equiv), CH<sub>2</sub>Cl<sub>2</sub>, TMSOTf (0.3 equiv), rt, 3 h, 80.4%. (f) i. PdCl<sub>2</sub>, CH<sub>3</sub>OH, rt, 4 h; ii. CH<sub>3</sub>OH satd with dry NH<sub>3</sub>, rt, 72 h, 70.2% (two steps).



ORDER		REPRINTS
-------	--	----------

methanol solution containing 0.5% HCl gave the glycosyl acceptor 15 in a high yield. The disaccharide 16 was prepared using the "inverse Schmidt" strategy.<sup>9</sup> Thus the glycosyl acceptor 15 and the catalyst TMSOT were mixed first in dry  $CH_2Cl_2$ , and after stirring for 15 min, the glycosyl donor 5 in dry dichloromethane was added dropwise within 30 min in order to give 16 in a high yield. Selective removal of the acetyl group of 16 gave the glycosyl acceptor 17, "inverse Schmidt" coupling of which with 2-O-acetyl-3,4,6-tri-O-benzoyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (13) afforded the trisaccharide 18 in 85.7% yield. The  $^{1}$ H NMR spectrum of **18** showed one acetyl signal ( $\delta$  2.14), one allyl signal, both signals characteristic of the structure of the trisaccharide 18. Selective removal of the 2-O-acetyl group of the trisaccharide 18 gave the glycosyl accepter 19. The fully protected tetrasaccharide 20 was smoothly obtained using the "inverse Schmidt" method, again by coupling with 2,3,4,6-tetra-O-benzoyl- $\alpha$ -D-mannopyranosyl trichloroacetimidate (7). Deprotection of **20** using PdCl<sub>2</sub> in CH<sub>3</sub>OH,<sup>10</sup> followed by treatment with NH<sub>3</sub> in CH<sub>3</sub>OH, gave the title mannotetraose 1. The bioassay of 1 is in process.

## **EXPERIMENTAL**

**General Methods.** Optical rotations were determined at 25°C with a Perkin-Elmer Model 241-Mc automatic polarimeter. Melting points were determined with a "Mel-Temp" apparatus. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded with Bruker ARX 400 spectrometers for solutions in CDCl<sub>3</sub>. Chemical shifts are given in parts per million (ppm) downfield from internal Me<sub>4</sub>Si. Mass spectra were recorded with a JMS-D300S mass spectrometer using a direct sample introduction technique. Thin-layer chromatography (TLC) was performed on Silica Gel HF<sub>254</sub> with detection by charring with 30% (v/v) H<sub>2</sub>SO<sub>4</sub> in MeOH or in some cases by UV detection. Column chromatography was conducted by elution of a column (16 × 240, 18 × 300, 35 × 400 mm) of silica gel (100–200 mesh) with EtOAc - petroleum ether (60–90°C) as the eluent. Solutions were concentrated at < 60°C under diminished pressure.

**1,6-Di-***O***-acetyl-2,3,4-tri-***O***-benzoyl-** $\alpha$ **-D-mannopyranose (3).** A solution of mannose **2** (15 g, 83.3 mmol) and trityl chloride (28 g, 100.5 mmol) in pyridine (100 mL) was stirred at 50°C for 32 h, at the end of which time TLC (4:1 EtOAc-methanol) indicated that the reaction was complete. The reaction mixture was cooled to 0°C, and then benzoyl chloride (46.5 mL, 400.9 mmol) was added dropwise for 30 min to keep the reaction temperature under 40°C. After 24 h, water (300 mL) was added to the reaction mixture, and stirring was continued for 30 min. The aq solution was extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 100 mL), the extract was washed with HCl (1 N) and saturated aqueous sodium bicarbonate, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to dryness. The residue without separation was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (50 mL), Ac<sub>2</sub>O (50 mL) and AcOH (30 mL), the solution cooled to 10°C in an ice bath, and H<sub>2</sub>SO<sub>4</sub> (8.8 mL) added dropwise over 20 min. After the addition





was complete, the ice bath was removed and the reaction was continued for 20 h at ambient temperature. The reaction solution was poured into ice water (400 mL), stirring was continued for an additional 15 min, and the aqueous solution was extracted with chloroform ( $3 \times 100$  mL). The combined chloroform extracts were carefully washed with 10% aq NaHCO<sub>3</sub> ( $3 \times 60$  mL), dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated to a syrup which was subjected to column chromatography with 4:1 petroleum ether-EtOAc as the eluent. Compound **3** was obtained as a syrup (34.2 g, 71.3%); [ $\alpha$ ]<sub>D</sub> + 9.5° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.11–7.28 (m, 15 H, 3 PhH), 6.36 (d, 1 H, J<sub>1,2</sub> = 2.0 Hz, H-1), 6.01 (t, 1 H, J<sub>3,4</sub> = J<sub>4,5</sub> = 10.2 Hz, H-4), 5.88 (dd, 1 H, J<sub>2,3</sub> = 3.2 Hz, J<sub>3,4</sub> = 10.2 Hz, H-3), 5.70 (dd, 1 H, J<sub>1,2</sub> = 2.0 Hz, J<sub>2,3</sub> = 3.2 Hz, H-2), 4.34 (m, 2 H, H-5, 6a), 4.27 (dd, 1 H, J<sub>6a,6b</sub> = 13.4 Hz, J<sub>5,6b</sub> = 4.2 Hz, H-6b), 2.28, 2.08 (2 s, 6 H, 2COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): 170.0, 167.7 (2 C, 2 COCH<sub>3</sub>), 165.1, 165.0, 164.6 (3 C, 3 COPh), 133.3–127.8 (Ph), 90.2 (C-1), 70.2, 69.3, 68.8, 65.9 (4 C, C-2, 3, 4, 5), 62.0 (C-6), 20.4, 20.1 (2 C, 2 COCH<sub>3</sub>).

Anal. Calcd for C<sub>31</sub>H<sub>28</sub>O<sub>11</sub>: C, 64.58; H, 4.89. Found: C, 64.67; H, 4.94.

**6-O-Acetyl-2,3,4-tri-O-benzoyl-α-D-mannopyranose** (**4**). A solution of compound **3** (5 g, 8.67 mmol) and benzylamine (3 mL, 27.4 mmol) in anhydrous THF (30 mL) was stirred at room temperature for 24 h, at the end of which time TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solution was concentrated. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound **4** as a syrup (4.0 g, 86.2%); [α]<sub>D</sub> + 16.8° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): δ 8.11–7.27 (m, 15 H, 3 PhH), 5.98 (m, 2 H, H-3, 4), 5.71 (dd, 1 H, J<sub>1,2</sub> = 1.6Hz, J<sub>2,3</sub> = 2.4 Hz, H-2), 5.53 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-), 4.55 (m, 1 H, H-5), 4.33 (m, 2 H, H-6a, 6b), 2.09( s, 3 H,COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCI<sub>3</sub>) ; 170.6 (COCH<sub>3</sub>), 166.1, 165.2, 165.2 (3 C, 3 *C*OPh), 133.1–127.9 (Ph), 91.7 (C-1), 70.6, 69.4, 68.0, 66.5 (4 C, C-2, 3, 4, 5), 62.4 (C-6), 20.3 (*C*OCH<sub>3</sub>).

Anal. Calcd for C<sub>29</sub>H<sub>26</sub>O<sub>10</sub>: C, 65.17; H, 4.90. Found: C, 65.42; H, 4.94.

**6-O-Acetyl-2,3,4-tri-O-benzoyl-α-D-mannopyranosyl trichloroacetimidate (5).** A mixture of **4** (4.0 g, 7.48 mmol), trichloroacetonitrile (2.1 mL, 20.9 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.25 mL, 1.67 mmol) in dry dichloromethane (25mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **5** (4.47 g, 88.1%) as crystals; mp 88–91°C;  $[\alpha]_D + 20.3°$  (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): δ 8.87 (s, 1 H, OC(NH)CCI<sub>3</sub>), 8.12–7.27 (m, 15 H, 3 PhH), 6.55 (d, 1 H, J<sub>1,2</sub> = 2.0 Hz, H-1), 6.06 (t, 1 H, J<sub>3,4</sub> = J<sub>4,5</sub> = 10.0 Hz, H-4), 5.93 (dd, 1 H, J<sub>2,3</sub> = 3.2 Hz, J<sub>3,4</sub> = 10.0 Hz, H-3), 5.90 (dd, 1 H, J<sub>1,2</sub> = 2.0 Hz, J<sub>2,3</sub> = 3.2 Hz, H-2), 4.49 (m, 1 H, H-5), 4.34 (m, 2 H, H-6a, 6b), 2.07 (s, 3 H, COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCI<sub>3</sub>): 170.0 (COCH<sub>3</sub>), 165.0, 164.6, 159.4 (4 C, 3 COPh, *C*(NH)CCI<sub>3</sub>), 133.3–127.9 (Ph), 94.1 (C-1), 90.1 (*C*CI<sub>3</sub>), 70.8, 69.2, 68.3, 65.7 (4 C, C-2, 3, 4, 5), 61.9 (C-6), 20.1 (*COCH<sub>3</sub>*).

Anal. Calcd for C<sub>31</sub>H<sub>26</sub>NO<sub>10</sub>CI<sub>3</sub>: C, 54.84; H, 3.86. Found: C, 54.72; H, 3.90.

Marcel Dekker, Inc.

270 Madison Avenue, New York, New York 10016

ORDER		REPRINTS
-------	--	----------

HENG, NING, AND KONG

**2,3,4,6-Tetra-***O***-benzoyl-** $\alpha$ **-D-mannopyranosyl trichloroacetimidate (7).** A mixture of 2,3,4,6-tetra-*O*-benzoyl- $\alpha$ -D-mannopyranose (6)<sup>11</sup> (3.5 g, 5.87 mmol), trichloroacetonitrile (1.9 mL, 18.9 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.20 mL, 1.34 mmol) in dry dichloromethane (30 mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give 7 (3.72 g, 85.6%) as crystals; mp 132–134°C; [ $\alpha$ ]<sub>D</sub> +48.3° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.86 (s, 1 H, OC(NH)CCI<sub>3</sub>), 8.10–7.26 (m, 20 H, 4 PhH), 6.57 (d, 1 H, J<sub>1,2</sub> = 1.3 Hz, H-1), 6.23 (t, 1 H, J<sub>3,4</sub> = J<sub>4,5</sub> = 10.0 Hz, H-4), 5.99–5.94 (, 2 H, H-2, 3), 4.72 (dd, 1 H, J<sub>5,6a</sub> = 2.4 Hz, J<sub>6b,6a</sub> = 12.3 Hz, H-6b), 4.62 (m, 1 H, H-5), 4.50 (dd, 1 H, J<sub>5,6b</sub> = 4.0 Hz, J<sub>6b,6a</sub> = 12.3 Hz, H-6b); <sup>13</sup>C NMR (100 MHz, CDCI<sub>3</sub>), 165.5, 165.0, 164.9, 164.6 (4 C, 4 COPh), 159.0 (*C*(NH)CCI<sub>3</sub>), 133.2–127.9 (Ph), 94.2 (C-1), 71.1, 69.4, 68.4, 65.6 (4 C, C-2, 3, 4, 5), 61.95 (C-6).

Anal. Calcd for C<sub>36</sub>H<sub>28</sub>NO<sub>10</sub>CI<sub>3</sub>: C, 58.35; H, 3.81. Found: C, 58.44; H, 3.76.

**3,4,6-Tri**-*O*-benzoyl-1,2-*O*-(*S*-ethylidene)-β-D-mannopyranose (9). 1.2-O-(S-ethylidene)- $\beta$ -D-mannopyranose (8)<sup>7</sup> (6.4 g, 31.1 mmol) in pyridine (30 ml) was cooled to 0°C, and then benzoyl chloride (11.9 mL, 102.5mmol) was added dropwise for 30 min to keep the reaction temperature under 40°C. After 24 h, water (300 mL) was added to the reaction mixture, and stirring was continued for 30 min. The aq solution was extracted with  $CH_2Cl_2$  (3  $\times$  100 mL), the extract was washed with HCl (1 N) and then saturated sodium bicarbonate, dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated to dryness. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **9** (15.3 g, 94.8%) as crystals; mp 123–125°C;  $[\alpha]_D$ + 25.9° (c 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400MHz, CDCI<sub>3</sub>):  $\delta$  8.04–7.25 (m, 15 H, 3 Ph*H*), 6.01 (t, 1 H,  $J_{3,4} = J_{5,4} = 10.2$  Hz, H-4), 5.80 (dd, 1 H,  $J_{3,2} = 3.6$  Hz,  $J_{4,3}$ =10.2 Hz, H-3), 5.51 (dd, 1 H, J<sub>1.2</sub> = 2.0 Hz, J<sub>3.2</sub> = 3.6 Hz, H-2), 5.45 (q, 1 H, J = 5.0 Hz, CH<sub>3</sub>CH), 5.36 (d, 1 H, J<sub>2.1</sub> = 2.0 Hz, H-1), 4.70 (m, 1 H, H-5), 4.50–4.44  $(m, 2 H, H-6, 6'), 1.57 (d, 3 H, J = 5.0 Hz, CH_3CH); {}^{13}C NMR (100 MHz, CDCl_3),$ 165.7, 165.5, 164.8 (3 C, 3 COPh), 133.0–127.8 (Ph), 104.4 (CHCH<sub>3</sub>), 96.3 (C-1), 77.06, 71.4, 71.0, 66.3 (4 C, C-2, 3, 4, 5), 62.9 (C-6), 21.1 (CHCH<sub>3</sub>).

Anal. Calcd for C<sub>29</sub>H<sub>26</sub>O<sub>9</sub>: C, 67.18; H, 5.05. Found: C, 67.33; H, 5.00.

**3,4,6-Tri-***O***-benzoyl-D-mannopyranose (10)**. A solution of compound **9** (7.2 g, 13.9 mmol) in 90% F<sub>3</sub>CCOOH (30 mL) was stirred at room temperature for 4 h, at the end of which time TLC (1:1 petroleum ether-EtOAc) indicated that the reaction was complete. The reaction mixture was diluted with toluene (50 mL) and then concentrated to dryness. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound 10 as a crystalline mixture of  $\alpha$  and  $\beta$  forms in a ratio of 2:1 (6.1 g, 89.0%); mp 99–101°C; [ $\alpha$ ]<sub>D</sub> + 40.8° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.04–7.27 (m, 15 H, 3 PhH), 6.00 (t, 0.66 H, J<sub>3,4</sub> = J<sub>5,4</sub> = 9.8 Hz, H-4 of  $\alpha$  anomer), 5.92 (t, 0.34 H, J<sub>3,4</sub> = J<sub>5,4</sub> = 9.9 Hz, H-4 of  $\beta$  anomer), 5.76 (dd, 0.66 H, J<sub>3,2</sub> = 3.2 Hz, J<sub>4,3</sub> = 9.8 Hz, H-2 of  $\alpha$  anomer), 5.44 (dd, 0.34 H, J<sub>3,2</sub> = 3.1 Hz, J<sub>4,3</sub> = 9.7 Hz, H-2 of  $\beta$  anomer), 5.43 (d, 0.66 H, J<sub>2,1</sub> = 1.0 Hz, H-1 of  $\alpha$  anomer), 5.1 (d, 0.34 H, J<sub>2,1</sub> = 1.1 Hz, H-1 of  $\beta$  anomer),

Downloaded by [Umeå University Library] at 20:18 18 November 2014





Downloaded by [Umeå University Library] at 20:18 18 November 2014

4.67–4.37 (m, 4 H, H-2, 5, 6, 6'); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): 166.1, 165.5, 165.3 (3 C, 3 COPh), 133.3–127.9 (Ph), 94.0, (C-1), 72.2, 69.2, 68.0, 66.5 (4 C, C-2, 3, 4, 5), 63.2 (C-6).

Anal. Calcd for C<sub>27</sub>H<sub>24</sub>O<sub>9</sub>: C, 65.85; H, 4.91. Found: C, 66.02; H, 4.95.

**1,2-Di-***O***-acetyl-3,4,6-tri-***O***-benzoyl-\alpha-D-mannopyranose (11)**. Acetylation of **10** (3 g, 6.1 mmol) with acetic anhydride (8 mL) in pyridine (10 mL) at room temperature for 4 h gave compound **11** in a quantitative yield as crystals; mp 115–117°C; [ $\alpha$ ]<sub>D</sub> + 35.9° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400MHz, CDCI<sub>3</sub>):  $\delta$  8.05–7.27 (m, 15 H, 3 Ph*H*), 6.24 (d, 1 H, J<sub>2,1</sub> = 2.0 Hz, H-1), 6.01 (t, 1 H, J<sub>3,4</sub> = J<sub>5,4</sub> = 10 Hz, H-4), 5.80 (dd, 1 H, J<sub>3,2</sub> = 3.6 Hz, J<sub>4,3</sub> = 10.2 Hz, H-3), 5.51 (dd, 1 H, J<sub>1,2</sub> = 2.0 Hz, J<sub>3,2</sub> = 3.6 Hz, H-2), 4.63 (m, 1 H, H-5), 4.49–4.43 (m, 2 H, H-6,6'), 2.26, 2.18 (2 s, 6 H, 2 COCH<sub>3</sub>); <sup>13</sup>C NMR (75 MHz, CDCI<sub>3</sub>): 169.5, 168.2 (2C, 2 COCH<sub>3</sub>), 166.0, 165.6, 165.3 (3C, 3 COPh), 133.6–128.3 (Ph), 90.6 (C-1), 70.8, 69.5, 68.7, 66.4 (4 C, C-2, 3, 4, 5), 62.8 (C-6), 21.0, 20.7 (2 C, 2 COCH<sub>3</sub>). Anal. Calcd for C<sub>31</sub>H<sub>28</sub>O<sub>11</sub>: C, 64.58; H, 4.89. Found: C, 64.70; H, 4.93.

**2-0-Acetyl-3,4,6-tri-***O***-benzoyl-α-D-mannopyranose** (**12**). A solution of compound **11** (8 g, 13.8 mmol) and benzylamine (6 mL, 54.9 mmol) in anhydrous THF (50 mL) was stirred at room temperature for 24 h, at the end of which time TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solution was concentrated. Purification by flash column chromatography on silica gel (3:1 petroleum ether-EtOAc) gave compound **12** as crystals (6.56g, 88.4%); mp 89–91°C;  $[\alpha]_D$  + 39.8° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): δ 8.07–7.34 (m, 15 H, 3 Ph*H*), 5.96 (t, 1 H, J<sub>3,4</sub> = J<sub>5,4</sub> = 10 Hz, H-4), 5.87 (dd, 1 H, J<sub>3,2</sub> = 3.2 Hz, J<sub>4,3</sub> = 10.0 Hz, H-3), 5.50 (dd, 1 H, J<sub>1,2</sub> = 2.0 Hz, J<sub>3,2</sub> = 3.2 Hz, H-2), 5.37 (d, 1 H, J<sub>2,1</sub> = 2.0 Hz, H-1), 4.66 (dd, 1 H, J<sub>6,6'</sub> = 12.0 Hz, J<sub>5,6</sub> = 2.8 Hz, H-6), 4.60 (m, 1 H, H-5), 4.44 (dd, 1 H, J<sub>6,6'</sub> = 12.0 Hz, J<sub>5,6'</sub> = 4.4 Hz, H-6), 2.18 (s, 3H, COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 169.6 (COCH<sub>3</sub>), 165.9, 165.1 (3 C, 3 COPh), 132.9–127.9 (Ph), 91.7 (C-1), 70.0, 69.1, 68.4, 66.6 (4 C, C-2, 3, 4, 5), 62.7 (C-6), 20.3 (COCH<sub>3</sub>).

Anal. Calcd for C<sub>29</sub>H<sub>26</sub>O<sub>10</sub>: C, 65.17; H, 4.90. Found: C, 65.02; H, 4.85.

**2-O-Acetyl-3,4,6-tri-O-benzoyl-α-D-mannopyranosyl trichloroacetimidate (13).** A mixture of **12** (4.9 g, 9.17 mmol), trichloroacetonitrile (3.0 mL, 30.0 mmol), and 1,8-diazabicyclo[5.4.0]undecene (DBU) (0.40 mL, 2.7 mmol) in dry dichloromethane (40 mL) was stirred under nitrogen for 5 h and then concentrated *in vacuo*. The residue was purified by flash chromatography (4:1 petroleum ether-EtOAc) to give **13** (5.37 g, 86.3%) as crystals; mp 87–90°C;  $[\alpha]_D$  + 36.7° (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): δ 8.82 (s, 1 H, OC(NH)CCI<sub>3</sub>), 8.04–7.36 (m, 15 H, 3 Ph*H*), 6.42 (d, 1 H, J<sub>1,2</sub> = 2.0 Hz, H-1), 6.03 (t, 1 H, J<sub>3,4</sub> = J<sub>4,5</sub> = 10.0 Hz, H-4), 5.85 (dd, 1 H, J<sub>2,3</sub> = 3.4 Hz, J<sub>3,4</sub> = 10.0 Hz, H-3), 5.71 (dd, 1 H, J<sub>1,2</sub> = 2.0 Hz, J<sub>2,3</sub> = 3.4 Hz, H-2), 4.64 (dd, 1 H, J<sub>6,6'</sub> = 12.8 Hz, J<sub>5,6</sub> = 7.0 Hz, H-6), 4.56 (m, 1 H, H-5), 4.46 (dd, 1 H, J<sub>6,6'</sub> = 12.8 Hz, J<sub>5,6'</sub> = 2.8 Hz, H-6'), 2.17 (s, 3H, COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCI<sub>3</sub>), 169.5 (COCH<sub>3</sub>), 165.9, 165.1, 165.0,

ORDER		REPRINTS
-------	--	----------

164.9(4 C, 3 COPh, *C*(NH)CCl<sub>3</sub>), 133.2–127.9 (Ph), 95.7 (C-1), 91.7 (C(NH)*C*Cl<sub>3</sub>), 69.9, 69.1, 68.4, 66.6 (4 C, C-2, 3, 4, 5), 62.7 (C-6), 20.3 (CO*C*H<sub>3</sub>). Anal. Calcd for C<sub>31</sub>H<sub>26</sub>NO<sub>10</sub>Cl<sub>3</sub>: C, 54.84; H, 3.86. Found: C, 54.69; H, 3.98.

Allyl 6-O-Acetyl-2,3,4-tri-O-benzoyl- $\alpha$ -D-mannopyranoside (14). A solution of 3 (2.5 g, 4.3 mmol) and allyl alcohol (0.6 mL, 8.8 mmol) in dry dichloromethane (40 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and then TMSOTf (0.2 mL, 1.1 mmol) was added dropwise. After 1 h the reaction mixture was diluted with dichloromethane (50 mL) and washed with a satd sodium hydrogen carbonate solution (15 mL). The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. Purification of the residue by flash chromatography (3:1 petroleum ether-EtOAc) gave 14 as a syrup (2.19 g, 88%);  $[\alpha]_{D}$  + 15.9° (c 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400MHz, CDCI<sub>3</sub>):  $\delta$  8.11–7.26 (m, 15 H, 3 PhH), 6.12–5.91 (m, 3 H, H-3, 4,  $CH_2CH=CH_2$ ), 5.68 (dd,  $J_{1,2} = 2.0$  Hz,  $J_{3,2} = 2.8$  Hz, H-2), 5.40 (dd, 1 H, <sup>2</sup>J = 1.6Hz, <sup>3</sup>J<sub>trans</sub> = 17.2 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.30 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{cis} = 10.4$  Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.14 (d, J<sub>1,2</sub> = 2.0 Hz, H-1), 4.47–4.27 (m, 4 H, H-6, 6', CH<sub>2</sub>CH=CH<sub>2</sub>), 4.17 (m, 1 H, H-5), 2.10 (s, 3 H,COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 170.1 (COCH<sub>3</sub>), 165.7, 165.6, 165.5 (3 C,3 COPh), 133.7–128.4 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.7 (CH<sub>2</sub>CH=CH<sub>2</sub>), 96.7 (C-1), 70.5, 70.0, 69.0, 68.7, 66.9 (5 C, C-2, 3, 4, 5, CH<sub>2</sub>CH=CH<sub>2</sub>), 62.8 (C-6), 20.8 (COCH<sub>3</sub>).

Anal. Calcd for C<sub>32</sub>H<sub>30</sub>O<sub>10</sub>: C, 66.89; H, 5.26. Found: C, 66.93; H, 5.31.

Allyl 2,3,4-Tri-O-benzoyl- $\alpha$ -D-mannopyranoside (15). A solution of 14 (1.6 g, 2.8 mmol) in methanol solution (80 mL) containing 0.5 % HCl was stirred at room temperature for 12 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and  $CH_2Cl_2$ , then the organic layer dried over  $Na_2SO_4$ , and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 15 as a syrup (1.42 g, 96%);  $[\alpha]_D$ + 14.2° (c 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400MHz, CDCI<sub>3</sub>):  $\delta$  8.12–7.24 (m, 15 H,3 Ph*H*), 6.03–5.98 (m, 2 H, H-3, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.85 (t, 1 H,  $J_{3,4} = J_{4,5} = 10.0$  Hz, H-4), 5.69 (dd,  $J_{1,2} = 1.6$ Hz,  $J_{2,3} = 3.2$ Hz, H-2), 5.40 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>trans</sub> = 17.2 Hz,  $CH_2CH=CH_2$ ), 5.29 (dd, 1 H, <sup>2</sup>J = 1.6Hz, <sup>3</sup>J<sub>cis</sub> = 10.4 Hz,  $CH_2CH=CH_2$ ), 5.16 (d,  $J_{1,2} = 1.6$  Hz, H-1), 4.34–4.16 (m, 2 H,  $CH_2CH=CH_2$ ), 4.11 (m, 1 H, H-5), 3.83–3.79 (m, 2 H, H-6,6'); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>). 166.6, 165.6, 165.5 (3 C, 3 COPh), 133.7–128.3 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.5 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 96.8 (C-1), 71.0, 70.7, 69.6, 68.8, 67.3 (5 C, C-2, 3, 4, 5,  $CH_2CH=CH_2$ ), 61.3 (C-6).

Anal. Calcd for C<sub>30</sub>H<sub>28</sub>O<sub>9</sub>: C, 67.66; H, 5.30. Found: C, 67.83; H, 5.37. .

Allyl 6-*O*-Acetyl-2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranosyl-(1 $\rightarrow$ 6)-2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranoside (16). A solution of 15 (0.37 g, 0.69 mmol) and TMSOTf (10  $\mu$ L, 0.055mmol) in dry dichloromethane (6 mL) was stirred with



Copyright © Marcel Dekker, Inc. All rights reserved



dried molecular sieves (4 A, 0.4 g) under nitrogen for 15 min, and then 5 (0.66 g, 0.97 mmol) in dichloromethane (4 mL) was added dropwise for 20 min. After 3 h the reaction mixture was diluted with dichloromethane (30 mL) and washed with satd sodium hydrogen carbonate solution (5 mL). The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 16 as a syrup (0.63 g, 86.5%);  $[\alpha]_D$  + 35.3° (c 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): δ 8.08–7.27 (m, 30 H, 6 PhH),  $6.07-5.91 \text{ (m, 5 H, H-3_A, 3_B, 4_A, 4_B, CH_2CH=CH_2)}, 5.77 \text{ (dd, 1 H, J}_{1,2} = 1.5 \text{ Hz},$  $J_{2,3} = 3.2 \text{ Hz}, \text{H-2}_{A}$ ), 5.74 (dd, 1 H,  $J_{1,2} = 1.5 \text{ Hz}, J_{2,3} = 3.2 \text{ Hz}, \text{H-2}_{B}$ ), 5.50 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{trans} = 17.2$  Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.35 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{cis} =$ 10.4 Hz,), 5.18 (d, 1 H,  $J_{1,2} = 1.5$  Hz, H-1<sub>A</sub>), 5.13 (d, 1 H,  $J_{1,2} = 1.5$  Hz, H-1<sub>B</sub>),  $4.45-3.76 (m, 8 H, H-5_A, 5_B, 6_A, 6_b, 6_A', 6_B', CH_2CH=CH_2), 1.92 (s, 3 H, COCH_3);$ <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 169.5 (COCH<sub>3</sub>), 164.6, 164.5, 164.2, 164.1 (6 C, 6 COPh), 132.6–127.3 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>). 117.6 (CH<sub>2</sub>CH=CH<sub>2</sub>), 96.4, 95.7 (2 C, C-1<sub>A</sub>, 1<sub>B</sub>), 69.5, 69.1, 68.8, 68.5, 67.9, 67.7, 65.9, 65.7, 61.5 (11 C, C-2<sub>A</sub>, 2<sub>B</sub>, 3<sub>A</sub>, 3<sub>B</sub>,  $4_A, 4_B, 5_A, 5_B, 6_A, 6_B, CH_2CH = CH_2$ ), 19.5 (COCH<sub>3</sub>).

Anal. Calcd for C<sub>59</sub>H<sub>52</sub>O<sub>18</sub>: C, 67.55; H, 5.00. Found: C, 67.34; H, 4.96.

Allyl 2,3,4-Tri-O-benzovl- $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-ben $zoyl-\alpha$ -D-mannopyranoside (17). A solution of 16 (0.55 g, 0.52 mmol) in methanol solution (25 mL) containing 0.5 % HCl was stirred at room temperature for 14 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and  $CH_2Cl_2$ , then the organic layer dried over  $Na_2SO_4$ , and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 17 as a syrup (0.5 g, 95.6%);  $[\alpha]_{\rm D}$  + 31.0° (c 1.5, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.17–7.26 (m, 30 H, 6 PhH), 6.06–5.80 (m, 5 H, H-3<sub>A</sub>, 3<sub>B</sub>, 4<sub>A</sub>, 4<sub>B</sub>, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.76–5.73 (m, 2 H, H-2<sub>A</sub>, 2<sub>B</sub>), 5.34 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{trans} = 17.2$  Hz, CH<sub>2</sub>CH=CH<sub>2</sub> ), 5.50 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{cis} = 10.4 \text{ Hz}$ , 5.17 (d, 1 H,  $J_{1,2} = 1.6 \text{ Hz}$ , H-1<sub>A</sub>), 5.14 (d, 1 H,  $J_{1,2} = 1.6 \text{ Hz}$ , H- $1_{\rm B}$ ), 4.44–3.55 (m, 8 H, H- $5_{\rm A}$ ,  $5_{\rm B}$ ,  $6_{\rm A}$ ,  $6_{\rm B}$ ,  $6_{\rm A}'$ ,  $6_{\rm B}'$ ,  $CH_2CH=CH_2$ ); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 166.7, 165.7, 165.5, 165.4, 165.2 (6 C, 6 COPh), 133.7–128.3 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.6 (CH<sub>2</sub>CH=CH<sub>2</sub>), 97.8, 96.7 (2 C, C-1<sub>A</sub>, 1<sub>B</sub>), 70.9, 70.6, 70.4, 70.2, 69.5, 68.9, 67.1, 67.0, 66.7, 61.0 (11 C, C-2<sub>A</sub>, 2<sub>B</sub>, 3<sub>A</sub>, 3<sub>B</sub>, 4<sub>A</sub>, 4<sub>B</sub>, 5<sub>A</sub>, 5<sub>B</sub>, 6<sub>A</sub>,  $6_{\rm B}, CH_2CH=CH_2$ ).

Anal. Calcd for C<sub>57</sub>H<sub>50</sub>O<sub>17</sub>: C, 67.99; H, 5.00. Found: C, 68.11; H, 5.02.

Allyl 2-*O*-Acetyl-3,4,6-tri-*O*-benzoyl- $\alpha$ -D-mannopyranosy- $(1\rightarrow 6)$ -2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-*O*-benzoyl- $\alpha$ -D-mannopyranoside (18). A solution of 17 (0.25 g, 0.25 mmol) and trimethylsilyl trifluoromethanesulfonate (8  $\mu$ L, 0.04 mmol) in dry dichloromethane (20 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and 13 (0.24 g, 0.35 mmol) in dry dichloromethane (10 mL) was added dropwise for 30 min. After 3 h the reaction mixture was diluted with dichloromethane (20 mL) and

Marcel Dekker, Inc.

270 Madison Avenue, New York, New York 10016

ORDER		REPRINTS
-------	--	----------

### HENG, NING, AND KONG

washed with satd sodium hydrogen carbonate solution (8 mL). The organic layer was dried with Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. Purification of the residue by column chromatography (2:1 petroleum ether-EtOAc) gave **18** as a syrup (0.32 g, 85.7%);  $[\alpha]_D + 28.7^{\circ}$  (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.10–7.25 (m, 45 H, 9 PhH), 6.21–5.88 (m, 7 H, H-3<sub>A</sub>, 3<sub>B</sub>, 3<sub>C</sub>, 4<sub>A</sub>, 4<sub>B</sub>, 4<sub>C</sub>, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.84 (dd, 1 H, J<sub>1,2</sub> = 1.6 Hz, J<sub>2,3</sub> = 3.2 Hz, H-2<sub>A</sub>), 5.79 (dd, 1 H, J<sub>1,2</sub> = 1.6 Hz, J<sub>2,3</sub> = 3.2 Hz, H-2<sub>B</sub>), 5.44 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>trans</sub> = 17.2 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.37 (dd, 1 H, J<sub>1,2</sub> = 1.6 Hz, J<sub>2,3</sub> = 3.2 Hz, H-2<sub>C</sub>), 5.26 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>cis</sub> = 10.4 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.20 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>A</sub>), 5.16 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>B</sub>), 4.73 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>C</sub>), 4.43–4.37 (m, 11 H, H-5<sub>A</sub>, 5<sub>B</sub>, 5<sub>C</sub>, 6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>, 6<sub>A</sub>', 6<sub>B</sub>', 6<sub>C</sub>', CH<sub>2</sub>CH=CH<sub>2</sub>), 2.07 (s, 3 H, COCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 169.5 (COCH<sub>3</sub>), 166.0, 165.7, 165.5, 165.3, 165.2 (9 C, 9 COPh), 133.4–128.3 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.8 (CH<sub>2</sub>CH=CH<sub>2</sub>), 98.0, 97.2, 96.9 (3 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>), 20.7 (COCH<sub>3</sub>).

Anal. Calcd for C<sub>86</sub>H<sub>74</sub>O<sub>26</sub>: C, 67.80; H, 4.90. Found: C, 68.04; H, 4.96.

Allyl 3,4,6-Tri-O-benzoyl- $\alpha$ -D-mannopyranosyl- $(1 \rightarrow 6)$ -2,3,4-tri-O-benzovl- $\alpha$ -D-mannopyranosyl- $(1 \rightarrow 6)$ -2,3,4-tri-O-benzovl- $\alpha$ -D-mannopyranoside (19). A solution of 18 (0.37 g, 0.24 mmol) in methanol solution (25 mL) containing 0.5 % HCl was stirred at room temperature for 14 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) indicated that the starting material had disappeared. The mixture was carefully neutralized with triethylamine, and then concentrated to dryness. The residue was partitioned between water and CH<sub>2</sub>Cl<sub>2</sub>, then the organic layer dried over  $Na_2SO_4$ , and concentrated to a syrup. Purification of the residue by flash chromatography (2:1 petroleum ether-EtOAc) gave 19 as a syrup (0.34 g, 93.5%);  $[\alpha]_{\rm D}$  + 23.0° (c 1.5, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>): $\delta$ 8.09–7.26 (m, 45 H, 9 PhH), 6.19–5.70 (m, 10 H, H-2<sub>A</sub>, 2<sub>B</sub>, 2<sub>C</sub>, 3<sub>A</sub>, 3<sub>B</sub>, 3<sub>C</sub>, 4<sub>A</sub>, 4<sub>B</sub>,  $4_{\rm C}$ , CH<sub>2</sub>CH=CH<sub>2</sub>), 5.48 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>trans</sub> = 17.2 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.29 (dd, 1 H,  ${}^{2}J = 1.6$  Hz,  ${}^{3}J_{cis} = 10.4$  Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.20 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>A</sub>), 5.17 (d, 1 H,  $J_{1,2} = 1.6$  Hz, H-1<sub>B</sub>), 4.75 (d, 1 H,  $J_{1,2} = 1.6$  Hz, H-1<sub>C</sub>), 4.43–4.37 (m, 11 H, H-5<sub>A</sub>, 5<sub>B</sub>, 5<sub>C</sub>, 6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>, 6<sub>A</sub>', 6<sub>B</sub>', 6<sub>C</sub>', CH<sub>2</sub>CH=CH<sub>2</sub>);  $^{13}$ C NMR (100 MHz, CDCl<sub>3</sub>), 166.1, 165.8, 165.7, 165.6, 165.5, 165.4, 165.3, 165.2, 165.1 (9 C, 9 COPh), 133.6–27.7 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.5 (CH<sub>2</sub>CH=CH<sub>2</sub>), 97.7, 96.8, 96.4 (3 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>).

Anal. Calcd for C<sub>84</sub>H<sub>72</sub>O<sub>25</sub>: C, 68.10; H, 4.90. Found: C, 67.99; H, 4.94.

Allyl 2,3,4,6-Tetra-O-benzoyl- $\alpha$ -D-mannopyranosyl- $(1\rightarrow 2)$ -3,4,6-tri-Obenzoyl- $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- $\alpha$ -D-mannopyranosyl- $(1\rightarrow 6)$ -2,3,4-tri-O-benzoyl- $\alpha$ -D-mannopyranoside (20). A solution of 19 (0.21 g, 0.14 mmol) and trimethylsilyl trifluoromethanesulfonate (8  $\mu$ L, 0.04 mmol) in dry dichloromethane (20 mL) was stirred with dried molecular sieves (4 A, 1 g) under nitrogen for 15 min, and then 7 (0.52 g, 0.70 mmol) in dry dichloromethane (10 mL) was added dropwise for 30 min. After 3 h the reaction mixture was diluted with dichloromethane (20 mL) and washed with satd sodium hydrogen carbonate solution (8 mL). The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and





concentrated *in vacuo*. Purification of the residue by column chromatography (2:1 petroleum ether-EtOAc) gave **20** as a syrup (0.23 g, 80.4%);  $[\alpha]_D + 14.7^{\circ}$  (*c* 1.0, CHCI<sub>3</sub>); <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>):  $\delta$  8.04–7.18 (m, 65 H, 13 Ph*H*), 6.21–5.80 (m, 13 H, H-2<sub>A</sub>, 2<sub>B</sub>, 2<sub>C</sub>, 2<sub>D</sub>, 3<sub>A</sub>, 3<sub>B</sub>, 3<sub>C</sub>, 3<sub>D</sub>, 4<sub>A</sub>, 4<sub>B</sub>, 4<sub>C</sub>, 4<sub>D</sub>, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.45 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>trans</sub> = 17.2 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.28 (dd, 1 H, <sup>2</sup>J = 1.6 Hz, <sup>3</sup>J<sub>cis</sub> = 10.4 Hz, CH<sub>2</sub>CH=CH<sub>2</sub>), 5.24 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>A</sub>), 5.22 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>B</sub>), 5.05 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>C</sub>), 4.87 (d, 1 H, J<sub>1,2</sub> = 1.6 Hz, H-1<sub>D</sub>), 4.43–4.37 (m, 14 H, H-5<sub>A</sub>, 5<sub>B</sub>, 5<sub>C</sub>, 5<sub>D</sub>, 6<sub>A</sub>, 6<sub>B</sub>, 6<sub>C</sub>, 6<sub>D</sub>, 6<sub>A</sub>', 6<sub>B</sub>', 6<sub>C</sub>', 6<sub>D</sub>', CH<sub>2</sub>CH=CH<sub>2</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>), 166.2, 165.9, 165.7, 165.6, 165.5, 165.4, 165.2, 165.0, 164.8 (13 C, 13 COPh), 133.4–128.3 (Ph, CH<sub>2</sub>CH=CH<sub>2</sub>), 118.4 (CH<sub>2</sub>CH=CH<sub>2</sub>), 99.7, 98.1, 98.0, 97.6 (4 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>).

Anal. Calcd for C<sub>118</sub>H<sub>98</sub>O<sub>34</sub>: C, 68.80; H, 4.79. Found: C, 68.97; H, 4.74.

α-D-Mannopyranosyl- $(1\rightarrow 2)$ -α-D-mannopyranosyl- $(1\rightarrow 6)$ -α-Dmannopyranosyl- $(1\rightarrow 6)$ -D-mannopyranose (1). A mixture of compound 20 (0.1 g, 0.048 mmol) and PdCl<sub>2</sub> (0.02 mg) in dry methanol (10 mL) was stirred vigorously for 4 h at room temperature, TLC (1:1 petroleum ether-EtOAc) indicated that the reaction was complete, then filtered through Celite. The filtrate was concentrated to dryness. The resulting compound was dissolved in methanol (20 mL) satd with dry NH<sub>3</sub> and stirred at room temperature for 72 h, at the end of which time TLC (1:1.5 Methanol-EtOAc) indicated that the reaction was complete. The solution was concentrated to dryness and was chromatographed (methanol) on a column of Sephadex LH-20 to afford 1 (0.023 g, 70.2%) as an amorphous mass; [α]<sub>D</sub> + 3.1° (*c* 0.1, MeOH); <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O): δ 5.09, 5.01, 4.91, 4.89 (4 d, 4 H, J<sub>1,2</sub> = 1.5 Hz, H-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>); <sup>13</sup>C NMR (100 MHz, D<sub>2</sub>O), δ 100.3, 100.1, 99.8, 98.6 (4 C, C-1<sub>A</sub>, 1<sub>B</sub>, 1<sub>C</sub>, 1<sub>D</sub>). ESMS Calcd for C<sub>24</sub>H<sub>42</sub>O<sub>21</sub>: 666.58 (M). Found: 665.4 (M-H).

## ACKNOWLEDGMENTS

This work was supported by the Chinese Academy of Sciences (KJ952J<sub>1</sub>510 and KIP-RCEES9904) and the National Natural Science Foundation of China (59973026 and 29905004). Mr. Linsen Heng is a visiting scholar from Da Xian Normal College, Si Chuan, P. R. China.

### REFERENCES

- Gorin, P.A.J.; Spencer, J.F.T. Structural Chemistry of Fungal Polysaccharides. Adv. Appl. Microbiol. 1970, 13, 25–42.
- Bovina, E.V.; Deryabin, V.V.; Gagloev, V.N.; Serebryakov, N.G. Carbon-13 NMR Study of Mannans Produced by *Candida maltosa* and *Candida tropicalis*. Prikl. Biokhim. Mikrobiol. **1988**, 24 (2), 218–225.
- Suzuki, M.; Fukazawa, Y. Immunochemical Characterization of *Candida albicans* Cell Wall Antigens: Specific Determinant of *Candida albicans* Serotype A Mannan. Microbiol. Immunol. **1982**, *26* (5), 387–402.

Marcel Dekker, Inc.

270 Madison Avenue, New York, New York 10016

ORDER		REPRINTS
-------	--	----------

- Kogan, G.; Pavliak, V.; Masler, L. Structural Studies of Mannans from the Cell Wall of the Pathogenic Yeasts *Candida albicans* Serotypes A and B and *Candida parapsilosis*. Carbohydr. Res. **1988**, *172* (2), 243–253.
- 5. Kogan, G.; Pavliak, V.; Sandula, J.; Masler, L. Novel Structure of the Cellular Mannan of the Pathogenic Yeast *Candida Krasei*. Carbohydr. Res. **1988**, *184*, 171–182.
- Jiménez-Barbero, J.; Prieto, A.; Gómez-Miranda, B.; Leal, J.A.; Bernabé, M. Chemical Structure of Fungal Cell-Wall Polysaccharides Isolated from *Microsporum gypseum* and Related Species of *Microsporum* and *Trychophyton*. Carbohydr. Res. 1995, 272 (1), 121–128.
- Betaneli, V.I.; Ovchinnikov, M.V.; Backinowsky, L.V.; Kochetkov, N.K. A Convenient Synthesis of 1,2-O-Benzylidene and 1,2-O-Ethylidene Derivatives of Carbohydrates. Carbohydr. Res. 1982, 107 (2), 285–291.
- 8. Helferich, B.; Schmitz-Hillebrecht, E. A New Method for the Synthesis of Glucosides of the Phenols. Ber. **1933**, *66B*, 378–383.
- 9. Schmidt, R.R.; Toepfer, A. Glycosylation with Highly Reactive Glycosyl Donors: Efficiency of the Inverse Procedure. Tetrahedron Lett. **1991**, *32* (28), 3353–3356.
- a) Boss, R.; Scheffold, R. Cleavage of Allyl Ether with Palladium/Carbon. Angew. Chem. **1976**, 88 (17), 578–579; b) Ogawa, T.; Nakabayashi, S. Synthetic Studies on Cell-Surface Glycans. Part 8. Synthesis of a Hexasaccharide Unit of a Complex Type of Glycan Chain of a Glycoprotein. Carbohydr. Res.**1981**, *93* (1), C1–C5.
- 11. Nikolaev, A.V.; Ivanova, I.A. Shibaev, V.N.; Kochetkov, N.K. Application of the Hydrogenphosphonate Approach in the Synthesis of Glycosyl Phosphosugar Linked through Secondary Hydroxyl Groups. Carbohydr. Res. **1990**, *204*, 65–78.

Received September 20, 2000 Accepted March 22, 2001

Copyright © Marcel Dekker, Inc. All rights reserved



296

## **Request Permission or Order Reprints Instantly!**

Interested in copying and sharing this article? In most cases, U.S. Copyright Law requires that you get permission from the article's rightsholder before using copyrighted content.

All information and materials found in this article, including but not limited to text, trademarks, patents, logos, graphics and images (the "Materials"), are the copyrighted works and other forms of intellectual property of Marcel Dekker, Inc., or its licensors. All rights not expressly granted are reserved.

Get permission to lawfully reproduce and distribute the Materials or order reprints quickly and painlessly. Simply click on the "Request Permission/Reprints Here" link below and follow the instructions. Visit the <u>U.S. Copyright Office</u> for information on Fair Use limitations of U.S. copyright law. Please refer to The Association of American Publishers' (AAP) website for guidelines on <u>Fair Use in the Classroom</u>.

The Materials are for your personal use only and cannot be reformatted, reposted, resold or distributed by electronic means or otherwise without permission from Marcel Dekker, Inc. Marcel Dekker, Inc. grants you the limited right to display the Materials only on your personal computer or personal wireless device, and to copy and download single copies of such Materials provided that any copyright, trademark or other notice appearing on such Materials is also retained by, displayed, copied or downloaded as part of the Materials and is not removed or obscured, and provided you do not edit, modify, alter or enhance the Materials. Please refer to our <u>Website</u> User Agreement for more details.

# **Order now!**

Reprints of this article can also be ordered at http://www.dekker.com/servlet/product/DOI/101081CAR100104864