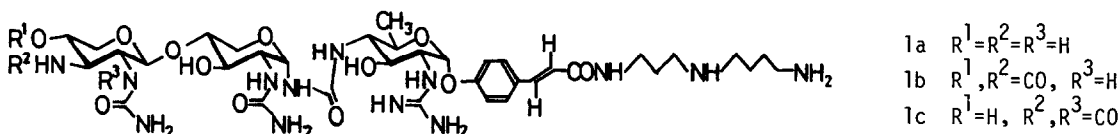


SYNTHETIC KEY INTERMEDIATES OF GLYCOCINNAMOYLSPERMIDINES ;
 DIAMINOHEXOSYLOXYCINNAMATE AND AMINOPENTOSE DISACCHARIDE MOIETIES

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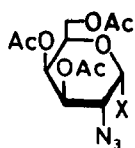
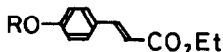
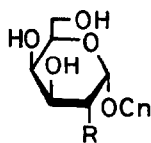
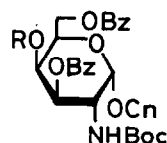
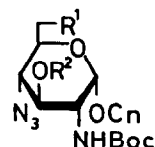
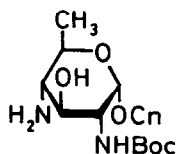
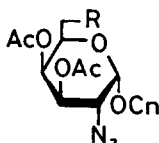
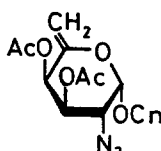
Summary: Two key intermediates for the total synthesis of glycocinnamoylspermidines were derived from the corresponding 2-azido sugars.

Glycocinnamoylspermidines, LL-BM 123β (1a), γ₁ (1b) and γ₂ (1c), are new broad-spectrum antibiotics whose structures were recently proposed.¹ These antibiotics are of special interest because of not only their biological activity against gram-negative organisms and their protective effects against infections produced in mice,² but also their unique structure including α-ureido linkage between amino sugar components. Here we would like to report the synthesis of two key intermediates for the components, i.e., ethyl p-[4-amino-2-t-(butoxycarbonyl)amino-2,4,6-trideoxy-α-D-glucopyranosyl]oxycinnamate (15) and methyl 4-O-(2-azido-2-deoxy-β-D-xylopyranosyl)-2-azido-2-deoxy-α-D-xylopyranoside derivative (31). Because of chemical stability and easy conversion into



various modified amino functions, 2-azido sugars were chosen as starting compounds.

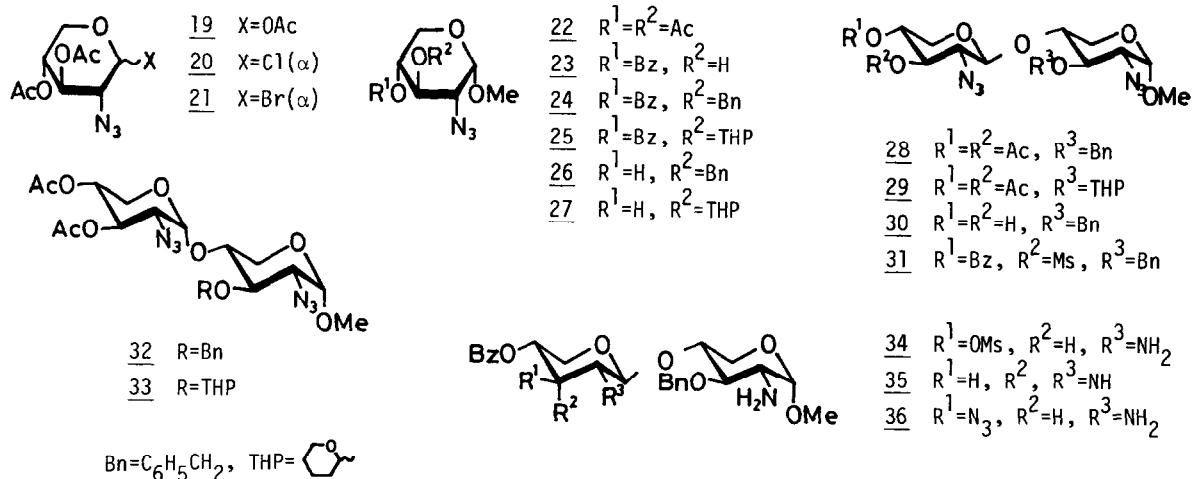
In order to convert 2-azido-2-deoxy-D-galactose into 15 the required synthetic operations are (A) glycosylation, (B) 4-amination and (C) 6-deoxygenation. Considering availability of starting 2-azido sugars and necessity of 2-azido group for the glycosylation, the sequence, A → B → C, seemed to be favorable. On glycosylation of ethyl p-hydroxycinnamate (4) with 3,4,6-tri-O-acetyl-2-azido-2-deoxy-α-D-galactopyranosyl chloride³ (2) the best result as to the ratio of α-anomer formed [α-(3*) : β-anomer = 5.5 : 1.0, yield 42%] was obtained in the presence of AgClO₄ (1.2 equiv.) and Ag₂CO₃ (2 equiv.) in dichloromethane at room temperature for 30 h. The yield was improved to 63% (α/β=5.5) by condensation of 2 with trimethylsilyl ether (5) of 4 under the same condition. This combination seems to be a new type of glycosidation of phenol derivatives⁴ and proved to be very effective. The azido group of its de-O-acetylated product (6) was selectively hydrogenolyzed with 5% Pd/BaSO₄ (about one forth of substrate in weight)-quinoline (one drop/50 mg catalyst) in methanol for 15-24 h to give the 2-amino derivative (7), which was converted into N-(t-butoxycarbonyl) derivative (8) with 2-(t-butoxycarbonyl)thio-4,6-dimethylpyrimidine and Et₃N in dioxane-water. Benzoylation of 8 in acetone with BzCl (3 equiv.) in the presence of Et₃N at -10°C for 16 h gave

2 X=Cl3 X=OCn4 R=H(CnOH)5 R=TMS6 R=N₃7 R=NH₂8 R=NHBoc9 R=H10 R=Ms11 R¹=OBz, R²=Bz12 R¹=OH, R²=H13 R¹=OTs, R²=H14 R¹=R²=H1516 R=OTs17 R=H18Ac=CH₃CO, Bz=C₆H₅COBoc=(CH₃)₃COCO, Ms =CH₃SO₂Ts=p-CH₃C₆H₄SO₂, TMS=(CH₃)₃Si

3,6-dibenzoate (9) in 75% yield from 6. Its 4-mesylate (10) was treated with NaN₃ (2 equiv.) in hexamethylphosphoric triamide (HMPT) at 80°C for 10 h to afford the corresponding 4-azide (11*) in 77% yield. Tosylation of its debenzoylated derivative (12) with TsCl (2 equiv.) in pyridine at room temperature for 20 h to give 6-tosylate (13) in 95% yield. Highly selective reduction of 6-tosyloxy group of 13, without reduction of CC double bond, was successfully carried out by NaBH₃CN (7.5 mol) in the presence of NaI⁵ (4 equiv.) in HMPT at 70-75°C for 15 h to yield the corresponding 6-deoxy derivative (14) in 79% yield. Finally, similar selective hydrogenolysis used for 6 gave ethyl p-[4-amino-2-(t-butoxycarbonyl)amino-2,4,6-trideoxy-α-D-glucopyranosyl]oxycinnamate (15*) in 75% yield.

In the course of above-mentioned transformation attempted 6-deoxygenation in advance of 4-amination, i.e., the operation sequence A → C → B, failed presumably due to the effect of axially oriented acyloxy group on C-4.⁶ For example, reduction of ethyl [3,4-di-O-acetyl-2-azido-2-deoxy-6-O-(p-tolylsulfonyl)-α-D-galactopyranosyl]oxycinnamate (16), which was derived from 6 by partial tosylation and successive acetylation in 64% yield, with NaBH₃CN in the presence of NaI in HMPT at 70°C for 70 h gave the desired 6-deoxygenated product (17) only in 20% yield together with 5-enopyranoside (18) in 10% yield.

On the other hand, the disaccharides (29 and 30) were chosen as feasible candidates of synthetic intermediate for aminopentose disaccharide moiety. Azidonitration³ of D-xylal diacetate followed by acetolysis gave peracetylated 2-azido-2-deoxy-D-xylopyranose (19) in 58% yield regio- and stereoselectively. Treatment of 19 with TiCl₄ or TiBr₄ in dichloromethane at room temperature gave the glycosyl chloride (20*) or bromide (21) in 77 and 63% yields, respectively. Furthermore, direct addition of methanol to the reaction mixture of azidonitration gave methyl 3,4-di-O-acetyl-2-azido-2-deoxy-α-D-xylopyranoside (22) in 84% yield. Deacetylation of 22 followed by benzylation with BzCl and Et₃N (each 2 equiv.) in acetone gave 4-benzoate (23) in 75% yield together with 3,4-dibenzoate (20%). Then, treatment of 23 with BnBr and Ag₂O in benzene and with 2,3-dihydro-4H-pyran and pyridinium p-toluenesulfonate in dichloromethane gave, respectively, 3-O-benzyl derivative (24) in 74% yield and 3-O-tetrahydropyranyl derivative (25) quantitatively. Treatment of 24 and 25 with sodium methoxide in methanol yielded de-O-benzoylated derivatives (26 and 27, respectively).



Several glycosylation methods⁸⁻¹¹ reported to favor 1,2-trans glycoside (β -glycoside in this case) formation were tried using 20 or 21 as donors and 26 and 27 as acceptors. The best result was obtained by coupling between 20 and 26 with a ratio of 1:3 in the presence of AgClO₄ and 2,4,6-trimethylpyridine (each 1.2 equiv.) in nitromethane¹¹ at -10°C for 60 h. The β -(28*) and α -disaccharide (32) was obtained in 73% yield with a ratio of 1.2:1.0. However, condensation of 20 with 27 under the same conditions afforded the β -(29) and α -disaccharide (33) in 47% yield with a ratio of 1.0:7.5 presumably due to decrease of reactivity of the acceptor because of the bulky tetrahydropyranyl group. On the other hand, trimethylsilyl trifluoromethanesulfonate (TMSOTf) was recently introduced as a new effective Lewis acid for glycosylation.¹² This reagent seems to be also promising in our case, giving almost equal stereoselectivity and yield with those obtained by AgClO₄. Even equimolar condensation of 19 and 26 in the presence of TMSOTf (1.2 equiv.) and molecular sieve 4A in dichloromethane at 0°C for 24 h gave 28 and 32 in 36% and 16% yields, respectively. The above mentioned results concerning both yield and stereoselectivity are not completely sufficient, but may be accepted as reasonable considering that peracetylated 2-azido sugars has in general very low reactivity as shown in the case of the corresponding aldohexose analogs.⁸

Moreover, on the synthesis of glycocinnamoylspermidines via the trisaccharides obtained by the coupling of diaminopentose moiety such as 28 with 15 the conversion of the non-reducing end into the corresponding 2,3-diamino-D-xylose derivative is necessary. Therefore, this conversion was examined with 28. Its 4-O-benzoyl-3-O-mesyl derivative (31*) obtained by partial benzylation followed by mesylation was hydrogenolyzed in the presence of PtO₂ to give 2,2'-diamino derivative (34). Treatment of 34 with NaOAc in N,N-dimethylformamide at 80°C for 3 h afforded quantitatively the epimino derivative (35), which could be converted into the desired precursor (36*) of 2',3'-diamino derivative by azidolysis with NaN₃ in 2-methoxyethanol at 110°C for 3 h in 63% yield with high regioselectivity.

The structures of all new compounds were confirmed by their spectroscopic data and elemental analyses. Physical properties and spectral data of some key compounds (the number with asterisk) are given in the end of this paper.¹³

References and Notes

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- 13) **5**: mp 95-97°C, $[\alpha]_D +164.9^\circ$; IR ν_{\max}^{KBr} 2125, 1755, 1740, 1715, and 1630 cm^{-1} ; NMR δ 5.65 (d, $J_{1,2}$ 3.8 Hz, H-1), 1.94, 2.10, and 2.17 (each s, 9 H, Ac), 1.33 (t) and 4.24 (q) (J 7.2 Hz, Et), 6.30 and 7.61 (each d, J 16.2 Hz, olefinic), and 7.07 and 7.45 (each d, 4 H, J 9.0 Hz, aromatic).
11: $[\alpha]_D +160.9^\circ$; IR ν_{\max}^{NaCl} 2130 cm^{-1} ; NMR δ 3.92 (t, $J_{3,4}=J_{4,5}$ 10.5 Hz, H-4).
15: mp 167-170°C, $[\alpha]_D +191.0^\circ$; IR ν_{\max}^{KBr} 3450, 1710, 1680, 1630, 1600, 1520, and 1505 cm^{-1} ; NMR δ 5.56 (d, $J_{1,2}$ 3.0 Hz, H-1), 2.63 (t, $J_{3,4}=J_{4,5}$ 9.3 Hz, H-4), 1.23 (d, 3 H, $J_{5,6}$ 7.0 Hz, H-6), 5.20 (d, $J_{\text{NH},2}$ 8.3 Hz), 1.44 (s, 9 H, Boc) and signals for the cinnamate portion like **3**.
20: mp 71-72°C, $[\alpha]_D +149.4^\circ$; NMR δ 6.10 (d, $J_{1,2}$ 3.3 Hz, H-1), and 3.78 (dd, $J_{2,3}$ 9.9 Hz, H-2).
28: $[\alpha]_D +10.2^\circ$; IR ν_{\max}^{NaCl} 2100, 1745, and 1605 cm^{-1} ; NMR δ 4.67 (d, $J_{1,2}$ 3.0 Hz, H-1), 4.35 (d, $J_{1',2'}$ 7.8 Hz, H-1'), 3.44 (s, 3 H, OMe), 2.04 and 2.12 (each s, 6 H, Ac).
31: mp 128-130°C, $[\alpha]_D -32.3^\circ$; NMR δ 4.73 (dd, $J_{2',3'}$ 9.8 Hz, H-3') and 5.20 (dt, $J_{3',4'}=J_{4',5'a}$ 9.0 Hz, $J_{4',5'e}$ 5.1 Hz, H-4').
36: $[\alpha]_D -7.7^\circ$; IR ν_{\max}^{NaCl} 3370, 3300, 2100, and 1725 cm^{-1} ; NMR δ 4.56 (d, $J_{1,2}$ 1.5 Hz, H-1), 4.31 (d, $J_{1',2'}$ 7.5 Hz, H-1'), 2.75 (dd, $J_{2',3'}$ 10.2 Hz, H-2'), 3.39 (s, 3 H, OMe).
 Optical rotations and NMR spectra at 100 MHz were recorded in CHCl_3 and CDCl_3 , respectively.