Total Synthesis of Agelagalastatin

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ABSTRACT



The total synthesis of agelagalastatin, an antineoplastic glycosphingolipid, has been achieved. The synthesis involved an α -selective glycosylation of the ceramide moiety with the trisaccharide fluoride. The trisaccharide component was constructed employing the CB glycoside method which permitted a completely α -stereoselective galactofuranosylation.

Agelagalastatin (1) was isolated from the Western Pacific marine sponge *Agelas* sp., and its structure was elucidated in 1991.¹ This compound is a member of a family of glycospingolipids found in *Agelas* sp. which include agelasphin-9b,² longiside,³ triglycosylceramide,⁴ and KRN7000.⁵ These glycosphingolipids, which commonly contain α -*O*-galactopyranosyl ceramide moieties as their integral parts, have shown immunomodulating activity.^{5,6} Among them, agelasphin-9b and KRN7000 exhibited immunostimulatory activity, which was suggested to be related to the interesting in vivo antitumoral properties of these compounds through an activation of the immune system,^{2,7} and thus, KRN7000 is in clinical trials as a novel anticancer agent.⁸ Agelagal-

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astatin (1) displayed significant in vitro inhibitory activities against human cancer cell growth, its GI₅₀ values ranging from 0.77 μ g/mL for lung NCI-H460 to 2.8 μ g/mL for the ovarian OVCAR-3.1 Besides its biological activity, the unique structure of 1 has attracted our attention as it is composed of two galactofuranosides having α - and β -glycosyl linkages and a galactopyranoside having an α -glycosyl linkage with ceramide, as shown in structure 1. The galactofuranoside moiety with α -configuration is quite rare in Nature and has been found only in a few microorganisms such as Penicillium varians,⁹ Leishmania major,¹⁰ and Talaromyces *flavus.*¹¹ In addition, agelagalastatin (1) was isolated as a mixture of two structural isomers (1a/1b = 4:1) in very low yield $(7.42 \times 10^{-6}\%)$ because of the difficulty in separation.¹ Herein, we describe the first total synthesis of pure 1a and 1b.

Retrosynthesis of the target compound 1 leads to three galactoside building blocks 5-7 and a ceramide moiety 3 (Scheme 1). For the successful synthesis of 1, it is essential to choose efficient glycosylation methods. Particularly challenging is its α -galactofuranosyl moiety, which is often a

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problematic subunit to incorporate stereoselectively. Attention needs to be paid to coupling the four building blocks, 3 and 5-7, in the proper and correct order. The stereospecific construction of 1,2-cis α -galactofuranosyl and α -galactopyranosyl linkages has been one of the great challenges of D-glycoside synthesis.¹² In particular, there are no reliable methods available for the stereospecific synthesis of α -galactofuranosides, although a few attempts have been made employing galactofuranosyl trichloroacetimidates13 and thiogalactofuranosides¹⁴ as glycosyl donors. Our original plan was to employ 2'-carboxybenzyl (CB) glycosides as glycosyl donors¹⁵ for the coupling of all four components of **1**. We envisioned that the construction of the α -galactopyranosyl linkage of 1 could be carried out by coupling between CB trisaccharide 2 and the ceramide moiety 3 in the later stages. We also believed that the crucial α -galactofuranosylation might be possible in the reaction between CB galactofuranoside 5 and 2'-(benzyloxycarbonyl)benzyl (BCB) disaccharide 4, which would be readily obtainable from 6 and 7.

Our synthesis commenced with the preparation of three building blocks **5**–**7** starting from D-galactose: compounds **5** and **6** were prepared via peracetylgalactofuranose **8**¹⁶ as shown in Scheme 2 and compound **7** via 2,3,4,6-tetra-*O*-acetyl- α -D-galactopyranosyl bromide (see the Supporting Information).

Anomeric chlorination of **8** followed by coupling of the resulting crude galactosyl chloride and benzyl 2-hydroxymethylbenzoate (**18**) afforded BCB tetraacetylgalactoside **9**. After conversion of **9** to **10** by deacetylation, the tetrol **10** was subjected to *O*-benzylation. Subsequent selective hydrogenolysis of the benzyl ester functionality¹⁵ in the resultant



BCB benzylgalactoside **11** thereafter provided the building block **5**. Protection of **10** with dimethoxypropane, on the other hand, gave the 5,6-*O*-isopropylidene derivative **12** and the subsequent protection of the diol **12** with TBDMSCI afforded the desired 2-*O*-silyl ether **13** along with a small amount of 3-*O*-silyl ether **14** (**13/14** = 10:1) in 90% yield. *O*-Benzylation of **13** followed by *O*-desilylation of the resulting **15** with Bu₄NF gave alcohol **16**. After *O*-pivaloylation of **16**, the resultant BCB galactoside **17** was converted into the building block **6**. Attempts to directly convert **12** into **17** by selective *O*-pivaloylation of the diol **12** by using PivCl in the presence of pyridine at 0 °C resulted in the formation of a mixture of 2-*O*-, 3-*O*- and 2,3-di-*O*-pivaloyl compounds.

To evaluate the exact reaction conditions needed and appropriate structures for glycosyl donors and acceptors in the crucial α -galactofuranosylation, we carried out a model study on the reaction of the donor 5 with the simpler acceptor 16 in the place of the disaccharide acceptor 4. The model study began with dropwise addition of a diluted solution of Tf₂O in CH₂Cl₂ to a solution of 5, 16, and 2,6-di-tert-butyl-4-methylpyridine (DTBMP) in CH₂Cl₂ at -78 °C to afford the desired α -disaccharide **19** exclusively in 68% yield along with self-condensed ester 20, which probably resulted from coupling between the carboxylate anion and the oxocarbenium ion generated from 5, in 24% yield (method A in Scheme 3). To suppress formation of the ester 20, the glycosylation was carried out with reversal of the order of the addition of reactants such that the concentration of 5 could be kept to a minimum during the glycosylation. Thus, slow addition of the donor 5 to a solution of acceptor 16, DTBMP, and Tf₂O in CH₂Cl₂ at -78 °C afforded only the α -galactosyl disaccharide **19** in 82% yield (method B). The stereochemistry at the newly generated anomeric center of the disaccharide 19 was determined on the basis of its ¹H and ¹³C NMR spectral data, especially the H1'-H2' coupling constant ($J_{\text{H1'-H2'}} = 4.5 \text{ Hz}$) and the C1' chemical shift (δ_{C1} ' 99.0).14

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With this promising result for the α -galactofuranosylation with **5**, the stage was set for the assembly of building blocks **5–7** to make the trisaccharide **2** (Scheme 4). Glycosylation



of **7** with **6** was carried out under the conditions of method B to give the β -disaccharide **21** exclusively in 79% yield. Removal of the *O*-pivaloyl group from **21** with NaOBn gave the alcohol **4**. Then, the crucial α -galactofuranosylation of the acceptor **4** with the donor **5** was successfully executed under the conditions of method B to afford the desired α -galactofuranosyl trisaccharide **22** ($J_{\text{H1'-H2'}} = 4.6$ Hz and $\delta_{\text{C1'}}$ 98.5) exclusively in 91% yield. No β -trisaccharide was detected at all in the reaction mixture. The BCB trisaccharide **22** was converted into the CB trisaccharide **2** by selective hydrogenolysis. It is noteworthy that the reaction of the CB galactofuranoside **5** with the disaccharide **4** provided only the α -trisaccharide **22** in excellent yield, which is the first example of a completely stereoselective α -galactofuranosyl-ation.

Our approach to the synthesis of ceramides 3a and 3b made use of the olefin cross-metathesis reaction to install the main carbon chain. Recently, the cross-metathesis olefination for the synthesis of sphingosines has become attractive due to the high *E*-selectivity and good yield of the process and its wide functional group tolerance.¹⁷ Wittig reaction of known D-lyxose derivative 23^{18} with ylide CH₂= PPh₃ gave olefin **24**. The olefin cross-metathesis of **24** with 11-methyldodec-1-ene (**39**) and with 12-methyltridec-1-ene (**40**) in the presence of Grubbs' catalyst **41**¹⁹ in refluxing CH₂Cl₂ provided the desired *trans*-olefins **25** and **26** in 75% and 76% yields, respectively (Scheme 5). After hydrogena-





tion of **25**, the hydroxy group in **27** was activated as an *O*-triflate, and this subjected to an S_N^2 reaction with tetramethylguanidinium azide $(TMGA)^{20}$ to give azido compound **29** with inverted configuration. Removal of the *O*-isopropylidene group of **29** by trifluoroacetic acid (TFA) followed by benzoylation of the resulting diol afforded

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compound **31** which, after hydrogenolysis, gave amine **33**. The homologue of **33**, compound **34**, was readily obtained in a like fashion from **26**. Coupling reaction of **33** with (*R*)-2-acetoxypentacosanoic acid (**35**) using 2-ethoxy-1-ethoxy-carbonyl-1,2-dihydroquinoline (EEDQ)²¹ afforded amide **37**, and same reaction of **34** with (*R*)-2-acetoxytetracosanoic acid (**36**) provided amide **38**. Finally, a desilylation of **37** and **38** with Bu₄NF gave alcohols **3a** and **3b**, respectively.

Glycosylation of the ceramide 3a with the CB trisaccharide 2 afforded the desired protected agelagalastatin 42a α (δ_{C1} 100.6) in 45% yield, along with undesired β -anomer 42a β $(\delta_{C1} 104.5)$ in 32% yield, and the reaction of ceramide **3b** with 2 also gave a mixture of α - and β -anomers 42b with the same ratio ($\alpha/\beta = 1.4:1$) in 75% yield. Poor stereoselectivity in the glycosylation of **3** with the CB glycoside **2** led us to examine other glycosyl donors that could be readily prepared from 2. We envisaged that sequential treatment of the CB glycoside 2 with Tf_2O and then with a fluoride reagent would give the glycosyl fluroride. Reaction of 2 with Tf₂O and then with (diethylamino)sulfur trifluoride (DAST) as a fluoride source afforded a mixture of α - and β -glycosyl fluoride 43 in 48% yield, whereas the same reaction with hydrogen fluoride pyridine as a fluoride source was quite efficient giving 43 in 83% ($\beta/\alpha = 20:1$) yield (Scheme 6).

Glycosylations of **3a** and **3b** with **43** as the glycosyl donor were carried out by dropwise addition of a solution of **43** in THF to a solution of acceptor **3a** or **3b**, SnCl₂, AgClO₄,²² and DTBMP in THF at -10 °C to afford desired α -glycosides **42a** α and **42b** α exclusively in 72% and 73% yield, respectively. The deprotection of **42a** α started with removal of the *O*-benzylidene and *O*-isopropylidene groups using TFA and was followed by hydrogenolysis to remove the benzyl groups. Finally, the *O*-acetyl and *O*-benzoyl protecting groups were removed using sodium methoxide in MeOH and CH₂Cl₂ to give the target compound **1a**. The other target compound **1b** was obtained from **42b** α by the same deprotection sequence under the same reaction conditions.

The ¹H NMR and ¹³C NMR spectra of the authentic agelagalastatin isolated by Pettit and of our synthetic agelagalastatin (1) were identical in all respects.²³ Also, the optical rotations of synthetic compounds **1a** and **1b** were measured to be $[\alpha]_D + 58.8 (c \ 0.65, CH_3OH)$ and $[\alpha]_D + 59.8 (c \ 0.45, CH_3OH)$, respectively, which is exactly matched with the reported value of $[\alpha]_D + 59 (c \ 0.65, CH_3OH)$ for the authentic agelagalastatin, which is a mixture of **1a** and **1b** with a ratio of 4:1.

We have thus achieved the first total synthesis of both the major component **1a** and the minor component **1b** of agelagalastatin (**1**). For the construction of the trisaccharide moiety **2**, we made use of the CB glycoside method.



Particularly, the glycosylation of **4** with the CB galactofuranoside **5** showed complete α -stereoselectivity, giving only the α -galactofuranosyl trisaccharide **22**. On the other hand, for the coupling of the trisaccharide moiety and the ceramide part, trisaccharyl fluoride **43** was utilized as the glycosyl donor to afford α -glycosides **42a** α and **42b** α exclusively. For the synthesis of ceramides **3a** and **3b**, we employed olefin cross metathesis to install the main carbon chain efficiently. The biological activity of each component of agelagalastatin, **1a** and **1b**, will be reported in a separate communication.

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Supporting Information Available: Experimental procedures and characterization data. This material is available free of charge via the Internet at http://pubs.acs.org.

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