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Synthesis and characterization of 8-ethynyl-1,3-dihydrobenzo[b][1,4]diazepin-2-one derivatives: Part 2. New potent non-competitive metabotropic glutamate receptor 2/3 antagonists

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Abstract—A series of 1,3-dihydro-benzo[*b*][1,4]diazepin-2-one derivatives was evaluated as non-competitive mGluR2/3 antagonists. Replacement of a cyano group by a five-membered heterocycle produced compounds inhibiting the binding of [3 H]-LY354740 to rat mGluR2 with low nanomolar affinity and consistent functional effect at both mGluR2 and mGluR3. Further modification to improve the physicochemical properties led eventually to compounds with the ability to reverse LY354740-mediated inhibition of field excitatory postsynaptic potentials in the rat dentate gyrus. © 2007 Elsevier Ltd. All rights reserved.

The glutamate receptors are divided into two major groups: ionotropic (iGluR: ligand-gated ion channels) and metabotropic (mGluR: G-protein coupled receptors) glutamate receptors.¹ From mGluRs eight subtypes were identified and grouped into three groups according to their sequence homology, agonist pharmacology, and second messenger coupling. Group I receptors (mGluR1 and 5) are positively coupled to phospholipase C, whereas group II (mGluR2 and 3) and group III (mGluR4, 6, 7, and 8) receptors are negatively coupled to the activity of adenyl cyclase. Targeting mGluRs has been recognized as a valuable approach for the treatment of CNS disorders, such as depression, anxiety, and schizophrenia.² It was shown that mGluR2/ 3 agonists exhibit antipsychotic properties and mGluR2/ 3 antagonists may be useful as antidepressants and cog-

Keywords: Metabotropic glutamate receptors; mGluR2; LY354740; Non-competitive antagonists; 1,3-Dihydro-benzo[*b*][1,4]diazepin-2ones; Excitatory postsynaptic potentials.

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nitive enhancers as demonstrated in different animal models.^{3,4}

Recently we reported about the development of the random screening hit 1—a compound from the chemical class of 1,3-dihydro-benzo[*b*][1,4]diazepin-2-ones—into compounds with low nanomolar affinity (Fig. 1) assayed by partial inhibition of [³H]-LY354740 binding to rat mGluR2⁵ (Fig. 2a).⁶ The in vitro binding characteristics of this selective mGluR2/3 radioligand with agonist properties were described by Schweitzer et al.^{5a} The specific binding of [³H]-LY354740 was dependent on the addition of divalent cations, it showed the presence of a single binding site, and was partially inhibited by GTP γ S. Saturation isothermal curves on [³H]-LY354740 binding to rat mGluR2 and functional studies carried out with 1 and derivatives on (1*S*,3*R*)-ACPD



Figure 1. Development of the random screening hit 1 into compounds, like, for example, 2, with low nanomolar affinity in inhibition of $[^{3}H]$ -LY354740 binding to rat mGluR2.⁶

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Figure 2. (a) 2 (\blacksquare), 150 (\bullet), and 15q (\blacktriangle) inhibition of specific [³H]-LY354740 binding to the recombinant rat mGluR2 permanently expressed in CHO cells. Inhibition curves are normalized as % of the maximum specific bound. All compounds partially inhibit the radioligand binding to mGluR2 with a residual of about 25% of specific bound ([³H]-LY354740, 10 nM). The binding of this radioligand to mGluR2 is GTP_YS sensitive and it is blocked by pertussin toxin. (b) (1*S*,3*R*)-ACPD-induced inhibition of cAMP production: the non-competitive mGluR2 antagonist 15b reduces maximal response and causes rightward shift of dose–response curve of (1*S*,3*R*)-ACPD [+ (1*S*,3*R*)-ACPD (*n* = 5); (\bullet) 15b 1 nM (*n* = 2); (\blacktriangle) 15b 3 nM (*n* = 3)] (also cf. Ref. 7).

inhibition of forskolin stimulated cAMP production (Fig. 2b), on the ³⁵S-GTP γ S binding induced by (1*S*,3*R*)-ACPD, and on the glutamate induced GIRK currents, respectively, showed the presence of a non-competitive antagonist activity. The conclusion that these mGluR2 antagonists do not interact with the orthosteric site of the receptor is also in agreement with the observation that they do not displace [³H]-LY341495, a competitive antagonist to mGluR2.⁷ The capacity of 1 and derivatives to partially displace [³H]-LY354740 is most likely due to an effect on coupling efficiency of mGluR2. This property, in the described experimental conditions, was used in association to functional studies to optimize the structure–activity relationship for this class of compounds.

The extension of the methyl group in 8-position to a phenylacetylene residue as well as the attachment of a cyano group in meta-position on the phenyl in 4-position resulted in the very potent compound **2**. Nevertheless these modifications led to very lipophilic molecules (**2**: clog P 4.44; calculated $log(c_{octanol}/c_{water})$) and therefore we sought for further alteration of the substituents enabling the evaluation of compounds from this class in different models. Our main focus was the search for a suitable nitrile replacement as well as the exploration of the 7-position.

The regioselective synthesis of 1,3-dihydro-benzo[b]-[1,4]diazepin-2-ones has already been described⁶ and is

here exemplified by the preparation of compound 150 (Scheme 1). The o-phenylenediamine part utilized 5chloro-4-iodo-2-nitroaniline 3^8 and the β -ketoester part used 3-imidazol-1-yl-benzoic acid 10⁹ as starting materials. Nucleophilic displacement of the Cl in 3 with allyl alcohol gave 4 wherein-after introduction of the Bocgroup—the allyl group was switched to *tert*-butyl by Pd-catalyzed deprotection with morpholine and introduction of the *t*-Bu group by the method of Widmer¹⁰ to give intermediate 7. Sonogashira-type coupling with 4-fluorophenylacetylene was followed by reduction of the nitro group and yielded the mono-Boc protected ophenylenediamine building block 9. For the β -ketoester part the known 3-imidazol-1-yl-benzoic acid 10⁹ was converted via its methyl ester 11 into the *t*-Bu β -ketoester 12 following a protocol of Ohta.¹¹ To increase the reactivity of the *t*-Bu β -ketoester **12** for the condensation with the *o*-phenylenediamine building block 9, it was transformed into the 2,2-dimethyl-[1,3]dioxin-4-one 13 by treatment with TFAA/TFA in acetone as described by Winkler.¹² Both pieces 9 and 13 were then condensed in refluxing toluene to the corresponding β -ketoamide 140, which was doubly deprotected and concomitantly cyclized by treatment with TFA yielding the unsymmetrically 7,8-substituted 1,3-dihydro-benzo[b][1,4]diazepin-2-one 150.

We found that five-membered heterocycles were suitable replacements for the cyano group, especially the employment of imidazoles (15a-c) and 1,2,3-triazoles



Scheme 1. Synthesis of compound 150. Reagents and conditions: (a) allyl alcohol, KOH, DMSO, rt, 95%; (b) Boc₂O, DMAP, THF, rt, 99%; (c) TFA, DCM, 0 °C, 75%; (d) morpholine, Pd(PPh₃)₄, THF, rt, 98%; (e) Me₂NCH(OBu^t)₂, toluene, 80 °C, 73%; (f) 4-F–C₆H₄C==CH, PdCl₂(PPh₃)₂, PPh₃, CuI, Et₃N, THF, 60 °C, 38%; (g) SnCl₂·2H₂O, pyridine, DMF, rt, 68%; (h) concd H₂SO₄, MeOH, reflux, 98%; (i) *t*-BuOAc, LDA, LiOBu^t, THF, -78 °C, 95%; (j) TFAA, TFA, acetone, -78 to 23 °C, 65%; (k) toluene, reflux, 68%; (l) TFA, DCM, rt, 65%.

Table 1. Activities of the 8-ethynyl-1,3-dihydro-benzo[b][1,4]diazepin-2-ones 15

Compound	R ^{3′}	R ⁸	R^7	Rat mGluR2 [3 H]-LY354740 binding ^a IC ₅₀ (μ M)	Rat mGluR2 (1 <i>S</i> ,3 <i>R</i>)-ACPD inhibition of forskolin stimulated $cAMP^{b}$ IC ₅₀ (μ M)
2	CN	Ph-C=C-	Н	0.034	0.017
15a		Ph-C=C-	Н	0.013	0.010
15b		4-F-C ₆ H ₄ -C=C-	Н	0.009	0.009
15c		2-F-C ₆ H ₄ -C=C-	Н	0.020	0.016
15d	N N Me	4-F-C ₆ H ₄ -C=C-	Н	0.098	nt
15e	Me N Me	Ph-C=C-	Н	0.170	nt
15f	Me	Ph-C=C-	Н	0.730	nt
15g		4-F-C ₆ H ₄ -C=C-	Н	0.098	nt
15h	N ^{×N} - N	4-F-C ₆ H ₄ -C=C-	Н	0.006	0.012
15i	N ^{=N} , N / N /	4-F-C ₆ H ₄ -C=C-	Н	0.072	nt
15j	N	Ph-C=C-	OCH ₂ CH ₂ OMe	0.023	0.015
15k		Ph-C=C-	OCH ₂ CH ₂ OH	0.064	nt
151		4-F-C ₆ H ₄ -C=C-	OCH ₂ CH ₂ OH	0.046	nt
15m		Ph-C=C-	OCH ₂ CN	0.018	0.013
15n		4-F-C ₆ H ₄ -C=C-	N(Me)CH ₂ CH ₂ OH	0.140	nt
150		4-F-C ₆ H ₄ -C=C-	ОН	0.026	0.011
15p	N ^{SN} N	Ph-C=C-	NMe ₂	0.036	nt
15q	N ^{×N} N	4-F-C ₆ H ₄ -C=C-	ОН	0.020	0.017
15r	N ^{×N} N OH	4-F-C ₆ H ₄ -C=C-	ОН	0.378	nt

 $R^{3'}$, R^7 , and R^8 refer to the positions of Rs in Figure 1. nt, not tested.

^a Values are means of at least two independent experiments. ^b Values are means of three independent experiments.

(15h,p,q) led to very potent mGluR2/3 antagonists (Table 1). Interestingly the *N*-linked 1-imidazoles (e.g., 15a or 15e) were far more active than its *C*-linked 2-isomer (15f). With regard to the methyl substitution on the 1-imidazoles it appeared only to some extent to be tolerated resulting a ca. 10-fold drop in affinity (15a vs 15e; 15b vs 15d). The order of activity depending on the heterocycles incorporated was determined as follows: 1-imidazole ~ 1,2,3-triazole > 1-tetrazole ~ 1,2,4-triazole. Overall, the unsubstituted 1-imidazole and 1,2,3-triazole proved to be most favorable (e.g., 15a, 15b, 15c, and 15h all exhibit IC₅₀ \leq 20 nM).

The structural class of the 1,3-dihydro-benzo[b][1,4]diazepin-2-ones offered in addition to the (2-aryl)-ethynylmoiety in 8-position and the meta-attached heterocycle on the phenyl group in 4-position a third exit vectorthe 7-position—where the introduction of further substituents was allowed as previously described.⁶ The most successful ones in altering physicochemical properties to reduce the overall lipophilicity without loss of mGluR2 affinity were small oxygen- or nitrogen-linked C2 units bearing polar head groups like OH (15k,l,n), OMe (15i) or CN (15m). It was found that a 2-hydroxyethyl residue is more active when O-linked (151) rather than *N*-linked (15n) to the core structure, but in general even the relatively small 2-hydroxyethyl group with the free OH (15k) resulted in loss of activity (ca. 3-fold) in comparison with the less polar OMe (15j) and CN (15m). Replacing the 2-hydroxyethyl group in 15n by a simple methyl group produced the rather non-polar 15p (bearing the NMe₂ group) but was accompanied with a regain in affinity. In analogy cutting off the 2-hydroxyethyl group in 15l yielded the free phenol 150-a very potent imidazole compound. The corresponding 1,2,3-triazole compound 15q proved to be equipotent.

Further increase in polarity by introduction of hydroxymethyl group in the 5-position on the 1,2,3-triazole in 15q (clog *P* 3.80) resulted in 15r (clog *P* 2.88) and ca. 20-fold loss of affinity.

In conclusion, only limited allowance for the introduction of more polar groups was found—the 1-imidazole or 1,2,3-triazole in combination with free phenol in 7position appeared to be the maximum in order to achieve highly potent compounds.

As described earlier **150** and **15q** partially inhibit [³H]-LY354740 binding to mGluR2 (Fig. 2a).¹³ To demonstrate non-competitive nature of the antagonism at mGluR2 by the newly generated compounds, we show hereby the effect of increasing concentrations of **15b** on the (1*S*,3*R*)-ACPD-induced inhibition of forskolin stimulated cAMP production (Fig. 2b).^{7,13}

The antagonist properties of **150** and **15q** were also evaluated electrophysiologically in CHO cells stably expressing the human Kir3.1 and Kir3.2c GIRK subunits and transiently transfected with either rat mGluR2 or rat mGluR3 (Fig. 3).¹³ In these cells, glutamate concentration-dependently induced a reversible inward K⁺ current, which was inhibited by both **150** and **15q**.



Figure 3. Concentration–response curves for the inhibition of GIRK currents by 150 (Δ) and 15q (\blacktriangle) in CHO cells stably expressing GIRKs and transiently transfected with: (a) rat mGluR2 (glutamate 10 μ M) and (b) rat mGluR3 (glutamate 1 μ M).



Figure 4. Antagonist activity of compound **150** on the LY354740mediated inhibition of synaptic transmission in the rat hippocampus: (a) Compound **150** reverses the LY354740-mediated inhibition of field excitatory postsynaptic potentials (fEPSPs) evoked by stimulation of the medial perforant path and recorded in the dentate gyrus (compound **150** (1 μ M), ---- LY354740 (1 μ M); mean ± SE, *n* = 4). The inset shows representative fEPSPs under control conditions, in the presence of 1 μ M LY354740, and in the presence of 1 μ M LY354740 and 1 μ M **150** at 180 min. (b) The concentration-dependent inhibition of fEPSPs by LY354740 (IC₅₀ = 0.23 μ M, slope = 1.2) was completely blocked following 3-h pre-incubation of hippocampal slices with **150** [• control (*n* = 7); o +30 nM **150** (*n* = 4)].

We assessed the selectivity of **150** and **15q** and these compounds neither activated nor inhibited glutamatestimulated rat mGluR1a and mGluR5a (using a Ca²⁺ mobilization functional assay, when tested at 30 μ M final concentration). Compound **150** was also inactive in the binding of [³H]-L-AP4 to rat mGluR8a, while **15q** showed an IC₅₀ of 5 μ M. In addition both compounds were devoid of any affinity at NMDA and GABA_A receptors (data not shown).

Using compound **150** we could also demonstrate antagonism at native group II metabotropic glutamate receptors by reversal of LY354740-mediated inhibition of fEPSPs in the dentate gyrus (Fig. 4).¹⁴

In summary, by replacement of a nitrile with a five-membered heterocycle we were able to develop a series of 1,3dihydro-benzo[b][1,4]diazepin-2-one derivatives which are selective and potent non-competitive antagonists at recombinant group II metabotropic glutamate receptors. Further optimization of physicochemical properties is required before approaching in vivo studies.

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References and notes

- Reviews: (a) Gasparini, F.; Spooren, W. Curr. Neuropharmacol. 2007, 5, 187; (b) Monn, J. A.; Johnson, M. P.; Schoepp, D. D. Handb. Contemp. Neuropharmacol. 2007, 1, 421; (c) Neugebauer, V. Handb. Exp. Pharmacol. 2007, 177, 217; (d) Oswald, R. E.; Ahmed, A.; Fenwick, M. K.; Loh, A. P. Curr. Drug Targets 2007, 8, 573; (e) Ossowska, K. Dopamine Glutamate Psychiatr. Disord. 2005, 117; (f) Ferraguti, F.; Shigemoto, R. Cell Tissue Res. 2006, 326, 483; (g) Schoepp, D. D.; Jane, D. E.; Monn, J. A. Neuropharmacology 1999, 38, 1431.
- (a) Récasens, M.; Guiramand, J.; Aimar, R.; Abdulkarim, A.; Barbanel, G. Curr. Drug Targets 2007, 8, 651; (b)

Swanson, C. J.; Bures, M.; Johnson, M. P.; Linden, A.-M.; Monn, J. A.; Schoepp, D. D. Nat. Rev. Drug Discov.
2005, 4, 131; (c) Higgins, G. A.; Miczek, K. A. Psychopharmacology 2005, 179, 1; (d) Mutel, V. Expert Opin. Ther. Patents 2002, 12, 1845; (e) Schoepp, D. D. J. Pharmacol. Exp. Ther. 2001, 299, 12.

- Spinelli, S.; Ballard, T.; Gatti-McArthur, S.; Richards, G. J.; Kapps, M.; Woltering, T.; Wichmann, J.; Stadler, H.; Feldon, J.; Pryce, C. R. *Psychopharmacology* 2005, *179*, 292.
- Higgins, G. A.; Ballard, T. M.; Kew, J. N. C.; Richards, J. G.; Kemp, J. A.; Adam, G.; Woltering, T.; Nakanishi, S.; Mutel, V. *Neuropharmacology* 2004, 46, 907.
- (a) Schweizer, C.; Kratzeisen, C.; Adam, G.; Lundstrom, K.; Malherbe, P.; Ohresser, S.; Stadler, H.; Wichmann, J.; Woltering, T.; Mutel, V. *Neuropharmacology* 2000, *39*, 1700; (b) Schaffhauser, H.; Richards, J. G.; Cartmell, J.; Chaboz, S.; Kemp, J. A.; Klingelschmidt, A.; Messer, J.; Stadler, H.; Woltering, T.; Mutel, V. *Mol. Pharmacol.* 1998, *53*, 228.
- Woltering, T. J.; Adam, G.; Alanine, A.; Wichmann, J.; Knoflach, F.; Mutel, V.; Gatti, S. *Bioorg. Med. Chem. Lett.* 2007, 17, 6811.
- Hemstapat, K.; Da Costa, H.; Nong, Y.; Brady, A. E.; Luo, Q.; Niswender, C. M.; Tamagnan, G. D.; Conn, P. J. *J. Pharmacol. Exp. Ther.* 2007, *322*, 254.
- (a) Adam, G.; Alanine, A.; Goetschi, E.; Mutel, V.; Woltering, T. J. World Patent WO01/029011, 2001; *Chem. Abstr.* 2001, 134, 311234; (b) Adam, G.; Goetschi, E.; Mutel, V.; Wichmann, J.; Woltering, T. J. World Patent WO02/083652, 2002; *Chem. Abstr.* 2002, 137, 325447.
- Hagedorn, A. A., III; Erhardt, P. W.; Lumma, W. C., Jr.; Wohl, R. A.; Cantor, E.; Chou, Y. L.; Ingebretsen, W. R.; Lampe, J. W.; Pang, D.; Pease, C. A.; Wiggins, J. J. Med. Chem. 1987, 30, 1342.
- 10. Widmer, U. Synthesis 1983, 135.
- 11. Ohta, S.; Shimabayashi, A.; Hayakawa, S.; Sumino, M.; Okamoto, M. Synthesis 1985, 45.
- 12. Winkler, J. D.; Hershberger, P. M.; Springer, J. P. *Tetrahedron Lett.* **1986**, *27*, 5177.
- Gatti, S.; Knoflach, F.; Kew. J.; Adam, G.; Goetschi, E.; Wichmann, J.; Woltering, T.; Kemp, J.; Mutel, V. *Abstract of Papers*, 31st Annual Meeting of the Society for Neuroscience, San Diego, CA; Society for Neuroscience: Washington, DC, 2001; Abstract 705.10.
- Kew, J. N. C.; Ducarre, J.-M.; Pflimlin, M.-C.; Mutel, V.; Kemp, J. A. *Neuropharmacology* 2001, 40, 20.