Synthesis of the Tetraketide Lactones from the Pikromycin **Biosynthetic Pathway**

Hong-Se Oh,^[a] Ji-Suk Yun,^[a] Ki-Hyun Nah,^[a] Han-Young Kang,^{*[a]} and David H. Sherman^[b]

Keywords: Polyketide / Pikromycin / Biosynthesis / Lactones / Asymmetric synthesis

Synthesis of tetraketide lactones 2 and 3, which are likely to be produced by a model pikromycin polyketide synthase (PKS), has been investigated. The tetraketide lactones with six-membered rings, 2a and 2b, were synthesized successfully by the asymmetric aldol reaction, allylation, and the Reformatsky reaction. The attempted synthesis of tetraketide lactones with eight-membered rings, 3a and 3b, led to the formation of the compounds 2a and 2b. The synthesis of an-

other tetraketide lactone compounds 35 was attempted with the hope that introducing an additional methyl group would lead to a change in thermodynamic stability. However, it produced the corresponding tetraketide lactone 34 with a sixmembered ring.

(© Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, Germany, 2007)

Introduction

The development of the modern synthetic methods has made important contributions to not only answering the emerging questions from chemistry but also explaining a variety of phenomena originating from the biological fields. Recently, studies on natural product chemistry based on genetic techniques have enjoyed a dramatic growth.^[1] The driving force of this is mainly attributed to the power of modern organic synthesis and the fast-developing genetic knowledge and techniques. Polyketide macrolides belong to a well-studied class of compounds in natural products, and the investigation of their biosynthesis is dependent on genetic techniques. For example, erythromycin is a typical macrolide that has been a target for examining various biosynthetic pathways using the newly developed genetic concepts.^[2] Polyketide synthase (PKS), an enzyme responsible for producing erythromycin, is made up of characteristic modular structures. Studies on 6-deoxyerythronolide B polyketide synthase have made a significant contribution to expanding our understanding of the bacterial type I PKS, which is organized of modules, each of which catalyzes one cycle of elongation of the chain.^[3]

Pikromycin is another polyketide macrolide that has attracted considerable attention in biosynthetic research on account of the remarkable ability of the PKS enzyme to produce this large ring-sized polyketide lactone. Pikromycin

Supporting information for this article is available on the

PKS is a gigantic enzyme that is involved in the synthesis of macrolide antibiotics including methymycin and pikromycin.^[3] This is fascinating because a single enzyme produces macrocyclic compounds with two different ring sizes, 12- and 14-membered rings (methymycin and pikromycin). The pikromycin PKS also possesses additional post-modification functions (i.e. glycosylation and oxidation) for the macrolides. These features make PKS an ideal system for generating a diverse range of macrolides through so-called combinatorial biosynthesis, which is a brilliant concept that involves cutting and reshuffling the genes responsible for individual biochemical transformations for the synthesis of iterating structural units.^[4] Figure 1 shows the genetic modular arrangement of the pikromycin PKS system. Whilst understanding the nature of the PKS is essential for investigating the biosynthetic pathway, the structural complexity of the macrolides has been an impediment to the progress of the biosynthetic research. Although, in principle, the concept of combinatorial biosynthesis for generating structurally new compounds by mixing and combining genetic modules is intriguing, biosynthetic research in this area has been hindered by the limited availability of the biosynthesized compounds due to the interrupted function of the resulting hybrid PKS.

A research program was, therefore, initiated with the aim of developing the systematic methods for the synthesis of the possible intermediates; particularly polyketide lactones from the pikromycin biosynthetic pathway with the view to understand further the importance of securing polyketide intermediates from the anticipated biosynthetic pathway. The structure of the products from the genetic work is so complex that we became interested in the simpler PKS system which could, in turn, generate intermediate polyketides with simpler structures. This concept has been demon-



[[]a] Department of Chemistry, Chungbuk National University, Cheongju, Chungbuk 361-763, Republic of Korea Fax: +82-43-267-2279 E-mail: hykang@chungbuk.ac.kr

[[]b] Department of Medicinal Chemistry, University of Michigan, Ann Arbor, MI 48109, USA

WWW under http://www.eurjoc.org or from the author.

FULL PAPER



Figure 1. Modular organization of the pikromycin PKS.

strated with smaller hybrid PKS systems such as DEBS1 (DEBS = 6-deoxyerythronolide B synthase) + TE (thioesterase),^[5] or DEBS1-TE^[6] truncated version of erythromycin PKS. Both these simpler PKSs have been used extensively to examine the biosynthesis of 6-deoxyerythrolide B. These hybrid PKSs have produced the expected smaller polyketide lactones.^[7]

Inspired by the simpler erythromycin PKS systems, a similar truncated version of the hybrid PKS system was designed for pikromycin biosynthesis. The model PKS system used in this study involved cutting the genes between the AT and DH domain in module 2 and between AT and KR domain in module 5 and combining the two modules. It was anticipated that this fused PKS would produce tri- and tetraketide lactones, i.e. six-membered and eight-membered lactones, as shown in Figure 2. Therefore, the hybrid model PKS could serve not only as a model system but also as a valuable tool for investigating the function and role of modules 5 and 6, which is expected to make a significant contribution to the understanding of the pikromycin PKS system.^[8]

In order to achieve a combinatorial biosynthesis using a polyketide synthase, it is essential to understand the biosynthetic pathways and experiences of handling the intermediates efficiently. Therefore, the aim of this study is to develop synthetic routes to prepare polyketide lactones through the pikromycin biosynthetic pathways. In connection with the generation of a simple hybrid PKSs derived from the pikro-



Figure 2. Model Pik PKS system for the synthetic targets. Fusion of pik AI mod::pik AIII mod 5.

mycin PKS system, the initial focus was on synthesizing the polyketide lactones 1, 2, and 3. The synthesis of simple sixmembered δ -lactones 1 is reported previously.^[9,10] Here, we describe the results of the synthetic studies of the tetrake-tide lactones 2 and 3.

Results and Discussion

The retrosynthetic analysis of the lactones shown in Scheme 1 indicates that the target tetraketide lactone **2**, which mainly exists as an enol form, could be synthesized from the linear intermediate **A** through a simple consecutive two-carbon chain elongation of the intermediate, which was previously prepared during the synthesis of the triketide δ -lactone **1**.^[9] Because it is preferable to secure all the possible intermediate polyketide lactones for future biosynthetic research, we believed that it would be more appropriate to develop a synthetic route to prepare not only the lactones **2** and **3** but also their stereoisomers. Therefore, an attempt was made to synthesize the target lactones and their diastereomers (i.e. C5-epimers, vide infra).



Scheme 1. Retrosynthetic scheme.

Scheme 2 shows the synthetic route according to the retrosynthetic analysis (Route a). Aldehyde **5** was obtained by the oxidation of the alcohol **4**.^[9] The aldol reaction proceeded without incident to give the aldol product **7**. Oxidation provided the 1,3-keto compound **8**, which was hydrolyzed to the corresponding carboxylic acid. However, cyclization of the resulting carboxylic acid was unsuccessful. Because this failure might be due to the enolization of the 1,3-keto compound **8**, cyclization was attempted using the aldol product **7**. Cyclization was achieved successfully and oxidation with the resulting lactone provided the keto lactone **9**. Unfortunately, the deprotection of the benzyl group was unsuccessful, even with various known methods. The difficulty in deprotection led us to examine other approaches for preparing the tetraketide lactones.

The synthetic route was re-analyzed and an attempt was made to devise a common route for synthesizing both tetraketide lactones 2 and 3 (a six-membered and a eight-membered lactone, respectively). Retrosynthetic analysis revealed that the paths that lead to the intermediates **B** and **C** in Scheme 1 (which would lead to compounds 2 and 3, respectively, route b) would have the advantage of employing similar intermediates for the synthesis of both tetraketide lactones. Moreover, the synthesis of a similar tetraketide lactone has been reported (Scheme 3).^[11] An intramo-



Scheme 2. Reagents and conditions: (a) Swern Oxidation (83%) (b) **6a**, Bu₂OTf, Et₃N, CH₂Cl₂ (68%) (c) Dess–Martin periodinane (DMP), CH₂Cl₂ (80%) (d) (1) H₂O₂, LiOH (2) 1 M HCl/THF, 5:1 (68%, two steps) (e) DMP, CH₂Cl₂ (85%).

lecular Claisen condensation was employed for the efficient construction of the six-membered lactones for a similar tetraketide lactone.



Scheme 3. An example of the Claisen condensation approach.

The tetraketide lactones synthesized in this study could be achieved using a similar procedure because the only difference between the tetraketide lactone in Scheme 3 and the target lactone **2a** is the existence of a methyl group. It was believed that this synthesis would be quite straightforward. Scheme 4 summarizes the synthesis according to the Claisen condensation route.



Scheme 4. Attempt to synthesize the tetraketide lactone according to the Claisen condensation route.

The acylated product **11a**, which was prepared by the propionation of the previously reported alcohol **10**, was treated with a strong base (KHMDS) with the aim of producing the desired lactone. However, the product formed is believed to be the α , β -unsaturated compound **12a**. The

FULL PAPER

anion formed by proton abstraction at the α position can lead to the elimination of the EtCO₂ group (Scheme 4). The absence of a methyl group at the α position created a difference in the course of the reaction. Hoping that the acidity of the α -proton might be altered by changing the chiral auxiliary with other group, the oxazolidinone moiety was switched to the Weinreb amide (10b). However, this change did not lead to the production of the desired lactone. The formation of the elimination product 12b was observed. Therefore, it was necessary to modify the synthetic route. The use of the intramolecular samarium(II) iodide-mediated Reformatsky reaction to produce various cyclic products including six- and eight-membered rings was considered (Scheme 5).^[12] The intramolecular Reformatsky reaction mediated by samarium(II) iodide has been used successfully by many researchers, including Molander and Mukaiyama, to prepare cyclic products.



Scheme 5. Intramolecular SmI₂-mediated Reformatsky reaction.

The samarium(II) iodide-mediated intramolecular Reformatsky reaction was used to synthesize the desired tetraketide lactones **2a** and **2b**, as summarized in Scheme 6. Although the biosynthetic mechanism suggests compound **2b** to be the expected product with the correct stereochemistry, a decision was made to synthesize both epimers for the future biosynthetic studies.



Scheme 6. Reagents and conditions: (a) $ref.^{[9]}$ [SmI₂, THF, -78 °C] (b) (1) LiBH₄, H₂O (78%) (2) imidazole, TBSCl (97%) (c) (1) CH₃CHBrCOOH, DCC, DMAP (96%) (2) CSA(cat.), MeOH (63%) (3) DMP (72%) (d) SmI₂, -78 °C (24:1 selectivity, 59%) (e) (1) DMP (68%), (2) HF/CH₃CN (74%) (f) (1) LiBH₄, H₂O (70%), (2) imidazole, TBSCl (99%) (g) (1) CH₃CHBrCOOH, DCC, DMAP (97%) (2) CSA(cat.), MeOH (88%) (3) DMP (89%) (h) SmI₂, THF, -78 °C (20:1 selectivity, 60%) (i) (1) DMP (68%) (2) HF/CH₃CN (70%).

The asymmetric intermolecular Reformatsky reaction mediated by samarium(II) iodide^[13] provided the epimeric aldol products 14a and 14b (with no diastereoselectivity). After chromatographic separation, the aldol product 14a (a series) was converted to a TBS-protected triol 15a, by reduction followed by silvlation of the newly formed primary hydroxy group. The required ester for the intramolecular Reformatsky reaction was prepared by esterification with DCC. The hydroxy group was deprotected under acidic conditions, and the aldehyde 16a, which is a precursor for cyclization, was secured. The key samarium(II) iodide-mediated intramolecular Reformatsky reaction was performed successfully (SmI₂, THF, -78 °C). The resulting lactone (ca. 24:1 diastereoselectivity) was oxidized and deprotected to afford the desired tetraketide lactone 2a. The epimeric lactone 2b was synthesized by an identical reaction sequence starting from the aldol product 14b (b series). Therefore, the Reformatsky reaction mediated by samarium(II) iodide with aldehyde 16b yielded the lactone 17b (diastereoselectivity = 20:1). Oxidation followed by deprotection produced the desired tetraketide lactone 2b.

The synthesis of the eight-membered tetraketide lactones 3 was attempted after successfully synthesizing the sixmembered ring tetraketide lactones 2a and 2b. The plan was to use the same protected triol intermediate, which was thought to give both target tetraketide lactones 2 and 3 depending on which hydroxy group was chosen for acylation. The asymmetric Reformatsky reaction of compound 6a with the aldehyde 13b gave a 13:1 mixture of the compounds 18a and 18b (Scheme 7).^[14] The same reaction with compound **6b** provided a lower ratio (3:1) as a consequence of a mismatched pair. After protecting the free hydroxy group at the 3-position of the major isomer 18a with a silyl group, the chiral auxilary was removed by reduction with LiBH₄. The resulting primary alcohol was protected with the TBS group. The secondary alcohol 19a was obtained by deprotecting the PMB group. It was expected that this alcohol would be converted into the desired eight-membered lactones 3a.

The free secondary hydroxy group of compound 19a was esterified with 2-bromopropanoic acid. The removal of the TBS group followed by oxidation gave the aldehyde 20a, which was subjected to the key samarium(II) iodide-mediated intramolecular Reformatsky reaction. ¹H NMR spectral analysis showed that the Reformatsky reaction gave the eight-membered lactone 21a as a mixture of two diastereomers with a ratio of 7:1. The precise stereochemistry of the products in the mixture was not determined. The mixture was further oxidized to produce compound 22a as an 8:1 mixture with a C-2 stereocenter. Therefore, the intramolecular Reformatsky reaction was expected to be highly stereoselective with respect to the chiral C-3 atom. It was hoped that the stereochemistry at C-2 would be controlled by the thermodynamic stability. The only remaining step to complete the synthesis of the eight-membered lactone 3a was the deprotection of the hydroxy group. Surprisingly, although understandable under the deprotection condition (HF/CH₃CN), the protected lactone 22a was converted into



Scheme 7. Reagents and conditions: (a) SmI_2 , THF, -78 °C (65%) [dr = 13:1 (**18a/18b**)] (b) SmI_2 , THF, -78 °C (74%) [dr = 3:1 (a:b)] (c) 2,6-Lutidine, TBSOTf (82%) (d) (1) LiBH₄, H₂O (94%) (2) imidazole, TBSCI (95%) (3) DDQ (74%) (e) (1) CH₃CHBrCOOH, DCC (97%) (2) CSA(cat.), MeOH (88%) (3) DMP (91%) (f) SmI₂, -78 °C (72%, 7:1 mixture) (g) DMP (88%, 8:1 mixture) (h) HF/ CH₃CN (58%).

the tetraketide δ -lactone **2a**. The attempted deprotection of the lactone removed the silyl group and caused the translactonization to produce the lactone **2a**, which was identical to the lactone prepared previously (Scheme 6). This translactonization is likely driven by the thermodynamic stability.^[15] Desilylation **22a** under acidic conditions (HCl, AcOH, or CSA) also tried but failed to produce the desired lactone **3a**. Only decomposition was observed. The lactone having the PMB-protected hydroxy group (**22c**) was also prepared to find out any difference upon changing the protecting group. However, only **2a** was formed under the standard deprotection condition (DDQ, CH₂Cl₂:H₂O = 10:1, 0 °C). The structure of the lactone **2a** was confirmed by ¹H NMR, ¹³C NMR, and ¹H-¹H COSY spectra.

It was believed that it would be preferable to secure the possible diastereomers of the target lactones for the future biosynthetic studies. Therefore, an attempt was made to synthesize the diastereomeric lactone 3b. The main interest in this attempt was to determine the effect of the stereochemistry at the C-5 position. The synthesis of the epimeric tetraketide lactone 3b was initiated using a modified procedure from that used in the attempted synthesis of 3a. It was hoped that the hydroxy group with the epimeric stereochemistry at C-5 would lead to an eight-membered ring structure or render the resulting oxygen-centered anion, which had been formed by the deprotection of the silyl group, unable to attack the carbonyl carbon of the lactone. At the time of performing this synthesis, a more practical synthetic route was adopted to prepare the required precursors for the key Reformatsky reaction (Scheme 8; b series). In other words, the allylation reaction was used instead of the aldol reaction based on the Evans' chiral auxiliaries. Although, the conditions for producing the desired allylated

FULL PAPER

product **23b** exclusively were determined,^[10] a simple, practical, substrate-controlled approach was adopted for the synthesis of compound 23b. The aldehyde 13b was allylated to give a 3:2 mixture of compounds 23a and 23b. The undesired isomer 23a was recycled to the desired isomer 23b by oxidation followed by reduction. The allylated alcohol 23b was obtained in 40% yield after this oxidation-reduction sequence. The alcohol 23b was converted into the aldehyde 25b through protection of the free hydroxy and oxidative cleavage of the vinyl group. The aldehyde 25b was then reduced, and the resulting primary hydroxy group was protected with the TBS group. The PMB ether was deprotected and acylated. After regenerating the primary hydroxy group followed by oxidation, the key intramolecular Reformatsky reaction was performed using the aldehyde 20b. The desired eight-membered ring was formed as a mixture of two diastereomers with a ratio of 1:2. The two products are believed to be the diastereomeric mixture of C-2 (and C-3) isomers. Furthermore, although not precisely identified, two diastereomeric products appeared to be mainly C-2 stereoisomers. This was supported by the fact that the β keto ester 22b, which was formed by the oxidation of compound 21b, maintained a similar isomeric ratio (2:1). Un-



Scheme 8. Reagents and conditions: (a) In, allyl bromide, THF:H₂O = 1:1 (70%) (b) TBSOTf, 2,6-lutidine, 0 °C, CH₂Cl₂ (98%) (c) (1) DMP, (2) NaBH₄ (2:3 mixture) (recycled, 40%) (d) (1) NMO-OsO₄ (2) NaIO₄ (*t*BuOH:THF:H₂O = 10:2:1) (92%, two steps) (e) (1) NaBH₄, MeOH, 0 °C (94%) (2) TBSOTf, 2,6-lutidine, 0 °C, CH₂Cl₂ (99%) (f) DDQ, DCM:pH7 buffer = 10:1 (96%) (g) (1) CH₃CHBrCOOH, DMAP(cat), DCC (95%) (2) CSA, MeOH (91%) (3) DMP (90%) (h) SmI₂, CH₂Cl₂ (84%, 2:1 mixture) (i) DMP (96%, 2:1 mixture) (j) HF/CH₃CN, room temp. (97%).

fortunately, it was found that deprotection of the silyl group of **22b** did not lead to isolate the desired eight-membered lactone **3b** but instead, produced the tetraketide lactone **2b**. Therefore, the translactonization to an eight-membered lactone by attacking the anionic oxygen atom, which was formed by deprotecting of the ester carbonyl group, was favored.

We were unable to obtain the desired eight-membered lactones with free hydroxy groups 3a and 3b, even though the synthesis of eight-membered lactone rings has been successful using the samarium(II) iodide-promoted intramolecular Reformatsky reaction. As a result, a decision was made to further examine the factors that influence the stability. We have experienced that the existence of a methyl group can make a critical difference in acidity that can enable the desired Claisen condensation to proceed. Therefore, it would be interesting to check the behavior of a similar structure in cyclization with the presence of the methyl group at the C-4 position. A lactone with a similar structure to the compound 2 could be prepared with an additional methyl group at the C-4 position. Scheme 9 shows the synthetic pathway of the lactone. The aldol reaction was performed with compounds 27 and 13b to provide compound **28**. This aldol product **28** was converted into the fully protected triol 29 in a straightforward manner. After deprotection of the PMB group, the same procedure as that used in the previous attempt to prepare compound 3 was applied. The aldehyde 31, which is the precursor for the Reformatsky reaction, was treated with samarium(II) iodide in THF. The desired eight-membered lactone (with a methyl group at C-4) 32 was formed efficiently as a diastereomeric mixture with a ratio of 5:1.



Scheme 9. Reagents and conditions: (a) **13b**, Bu_2BOTf , Et_3N (80%) (b) (1) 2,6-lutidine, TBSOTf, DCM, 0 °C (89%) (2) LiAlH₄, H₂O (86%) (3) 2,6-lutidine, TBSOTf (94%) (c) DDQ, DCM:pH 7 buffer = 10:1 (99%) (d) (1) CH₃CHBrCOOH, DMAP(cat), DCC, CH₂Cl₂, 0 °C (89%) (2) CSA, MeOH, 0 °C (85%) (3) DMP, CH₂Cl₂ (91%) (e) SmI₂, THF, -78 °C (62%) (f) DMP, CH₂Cl₂, room temp. (77%) (g) HF, CH₃CN (69%).

The major isomer of **32** was oxidized to a keto ester **33** using Dess–Martin periodinane to give a mixture of diastereomers (2:1, C-2 isomers). Deprotection of the TBS ether also afforded the δ -lactone **34** instead of the desired ξ -lactone **35**. The hydroxy group of compound **32** was deprotected and treated with HF in order to determine the effect of the keto group. In this case, the corresponding δ lactone was also formed but the formation of the eightmembered lactone was not observed.

Conclusions

The overall aim of this study was to synthesize the intermediates and products to investigate various biosynthetic pathways. The main focus was on developing synthetic routes to prepare polyketide lactones, which are involved in the pikromycin biosynthetic pathway. A model PKS system was devised to simplify the investigation. This study examined the synthetic routes for the simpler tetraketide lactones with the six- or eight-membered rings anticipated by this model PKS. The routes were developed based on an identical key intermediate used to synthesize both tetraketide lactones with six- and eight-membered rings. The tetraketide lactones 2a and 2b with six-membered rings were synthesized efficiently utilizing the asymmetric aldol reaction and the samarium(II) iodide-mediated Reformatsky reaction as the key reactions. The formation of eight-membered lactone derivatives was also successfully achieved using the samarium(II) iodide-mediated Reformatsky reactions. Therefore, a synthetic route for the tetraketide lactone derivatives with eight-membered rings has been developed. However, the synthesis of the final target tetraketide lactones with a free hydroxy group was unsuccessful. The hydroxy lactones with an eight-membered ring, which were obtained from the final deprotection of the silvl groups, spontaneously converted into the tetraketide lactones 2a and 2b prepared previously. This indicates that the tetraketide hydroxy lactones with six-membered ring are thermodynamically more stable than those with eight-membered rings. The results of this study strongly indicate that the model PKS would produce only the tetraketide lactones with six-membered rings from the future incubation study. The eight-membered hydroxy lactone with an additional methyl group was also transformed to the corresponding six-membered lactone 34. This confirms the stability of the tetraketide lactones with six-membered rings, which is in contrast to the corresponding lactones with eight-membered rings.

The results from this synthetic investigation are expected to provide important information for the future biosynthetic research with the model PKS. We are planning to carry out an incubation study with the model PKS to elucidate the pikromycin biosynthetic mechanisms. The tetraketide lactones prepared in this study will facilitate the investigation by resolving the difficulty in identifying the key intermediates formed by the model PKS. This synthetic study would also provide valuable experience and knowledge to prepare related and more complex polyketide lactones from the pikromycin biosynthetic pathway. Further synthetic investigations into more complicated polyketide lactones will be reported in the future.

Experimental Section

¹H NMR and ¹³C NMR spectra were recorded with a Bruker DPX-300 and Bruker Avance 500 NMR Spectrometer. The chemical shifts are reported in ppm on scale downfield from TMS, and signal patterns are indicated as follows: s, singlet; d, doublet; t, triplet; m, multiplet; br, broad peak. IR spectra were recorded with a JASCO FT/IR-300E. Optical rotations were measured by JASCO DIP-1000 digital polarometer in solution in a 1-dm cell. Mass spectra (and HRMS) were obtained with a VG AUTOSPEC Ultma GC/MS system using direct insertion probe (DIP) and electronimpact method (EI, 70 eV). All reagent and solvents were reagent grade and used without further purification unless specified otherwise. Technical grade ethyl acetate, hexane, and pentane used for column chromatography were distilled prior to use. Tetrahydrofuran (THF) and diethyl ether, when used as solvents for reactions, were freshly distilled from sodium/benzophenone ketyl. Dimethylformamide (DMF) was stored over 4-Å molecular sieves, and triethylamine was distilled before use. Flash chromatography was carried out on Woelm 32-64 µm silica packed in glass columns.^[16]

(3S,4R,5R)-1,5-Bis(tert-butyldimethylsilyloxy)-4-methyl-3-heptanol (15a): Distilled water (41 µL, 2.3 mmol) was added to a solution of (4S,3'S,4'R,5'R)-4-benzyl-3-[5'-(tert-butyldimethylsilyloxy)-3'-hydroxy-4'-methylheptanoyl]-2-oxazolidinone (14a)^[9] (689 mg, 1.53 mmol) in diethyl ether (5 mL). After the solution was cooled to 0 °C, lithium borohydride (1.10 mL of a 2.0 M solution in THF, 2.30 mmol) was added slowly with stirring. After 10 min, the temperature of the solution was raised to room temperature, and stirred for additional 2 h. The reaction was terminated with addition of aqueous NaOH solution (1.0 m, 10 mL) and extracted with diethyl ether $(3 \times 10 \text{ mL})$. After the organic layer was washed with saturated sodium chloride solution (20 mL), the ethereal solution was dried (MgSO₄) and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 2:1) provided the desired diol as a colorless liquid (329 mg, 78%). IR (thin film): \tilde{v} = 3387, 2957, 1641, 1462, 1256, 1057 cm⁻¹. ¹H NMR (300 MHz, $CDCl_3$): $\delta = 3.92$ (dt, J = 9.8, 2.7 Hz, 1 H, CHOSi), 3.74 (m, 3 H, CH₂OH, CHOH), 3.51 (s, 2 H, CH₂OH, CHOH), 1.75 (m, 1 H, CHCH₃), 1.50 (m, 4 H, CH₂CHOH, CH₂CH₃), 0.85 (d, J = 3.6 Hz, 3 H, CHCH₃), 0.83 [s, 9 H, SiC(CH₃)₃], 0.76 (t, J = 7.3 Hz, 3 H, CH₂CH₃), 0.03 [s, 6 H, Si(CH₃)₂] ppm. ¹³C NMR (75 MHz, $CDCl_3$): $\delta = 79.2, 75.5, 62.2, 40.5, 37.4, 27.9, 27.0, 18.6, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4, 6.8, 10.4,$ -3.1, -4.0 ppm. $[a]_{D}^{26.2} = +19.7$ (c = 1.61, CHCl₃). MS (EI): m/z(%) = 276 [M⁺], 219, 202, 175, 145, 134, 117, 109, 77, 71, 68 (100), 58. HRMS: *m*/*z* calcd. for C₁₄H₃₂O₃Si: 276.2120, found 276.2117.

The diol (329 mg, 1.19 mmol) obtained above was dissolved in CH_2Cl_2 (4 mL). To this solution was added a solution of imidazole (122 mg, 1.79 mmol) and *tert*-butyldimethylsilyl chloride (215 mg, 1.43 mmol) in CH_2Cl_2 (1 mL) at 0 °C under nitrogen. After it was stirred for 10 min at 0 °C, the solution was warmed to room temperature and stirred for additional 1 h at room temperature. After the reaction was completed, aqueous saturated NH_4Cl (5 mL) was added and the mixture was extracted with CH_2Cl_2 (3 × 10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexan/EtOAc, 10:1) offered the desired TBS-protected alcohol **15a** (453 mg, 97%)

as a colorless oil. IR (film): $\tilde{v} = 3453$, 2957, 2857, 1641, 1471, 1389, 1361, 1255 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 3.88$ (m, 1 H, CHOH), 3.65–3.80 (m, 3 H, CH₂OSi, CHOSi), 3.25 (s, 1 H, CHOH), 1.68 (m, 1 H, CHCH₃), 1.58–1.62 (m, 2 H, CH₂CHOH), 1.51 (m, 2 H, CH₂CH₃), 0.86 [m, 21 H, SiC(CH₃)₃, SiC(CH₃)₃, CHCH₃] 0.78 (t, J = 7.4 Hz, 3 H, CH₂CH₃), 0.03 [s, 12 H, Si-(CH₃)₂, Si(CH₃)₂] ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = 77.6$, 72.9, 61.9, 40.4, 37.4, 27.2, 25.9, 25.9, 18.2, 18.0, 9.8, 7.1, –3.8, –4.6, –5.5 ppm. [a]_E^{7.6} = +8.1 (c = 1.15, CHCl₃). MS (EI): m/z (%) = 390 [M⁺], 343, 333, 241, 201, 189, 173, 145, 133, 109, 89, 75 (100), 73. HRMS: m/z calcd. for C₂₀H₄₆O₃Si₂: 390.2986, found 390.2968.

(1S,2R,3R)-3-(tert-Butyldimethylsilyloxy)-2-methyl-1-(2-oxoethyl)pentyl 2-Bromopropanoate (16a): 2-Bromopropionic acid (313 µL, 3.48 mmol) and DMAP (114 mg, 0.93 mmol) was added to a solution of 15a (453 mg, 1.16 mmol) in CH₂Cl₂ (5 mL) at room temperature. After the solution was cooled to 0 °C, N,N-dicyclohexylcarbodiimide (264 mg, 1.28 mmol) was added. The resulting solution was stirred for 5 min at 0 °C before it was warmed to room temperature. After additional stirring for 30 min at room temperature, the precipitate was filtered. The filtrate was concentrated and purified by flash chromatography (hexane/EtOAc, 10:1). The desired bromo ester was obtained as a colorless liquid (582 mg, 96%). IR (film): $\tilde{v} = 2928, 2857, 1738, 1471, 1381, 1256 \text{ cm}^{-1}$. ¹H NMR (300 MHz, CDCl₃): δ = 5.12 [m, 1 H, CHO(C=O)], 4.32 [q, J = 7.0 Hz, 1 H, -C(O)CH], 3.60 (m, 3 H, CH₂OSi, CHOSi), 1.89 [m, 3 H, $CH_2CH_2CH(O)$, $CHCH(CH_3)CH$], 1.78 (d, J = 6.9 Hz, 3 H, BrCHCH₃), 1.5 (m, 2 H, CH₂CH₃), 0.90 (m, 6 H, CHCH₃, CH₂CH₃), 0.86 [s, 18 H, SiC(CH₃)₃, SiC(CH₃)₃], 0.02 (s, 6 H, $2 \times \text{SiC}H_3$), 0.00 (s, 6 H, $2 \times \text{SiC}H_3$) ppm. ¹³C NMR (75 MHz, $CDCl_3$): $\delta = 170.5, 75.2, 75.1, 60.6, 41.9, 41.5, 36.9, 27.8, 27.0,$ 22.8, 19.3, 11.3, 10.4, -2.94, -3.38, -4.33 ppm. $[a]_{D}^{24.5} = +5.9$ (c = 1.21, CHCl₃). MS (EI): m/z (%) = 524 [M⁺], 526, 343, 290, 241, 211, 173, 121 (100), 109, 89, 73. HRMS: m/z calcd. for C₂₃H₄₉BrO₄Si₂: 524.2353, found 524.2335.

The bromo ester (582 mg, 1.11 mmol) obtained as described above was dissolved in MeOH (3 mL). To this solution DL-10-camphorsulfonic acid (51 mg, 0.22 mmol) was added. The resulting solution was stirred at 0 °C for 1 h. The reaction was terminated by addition of Et_3N (162 µL, 1.17 mmol). After the solution was concentrated, purification of the residue by flash chromatography (hexane/EtOAc, 3:1) gave the desired primary alcohol (285 mg, 63%) as a colorless liquid. IR (film): $\tilde{v} = 3425, 2932, 2856, 1737,$ 1471, 1380, 1257 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 5.20 [m, 1 H, CHO(C=O)], 4.38 [q, J = 6.9 Hz, 1 H, C(=O)CHBr], 3.60 (m, 3 H, CH₂OH, CHOSi), 2.23 (s, 1 H, CH₂OH), 1.92 (m, 1 H, CHCH₃), 1.86 (d, J = 6.9 Hz, 3 H, BrCHCH₃) 1.46 (dq, J = 7.7, 5.9 Hz, 2 H, CH₂CH₃), 0.96 (d, J = 6.9 Hz, 3 H, CHCH₃), 0.89 [s, 9 H, SiC(CH₃)₃], 0.84 (t, J = 7.5 Hz, 3 H, CH₂CH₃), 0.05 (s, 3 H, SiCH₃), 0.04 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 171.6, 74.9, 74.4, 59.2, 41.9, 40.9, 36.6, 27.2, 26.6, 22.4, 18.8, 11.0, 10.2, -3.40, -3.78 ppm. $[a]_{D}^{23.9} = +4.4$ (c = 1.43, CHCl₃). MS (EI): m/z (%) = 410 [M⁺], 351, 267, 229, 201, 173 (100), 153, 127, 109, 73, 55. HRMS: *m*/*z* calcd. for C₁₇H₃₅BrO₄Si: 410.1488, found 410.1486.

The alcohol (285 mg, 0.69 mmol) obtained as describe above was dissolved in CH_2Cl_2 (5 mL). To this solution was added Dess–Martin periodinane (DMP) (352 mg, 0.83 mmol). The resulting solution was stirred for 30 min at room temperature. After the reaction was completed, aqueous saturated NaHCO₃ (10 mL) was added and the mixture was extracted with CH_2Cl_2 (3 × 10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc,

H.-Y. Kang et al.

7:1) offered the desired aldehyde **16a** (204 mg, 72%) as a yellow liquid. ¹H NMR (300 MHz, CDCl₃): δ = 9.67 [s, 1 H, C(O)*H*], 5.40 [dt, *J* = 6.9, 4.2 Hz, 1 H, CHO(C=O)], 4.27 [q, *J* = 6.8 Hz, 1 H, C(=O)CHBr], 3.56 (m, 1 H, CHOSi), 2.79 [dd, *J* = 16.6, 4.0 Hz, 1 H, one of CH₂C(=O)H], 2.66 (ddd, *J* = 20.7, 16.6, 3.0 Hz, 1 H, BrCHCH₃), 1.86 (m, 1 H, CHCH₃), 1.74 (d, *J* = 6.9 Hz, 3 H, BrCHCH₃), 1.47 (dq, *J* = 13.7, 6.3 Hz, 2 H, CH₂CH₃), 0.90 (d, *J* = 7.8 Hz, 3 H, CHCH₃), 0.84 [s, 9 H, SiC(CH₃)₃], 0.80 (t, *J* = 7.4 Hz, 3 H, CH₂CH₃), 0.05 (s, 3 H, SiCH₃), 0.03 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 200.1, 170.3, 74.9, 72.4, 47.5, 41.3, 40.4, 27.2, 26.6, 22.2, 18.8, 11.1, 10.5, -3.30, -3.71 ppm.

(6S)-6-[(1R,2R)-2-(tert-Butyldimethylsilyloxy)-1-methylbutyl]-4hydroxy-3-methyltetrahydro-2H-pyran-2-one (17a): Diiodomethane (79 µL, 0.98 mmol) was added to a mixture of samarium (powder, 162 mg, 1.07 mmol) in THF (5 mL) at room temperature under nitrogen. Samarium(II) iodide was obtained as a blue-colored solution which was used after stirring for 2 h. After the solution was cooled to -78 °C, a solution of aldehyde 16a (200 mg, 0.49 mmol) in THF (1 mL) was added under nitrogen. After being stirred for 1 h at -78 °C, the solution was warmed to room temperature and 0.1 M HCl (8 mL) was added. The mixture was extracted with diethyl ether $(3 \times 20 \text{ mL})$. After the organic layer was washed with saturated sodium thiosulfate solution (15 mL) and sodium chloride solution (15 mL), the ethereal solution was dried (MgSO₄) and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 3:1) provided the desired lactone 17a as a yellow liquid [92 mg (57%) and 3.9 mg (2.4%), selectivity 24:1]; $R_{\rm f} = 0.57$ and 0.50 (hexane/EtOAc, 1:1), respectively. Spectroscopic data for the major isomer is as follows. IR (film): $\tilde{v} = 3442$, 2933, 2856, 1713, 1462, 1380, 1258, 1197 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 4.79$ [m, 1 H, CHO(C=O)], 4.14 (s, 1 H, CHOH), 3.61 (m, 1 H, CHOSi), 2.44 [qd, J = 7.2, 3.2 Hz, 1 H, C(=O)CHCH₃], 2.04 (m, 1 H, CHCH₃), 1.82 (m, 2 H, CH₂CHOH), 1.67 (m, 2 H, CH_2CH_3), 1.28 (d, J = 7.2 Hz, 3 H, $CHCH_3$), 0.96 (d, J = 6.9 Hz, 3 H, CHCH₃), 0.83 [m, 12 H, CH₂CH₃, Si(CH₃)₃], 0.00 (s, 6 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 174.9, 75.1, 68.5, 42.5, 37.1, 27.1, 26.6, 18.8, 11.7, 10.9, 10.3, -3.3, -3.8 ppm. MS (EI): m/z (%) = 330 [M⁺], 301, 215, 197, 173, 73 (100), 57. HRMS: m/z calcd. for C₁₇H₃₄O₄Si: 330.2226, found 330.2229.

(6S)-4-Hydroxy-6-[(1S,2R)-2-hydroxy-1-methylbutyl]-3-methyl-5,6dihydro-2H-pyran-2-one (2a): Lactone 17a (50 mg, 0.15 mmol) (a 24:1 mixture of diastereomers) obtained as described above was dissolved in CH₂Cl₂ (1 mL). To this solution was added Dess-Martin periodinane (DMP) (127 mg, 0.30 mmol) and the resulting solution was stirred for 30 min at room temperature. After the reaction was completed, aqueous saturated NaHCO3 (5 mL) was added and the mixture was extracted with CH_2Cl_2 (3×10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 3:1) offered the desired ketone (34 mg, 68%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃): δ = 4.76 [ddd, J = 11.8, 6.1, 2.8 Hz, 1 H, CHO(C=O)], 3.59 [dt, J = 6.4, 3.5 Hz, 1 H, $C(=O)CHCH_3$], 3.54 [q, J = 6.6 Hz, 1 H, C(=O)CHCH₃], 2.73 [dd, J = 19.0, 2.9 Hz, 1 H, one of $CH_2C(=O)$], 2.49 [dd, J = 19.0, 11.8 Hz, 1 H, one of CH₂C(=O)], 1.83 (m, 1 H, CHCHCH₃), 1.36–1.61 (m, 2 H, CH_2CH_3), 1.32 [d, J = 6.6 Hz, 3 H, $C(=O)CHCH_3$], 1.04 (d, J =6.9 Hz, 3 H, CH₂CH₃), 0.87 [m, 12 H, SiC(CH₃)₃, CH₂CH₃], 0.04 (s, 3 H, SiCH₃), 0.00 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, $CDCl_3$): $\delta = 202.7, 170.7, 75.8, 75.2, 52.3, 42.5, 42.2, 26.9, 26.5,$ 18.8, 11.1, 10.9, 8.42, -3.30, -3.80 ppm.

The lactone (29 mg, 0.09 mmol) prepared as described above was dissolved in HF/CH₃CN [48% HF/CH₃CN, 1:19 (v/v)] (1 mL) was

stirred for 1 h at room temperature. The reaction was terminated by addition of solid CaCO₃ (30 mg). After the mixture was filtered with aid of Celite and washed with diethyl ether $(3 \times 10 \text{ mL})$ and the filtrate was concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 1:1 to 1:2) provided the desired lactone 2a (14 mg, 74%) as a white solid: M.p. 108-110 °C. IR (film): $\tilde{v} = 3427, 2970, 1724, 1255 \text{ cm}^{-1}$. ¹H NMR (500 MHz, MeOH): $\delta = 4.24$ [ddd, J = 12.3, 6.4, 3.8 Hz, 1 H, CHOC(=O)], 3.53 (dt, J = 6.6, 3.4 Hz, 1 H, CHOH), 2.59–2.65 (m, 1 H, one of CH_2COH), 2.38 (dd, J = 16.9, 3.3 Hz, 1 H, one of CH_2COH), 1.69 (quintet of d, J = 6.8, 3.4 Hz, 1 H, CHCH₃), 1.63 (s, 3 H, $-C=CCH_3$, 1.43 (quintet, J = 7.2 Hz, 2 H, CH_2CH_3), 0.95 (d, J =6.9 Hz, 3 H, CHCH₃), 0.87 (t, J = 7.4 Hz, 3 H, CH₂CH₃) ppm. ¹³C NMR (75 MHz, MeOH): δ = 173, 169, 99.2, 79.1, 73.9, 43.3, 32.6, 28.9, 11.4, 9.78, 8.95 ppm. $[a]_{D}^{26.5} = 13.53$ (c = 0.82, MeOH). MS (EI): m/z (%) = 214 [M⁺], 167, 157, 141, 129, 116, 111, 101, 83, 71, 55 (100). HRMS: m/z calcd. for C₁₁H₁₈O₄: 214.1205, found 214.1205.

(6R)-4-Hydroxy-6-[(1S,2R)-2-hydroxy-1-methylbutyl]-3-methyl-5,6dihydro-2H-pyran-2-one (2b): Lactone 17b (83 mg, 0.25 mmol) (a 20:1 mixture of diastereomers) obtained above was dissolved in CH_2Cl_2 (2 mL). To this solution was added Dess-Martin reagent (213 mg, 0.50 mmol) and the resulting solution was stirred for 30 min at room temperature. After the reaction was completed, aqueous saturated NaHCO3 (5 mL) was added and mixture was extracted with CH_2Cl_2 (3×10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 3:1) offered the desired ketone (56 mg, 68%) as a colorless liquid. ¹H NMR (300 MHz, CDCl₃): δ = 7.99 (br., 1 H, C-OH), 4.22 [dd, J = 16.8, 8.7 Hz, 1 H, CHOC(=O)], 4.04 (ddd, J = 9.7, 6.0, 1.6 Hz, 1 H, CHOSi), 2.50 $(d, J = 7.6 \text{ Hz}, 2 \text{ H}, CH_2C\text{-OH}), 1.84 (m, 1 \text{ H}, CHCH_3), 1.79 (s, 3)$ H, C=CCH₃), 1.49 (m, 2 H, CH₂CH₃), 0.84 [m, 15 H, CHCH₃, CH₂CH₃, SiC(CH₃)₃], 0.04 (s, 3 H, SiCH₃), 0.00 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 202.5, 75.6, 71.5, 52.3, 42.4, 41.1, 32.4, 28.3, 26.5, 18.7, 10.7, 8.97, 8.46, 8.23, -3.55, -4.04 ppm.

The lactone (45 mg, 0.14 mmol) prepared as described above was dissolved in HF/CH₃CN [48% HF/CH₃CN, 1:19 (v/v)] (1 mL) and the resulting solution was stirred for 1 h at room temperature. The reaction was terminated by addition of solid CaCO₃ (30 mg) After the mixture was filtered with diethyl ether $(3 \times 10 \text{ mL})$ and concentrated. Purification by flash chromatography (hexane/EtOAc, 1:1 to 1:2) provided the desired lactone **2b** (20.5 mg, 70%) as a white solid. m.p. 106–107 °C. IR (film): v = 3431, 2931, 1728, 1255, 1138 cm⁻¹. ¹H NMR (300 MHz, MeOH): $\delta = 4.24$ [ddd, J = 11.6, 8.5, 4.8 Hz, 1 H, CHOC(=O)], 3.80 (ddd, J = 11.6, 5.1, 2.0 Hz, 1 H, CHOH), 2.49 (dd, J = 11.7, 1.7 Hz, 1 H, one of CH₂C-OH), 2.34-2.52 (m, 1 H, CH₂C-OH), 1.72 (m, 1 H, CHCH₃), 1.62 (s, 3 H, $-C=CCH_3$), 1.28–1.52 (m, 2 H, CH_2CH_3), 0.87 (t, J = 7.4 Hz, 3 H, CH₂CH₃), 0.79 (d, J = 7.0 Hz, 3 H, CHCH₃) ppm. ¹³C NMR (75 MHz, MeOH): δ = 172.8, 168.8, 99.4, 78.2, 72.4, 43.4, 32.8, 29.1, 11.5, 8.9, 8.6 ppm. $[a]_{D}^{23.8} = 4.17$ (c = 0.42 MeOH). MS (EI): m/z (%) = 214 [M⁺], 179, 167, 156, 141, 127, 121, 111, 101 (100), 95, 83, 71, 55. HRMS: m/z calcd. for C₁₁H₁₈O₄: 214.1205, found 214.1206.

(4R,3'S,4'R,5'R)-4-Benzyl-3-[5'-[(4-methoxybenzyl)oxy]-3'hydroxy-4'-methylheptanoyl]-2-oxazolidinone (18a) and (4R,3'R,4'R,5'R)-4-Benzyl-3-[5'-[(4-methoxybenzyl)oxy]-3'-hydroxy-4'-methylheptanoyl]-2-oxazolidinone (18b): Diiodomethane $(256 \ \mu L, 3.17 \ mmol)$ was added to a mixture of samarium (powder, 573 mg, 3.81 mmol) in THF (5 mL) at room temperature under nitrogen. Samarium(II) iodide was obtained as a greenish blue solution which was used after stirring for 2 h. After the solution was cooled to -78 °C, a solution of aldehyde **13b** (300 mg, 1.27 mmol) and oxazolidinone **6b** (379 mg, 1.27 mmol) in THF (5 mL) was added under nitrogen. After it was stirred for 1 h at -78 °C, the solution was warmed to room temperature and 0.1 M HCl (10 mL) was added. The mixture was extracted with diethyl ether (3×15 mL). After the organic layer was washed with saturated sodium thiosulfate solution (20 mL) and sodium chloride solution (20 mL), the ethereal solution was dried (MgSO₄) and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 3:1) provided the desired **18a** and **18b** as yellow liquids 344 mg (60%) and 26 mg (4.5%) (selectivity 13:1) [minor compound **18b**: $R_{\rm f} = 0.38$ and major compound **18a**: $R_{\rm f} = 0.31$. (hexane/EtOAc, 2:1)].

18a: IR (film): $\tilde{v} = 3453, 3058, 2930, 1781, 1700, 1613, 1514, 1455,$ 1390 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 7.27–7.12 (m, 7 H, PhCH₂CHN-, -OCH₂ArOCH₃), 6.78 (d, J = 8.6 Hz, 2 H, OCH₂- $ArOCH_3$), 4.62 (ddt, J = 10.4, 7.0, 3.3 Hz, 1 H, PhCH₂CHN–), 4.46 (dd, J = 17.6, 10.4 Hz, 2 H, $-OCH_2ArOCH_3$), 4.10 (m, 3 H, -CHOH, PhCH₂CHCH₂O–), 3.80 (d, J = 3.6 Hz, 1 H, $-CHOCH_{2}$ -ArOCH₃), 3.70 (s, 3 H, -OCH₂ArOCH₃), 3.52 (m, 1 H, -CHOH), $3.21 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, \text{PhC}H_2\text{CHCH}_2\text{O}-\text{)}, 3.04 \text{ (dd, } J = 13.4, 3.2 \text{ Hz}, 1 \text{ H}, 1 \text{ H},$ 6.2, 3.5 Hz, 2 H, $-COCH_2CH$), 2.69 (dd, J = 13.4, 9.6 Hz, 1 H, PhC H_2 CHCH $_2$ O-), 1.84 (quintet of d, J = 7.1, 2.6 Hz, 1 H, OHCHCHCH₃), 1.66 (m, 1 H, -CH₂CH₃), 1.45 (m, 1 H, -CH₂CH₃), 0.86 (m, 6 H, -CH₂CH₃, -CHCH₃) ppm. ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3)$: $\delta = 172.1, 159.1, 153.5, 135.2, 130.4, 129.4,$ 129.4, 128.9, 127.2, 113.7, 82.5, 71.4, 70.6, 66.2, 55.2, 41.3, 39.3, 37.8, 23.0, 11.6, 10.7 ppm. $[a]_{D}^{28.0} = -76.5$ (c = 1.92, CHCl₃). MS (EI): m/z (%) = 455 [M⁺], 302, 249, 219, 178, 137, 121 (100), 86, 69. HRMS: *m*/*z* calcd. for C₂₆H₃₃NO₆: 455.2308, found 455.2308.

18b: IR (film): \tilde{v} = 3459, 3054, 2926, 2852, 1782, 1695, 1612, 1514, 1457, 1384 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 7.29–7.12 (m, 7 H, $PhCH_2CHN_{-}$, $-OCH_2ArOCH_3$), 6.78 (d, J = 8.6 Hz, 2 H, $-OCH_2ArOCH_3$, 4.60 (ddt, J = 10.6, 6.8, 3.2 Hz, 1 H, PhCH₂CHN–), 4.49 (d, J = 11.1 Hz, 1 H, –OCH₂ArOCH₃) 4.31 (d, J = 11.1 Hz, 1 H, $-OCH_2ArOCH_3$), 4.21 (br. t, J = 4.3 Hz, 1 H, -CHOH), 4.11 (m, 2 H, PhCH₂CHCH₂O -), 3.71 (s, 3 H, -OCH₂ArOCH₃), 3.42 (m, 1 H, -CHOCH₂ArOCH₃), 3.32 (s, 1 H, -CHOH), 3.23 (dd, J = 13.4, 3.2 Hz, 1 H, -PhCH₂CHCH₂-O–), 3.10 (dd, J = 16.8, 9.2 Hz, 1 H, COCH₂CH), 2.84 (dd, J =16.7, 3.3 Hz, 1 H, $-COCH_2CH$), 2.68 (dd, J = 13.4, 9.6 Hz, 1 H, PhCH₂CHCH₂O-), 1.73 (m, 2 H, -OHCHCHCH₃, -CH₂CH₃), 1.50 (m, 1 H, $-CH_2CH_3$), 0.95 (d, J = 7.1 Hz, 3 H, $-CH CH_3$), 0.8 $(t, J = 7.5 \text{ Hz}, 3 \text{ H}, -CH_2CH_3) \text{ ppm}.$ ¹³C NMR (100 MHz, CDCl₃): $\delta = 172.4, 159.1, 153.4, 135.2, 130.6, 129.4, 128.9, 127.3, 113.8,$ 83.5, 70.7, 70.6, 66.2, 55.2, 55.1, 41.0, 39.1, 37.7, 23.1, 10.0, 7.4 ppm. $[a]_{D}^{20.9} = -54.1$ (c = 1.23, CHCl₃). MS (EI): m/z (%) = 455 [M⁺], 301, 248, 219, 178, 137, 121 (100), 86, 77, 65. HRMS: m/z calcd. for C₂₆H₃₃NO₆: 455.2308, found 455.2304.

(3*S*,4*S*,5*R*)-1,3-Bis(*tert*-butyldimethylsilyloxy)-4-methylheptan-5-ol (19a): 2,6-Lutidine (19 μ L, 0.17 mmol) and TBSOTf (38 μ L, 0.17 mmol) was added to a solution of **18a** (50.0 mg, 0.11 mmol) in CH₂Cl₂ (5 mL) at 0 °C. The resulting solution was stirred at 0 °C for 1 h. The reaction was terminated by addition of NaHNO₃ (5 mL) and extracted with CH₂Cl₂ (3 × 10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 7:1) offered the desired TBS-protected compound (51 mg, 82%) as a colorless liquid. IR (film): $\tilde{v} = 2958$, 1784, 1699, 1513, 1462, 1384, 1248, 1198 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 7.20$ (m, 5 H, *Ph*CH₂CHN–), 7.11 (d, *J* = 7.9 Hz, 2 H, –OCH₂*A*rOCH₃), 6.78 (d, $J = 8.6 \text{ Hz}, 2 \text{ H}, -\text{OCH}_2 Ar \text{OCH}_3), 4.54 \text{ (m, 1 H, PhCH}_2$ -CHN-), 4.44 (d, J = 11.0 Hz, 1 H, $-OCH_2ArOCH_3$), 4.30 (m, 2) H, $-OCH_2ArOCH_3$, $-CH_2CHOTBS$), 4.03 (d, J = 5.0 Hz, 2 H, PhCH₂CHCH₂O–), 3.69 (s, 3 H, $-OCH_2ArOCH_3$), 3.43 (dd, J =16.0, 9.2 Hz, 1 H, -CHCHOCH₂Ar), 3.18 (m, 2 H, PhC H_2 CHCH $_2$ O–), 2.67 (dd, J = 16.0, 2.6 Hz, 1 H, COC H_2 CH), 2.56 (dd, J = 13.3, 9.9 Hz, 1 H, COC H_2 CH), 1.85 (m, 1 H, TBSOCHCHCHO-), 1.56 (m, 2 H, -CH₂CH₃), 0.95 (d, J = 6.1 Hz, 3 H, -CH CH₃), 0.84 [m, 12 H, -CHCH₃, -Si(CH₃)₂C(CH₃)₃], -0.01 [d, J = 3.6 Hz, 6 H, $-Si(CH_3)_2C(CH_3)_3$] ppm. ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3)$: $\delta = 172.1, 158.9, 153.3, 135.4, 131.0, 129.4,$ 129.0, 128.9, 127.3, 113.7, 81.7, 71.2, 71.1, 65.9, 55.3, 55.2, 42.1, 38.4, 37.8, 25.7, 25.6, 23.8, 9.5, 9.0, -3.6, -4.6, -4.8 ppm. $[a]_{\rm D}^{28.1} =$ -62.44 (c = 1.27, CHCl₃). MS (EI): m/z (%) = 569 [M⁺], 374, 362, 301, 276, 252, 241, 218, 185, 121 (100), 91, 73, 59. HRMS: m/z calcd. for C₃₂H₄₇NO₆Si: 569.3173, found 569.3171.

To a solution of the TBS-protected compound (381 mg, 0.65 mmol) as described above in diethyl ether (10 mL) was added distilled water (21 µL, 1.21 mmol). After the solution was cooled to 0 °C, lithium borohydride (810 µL of a 2.0 M solution in THF, 1.62 mmol) was added slowly with stirring. After 10 min, the temperature of the solution was raised to room temperature, and the resulting solution was stirred for additional 1 h. The reaction was terminated with addition of aqueous NaOH solution (1.0 M, 15 mL) and extracted with diethyl ether $(3 \times 10 \text{ mL})$. After the organic layer was washed with saturated sodium chloride solution (15 mL), the ethereal solution was dried (MgSO₄) and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 7:1) provided the desired alcohol compound as a colorless liquid (250 mg, 94%). IR (film): $\tilde{v} = 3429$, 2955, 1612, 1514, 1461, 1249, 1060 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 7.15 (m. 2 H, ArH), 6.77 (m, 2 H, ArH), 4.42 (d, J = 11.1 Hz, 1 H, -CO-CH₂Ar), 4.26 (d, J = 11.1 Hz, 1 H, -COCH₂Ar), 3.76 (q, J = 5.8 Hz, 1 H, -CH₂CHCHCH₃), 3.70 (s, 3 H, -ArOCH₃), 3.63-3.57 (m, 2 H, HOCH₂-), 3.29 (m, 1 H, -HCCHCH₂), 1.91 (s, 1 H, -OH), 1.72 (m, 2 H, OHCH₂CH₂-), 1.68-1.42 (m, 3 H, $-CH_2CH_3$, $-CHCHCH_3$), 0.84 (d, J = 7.1 Hz, 3 H, $-CHCHCH_3$), 0.81 (t, J = 7.2 Hz, 3 H, $-CH_2CH_3$), 0.81 [s, 9 H, $-Si(CH_3)_2C$ - $(CH_3)_3$], 0.00 [d, J = 9.5 Hz, 6 H, Si $(CH_3)_2$ C $(CH_3)_3$] ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 159.7, 131.9, 129.9, 114.4, 80.6, 73.6, 71.7, 61.0, 56.0, 40.8, 36.3, 26.6, 24.7, 18.7, 11.3, 10.8, -3.7, -3.7 ppm. $[a]_{D}^{23.4} = -23.5$ (c = 1.75, CHCl₃). MS (EI): m/z (%) = 396 [M⁺], 260, 201, 189, 145, 137, 122 (100), 89, 75, 57. HRMS: m/z calcd. for C₂₂H₄₀O₄Si: 396.2696, found 396.2693.

The alcohol (250 mg, 0.63 mmol) obtained as described above was dissolved in DMF (5 mL). To this solution was added a solution of imidazole (61 mg, 0.90 mmol) and tert-butyldimethylsilyl chloride (118 mg, 0.78 mmol) in DMF (2 mL) at 0 °C under nitrogen. After it was stirred for 10 min at 0 °C, the solution was warmed to room temperature and stirred for additional 2 h at room temperature. After the reaction was completed, aqueous saturated NH₄Cl (10 mL) was added and the mixture was extracted with CH₂Cl₂ $(3 \times 20 \text{ mL})$. The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 20:1) offered the desired compound (305 mg, 95%) as a colorless oil. IR (film): v = 2954, 1614, 1587, 1513, 1462, 1388, 1360 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 7.30 (m, 2 H, ArH), 6.88 (m, 2 H, ArH), 4.53 (d, J = 10.9 Hz, 1 H, -COCH₂Ar), 4.26 $(d, J = 10.9 Hz, 1 H, -COCH_2Ar), 3.90 (m, 1 H,$ -CH₂CHCHCH₃), 3.81 (s, 3 H, -ArOCH₃), 3.66 (m, 2 H, TBSOC H_2 CH₂-), 3.45 (q, J = 5.7 Hz, 1 H, -HCCHCH₂-), 1.86-1.60 (m, 5 H, -CH₂CH₃, -CHCHCH₃ TBSOCH₂CH₂CH₂-), 0.98 (d, J = 6.9 Hz, 3 H, $-CHCHCH_3$), 0.95 (t, J = 7.3 Hz, 3 H,

FULL PAPER

-CH₂CH₃), 0.94 [s, 9 H, -Si(CH₃)₂C(CH₃)₃], 0.93 [s, 9 H, -Si-(CH₃)₂C(CH₃)₃], 0.07 [s, 6 H, -Si(CH₃)₂C(CH₃)₃], 0.06 [s, 6 H, -Si(CH₃)₂C(CH₃)₃] ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 159.0, 131.3, 129.2, 129.0, 128.9, 113.6, 80.3, 71.2, 70.5, 60.0, 55.1, 40.6, 37.5, 25.9, 25.9, 25.7, 24.0, 18.2, 18.1, 10.1, 9.6, -3.0, -4.2, -4.4, -5.3 ppm. [a]₂^{25.9} = -21.85 (*c* = 1.33, CHCl₃). MS (EI): *m/z* (%) = 510 [M⁺], 331, 321, 303, 242, 218, 185, 173, 147, 133, 121 (100), 101, 89, 73, 57. HRMS: *m/z* calcd. for C₂₈H₅₄O₄Si₂: 510.3561, found 510.3578.

The compound (305 mg, 0.60 mmol) obtained was dissolved in a solution of CH₂Cl₂ and pH 7 buffer solution (10:1)(11 mL). To this solution dichlorodicyanoquinone (DDQ) (198 mg, 0.87 mmol) was added at 0 °C. The resulting solution was stirred at 0 °C for 1 h. After filtration through the pad of Celite with CH_2Cl_2 (3 × 10 mL), the solution was concentrated. Purification by flash chromatography (hexane/EtOAc, 9:1) gave the desired alcohol 19a (173 mg, 74%) as a colorless liquid. IR (film): $\tilde{v} = 3438, 2955, 1471, 1389,$ 1360, 1256 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 4.00 (td, J = 6.0, 3.0 Hz, 1 H, CHOH), 3.64 (m, 2 H, CH₂OSi) ppm. 3.58 (td, 1 H, J = 5.9, 3.0 Hz, CHOSi), 2.66 (br., 1 H, CHOH), 1.72 (q, 2 H, J = 6.3 Hz, CH_2CH_2OSi), 1.59 (m, 1 H, $CHCH_3$), 1.38–1.52 (m, 2 H, CH_2CH_3), 0.90 (t, 3 H, J = 7.4 Hz, $CHCH_3$), 0.86 [s, 21 H, SiC(CH₃)₃, SiC(CH₃)₃, CHCH₃], 0.06 [s, 6 H, Si(CH₃)₂], 0.00 [s, 6 H, Si(CH₃)₂]. ¹³C NMR (75 MHz, CDCl₃): δ = 76.9, 75.4, 60.7, 40.2, 38.0, 28.6, 26.5, 18.8, 18.6, 11.1, 6.93, -3.20, -3.97, -4.82 ppm. $[a]_{D}^{25.5} = -12.94$ (c = 1.48, CHCl₃). MS (EI): m/z (%) = 390 [M⁺], 333, 303, 263, 241, 201, 189, 171, 147, 133, 109, 89, 73, 59. HRMS: m/z calcd. for C₂₀H₄₆O₃Si₂: 390.2986, found 390.2987.

(1R,2S,3S)-3-(tert-Butyldimethylsilyloxy)-1-ethyl-2-methyl-5-oxopentyl 2-Bromopropanoate (20a): 2-bromopropionic acid (111 µL, 1.23 mmol) and DMAP (40 mg, 0.33 mmol) was added to a solution of 19a (160 mg, 0.41 mmol) in CH₂Cl₂ (5 mL) at room temperature. After the solution was cooled to 0 °C, N,N-dicyclohexylcarbodiimide (93 mg, 0.45 mmol) was added. The resulting solution was stirred for 5 min at 0 °C before it was warmed to room temperature. After additional stirring for 30 min at room temperature, the precipitate was filtered. The filtrate was concentrated and purified by flash chromatography (hexane/EtOAc, 10:1). The desired bromoester was obtained as a colorless liquid (210 mg, 97%). IR (film): $\tilde{v} = 2928$, 2857, 1737, 1471, 1387, 1256, 1222 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 5.04 [m, 1 H, CHOC(=O)], 4.33 (m, 1 H, BrCHCH₃), 3.82 (m, 1 H, CHOSi), 3.60 (m, 2 H, CH₂OSi), 1.84 (m, 1 H, CHCH₃), 1.79 (d, J = 6.9 Hz, 3 H, BrCHCH₃), 1.62 (m, 4 H, CH₂CH₃, CH₂CHOSi), 0.86–0.91 [m, 24 H, CHCH₃, CH₂CH₃, SiC(CH₃)₃, SiC(CH₃)₃], 0.03 (s, 3 H, SiCH₃), 0.01 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 170.0, 78.3, 70.6, 59.9, 41.3, 41.1, 26.2, 25.9, 22.1, 18.5, 10.6, 10.0, -4.02, -4.99 ppm. $[a]_{D}^{24.3} = -7.13$ (c = 1.73, CHCl₃). MS (EI): m/z (%) = 524 [M⁺], 337, 315, 303, 287, 253, 241, 211, 189, 171, 145, 109, 89, 75 (100), 59. HRMS: m/z calcd. for C₂₃H₄₉BrO₄Si₂: 524.2353, found 524.2334.

The bromoester (210 mg, 0.40 mmol) obtained was dissolved in MeOH (2 mL). To this solution DL-10-Camphorsulfonic acid (19 mg, 0.08 mmol) was added. The resulting solution was stirred at 0 °C for 1 h. The reaction was terminated by addition of Et₃N (56 μ L, 0.42 mmol). After the solution was concentrated, purification of the residue by flash chromatography (hexane/EtOAc, 5:1) gave the desired primary alcohol (145 mg, 88%) as a colorless liquid. IR (film): $\tilde{v} = 3431$, 2928, 1737, 1257, 1163, 1060 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 5.02$ [m, 1 H, CHOC(=O)], 4.36 (m, 1 H, BrCHCH₃), 3.82 (m, 1 H, CHOSi), 3.73 (m, 2 H, CH₂OH), 1.93 (m, 2 H, CH₂CH₂CHOSi), 1.82 (dd, J = 7.0, 1.9 Hz, 3 H,

BrCHC*H*₃), 1.65 (m, 3 H, C*H*₂C*H*₃, C*H*C*H*₃), 0.98 (d, *J* = 6.9 Hz, 3 H, CHC*H*₃), 0.90 [m, 12 H, C*H*₂C*H*₃, SiC(C*H*₃)₃], 0.09 (s, 3 H, SiC*H*₃), 0.07 (s, 3 H, SiC*H*₃) ppm. ¹³C NMR (75 MHz, CDC*I*₃): δ = 170.4, 72.3, 60.1, 41.3, 41.1, 36.2, 26.5, 26.2, 22.2, 18.6, 10.9, 10.5, -3.75, -3.85 ppm. [*a*]_D^{24.4} = -0.80 (*c* = 1.08, CHC*I*₃). MS (EI): *m/z* (%) = 410 [M⁺], 213, 201, 189, 173, 145, 127 (100), 109, 89, 75, 55. HRMS: *m/z* calcd. for C₁₇H₃₅BrO₄Si: 410.1488, found 410.1500.

The alcohol (145 mg, 0.35 mmol) obtained above was dissolved in CH₂Cl₂ (2 mL). To this solution was added Dess-Martin periodinane (DMP) (179 mg, 0.42 mmol) and stirred for 30 min at room temperature. After the reaction was completed, aqueous saturated NaHCO₃ (5 mL) was added and mixture was extracted with CH_2Cl_2 (3 × 10 mL). The organic layer was separated, dried (MgSO₄), and concentrated. Purification by flash chromatography (hexane/EtOAc, 7:1) offered the desired aldehyde 20a (131 mg, 91%) as a yellow liquid. ¹H NMR (300 MHz, CDCl₃): δ = 9.77 [m, 1 H, C(=O)H], 5.02 (m, 1 H, CHCH₂CH₃), 4.36 [m, 1 H, C(=O) CHBr], 4.15 (m, 1 H, CHOSi), 2.59 (m, 2 H, CH₂CHOSi), 1.86 (m, 1 H, CHCH₃), 1.80 (dd, J = 6.9, 1.3 Hz, 3 H, BrCHCH₃), 1.64 (m, 2 H, CH_2CH_3), 0.95 (d, J = 7.0 Hz, 3 H, BrCHCH₃), 0.85– 0.89 [m, 12 H, CH₂CH₃, SiC(CH₃)₃], 0.06 (s, 3 H, SiCH₃), 0.02 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 202, 170, 77.1, 69.7, 48.8, 42.5, 41.0, 26.4, 26.2, 22.2, 18.6, 10.9, 10.5, -3.82, -3.92 ppm.

(6S,7S,8R)-6-(tert-Butyldimethylsilyloxy)-8-ethyl-4-hydroxy-3,7-dimethyl-1-oxacyclooctan-2-one (21a): Diiodomethane (51 µL, 0.64 mmol) was added to a mixture of samarium (powder, 105 mg, 0.70 mmol) in THF (3 mL) at room temperature under nitrogen. Samarium(II) iodide was obtained as a blue-colored solution which was used after stirring for 2 h. After the solution was cooled to -78 °C, a solution of the aldehyde 20a (130 mg, 0.32 mmol) in THF (2 mL) was added under nitrogen. After the solution was stirred for 1 h at -78 °C, it was warmed to room temperature and 0.1 M HCl (5 mL) was added. The mixture was extracted with diethyl ether $(3 \times 20 \text{ mL})$. After the organic layer was washed with saturated sodium thiosulfate solution (15 mL) and saturated sodium chloride solution (15 mL), the ethereal solution was dried (MgSO₄) and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 3:1) provided the desired lactone 21a as a yellow liquid [a 7:1 mixture of diastereomers; major compound: $R_{\rm f}$ = 0.24 (hexane/EtOAc, 1:1; 66 mg, 63%), minor compound: $R_{\rm f}$ = 0.15 (hexane/EtOAc, 1:1; 9.4 mg, 9.0%)]. The following data were obtained from the major compound. ¹H NMR (500 MHz, CDCl₃): δ = 4.87 [dd, J = 8.4, 5.3 Hz, 1 H, CHOC(=O)], 4.04 (dd, J = 9.1, 4.7 Hz, 1 H, CHOSi), 3.57 (m, 1 H, CHOH), 3.44 (br., 1 H, CHO*H*), 3.15 [dq, *J* = 9.4, 6.3 Hz, 1 H, C(=O)C*H*CH₃], 2.15 (ddd, J = 15.9, 9.2, 3.5 Hz, 1 H, SiOCHCHCH₃), 1.89 (m, 1 H, one of CH₂CH₃), 1.84 (m, 2 H, CH₂CHOH), 1.49 (m, 1 H, one of CH_2CH_3 , 1.34 [d, J = 6.3 Hz, 3 H, $C(=O)CHCH_3$], 0.96 (m, 3 H, SiOCHCHCH₃), 0.92 [s, 9 H, SiC(CH₃)₃], 0.89 (m, 3 H, CH₂CH₃), 0.12 (s, 3 H, SiCH₃), 0.10 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, $CDCl_3$): $\delta = 176.0, 78.8, 76.9, 76.0, 44.5, 42.6, 27.4, 26.4, 18.5,$ 16.1, 11.5, 10.8, -4.1, -4.3 ppm. MS (EI): m/z (%) = 330 [M⁺], 287, 257, 213, 201, 173, 159, 139, 115 (100), 99, 85, 75, 57. HRMS: m/z calcd. for C17H34O4Si: 330.2226, found 330.2244.

(6S,7S,8R)-6-(*tert*-Butyldimethylsilyloxy)-8-ethyl-3,7-dimethyl-1-oxacyclooctane-2,4-dione (22a): Lactone 21a (55 mg, 0.17 mmol) obtained as described in the previous procedure (a 7:1 mixture of diastereomers) was dissolved in CH₂Cl₂ (2 mL). To this solution was added Dess–Martin periodinane (DMP) (141 mg, 0.33 mmol). The resulting solution was stirred for 30 min at room temperature. After the reaction was completed, aqueous saturated NaHCO₃ (5 mL) was added and mixture was extracted with CH₂Cl₂ $(3 \times 10 \text{ mL})$. The organic layer was separated, dried (MgSO₄), and concentrated. Purification of the residue by flash chromatography (hexane/EtOAc, 7:1) offered the desired compound 22a (48 mg, 88%) as a yellow liquid (an 8:1 mixture of diastereomers). The major compound showed the following spectroscopic behavior. ¹H NMR (300 MHz, CDCl₃): δ = 5.04 [ddd, J = 8.6, 4.6, 4.4 Hz, 1 H, CHOC(=O)], 4.05 (m, 1 H, CHOSi), 3.49 [q, J = 6.7 Hz, 1 H, C(O)- $CHCH_3$], 3.23 [d, J = 11.8 Hz, 1 H, one of $CH_2C(=O)$], 2.71 [t, J $= 11.2 \text{ Hz}, 1 \text{ H}, \text{ one of } CH_2C(=O)$], 1.89 (m, 1 H, SiOCHCHCH₃), 1.55–1.72 (m, 2 H, CH_2CH_3), 1.19 [d, J = 6.7 Hz, 3 H, C(=O)- $CHCH_3$], 1.02 (t, J = 7.4 Hz, 3 H, CH_2CH_3), 0.94 (d, J = 4.9 Hz, 3 H, SiOCCHCH₃), 0.81 [s, 9 H, SiC(CH₃)₃], 0.03 (s, 3 H, SiCH₃), 0.00 (s, 3 H, SiCH₃) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 202, 172. 80.5, 71.7, 57.8, 45.5, 42.4, 26.2, 25.1, 18.6, 11.5, 11.1, 10.1, -4.58, -4.64 ppm.

(6*S*)-4-Hydroxy-6-[(1*S*,2*R*)-2-hydroxy-1-methylbutyl]-3-methyl-5,6dihydro-2*H*-pyran-2-one (2a): A solution of lactone 22a (41 mg, 0.12 mmol) as obtained previously (an 8:1 mixture of diastereomers) in HF/CH₃CN [48% HF/CH₃CN, 1:19 (v/v)] (1 mL) was stirred for 1 h at room temperature. The reaction was terminated by addition of solid CaCO₃ (30 mg). After the mixture was filtered with aid of Celite and washed by ether (3×10 mL), the filtrate was concentrated. Purification by flash chromatography (hexane/EtOAc, 1:1 to 1:2) provided the desired lactone 2a (15.5 mg, 58%) as a white solid. Spectroscopic data were matched with those obtained previously.

Supporting Information (see also the footnote on the first page of this article): Experimental procedure for compounds 15b–17b, 23a, 19b–26b, 28–34.

Acknowledgments

This work is supported by the Korea Research Foundation Grant (KRF-2004-015-C00267).

- [1] D. A. Hopwood, Chem. Rev. 1997, 94, 2465–2497.
- [2] J. Staunton, B. Wilkinson, Chem. Rev. 1997, 97, 2611-2629.
- [3] Y. Xue, D. H. Sherman, Nature 2000, 403, 571-574.
- Y. J. Yoon, B. J. Beck, B. S. Kim, H.-Y. Kang, K. A. Reynolds, D. H. Sherman, *Chem. Biol.* 2002, *9*, 203–214.
- [5] C. M. Kao, G. Luo, L. Katz, D. E. Cane, C. Khosla, J. Am. Chem. Soc. 1995, 117, 9105–9106.
- [6] J. Cortés, K. E. H. Wiesmann, G. A. Roberts, M. J. B. Brown, J. Staunton, P. F. Leadlay, *Science* **1995**, *268*, 1487–1489.
- [7] a) C. M. Kao, G. Luo, L. Katz, D. E. Cane, C. Khosla, J. Am. Chem. Soc. 1994, 116, 11612–11613; M. J. B. Brown, J. Cortés,

A. L. Cutter, P. F. Leadlay, J. Chem. Soc., Chem. Commun.
1995, 1517–1518; b) K. E. H. Wiesmann, J. Cortés, M. J. B.
Brown, A. L. Cutter, J. Staunton, P. F. Leadlay, Chem. Biol.
1995, 2, 583–589; c) R. Piper, S. Ebert-Khosla, D. E. Cane, C.
Khosla, Biochemistry 1996, 35, 2054–2060; d) G. Luo, R.
Pieper, A. Rosa, C. Khosla, D. E. Cane, Bioorg. Med. Chem.
1996, 4, 995–999; e) . A. L. Wilkinson, U. Hanefeld, B. Wilkinson, P. F. Leadlay, J. Staunton, Tetrahedron Lett. 1998, 39, 9827–9830.

- [8] B. J. Beck, Y. J. Yoon, K. A. Reynolds, D. H. Sherman, *Chem. Biol.* 2002, 9, 575–583.
- [9] S.-J. Kim, H.-Y. Kang, D. H. Sherman, Synthesis 2001, 1790– 1793.
- [10] H.-H. Yang, E.-S. Kim, Y. J. Yoon, H.-Y. Kang, Bull. Korean Chem. Soc. 2006, 27, 473–474.
- [11] B. J. Beck, C. C. Aldrich, R. A. Fecik, K. A. Reynolds, D. H. Sherman, J. Am. Chem. Soc. 2003, 125, 12551–12557.
- [12] a) G. A. Molander, J. B. Etter, L. S. Harring, P. Thorel, J. Am. Chem. Soc. 1991, 113, 8036-8045; b) H. B. Kagan, J. L. Namy, P. Girard, Tetrahedron 1981, 37, 175-180; c) T. Mukaiyama, I. Shina, H. Iwadare, M. Saitoh, T. Nishimura, N. Ohkawa, H. Satoh, K. Nishimura, Y. Tani, M. Hasegawa, K. Yamada, K. Saitoh, Chem. Eur. J. 1999, 5, 121-161; d) S. Inoue, Y. Iwabuchi, H. Irie, S. Hatakeyama, Synlett 1998, 735-736; e) T. Takemura, Y. Nishii, S. Takahashi, J. Kobayashi, T. Nakata, Tetrahedron 2002, 58, 6359-6365; f) P. P. Reddy, K. F. Yen, B. J. Uang, J. Org. Chem. 2002, 67, 1034-1035; g) M. Inoue, M. Sasaki, K. Tachibana, J. Org. Chem. 1999, 64, 9416-9429; h) M. Inoue, M. Sasaki, K. Tachibana, Tetrahedron 1999, 55, 10949-10970; i) M. Inoue, M. Sasaki, K. Tachibana, Tetrahedron Lett. 1997, 38, 1611-1614; j) S. Ichikawa, S. Shuto, N. Minakawa, A. Matsuda, J. Org. Chem. 1997, 62, 1368-1375; k) M. Inoue, M. Sasaki, K. Tachibana, Angew. Chem. Int. Ed. Engl. 1998, 37, 965-969; 1) M. Inoue, M. Tachibana, K. Sasaki, J. Org. Chem. 1999, 64, 9416-9429.
- [13] a) S.-i. Fukuzawa, H. Matsuzawa, S.-i. Yoshimitsu, J. Org. Chem. 2000, 65, 1702–1706; b) S.-i. Fukuzawa, M. Tatsuzawa, K. Hirano, Tetrahedron Lett. 1998, 39, 6899–6900.
- [14] We first tried the protection of the free hydroxy group of 14a. However, we were not able to protect the free secondary hydroxy group with the PMB group. Therefore, we have decided to use the aldehyde 13b for the asymmetric Reformatsky reaction to prepare 3.
- [15] For reported examples for translactonization by deprotection of the silyl groups, see: a) P. T. O'Sullivan, W. Buhr, M. A. M. Fuhry, J. R. Harrison, J. E. Davies, N. Feeder, D. R. Marshall, J. W. Burton, A. B. Holmes, J. Am. Chem. Soc. 2004, 126, 2194–2207; b) M. S. Congreve, A. B. Holmes, A. B. Hughes, M. G. Looney, J. Am. Chem. Soc. 1993, 115, 5815–5816.
- [16] W. C. Still, M. Kahn, A. Mitra, J. Org. Chem. 1978, 43, 2923– 2925.

Received: March 22, 2007 Published Online: May 11, 2007