

# Photochemistry of Tiaprofenic Acid, a Nonsteroidal Anti-Inflammatory Drug with Phototoxic Side Effects

FRANCISCO BOSCA, MIGUEL A. MIRANDA<sup>x</sup>, AND FRANKLIN VARGAS

Received August 6, 1990, from the *Departamento de Química and Instituto de Tecnología Química UPV-CSIC, Universidad Politécnica de Valencia, Camino de Vera s/n, Apartado 22012, 46071 Valencia, Spain.* Accepted for publication March 26, 1991.

**Abstract** □ The phototoxic nonsteroidal anti-inflammatory drug tiaprofenic acid (1) is photolabile under aerobic conditions. Irradiation of a methanol solution of 1 under oxygen produces the photoproducts 2, 3, 4, and 5, and also produces a singlet oxygen as evidenced by trapping with 2,5-dimethylfuran.

Nonsteroidal anti-inflammatory drugs (NSAIDs) have gained prominence in the last 10 years because of their efficiency in the treatment of inflammation diseases and the lack of the long-term side effects associated with corticosteroids. Tiaprofenic acid (1) is a NSAID with a balanced benefit:risk ratio, but whose use has been associated in some patients with the appearance of phototoxic effects such as erythema, flaring, and urticarial weal.<sup>1-3</sup> In a photopatch test for skin photosensitization to UV-A plus UV-B light, a significant portion of the patients gave a positive reaction to 1.<sup>4,5</sup> In vitro experiments also point to the potential phototoxicity of this drug.<sup>6</sup>

It appears that there may be a relationship between photochemical behavior and phototoxicity. In this context, nothing is as yet known about the photochemistry of 1. This has prompted us to examine the photolysis of 1 under a variety of conditions with two main goals: to establish the structures of the different photoproducts and to evidence the role of oxygen in these photoprocesses.

## Experimental Section

Tiaprofenic acid (1) was irradiated for 24 h in methanol (1.00 g in 200 mL) at 20 °C by means of a pyrex immersion-well photoreactor (Applied Photophysics, parts no. 3230 + 3307), with a OSRAM HQL 400 W medium-pressure Hg lamp, under oxygen or argon atmosphere. The course of the reaction was followed by UV-Vis spectrophotometry using a Perkin-Elmer Lambda 15 instrument, as well as by gas chromatography (GC) using a Hewlett-Packard 5890 instrument, with a 25 m × 0.32 mm × 0.52 μm capillary column of cross-linked 5% phenylmethylsilicone. The solvent was then evaporated at reduced pressure, and the residue was analyzed by column chromatography (silica gel). Elution was carried out with a mixture of hexane:ethyl acetate (5:1, v/v).

The products 2-5 (see structure) were isolated and analyzed by <sup>1</sup>H NMR spectrometry (Varian 360 EM spectrometer), IR spectrophotometry (Perkin-Elmer 781 spectrophotometer), and mass spectrometry (Hewlett-Packard 5988 A spectrometer). Some chemical correlations were established which allowed structural assignment. Thus, benzylic bromination of 5 with *N*-bromosuccinimide in boiling carbon tetrachloride, followed by treatment of the resulting bromide with

silver nitrate in aqueous acetone, gave the alcohol 4. Alternatively, heating of the same intermediate bromide in methanol produced the methyl ether 3.

Compound 2 had a mp of 115–116 °C; IR (KBr):  $\nu$  = 3100, 2900, 2710, 2550, 1690, 1640, 1500, 1290, 1260, 870, and 740 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 2.50 (s, 3 H, CH<sub>3</sub>) and 7.30–8.00 (m, 7 H, aromatic); MS (70 ev):  $m/z$  (%) = 230 (67, M<sup>+</sup>), 215 (100, M<sup>+</sup>–CH<sub>3</sub>), 187 (8), 153 (25), 105 (78), 77 (41), 51 (6).

Compound 3 showed a mp of 104–106 °C; IR (KBr):  $\nu$  = 3200, 3000, 2990, 2820, 1640, 1520, 1450, 1300, 1250, 1110, 880, 700, and 620 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 1.60 (d, 3 H, CH<sub>3</sub>), 3.30 (s, 3 H, –OCH<sub>3</sub>), 4.60 (q, 1 H, –CH), 7.00 (d, 1 H, thiophene-CH), and 7.20–7.80 (m, 6 H, aromatic); MS (70 ev):  $m/z$  (%) = 246 (23, M<sup>+</sup>), 231 (100, M<sup>+</sup>–CH<sub>3</sub>), 215 (27, M<sup>+</sup>–OCH<sub>3</sub>), 141 (35), 105 (43), 77 (28), 57 (10).

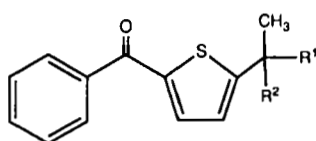
Compound 4: IR (KBr):  $\nu$  = 3600, 3500–3100, 2980, 1640, 1450, 1300, 1260, 1100, 1000, 870, and 600 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 1.65 (d, 3 H, CH<sub>3</sub>), 4.20 (s, 1 H, –OH), 5.20 (q, 1 H, –CH–), 6.80 (d, 1 H, thiophene-CH), and 7.10–7.60 (m, 6 H, aromatic); MS (70 ev):  $m/z$  (%) = 232 (16, M<sup>+</sup>), 217 (100, M<sup>+</sup>–CH<sub>3</sub>), 215 (65, M<sup>+</sup>–OH), 214 (30), 199 (22), 187 (13), 139 (52), 105 (25), 77 (30), 51 (5).

Compound 5: IR (KBr):  $\nu$  = 3100, 3000, 2990, 1640, 1450, 1300, 1250, 1100, 1000, 860, and 600 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  = 1.20 (t, 3 H, CH<sub>3</sub>), 2.70 (q, 2 H, CH<sub>2</sub>), 6.60 (d, 1 H, thiophene-CH), and 7.10–7.60 (m, 6 H, aromatic); MS (70 ev):  $m/z$  (%) = 216 (60, M<sup>+</sup>), 201 (32, M<sup>+</sup>–CH<sub>3</sub>), 187 (9), 173 (13), 139 (100, M<sup>+</sup>–C<sub>6</sub>H<sub>5</sub>), 105 (28), 77 (25), 51 (4).

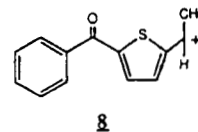
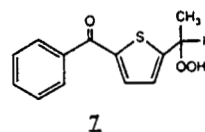
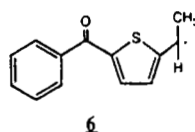
## Results and Discussion

The major photoproduct of 1 in oxygenated media was the ketone 2 (yield = 94%). Its formation, together with traces of the alcohol 4, can be explained as the result of oxygen trapping by a decarboxylated radical 6 and subsequent breakdown of an unstable hydroperoxide 7 (see structure). A similar mechanism has been proposed in the case of the closely related 2-arylpropionic acids ibuprofen,<sup>7</sup> naproxen,<sup>8-10</sup> and ketoprofen,<sup>11,12</sup> whose irradiation under oxygen also produces the analogous ketones as major products. Irradiation of 1 under anaerobic conditions resulted in the formation of 5 (yield = 96%), which can be explained as occurring from intermediate 6 via hydrogen abstraction from the medium. The small amounts of the methyl ether 3 might arise from the cation 8 after photoionization and subsequent homolysis of the C–C bond  $\alpha$  to the carboxyl group.<sup>9</sup>

In a separate experiment, 1 was found to be capable of producing singlet oxygen. Studies repeated in the presence of 2,5-dimethylfuran (<sup>1</sup>O<sub>2</sub> scavenger)<sup>8,12,13</sup> showed the formation of hexene-2,5-dione, detected by GC-MS. Thus, in principle, it could be possible that 1 photosensitizes its own oxidation, through singlet oxygenation of its enol form and subsequent decarboxylation of an intermediate,  $\alpha$ -peroxylac-



- 1: R<sup>1</sup> = COOH, R<sup>2</sup> = H  
2: R<sup>1</sup>, R<sup>2</sup> = O  
3: R<sup>1</sup> = OCH<sub>3</sub>, R<sup>2</sup> = H  
4: R<sup>1</sup> = OH, R<sup>2</sup> = H  
5: R<sup>1</sup> = R<sup>2</sup> = H



tone or  $\alpha$ -hydroperoxyacid, to give the ketone 2.<sup>9</sup> However, this possibility was easily ruled out by irradiating 1 in the presence of tetraphenylporphin (TPP) as source of singlet oxygen, using a potassium chromate solution (100 mg/L) as filter and allowing  $\lambda > 400$  nm (to ensure light absorption by TPP alone and avoid direct irradiation of 1) and maintaining all other conditions the same. In this experiment, no photodegradation of 1 was observed, which means that 1 is not consumed by reaction with  $^1\text{O}_2$  independently generated by TPP-sensitized photolysis.

In light of these data, the phototoxicity mechanism for 1 must involve reactions of free radical 6, stable photoproduct 2 or 5, or singlet oxygen with cell membranes following in vivo photoactivation.

## References and Notes

1. Diffey, B. L.; Daymond, T. J.; Fairgreaves, H. *Brit. J. Rheumatol.* 1983, 22, 239–242.
2. Neumann, R. A.; Knobler, R. M.; Lindenay, H. *Contact Dermatitis* 1989, 20, 270–273.
3. Valsecchi, O.; Dilandro, A.; Pigatto, P.; Cainelli, T. *Contact Dermatitis* 1989, 21, 345–346.
4. Przybilla, B.; Ring, J.; Schwab, U.; Galosi, A.; Dorn, M.; Braun-Falco, O. *Hautarzt* 1987, 38, 18–25.
5. Vonkries, R.; Holzle, E.; Lehmann, P.; Plewig, G. *Photodermatology* 1987, 4, 306–307.
6. Przybilla, B.; Schwab-Przybilla, U.; Ruzicka, T.; Ring, J. *Photodermatology* 1987, 4, 73–78.
7. Castell, J. V.; Gomez, M. J.; Miranda, M. A.; Morera, I. M. *Photochem. Photobiol.* 1987, 46, 991–996.
8. Moore, D. E.; Chappuis, P. P. *Photochem. Photobiol.* 1988, 47, 173–180.
9. Bosca, F.; Miranda, M. A.; Vaño, L.; Vargas, F. *J. Photochem. Photobiol., A. Chem.* 1990, 54, 131–134.
10. Costanzo, L. L.; De Guidi, G.; Condorelli, G.; Cambria, A.; Fama, M. *J. Photochem. Photobiol. B. Biol.* 1989, 3, 223–235.
11. Pietta, P.; Manera, E.; Ceva, P. *J. Chromatogr.* 1987, 390, 454–457.
12. Costanzo, L. L.; De Guidi, G.; Condorelli, G.; Cambria, A.; Fama, M. *Photochem. Photobiol.* 1989, 50, 359–365.
13. Spikes, J. D. In *The Science of Photobiology*; Smith, K. C., Ed.; Plenum; New York, 1977; pp 87–112.

## Acknowledgments

F. Vargas thanks the DGICYT (Spain) for a postdoctoral fellowship.