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## Interaction of chiral MS-245 analogs at h5-HT<sub>6</sub> receptors

Carmen Abate,<sup>a</sup> Renata Kolanos,<sup>a</sup> Malgorzata Dukat,<sup>a</sup> Vince Setola,<sup>b</sup> Bryan L. Roth<sup>b,c</sup> and Richard A. Glennon<sup>a,\*</sup>

<sup>a</sup>Department of Medicinal Chemistry, School of Pharmacy, Virginia Commonwealth University, Richmond, VA 23298-0540, USA <sup>b</sup>Department of Biochemistry, Case Western Reserve University, USA

<sup>c</sup>Department of Psychiatry and Neurosciences, School of Medicine, Case Western Reserve University, USA

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**Abstract**—Optically active pyrrolidinylmethylindole analogs related in structure to the benzenesulfonyltryptamine 5-HT<sub>6</sub> receptor antagonist MS-245 were evaluated and their *R*-isomers were found to bind with affinity higher than their *S*-enantiomers. © 2005 Elsevier Ltd. All rights reserved.

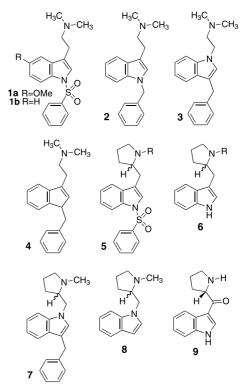
5-HT<sub>6</sub> receptors are members of the serotonin receptor family (5-HT<sub>1</sub>-5-HT<sub>7</sub>) and are of potential therapeutic interest due to their possible involvement in neuropsychiatric disorders, such as depression and pyschosis.<sup>1–6</sup> This receptor population was first identified about 10 years ago, but only in the past few years have 5-HT<sub>6</sub> receptor antagonists been described (reviewed<sup>5,6</sup>). We identified MS-245 (1a;  $K_i$  ca. 2 nM) as one of the first examples of a 5-HT<sub>6</sub> receptor antagonist.<sup>7,8</sup> Subsequent pharmacophoric studies have revealed that the 5-methoxy group of **1a** is not required for binding (i.e., **1b**;  $K_i = 4$  nM), that the benzenesulfonyl moiety can be replaced by a benzyl group (2;  $K_i = 6 \text{ nM}$ ) with retention of antagonist action,<sup>5</sup> and that the indole  $N_1$  nitrogen atom is not required for binding, as evidenced by the high affinity of, for example, isotryptamine 3 ( $K_i = 32 \text{ nM}$ ) and indene 4 ( $K_i = 3 \text{ nM}$ ) for this receptor population.

Various tryptamines lacking an  $N_1$  substituent generally bind with reduced affinity and, like 5-hydroxytryptamine (i.e., serotonin;  $K_i$  ca. 100 nM), can display agonist character.<sup>5</sup> On the basis of comparative structure–affinity and receptor modeling studies,<sup>8,10</sup> we have proposed that tryptamine analogs, although likely utilizing a common aspartate moiety, bind at 5-HT<sub>6</sub> receptors differently depending upon whether or not they possess an  $N_1$ (e.g., a benzenesulfonyl or benzyl) substituent.

If the presence or absence of an  $N_1$  substituent dictates how tryptamine analogs interact with 5-HT<sub>6</sub> receptors,

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it was reasoned that chiral analogs of  $N_1$ -unsubstituted tryptamines might bind differently than  $N_1$ -substituted tryptamines and that isomeric comparisons could prove to be informative. In an extreme case, optical isomers of  $N_1$ -substituted tryptamines might even display opposite enantioselectivity for binding, as compared to their corresponding  $N_1$ -unsubstituted counterparts.



Keywords: Serotonin; Pyrrolidinylmethylindoles.

<sup>\*</sup> Corresponding author. Tel.: +1 804 828 8487; fax: +1 804 828 7404; e-mail: glennon@hsc.vcu.edu

Introduction of substituents  $\alpha$  to the terminal amine of tryptamine analogs (e.g., an  $\alpha$ -methyl or  $\alpha$ -ethyl group) to create a chiral center has been shown to result in somewhat reduced 5-HT<sub>6</sub> receptor affinity.<sup>11</sup> Nevertheless, it was thought that reduced affinity might be an acceptable trade-off if it allowed enantiomeric potency comparisons to be made. Rather than examining isomers of  $\alpha$ -methyl- or  $\alpha$ -ethyltryptamines, however, we opted to examine their cognate pyrrolidinylmethylindole counterparts. That is, because necessary optically active starting materials with defined absolute configuration were readily available, and because pyrrolidinylmethylindoles have been previously shown to bind at other (i.e., 5-HT<sub>1</sub>) serotonin receptor populations<sup>12,13</sup>, we prepared the optical isomers of 5 (where R = H or Me) for binding comparisons with their N1-unsubstituted analogs 6. To evaluate this hypothesis further, we also prepared the isomers of isotryptamine 7 for comparison with their  $(C_3)$  unsubstituted counterparts 8.

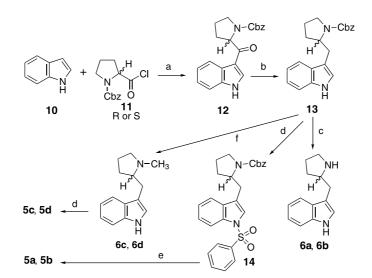
Compounds **6a** and **6b** were previously reported in the patent literature as 5-HT<sub>1</sub> ligands;<sup>12</sup> and the synthesis of their 5-methoxy analogs has also been described by Macor and co-workers.<sup>13,14</sup> Compounds **6a** and **6b** were prepared using the same methodology (Scheme 1). Ketones **12** were obtained by the reaction of indole (**10**) with either the commercially available *R*- or *S*-isomer of **11**; isomers **12** were reduced with LiBH<sub>4</sub> to protected amine **13**, which was deprotected via catalytic hydrogenation to the desired targets **6a** and **6b** (Table 1). In one instance, (-)**12** was subjected to direct catalytic reduction and resulted in ketone derivative S(-)9.<sup>15</sup>

Both isomers of 13 were treated with sodium bis(trimethylsilyl)amide to form the corresponding anions that were allowed to react with benzenesulfonyl chloride to afford intermediates 14; deprotection of 14 was performed, as described for 13, to afford isomers 5a and 5b (Scheme 1). Direct treatment of 13 with lithium aluminum hydride led to 6c and 6d, and these were subsequently converted to their corresponding benzenesulfonyl analogs **5c** and **5d**.

The isomers of 7 and 8 (Table 1) were conveniently synthesized by reaction of the indolyl anion (generated by treatment with NaH in DMF) of indole or 3-benzylindole<sup>16</sup>, with the known optical isomers of 2-(chloromethyl)-1-methylpyrrolidine hydrochloride.<sup>17</sup>

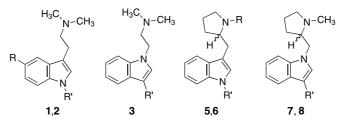
Radioligand binding<sup>18</sup> data for the target compounds are shown in Table 1. As with structurally simpler N,N-dialkyltryptamines,<sup>5</sup> introduction of an N<sub>1</sub>-benzenesulfonyl moiety led to enhanced affinity (comparing pyrrolidinylmethylindoles **5a** with **6a**, **5b** with **6b**, **5c** with **6c**, and **5d** with **6d**) (Table 1). Likewise, in the isotryptamine series, benzyl analogs **7a** and **7b** displayed affinity higher than those of their parents, **8a** and **8b**, respectively. In fact, the pyrrolidinylmethylindoles bind with affinities higher than those of their simpler N,N-dimethyltryptamine counterparts; for example, compound **5d** binds with more than 10 times the affinity of **1b** and **7b** binds with 3 times the affinity of **3**.

In the absence of enantioselectivity reversal, it is not possible to conclude that the N<sub>1</sub>-substituted and N<sub>1</sub>unsubstituted compounds are binding in a different fashion. Nevertheless, there is some tantalizing evidence in support of this possibility. For example, there is greater enantioselectivity in the absence of the benzenesulfonyl group (40- and 70-fold for **6a/6b** and **6c/6d**, respectively) than in its presence (<6-fold for 5a/5b and 5c/5d). A similar, but somewhat less pronounced, trend is seen with the isotryptamines. Furthermore, N-methylation has dissimilar effects in the unsubstituted and substituted series; whereas N-methylation enhanced the affinity of **6a**  $(\rightarrow 6c)$  and **6b**  $(\rightarrow 6d)$  by about 5-fold, methylation enhanced the affinity of 5a ( $\rightarrow$ 5c) and 5b ( $\rightarrow$ 5d) by 25-fold. Clearly, there are differences between the N<sub>1</sub>-unsubstituted and N<sub>1</sub>-substituted series.



Scheme 1. (a) EtMgBr; (b) LiBH<sub>4</sub>,  $\Delta$ ; (c) H<sub>2</sub>, 10% Pd/C, absolute EtOH, 35–40 psi/3 h; (d) i. [(CH<sub>3</sub>)<sub>3</sub>Si]<sub>2</sub>NNa, THF, -78 °C, ii. PhSO<sub>2</sub>Cl, rt; (e) H<sub>2</sub>, 10% Pd/C, absolute EtOH/MeOH, 35 psi/12 h; and (f) LiAlH<sub>4</sub>, THF,  $\Delta/4$  h.

Table 1. Radioligand binding data for reference compounds 1-3, and physicochemical properties and 5-HT<sub>6</sub> receptor affinities of the target compounds



	R	R′	Isomer	Melting point (°C)	Empirical formula <sup>a</sup>	h5-HT <sub>6</sub> $K_i$ , nM (±SEM) <sup>b</sup>
1a	OMe	SO <sub>2</sub> Ph	_	_		2
1b	Н	$SO_2Ph$	_			4
2	Н	CH <sub>2</sub> Ph			_	6
3		CH <sub>2</sub> Ph				32
6a	Н	Н	S(+)	203–205 <sup>c</sup>	$C_{13}H_{16}N_2$ HCl	2400 (±400)
6b	Н	Н	R(-)	204–205 <sup>°</sup>	$C_{13}H_{16}N_2$ HCl	60 (±10)
5a	Н	$SO_2Ph$	S(+)	182–184 <sup>c</sup>	C <sub>19</sub> H <sub>20</sub> N <sub>2</sub> SO <sub>2</sub> C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	46 (±6)
5b	Н	$SO_2Ph$	R(-)	179–181 <sup>°</sup>	$C_{19}H_{20}N_2SO_2 C_2H_2O_4^{d}$	7.8 (±1.4)
6c	Me	Н	S(-)	216–218 <sup>c</sup>	$C_{14}H_{18}N_2$ HCl	640 (±110)
6d	Me	Н	R(+)	216–217 <sup>°</sup>	$C_{14}H_{18}N_2$ HCl	9.1 (±0.8)
5c	Me	$SO_2Ph$	S(-)	145–147 <sup>°</sup>	C <sub>20</sub> H <sub>22</sub> N <sub>2</sub> SO <sub>2</sub> C <sub>2</sub> H <sub>2</sub> O <sub>4</sub>	1.7 (±0.3)
5d	Me	$SO_2Ph$	R(+)	145–147 <sup>°</sup>	$C_{20}H_{22}N_2SO_2 C_2H_2O_4{}^d$	0.3 (±0.1)
8a		Н	S(+)	157–158 <sup>e</sup>	$C_{14}H_{18}N_2$ HCl	8750 (±2200)
8b		Н	R(-)	157–158 <sup>e</sup>	$C_{14}H_{18}N_2$ HCl	550 (±110)
7a		CH <sub>2</sub> Ph	S(+)	124–125 <sup>e</sup>	C <sub>21</sub> H <sub>24</sub> N C <sub>4</sub> H <sub>4</sub> O <sub>4</sub>	29 (±5)
7b		CH <sub>2</sub> Ph	R(-)	124–125 <sup>e</sup>	C <sub>21</sub> H <sub>24</sub> N C <sub>4</sub> H <sub>4</sub> O <sub>4</sub>	9.9 (±1.3)

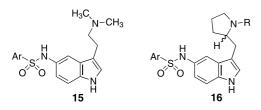
<sup>a</sup> All compounds analyzed within 0.4% of theory for C, H, and N.  $C_2H_2O_4$ , oxalate salt;  $C_4H_4O_4$ , maleate salt. <sup>b</sup>  $K_i$  values were determined, at least in triplicate, as previously described.<sup>18,22</sup> Binding data for **1–3** have been previously reported<sup>5,8,9</sup> and are shown here for comparison.

<sup>c</sup> Recrystallization solvent = anhydrous MeOH/absolute EtOH.

<sup>d</sup> Crystallized with 0.25 mol H<sub>2</sub>O.

<sup>e</sup> Recrystallization solvent =  $CH_2Cl_2/Et_2O$ .

It was shown that R-pyrrolidinylmethylindoles bind with higher affinity than their S-enantiomers and that **5d** ( $K_i = 0.3 \text{ nM}$ ) binds with nearly 10-fold higher affinity than MS-245 (1). Interestingly, a recent review of the patent literature revealed that 5d had been previously described as a 5-HT<sub>6</sub> antagonist; although the compound was not physically characterized and though no enantiomeric potency comparison was provided (because its S-enantiomer was not prepared), 5d was reported to bind at 5-HT<sub>6</sub> receptors with a  $K_{\rm i} < 10 \ {\rm nM}^{-19}$ 



Although the present results cannot be used as evidence for different modes of binding, they are not inconsistent with the concept that N<sub>1</sub>-substituted tryptamine-related analogs might bind differently than their N<sub>1</sub>-unsubstituted counterparts. The results further identify R-pyrrolidinylmethylindole analogs of MS-245 (1a) as binding with affinity higher than their S-enantiomers. While this manuscript was under preparation, Holenz et al.<sup>20</sup> showed that 5-arylsulfonamido analogs of tryptamines (e.g., 15) bind at 5-HT<sub>6</sub> receptors, and that some are 5-HT<sub>6</sub> antagonists, whereas others are agonists. At this time, however, it is not known how 15 binds relative to the compounds described herein. Nevertheless, Cole et al.,21 coincidentally, reported on the pyrrolidinylmethylindole counterparts of such agents (i.e., 16). In the latter investigation, there was a trend for the R-isomers of 16 (where R = Me) to bind with about 4-fold higher affinity, relative to the corresponding NH analogs, and even more fascinating is that the S-enantiomers generally behaved as  $5-HT_6$  antagonists, whereas the R-isomers displayed agonist action.<sup>21</sup>

## Acknowledgment

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## **References and notes**

- 1. Hoyer, D.; Hannon, J. P.; Martin, G. R. Pharmacol. Biochem. Behav. 2002, 71, 533.
- Humphrey, P. P. A.; Hartig, P. R.; Hoyer, D. Trends 2. Pharmacol. Sci. 1993, 14, 233.

- 3. Kroeze, W. K.; Kristiansen, K.; Roth, B. L. Curr. Top. Med. Chem. 2002, 2, 507.
- Meltzer, H. Y.; Li, Z.; Kaneda, Y.; Ichikawa, J. Prog. Neuropsychopharmacol. Biol. Psychiatry 2003, 27, 1159.
- 5. Glennon, R. A. J. Med. Chem 2003, 46, 2795.
- Woolley, M. L.; Marsden, C. A.; Fone, K. C. F. Curr. Drug Top. 2004, 3, 59.
- Glennon, R. A.; Lee, M.; Rangisetty, J. B.; Dukat, M.; Roth, B. L.; Savage, J. E.; McBride, A.; Rauser, L.; Hufesien, L.; Lee, D. K. H. J. Med. Chem. 2000, 43, 1011.
- Tsai, Y.; Dukat, M.; Slassi, A.; MacLean, N.; Demchyshyn, L.; Savage, J. E.; Roth, B. L.; Hufesein, S.; Lee, M.; Glennon, R. A. *Bioorg. Med. Chem. Lett.* 2000, 10, 2295.
- Kolanos, R.; Siripurapu, U.; Pullagurla, M.; Riaz, M.; Setola, V.; Roth, B. L.; Dukat, M.; Glennon, R. A. *Bioorg. Med. Chem. Lett.* 2005, 15, 1987.
- Pullagurla, M.; Westkaemper, R. B.; Glennon, R. A. Bioorg. Med. Chem. Lett. 2004, 14, 4569.
- Glennon, R. A.; Bondarev, M.; Roth, B. L. Med. Chem. Res. 1999, 9, 108.
- 12. Macor, J. E.; Whytes, M. J. U. S. Patent 5,607,951, 1997.
- Macor, J. E.; Blake, J.; Fox, C. B.; Jonhson, C.; Koe, B. K.; Lebel, L. A.; Morrone, J. M.; Ryan, K.; Schmidt, A. W.; Schulz, D. W.; Zorn, S. H. *J. Med. Chem.* **1992**, *35*, 4503.
- Macor, J. E.; Chenard, B. L.; Ronald, J. P. J. Org. Chem. 1994, 59, 7496.
- 15. Compound S(-)9 was characterized as its hydrogen oxalate salt, mp 205–206 °C, following recrystallization from methanol and analyzed within 0.4% of theory for  $C_{13}H_{14}N_2O \cdot C_2H_2O_4$ . S(-)9 was found to bind  $(K_i = 2700 \pm 600 \text{ nM})$  with an affinity similar to that of **6a**.
- Swaminathan, S.; Ranganathan, S.; Sulochana, S. J. Org. Chem. 1958, 23, 707.
- 17. Floyd, D. M. J. Med. Chem. 1992, 35, 756.

- 18. The h5-HT<sub>6</sub> radioligand binding assay was performed, as previously described.<sup>22</sup> In brief, h5-HT<sub>6</sub> cDNA was transiently expressed in HEK-293 cells using Fugene6 according to the manufacturer's recommendations; 24 h after transfection, the medium was replaced, and 24 h later, medium containing dialyzed serum (to remove 5-HT) was added. At 72 h after transfection, cells were harvested by scraping and centrifugation. Cells were then washed by centrifugation and resuspension in phosphatebuffered saline (pH 7.40; PBS), and frozen as tight pellets at -80 °C until use. Binding assays were performed at room temperature for 90 min in binding buffer (50 mM Tris-Cl, 10 mM MgCl<sub>2</sub>, and 0.1 mM EDTA, pH 7.40) with  $[^{3}H]LSD$  (1 nM final concentration) using 10  $\mu$ M clozapine for nonspecific binding. Concentrations of unlabeled test agent (1-10,000 nM) were used for  $K_i$ determinations with  $K_i$  values calculated using the program GraphPad Prism (V4.0). Specific binding represented 80–90% of total binding.  $K_i$  values are the result of triplicate determinations.
- Slassi, A.; Edwards, L.; O'Brien, A.; Xin, T.; Tehim, A. U. S. Patent 6,100,291, 2000.
- Holenz, J.; Merce, R.; Diaz, J. L.; Guitart, X.; Codony, X.; Dordal, A.; Romero, G.; Torrens, A.; Mas, J.; Andaluz, B.; Hernandez, S.; Monroy, X.; Sanchez, E.; Hernandez, E.; Perez, R.; Cubi, R.; Sanfeliu, O.; Buschmann, H. J. Med. Chem. 2005, 48, 1781.
- Cole, D. C.; Lennox, W. J.; Lombardi, S.; Ellingboe, J. W.; Bernotas, R. C.; Tawa, G. J.; Mazandarani, H.; Smith, D. L.; Zhang, G.; Coupet, J.; Schechter, L. E. J. Med. Chem. 2005, 48, 353.
- Kohen, R.; Metcalf, M. A.; Khan, N.; Druck, T.; Huebner, K.; Lachowicz, J. E.; Meltzer, H. Y.; Sibley, D. R.; Roth, B. L.; Hamblin, M. W. *J. Neurochem.* 1996, 66, 47.