

Total Synthesis of (–)-Ardeemin

Bin He, Hao Song, Yu Du, and Yong Qin*

Department of Chemistry of Medicinal Natural Products, Key Laboratory of Drug Targeting and Novel Delivery System of Ministry of Education, West China School of Pharmacy, and State Key Laboratory of Biotherapy, Sichuan University, Chengdu 610041, P. R. China

yongqin@scu.edu.cn

Received October 5, 2008



Total synthesis of potent anti-MDR indole alkaloids (–)-ardeemin and its *N*-acyl analogues has been accomplished from L-tryptophan with about 2% overall yield in 20 steps. The key step depended on the newly developed three-step one-pot cascade reaction of **7** with diazoester **8** via intermolecular cyclopropanation, ring opening, and ring closure to assemble the chiral 3-substituted hexahydropyrrolo[2,3-b]indole **4a**.

Introduction

Indole alkaloid ardeemins (Figure 1, 1-3), isolated from the fermentation of a strain of Aspergillus fischeri by McAlpine and co-workers in 1993,¹ have demonstrated a potent ability to reverse multi-drug resistance (MDR).² For example, combining the use of N-acetylardeemin 2 (10 μ M) with vinblastine, the cytotoxicity of vinblastine against a drug-resistant KBV-1 tumor cell line was enhanced more than 1000-fold compared with using vinblastine alone. Notably, 2 was 10-fold more effective than a well-known MDR modulator of verapamil³ in in vitro experiments. The promising MDR reversal activity of ardeemins has attracted the interest of synthetic chemists. Structurally, ardeemins contain a fundamental skeleton of 3-substituted hexahydropyrrolo[2,3-b]indole (4). Although a variety of methods have been reported for preparing racemic⁴ and chiral⁵ 3-substituted hexahydropyrrolo[2,3-b]indole in the past 2 decades, up to now, only Danishefsky's group accomplished an elegant total synthesis of (-)-ardeemin by using the *N*-phenylselenophthalimide induced selenocyclization of tryptophan as a key step to construct the chiral 3-selenenylated hexahydropyrrolo[2,3-*b*]indole.⁶

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5-*N*-Acetylardeemin (2), $R^{1} = Ac$, $R^{2} = H$; 15b-Hydroxy-5-*N*-acetylardeemin (3), $R^{1} = Ac$, $R^{2} = OH$

FIGURE 1. Structures of ardeemins.

SCHEME 1. Intermolecular Cyclopropanation of the Three-Step One-Pot Cascade Reaction^{*a*}



^{*a*} Reagents and conditions: (a) LiAlH₄, THF, 90%; (b) triphosgene, aqueous KOH, THF, 84%; (c) Boc₂O, Et₃N, 75%; (d) Cs₂CO₃, Bu₄NHSO₄, MeI, 91%; (e) TFA, 85%; (f) 1 mmol of **7**, 4 equiv of **8**, 5 mol % of Cu(OTf)₂, -35 °C in CH₂Cl₂, 30 h, 82% of **4a** and 5% of **4b**.

We have recently reported the total synthesis of the complex polycyclic indole alkaloids (\pm) -communesin F^7 and (\pm) -minfiensine⁸ by employing an intramolecular cyclopropanation strategy. As a continuation of our methodology development and application, we demonstrated an approach to chiral 3-substituted hexahydropyrrolo[2,3-*b*]indoles **4a** and **4b** from oxazo-lidinone **7** via a three-step one-pot cascade reaction of intermolecular cyclopropanation, ring opening, and ring closure in a high yield and in a high diastereoselectivity (Scheme 1).⁹ Through this approach, three stereocenters of the major isomer **4a** corresponding to the C5a, C15b and C16a of (–)-ardeemins were created. Herein, we wish to report our further efforts on the application of the intermolecular cyclopropanation of a three-step one-pot cascade reaction to the total synthesis of (–)-ardeemin, (–)-*N*-formalardeemin, and (–)-*N*-acetylardeemin.

Results and Discussion

Previously, a preparation of oxazolidinone 7 from L-tryptophan through a five-step reaction resulting in a 44% overall yield was not straightforward and required chromatographic separation and the use of expensive Cs_2CO_3 (Scheme 1).⁹ In order to make the procedure more practical, a new synthetic

SCHEME 2. Improved Synthesis of Oxazolidinone 7 and $4a^{a}$



^{*a*} Reagents and conditions: (a) NH₃/Na, MeI, $-60 \degree C$, 92%; (b) LiAlH₄, THF, rt; (c) triphosgene, 10% aqueous KOH, THF, 78% for two steps; (d) Cu(OTf)-tolulene, toluene, 25 $\degree C$, 45% yield of **4a** and 28% yield of **4b**.

approach to **7** from L-tryptophan was developed via a threestep reaction. As shown in Scheme 2, direct alkylation of L-tryptophan with MeI under a condition of liquid NH_3/Na gave the *N*-methyl-protected **11** in a 92% yield.¹⁰ Reduction of **11** with LiAlH₄ gave amino alcohol **12** in high yield. Without purification, compound **12** was treated with triphosgene under an aqueous basic condition in THF, followed by recrystallization from ethyl acetate to afford **7** in a 78% yield in two steps. By this new approach, the overall yield of **7** from L-tryptophan increased by 28% compared to the previously used five-step procedure. The three-step procedure was readily implemented on a 200 g scale without chromatographic separation.

For the synthesis of (-)-ardeemin, a large quantity of the key intermediate 4a was expected to be produced from 7. Therefore, the efficiency of the three-step one-pot cascade reaction of 7 with diazoester 8 was reevaluated at a larger scale rather than the previous small scale (1 mmol). Unfortunately, although a similar ratio of 4a to 4b (20:1) was observed,⁹ direct application of the previous condition (4 equiv of diazoester 8, 0.05 equiv of Cu(OTf)₂, -35 °C in CH₂Cl₂) was not successful at a 50 g scale. A low yield of 4a (22% yield) was obtained as a result of the formation of a large quantity of dark precipitates and rapid non-productive decomposition of diazoester 8. Because the first step of the cyclopropanation reaction was very sluggish at low temperatures in CH₂Cl₂, to efficiently promote the first step of the cyclopropanation reaction and to avoid the formation of precipitates, the cascade reaction has to be performed at room temperature. To our delight, in the presence of 0.2 equiv of freshly made CuOTf-toluene complex in toluene, the cascade reaction proceeded smoothly for 6 h to give 4a in a 45 % yield and $4b^{11}$ in a 28 % yield at a 50 g scale when 4 equiv of 8 was used (Scheme 2).

With easily available pyrroloindole **4a** in hand, our next task was to transfer the ethyl acetate group in **4a** to an isoprenyl group (Scheme 3). Thus, double α -alkylation of the ethyl acetate group with LDA/MeI resulted in dimethylated **13** in a 72% yield. Unfortunately, attempts to directly reduce the ester group in **13** to aldehyde by DIBAL failed due to a lack of functional group selectivity between the ester group and the oxazolidinone group. After screening several reducing reagents, LiBH₄ was found to be a good choice for reducing reagent to produce alcohol **14** in a 61% yield. Oxidation of the hydroxyl group in **14** with Dess–Martin reagent, followed by Wittig reaction, furnished olefin **16** in a 93% yield.

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^{*a*} Reagents and conditions: (a) LDA, MeI, THF, -78 °C to rt; (b) LDA, MeI, THF, -78 °C to rt, 72% from **4a**; (c) LiBH₄, THF/MeOH, 0 °C, 61%; (d) DMP, CH₂Cl₂, rt, 92%; (e) Ph₃P⁺MeI⁻, LiHMDS, THF, -78 °C to rt, 93%; (f) 'BuOK, aq 'BuOH, rt, quantitative yield; (g) (Boc)₂O, CH₂Cl₂, rt, 95%, (h) DMP, CH₂Cl₂, rt, 92%; (i) PCC, CH₂Cl₂, rt, 76%; (j) NaClO₂, NaH₂PO₄ buffer, rt, quantitative yield; (k) ClCOO'Bu, Et₃N, D-Ala-OMe, CH₂Cl₂, 0 °C, 81%.

Construction of the D-ring by condensation with D-alanine required transformation of oxazolidinone group in 16 to a protected aminocarboxy group (Scheme 3). Opening of the oxazolidinone ring in 16 with 'BuOK and aqueous 'BuOH, followed by protection of the resulting amino group with Boc₂O, resulted in alcohol 17 in an excellent yield for the two steps. Initially, it was planned to oxidize both the hydroxyl group and the methyl group in 17 into an acidic group and a formamido group (20) in a single-step reaction. However, a number of oxidation reagents and conditions were tried, but all attempts to realize this direct transformation failed because of the easy formation of an N-oxide functionality on the indoline nitrogen under the oxidation conditions tested. Therefore, a stepwise oxidation of 17 had to be conducted. As a result, the hydroxyl group in 17 was first oxidized with Dess-Martin reagent to give a mixture of two rotamers 18a and 18b in a 92% yield and a 1:2 ratio. The *N*-methyl group in **18a** and **18b** was then oxidized with PCC to an N-formal group to give rotamers 19a and 19b in a 76% yield. Treatment of the mixture of 19a and 19b with NaClO₂ in a NaH₂PO₄ buffer provided a mixture of two rotamers 20a and 20b in a quantitative yield. Condensation of 20a and 20b with D-alanine methyl ester in the presence of isobutyl chloroformate and triethylamine in CH2Cl2 afforded rotamers 21a and 21b in an 81% yield.

After the Boc group was removed by TMSI,^{6a} the ratio of rotamers changed from 1:2 in **21** to 1:5 in **22** because of partial release of the aminal rigidity (Scheme 4). With a congested formal group on the indoline nitrogen of **22**, cyclization of the D-ring by using a solution of saturated methanolic ammonia and a catalytic quantity of DMAP exclusively yielded the amide

SCHEME 4. Synthesis of Compounds 26a and 26b^a



^{*a*} Reagents and conditions: (a) TMSI, CH₃CN, 0 °C, 98%; (b) MeOH/ NH₃, DMAP, 0 °C to rt, 86%; (c) LiOH, aq. MeOH, 95% yield; (d) ClCOO/Bu, Et₃N, D-Ala-OMe, CH₂Cl₂, 0 °C to rt, 71% of **23a** and **23b**.

SCHEME 5. Synthesis of Ardeemins^a



^{*a*} Reagents and conditions: (a) 2 equiv of ^{*n*}BuLi, *o*-azidobenzoic anhydride, THF, -78 °C, 86% of **27** and **28** in a 1:1 ratio; (b) 2 equiv of ^{*n*}BuLi, *o*-azidobenzoic anhydride, THF, -78 °C, 33% of **27** and **28** in a 1:1 ratio; (c) ^{*n*}Bu₃P, benzene, rt, 93%; (d) Ac₂O, DIPEA, 60 °C, benzene.⁶

23, only trace amount of **26** was formed. We then turned our efforts of D-ring cyclization to the aid of coupling agents. After hydrolysis of **22** with LiOH in aqueous MeOH, the resulting acid **24** was treated with ClCOO'Bu/Et₃N to afford two separable diketopiperazines **26a** and C8-*epi*-**26b** in a 71% yield and a 1:1 ratio. Rotamerism caused by the formal group in **26** was eliminated after the D-ring was formed. Epimerization at C8 for **26b** probably proceeded through a ketene intermediate **25**. The relative configuration between C8 and C15b in **26a** and **26b** was unambiguously confirmed by noe experiments.

A strategy similar to Danishefsky's E-ring construction was adopted (Scheme 5).⁶ Condensation of diketopiperazine **26a** with *o*-azidobenzoic anhydride under a strong basic condition at low temperature provided azido **27** and deformal azido **28** in an 86% yield and 1:1 ratio. Interestingly, although the yield was low (33% yield), the epimerized **26b** also afforded azido **27** and **28** in a 1:1 ratio under the same condensation condition. Obviously,

thermodynamically favorable products **27** and **28** were produced from **26b** via an enolate intermediate **29** after quenching the reaction. Treatment of **27** and **28** with ^{*n*}Bu₃P accomplished the total synthesis of (–)-*N*-formalardeemin **30** and (–)-ardeemin **1**. (–)-*N*-Acetylardeemin **2** was prepared from (–)-ardeemin **1** by using the published procedure.^{6b}

Conclusion

In summary, total synthesis of indole alkaloid (-)-ardeemin, (-)-*N*-formalardeemin, and (-)-*N*-acetylardeemin has been accomplished from L-tryptophan with about 2% overall yield in 20 steps. The key step depended on our recently developed three-step one-pot cascade reaction of intermolecular cyclopropanation, ring opening, and ring closure to assemble the chiral 3-substituted hexahydropyrrolo[2,3-*b*]indole with three stereocenters corresponding to (-)-ardeemin. Current synthesis has provided a practicable route to prepare analogues of (-)-ardeemin for further SAR studies of anti-MDR activity.

Experimental Section

Improved Synthesis of Oxazolidinone 7 and 4a. Oxazolidinone 7.⁹ Compound 11 (150 g, 0.69 mol) was suspended in anhydrate THF. The resulting mixture was cooled to 0 °C in an ice—water bath, and then LiAlH₄ (78.7 g, 2.07 mol) was slowly added within 4 h. After reflux for 2 h, the reaction was cooled to 0 °C and quenched by saturated aqueous Na₂SO₄, filtered, and extracted with EtOAc (500 mL \times 6). The combined organic phases were dried over Na₂SO₄ and concentrated to give the red residue.

The above residue was dissolved in THF (200 mL) and a solution of KOH (200 g, 3.6 mol) in H₂O (2,000 mL) was added. At 0 °C, a solution of triphosgene (205 g, 0.69 mol) in THF (500 mL) was slowly added to the above mixture. The reaction was then allowed to warm to room temperature and stirred for 8 h. The mixture was diluted with water (500 mL), extracted with CH₂Cl₂ (500 mL × 3), dried over Na₂SO₄, concentrated, and recrystallized from EtOAc to give oxazolidinone **7** as a white solid (124 g, 78% yield): $[\alpha]^{20}_{\rm D} = -50^{\circ}$ (*c* 1.0, CHC1₃); ¹H NMR (400 MHz, CDCl₃) δ 3.78 (s, 3H), 3.04–2.97 (m, 2H), 4.22–4.15 (m, 2H), 4.52–4.46 (m, 1H), 5.29 (s, 1H), 6.93 (s, 1H), 7.16–7.12 (m, 1H), 7.27 (td, *J* = 7.2, 1.2 Hz, 1H), 7.34–7.32 (m, 1H), 7.56–7.54 (m, 1H).

Hexahydropyrroloindoles 4a and 4b.⁹ Under N₂, to a solution of **7** (50 g, 0.22 mol) and a freshly made CuOTf-toluene complex (25 g, 0.043 mol)¹² in dry toluene (200 mL) was slowly added a solution of diazo **8** (100 g, 0.88 mol) in dry CH₂Cl₂ (1000 mL) at 25 °C. The reaction mixture was stirred at room temperature for 6 h and then was concentrated. The residue was purified by flash chromatography (17% EtOAc/petroleum) to afford **4a** (31 g, 45% yield) and **4b** (19 g, 28% yield).⁹

4a: $[\alpha]^{20}{}_{\rm D} = -175^{\circ}$ (*c* 1.0, CHC1₃); ¹H NMR (400 MHz, CDCl₃) δ 1.21 (t, *J* = 7.6 Hz, 3H), 2.05 (t, *J* = 6.8 Hz, 1H), 2.56 (dd, *J* =

(11) The minor isomer **4b** has the same absolute configurations at C2, C3, and C11 corresponding to natural indole alkaloids epi-aszonalenins A, B and C; see: Rank, C.; Phipps, R. K.; Harris, P.; Frisvad, J. C.; Gotfredsen, C. H.; Larsen, T. O. *Tetrahedron Lett.* **2006**, *47*, 6099.



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12.0, 5.2 Hz, 1H), 2.74 (d, J = 15.6 Hz, 1H), 2.80 (d, J = 16.0 Hz, 1H), 2.92 (s, 3H), 3.77–3.74 (m, 1H), 4.19–4.09 (m, 3H), 4.40 (t, J = 8.0Hz, 1H), 5.38 (s, 1H), 6.42 (d, J = 8.0 Hz, 1H), 6.69 (td, J = 7.6, 0.8 Hz, 1H), 7.08 (dd, J = 7.6, 0.8 Hz, 1H), 7.15 (td, J = 7.6, 1.2 Hz, 1H).

4b: $[\alpha]^{20}{}_{\rm D} = +22^{\circ} (c \ 1.0, \text{CHC1}_3)$; ¹H NMR (400 MHz, CDCl₃) δ 1.17 (t, J = 6.8 Hz, 3H), 1.93 (dd, J = 12.4, 9.6 Hz, 1H), 2.50–2.46 (m, 1H), 2.72 (d, J = 16.0 Hz, 1H), 2.78 (d, J = 15.6 Hz, 1H), 3.23 (s, 3H), 3.94 (dd, J = 8.0, 6.8 Hz, 1H), 4.08 (q, J = 7.2Hz, 2H), 4.37–4.31 (m, 1H), 4.43 (t, J = 8.0 Hz, 1H), 5.16 (s, 1H), 6.52 (d, J = 8.0Hz, 1H), 6.71 (t, J = 7.6 Hz, 1H), 7.02–7.00 (m, 1H), 7.16 (td, J = 8.0, 1.2 Hz, 1H).

Synthesis of Compounds 21a and 21b. Dimethylated 13. Under N₂, LDA (lithium diisopropylamide, 2.5 M in THF, 19 mL) was slowly dropped to a stirred solution of 4a (5 g, 15.8 mmol) in THF (100 mL) at -78 °C. The resulting mixture was stirred at -78 °C for 1 h, and then MeI (5.1 mL, 79.1 mmol) was added. After the mixture was stirred for 8 h, and the reaction was quenched by the addition of a saturated aqueous solution of NH₄Cl (50 mL) and extracted with EtOAc (50 mL × 3). The combined organic phases were washed with brine (100 mL), dried over Na₂SO₄, and evaporated. The residue was purified by flash chromatography (17% EtOAc/petroleum) to give a yellowish residue.

Under N2, the above residue was dissolved in THF (100 mL) at -78 °C. LDA (2.5 M in THF, 19 mL) was slowly dropped to the solution. After the mixture was stirred at -78 °C for 2 h, MeI (5.1 mL, 79.1 mmol) was added, and then the reaction mixture was stirred at -78 °C for another 1 h. The reaction was allowed to warm to room temperature and stirred overnight. The reaction was quenched by addition of a saturated aqueous solution of NH₄Cl (50 mL) and extracted with EtOAc (50 mL \times 3). The combined organic phases were washed with brine (100 mL), dried over Na₂SO₄, and evaporated to give a yellowish residue. Flash chromatography (17% EtOAc/petroleum) of the residue yielded dimethylated 13 as a pale yellow solid (3.91 g, 72% yield), mp $110-111 \text{ °C}; [\alpha]^{20}_{\text{D}} = -241^{\circ} (c \ 1.0, \text{CHCl}_3); \text{ }^{1}\text{H NMR} (400 \text{ MHz},$ CDCl₃) δ 1.17 (s, 3H), 1.28 (t, J = 7.2, 3H), 1.30 (s, 3H), 2.21 (dd, J = 22.8, 10.8 Hz, 1H), 2.35 (dd, J = 12.0, 5.6 Hz, 1H), 2.88(s, 3H), 3.70-3.64 (m, 1H), 4.19-4.12 (m, 3H), 4.36 (dd, J =8.8, 7.6 Hz, 1H), 5.54 (s, 1H), 6.40 (d, J = 7.6, 1H), 6.67 (td, J =7.2, 0.8 Hz, 1H), 7.02 (dd, *J* = 7.6, 1.2 Hz, 1H), 7.15 (td, *J* = 7.6, 1.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 13.9, 21.9, 22.0, 30.8, 40.7, 47.0, 58.6, 61.0, 64.1, 66.9, 85.8, 106.0, 117.2, 124.5, 128.8, 129.3, 151.6, 160.3, 175.7; HRMS-ESI calcd for C19H24N2O4Na $(M + Na)^+$ 367.1628, found 367.1630; IR (KBr) 1759, 1603, 1472, 1045 cm⁻¹.

Alcohol 14. To a solution of dimethylated 13 (3.441 g, 10 mmol) in THF (100 mL) and MeOH (0.8 mL, 20 mmol) was added LiBH₄ (436 mg, 20 mmol) at 0 °C under N₂. After stirring at 0 °C for 4 h, the reaction was quenched by an ice-cold saturated NH₄Cl solution (100 mL) and extracted with EtOAc (100 mL \times 3). The combined organic layers were washed with brine (200 mL), dried over Na₂SO₄, and concentrated in vacuum to give a yellow residue. Purification of the residue by column chromatography (67% EtOAc/ petroleum) afforded alcohol 14 as a colorless solid (1.844 g, 61% yield), mp 196–198 °C; $[\alpha]^{20}_{D} = -273^{\circ} (c \ 1.0, \text{CHCl}_3); {}^{1}\text{H NMR}$ (400 MHz, CDCl₃) δ 1.02 (s, 3H), 1.12 (s, 3H), 2.07 (dd, J =12.0, 10.8 Hz, 1H), 2.33 (dd, J = 12.0, 5.2 Hz, 1H), 2.60 (s br, 1H), 2.89 (s, 3H), 3.14 (s, 2H), 3.60–3.53 (m, 1H), 4.17 (dd, J = 8.8, 3.2 Hz, 1H), 4.38 (dd, J = 8.8, 8.0 Hz, 1H), 5.43 (s, 1H), 6.54 (d, J = 8.0, 1H), 6.79 (td, J = 7.6, 1.2 Hz, 1H), 7.12 (dd, J = 7.8, 1.2 Hz), 7.12 (dd,0.8 Hz, 1H), 7.20 (td, J = 7.6, 1.2 Hz, 1H); ¹³C NMR (50 MHz, CDCl₃) & 21.1, 21.6, 29.7, 31.6, 39.9, 58.4, 64.8, 67.0, 70.4, 85.5, 107.9, 118.8, 124.4, 129.1, 130.7, 151.2, 160.6; HRMS-ESI calcd for C₁₇H₂₂N₂O₃Na (M + Na)⁺ 325.1523, found 325.1516; IR (KBr) 3471, 1740, 1402, 1246, 1048 cm⁻¹.

Aldehyde 15. To a solution of alcohol 14 (1.512 g, 5 mmol) in CH₂Cl₂ (100 mL) was added Dess-Martin reagent (Dess-Matrin periodinane, 2.544 g, 6 mmol) at room temperature. After stirring

at room temperature for 1 h, the reaction was quenched by saturated Na₂S₂O₃ solution (100 mL). The mixture was extracted with CH₂Cl₂ (50 mL \times 3). The combined organic layers were washed with saturated NaHCO3 solution (100 mL) and brine (100 mL), dried over Na₂SO₄, filtered, and concentrated in vacuum. The residue was purified by flash chromatography (20% EtOAc/petroleum) to give aldehyde **15** (1.382 g, 92% yield), mp 115–116 °C; $[\alpha]^{20}_{D} =$ -85° (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.12 (s, 3H), 1.19 (s, 3H), 2.08 (dd, J = 12.0, 10.8 Hz, 1H), 2.34 (dd, J = 12.0, 5.2 Hz, 1H), 2.89 (s, 3H), 3.73-3.66 (m, 1H), 4.18 (dd, J = 9.2, 2.8 Hz, 1H), 4.39 (dd, J = 9.2, 7.6 Hz, 1H), 5.39 (s, 1H), 6.43 (d, J = 8.0, 1H), 6.70 (td, J = 7.2, 0.8Hz, 1H), 7.05 (dd, J = 7.6, 0.8Hz, 1H), 7.17 (td, J = 8.0, 1.2 Hz, 1H), 9.48 (s, 1H); ¹³C NMR (50 MHz, CDCl₃) δ 18.8, 18.9, 31.0, 39.7, 49.5, 58.5, 62.9, 66.9, 85.1, 106.5, 117.8, 124.5, 128.5, 129.5, 151.4, 160.4, 204.3; HRMS-ESI calcd for $C_{17}H_{20}N_2O_3Na (M + Na)^+ 323.1366$, found 323.1361; IR (KBr) 3441, 1746, 1389, 1190 cm⁻¹.

Olefin 16. To a solution of Ph₃P⁺MeI⁻ (835 mg, 2.07 mmol) in THF (50 mL) at -78 °C was dropped LiHMDS (lithium hexamthyldisilazide, 1 M in THF, 2.3 mL). After stirring at 0 °C for 1 h, the solution was cooled to -78 °C again and was added dropwise a solution of adehyde 15 (400 mg, 1.33 mmol) in THF. The reaction was stirred overnight and allowed to warm to room temperature slowly. The reaction was quenched by addition of a saturated aqueous solution of NH₄Cl (50 mL) and extracted with EtOAc (50 mL \times 3). The combined organic phases were washed with brine (100 mL), dried over Na₂SO₄, filtered, and evaporated to give a residue. Flash chromatography (17% EtOAc/petroleum) of the residue yielded olefin 16 as a white solid (368 mg, 93% yield), mp 83-85 °C; $[\alpha]^{20}_{D} = -348^{\circ}$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.99 (s, 3H), 1.09 (s, 3H), 1.99 (dd, J = 12.0, 10.8 Hz, 1H), 2.19 (dd, J = 12.0, 5.6 Hz, 1H), 2.88 (s, 3H), 3.68-3.61 (m, 1H), 4.18 (dd, J = 8.8, 2.8 Hz, 1H), 4.35 (dd, J = 8.8, 7.6 Hz, 1H), 5.07 (dd, J = 17.6, 1.6 Hz, 1H), 5.13 (dd, J = 7.2, 1.2 Hz, 1H), 5.29 (s, 1H), 5.94 (dd, J = 17.2, 10.8 Hz, 1H), 6.39 (d, J = 8.0, 1H), 6.67 (td, J = 7.6, 0.8 Hz, 1H), 7.08 (dd, J = 7.6, 0.8 Hz, 1H), 7.15 (td, J = 7.6, 1.2 Hz, 1H); ¹³C NMR (50 MHz, CDCl₃) δ 22.5, 23.3, 30.9, 39.9, 40.7, 58.6, 65.5, 66.8, 85.3, 105.9, 114.3, 117.1, 124.7, 128.9, 129.8, 143.7, 151.7, 160.3; HRMS-ESI calcd for $C_{18}H_{22}N_2O_2Na (M + Na)^+$ 321.1574, found 321.1570; IR (KBr) 1755, 1746, 1604, 1190 cm⁻¹.

Alcohol 17. To a solution of olefin 16 (300 mg, 1.01 mmol) in aqueous 'BuOH (20 mL) was added 'BuOK (452 mg, 4.04 mmol) in one portion at room temperature. After stirring for 4 h, the mixture was diluted with EtOAc (50 mL), washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated in vacuum. The residue was purified by column chromatography (9% MeOH/ CH_2Cl_2) to yield amino alcohol as viscous liquid.

To a solution of the above amino alcohol in CH₂Cl₂ (20 mL) was added Boc₂O (di-tert-butyl dicarbonate, 872 mg, 4.00 mmol). After stirring at room temperature overnight, the mixture was concentrated and purified by column chromatography (17% EtOAc/ petroleum) to give alcohol 17 (353 mg, 95% yield) as a white solid, mp 88–90 °C; $[\alpha]^{20}_{D} = -232^{\circ} (c \ 1.0, \text{CHCl}_{3}); ^{1}\text{H NMR} (400 \text{ MHz},$ CDCl₃) & 0.93 (s, 3H), 1.04 (s, 3H), 1.73 (s br, 1H), 1.95-1.89 (m, 2H), 2.97 (s, 3H), 3.46-3.40 (m, 1H), 3.74-3.61 (m, 2H), 5.09 (dd, J = 17.2, 1.2 Hz, 1H), 5.09 (dd, J = 10.8, 0.8 Hz, 1H),5.25 (s, 1H), 5.89 (dd, J = 17.2, 10.8 Hz, 1H), 6.35 (d, J = 7.6, 1H), 6.65 (t, J = 7.2, 1H), 7.06 (d, J = 7.6 Hz, 1H), 7.11 (td, J = 7.6, 0.8 Hz, 1H); ¹³C NMR (50 MHz, CDCl₃) δ 22.2, 23.0, 28.5, 34.3, 37.5, 40.9, 60.1, 61.9, 63.9, 81.0, 85.6, 106.0, 113.6, 117.1, 124.4, 128.6, 130.8, 144.2, 152.1, 154.9; HRMS-ESI calcd for $C_{22}H_{32}N_2O_3Na (M + Na)^+$ 395.2311, found 395.2305; IR (KBr) 3422, 1680, 1604, 1409, 1075, 750 cm⁻¹.

Aldehydes 18a and 18b. To a solution of alcohol 17 (300 mg, 0.81 mmol) in CH_2Cl_2 (20 mL) was added Dess-Martin reagent (512 mg, 1.21 mmol) at room temperature. After stirring at room temperature for 2 h, the reaction was quenched by saturated $Na_2S_2O_3$ solution (50 mL) and extracted with CH_2Cl_2 (50 mL \times 3). The

combined organic layers were washed with saturated NaHCO₄ solution (100 mL) and brine (100 mL), dried over Na₂SO₄, filtered, and concentrated in vacuum. The residue was purified by flash chromatography (17% EtOAc/petroleum) to give a mixture of two aldehydes **18a** and **18b** (274 mg, 92% yield), $[\alpha]^{20}_{D} = -242^{\circ}$ (c 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) δ 0.94 (rotamer A and B; s, 3H), 1.08 (rotamer A and B; s, 3H), 1.40 (rotamer B; s, 9H), 1.54 (rotamer A; s, 9H), 2.23-2.04 (rotamer A and B; m, 2H), 3.02 (rotamer A; s, 3H), 3.08 (rotamer B; s, 3H), 3.78-3.72 (rotamer A and B; m, 1H), 5.15-5.05 (rotamer A and B; m, 2H), 5.17 (rotamer A; s, 1H), 5.37 (rotamer B; s, 1H), 5.91 (rotamer A and B; dd, *J* = 17.2, 7.2 Hz, 1H), 6.40-6.37 (rotamer A and B; m, 1H), 6.71-6.66 (rotamer A and B; m, 1H), 7.05 (rotamer A and B; d, J = 7.6 Hz, 1H), 7.17–7.13 (rotamer A and B; m, 1H), 9.22 (rotamer B; d, J = 4.4 Hz, 1H), 9.33 (rotamer A; d, J = 4.4 Hz, 1H); ¹³C NMR (50 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) rotamer B: δ 14.1, 22.6, 22.8, 28.1, 29.6, 33.6, 41.1, 62.0, 64.7, 79.1, 82.0, 109.2, 114.4, 118.4, 124.7, 128.8, 143.6, 149.8, 153.5, 196.5; rotamer A: δ 14.1, 22.6, 22.8, 28.5, 29.3, 33.1, 41.1, 63.4, 64.9, 78.3, 82.0, 109.1, 114.4, 118.9, 123.1, 129.3, 143.1, 149.3, 153.5, 196.5; HRMS-ESI calcd for $C_{22}H_{30}N_2O_3Na (M + Na)^+$ 393.2154, found 393.2149; IR (KBr) 1740, 1696, 1464, 1367, 1260, 803 cm⁻¹.

N-Formals 19a and 19b. To a solution of aldehydes 18a and 18b (200 mg, 0.54 mmol) in CH₂Cl₂ (20 mL) was added PCC (pyridinium chlorochromate, 233 mg, 1.08 mmol) at room temperature. After stirring at room temperature for 4 h, the reaction was concentrated and purified by flash chromatography (50% EtOAc/petroleum) to give a mixture of N-formals 19a and 19b (158 mg, 76% yield), $[\alpha]^{20}_{D} = -72^{\circ} (c \ 0.1, \text{CHCl}_3); {}^{1}\text{H NMR} (400 \text{ MHz},$ CDCl₃; a mixture of two rotamers in a 1:2 ratio) δ 0.97 (rotamer B; s, 3H), 1.11 (rotamer B; s, 3H), 1.13 (rotamer A; s, 3H), 1.25 (rotamer A; s, 3H), 1.39 (rotamer B; s, 9H), 1.54 (rotamer A; s, 9H), 2.27-2.22 (rotamer A and B; m, 2H), 3.73-3.68 (rotamer A and B; m, 1H), 5.10 (rotamer A and B; d, J = 17.2 Hz, 1H), 5.17 (rotamer A and B; d, J = 10.8 Hz, 1H), 5.68 (rotamer A; s, 1H), 5.88 (rotamer B; s, 1H), 5.90-5.80 (rotamer A and B; m, 1H), 7.16 (rotamer A and B; q, J = 8.0 Hz, 1H), 7.30–7.24 (rotamer A and B; m, 1H), 7.35 (rotamer A and B; t, J = 8.0 Hz, 1H), 7.92 (rotamer A; d, J = 8.0 Hz, 1H), 8.01 (rotamer B; d, J = 8.0 Hz, 1H), 8.68 (rotamer A; s, 1H), 8.93 (rotamer B; s, 1H), 9.26 (rotamer B; d, J = 4.4 Hz, 1H), 9.35 (rotamer A; d, J = 4.4 Hz, 1H); ¹³C NMR (50 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) rotamer B: δ 14.1, 22.1, 23.1, 29.3, 29.9, 31.9, 33.3, 41.1, 64.4, 78.7, 83.0, 115.2, 115.7, 116.5, 117.6, 124.7, 129.3, 142.6, 161.8, 196.5; rotamer A: δ 14.1, 22.1, 22.7, 28.0, 28.3, 32.1, 33.3, 41.1, 64.8, 78.2, 83.3, 115.0, 115.7, 117.6, 116.5, 125.1, 129.3, 142.1, 161.1, 196.7; HRMS-ESI calcd for $C_{22}H_{28}N_2O_4Na$ (M + Na)⁺ 407.1947, found 407.1941; IR (KBr) 1687, 1584, 1439, 1216, 758 cm^{-1} .

Acids 20a and 20b. To a solution of N-formals 19a and 19b (96 mg, 0.25 mmol) in a mixture of CH₃CN/^{*t*}BuOH/H₂O (2:2:1, 10 mL) were added NaH2PO4·2H2O (117 mg, 0.75 mmol) and NaClO₂ (136 mg, 1.5 mmol) sequentially. After stirring at room temperature for 0.5 h, the mixture was diluted with EtOAc (50 mL), washed with saturated Na₂S₂O₃ solution (50 mL) and brine (50 mL), dried over Na₂SO₄, and concentrated in a vacuum to give a residue. The residue was purified by column chromatography (9% MeOH/CH₂Cl₂) to yield a mixture of acids 20a and 20b (100 mg, 99% yield), $[\alpha]^{20}_{D} = -158^{\circ} (c \ 0.1, \text{ CHCl}_3); {}^{1}\text{H NMR} (400 \text{ MHz},$ CDCl₃; a mixture of two rotamers in a 1:2 ratio) δ 0.97 (rotamer A and B; s, 3H), 1.08 (rotamer B; s, 3H), 1.10 (rotamer A; s, 3H), 1.37 (rotamer B; s, 9H), 1.51 (rotamer A; s, 9H), 2.55-2.31 (rotamer A and B; m, 2H), 3.92 (rotamer A and B; dd, *J* = 9.6, 7.2 Hz, 1H), 5.06 (rotamer A and B; d, J = 17.2 Hz, 1H), 5.13 (rotamer A and B; d, J = 10.8 Hz, 1H), 5.70 (rotamer A; s, 1H), 5.87–5.77 (rotamer A and B; m, 1H), 5.90 (rotamer B; s, 1H), 7.19-7.13 (rotamer A and B; m, 1H), 7.27-7.24 (rotamer A and B; m, 1H), 7.34 (rotamer A and B; d, J = 7.6 Hz, 1H), 7.89 (rotamer A; d, J

= 8.0 Hz, 1H), 7.99 (rotamer B; d, J = 8.0 Hz, 1H), 8.65 (rotamer A; s, 1H), 8.91 (rotamer B; s, 1H); ¹³C NMR (50 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) rotamer B: δ 22.1, 23.1, 29.6, 31.8, 36.5, 41.0, 59.3, 61.1, 78.8, 82.1, 114.8, 117.7, 124.7, 129.2, 133.3, 140.6, 142.6, 162.1, 152.7, 177.4; rotamer A: 22.1, 23.1, 29.6, 31.8, 34.8, 41.0, 59.3, 62.5, 78.3, 83.1, 114.8, 117.7, 125.1, 129.2, 133.3, 140.2, 142.6, 153.1, 161.3, 177.0; HRMS-ESI calcd for C₂₂H₂₇N₂O₅ (M - H)⁻ 399.1926, found 399.1920; IR (KBr) 3460, 1688, 1596, 1367, 1150, 755 cm⁻¹.

Amides 21a and 21b. To a solution of acids 20a and 20b (50 mg, 0.12 mmol) and Et₃N (35 μ L, 0.25 mmol) in CH₂Cl₂ (5 mL) was added ClCOO'Bu (33 µL, 0.25 mmol) at 0 °C by using a syringe. After stirring for 15 min, to the mixture was added D-Ala-OMe (methyl D-alaninate, 25.8 mg, 0.25 mmol), and the mixture was allowed to stir for 1 h at 0 °C. The mixture was then quenched by saturated NaHCO₃ (10 mL) and CH₂Cl₂ (10 mL). The organic layer was separated, dried over Na₂SO₄, and concentrated in vacuum. The residue was purified by column chromatography (25% EtOAc/petroleum) to yield a mixture of amides 21a and 21b (49 mg, 81% yield), $[\alpha]^{20}_{D} = -135^{\circ}$ (c 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) δ 0.98 (rotamer A and B; s, 3H), 1.01 (rotamer A and B; s, 3H), 1.33 (rotamer B; s br, 9H), 1.45-1.39 (rotamer A and B; m, 3H), 1.50 (rotamer A; s br, 9H), 2.48-2.44 (rotamer A and B; m, 2H), 3.75 (rotamer B; s, 3H), 3.76 (rotamer A; s, 3H), 3.77-3.73 (rotamer A and B; m, 1H), 4.63-4.58 (rotamer A and B; m, 1H), 5.06 (rotamer A and B; d, J = 17.2 Hz, 1H), 5.14 (rotamer A and B; d, J = 10.8 Hz, 1H), 5.70 (rotamer A; s, 1H), 5.86-5.82 (rotamer A and B; m, 1H), 5.90 (rotamer B; s, 1H), 6.24 (rotamer A and B; s br, 1H), 7.16 (rotamer A and B; t, J = 7.2 Hz, 1H), 7.28–7.25 (rotamer A and B; m, 1H), 7.33 (rotamer A and B; t, J = 7.6 Hz, 1H), 7.89 (rotamer A; s br, 1H), 7.97 (rotamer B; d, J = 7.2 Hz, 1H), 8.65 (rotamer A; s, 1H), 8.89 (rotamer B; s, 1H); ¹³C NMR (50 MHz, CDCl₃; a mixture of two rotamers in a 1:2 ratio) rotamer B: δ 18.8, 22.2, 23.0, 28.0, 29.6, 37.3, 40.9, 47.9, 52.5, 61.2, 79.2, 82.2, 114.7, 117.6, 124.7, 129.1, 133.6, 140.9, 142.8, 153.0, 162.0, 170.8, 173.1; rotamer A: δ 18.6, 22.2, 23.0, 28.0, 29.6, 35.5, 40.9, 47.9, 52.5, 60.8, 79.2, 82.2, 114.7, 117.6, 124.5, 129.1, 133.6, 140.9, 142.8, 153.0, 161.5, 170.4, 173.1; HRMS-ESI calcd for C₂₆H₃₅N₃O₆Na (M + Na)⁺ 508.2418, found 508.2419; IR (KBr) 3322, 1774, 1681, 1460, 1151, 756 cm⁻¹.

Synthesis of Compounds 26a and 26b. Compounds 22a and 22b. A mixture of amides 21a and 21b (33 mg, 0.068 mmol) was dissolved in freshly distilled dry MeCN (5 mL), chilled to 0 °C under N2. To the solution was added TMSI (iodotrimethylsilane, 39 μ L, 0.27 mmol) dropwise.⁶ After 30 min, the reaction mixture was poured into saturated NaHCO3 (10 mL) and extracted with CH_2Cl_2 (10 mL \times 3). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to give a mixture of **22a** and **22b** (26 mg, 98% yield). $[\alpha]^{20}_{D} =$ -47° (c 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃; a mixture of two rotamers in 1:5 ratio) δ 1.03 (rotamer A; s, 3H), 1.07 (rotamer B; s, 6H), 1.10 (rotamer A; s, 3H), 1.40 (rotamer A and B; d, J =7.2 Hz, 3H), 2.05 (rotamer A and B; s br, 1H), 2.33-2.22 (rotamer A and B; m, 2H), 3.57-3.52 (rotamer A and B; m, 1H), 3.76 (rotamer A and B; s, 3H), 4.61-4.53 (rotamer A and B; m, 1H), 5.06 (rotamer A and B; d, J = 17.6 Hz, 1H), 5.13 (rotamer A and B; d, J = 10.8 Hz, 1H), 5.43 (rotamer A; s, 1H), 5.66 (rotamer B; s, 1H), 5.90 (rotamer A; dd, J = 12.0, 10.8 Hz, 1H), 5.94 (rotamer B; dd, J = 12.0, 10.8 Hz, 1H), 7.14–7.07 (rotamer A and B; m, 2H), 7.30-7.21 (rotamer A and B; m, 1H), 7.35 (rotamer B; d, J = 7.6 Hz, 1H), 8.04 (rotamer A; d, J = 7.6 Hz, 1H), 8.59 (rotamer A; s, 1H), 8.94 (rotamer B; s, 1H); ¹³C NMR (50 MHz, CDCl_{3;} a mixture of two rotamers in 1:5 ratio) rotamer B: δ 18.2, 22.5, 23.5, 29.7, 40.4, 47.2, 52.4, 60.0, 62.2, 78.3, 108.4, 114.4, 124.6, 126.5, 128.6, 131.1, 134.3, 143.4, 158.6, 172.6, 173.0; rotamer A: δ 19.1, 22.6, 23.3, 29.2, 39.4, 47.5, 52.4, 60.9, 62.2, 80.4, 108.4, 116.2, 124.9, 126.5, 128.8, 131.1, 134.3, 141.0, 159.4, 172.6, 173.0; HRMS-ESI calcd for $C_{21}H_{27}N_3O_4Na (M + Na)^+$ 408.1894, found 408.1899; IR (KBr) 1740, 1696, 1464, 1367, 1260, 803 cm⁻¹.

Amides 23a and 23b. To a mixture of amides 22a and 22b (2.5 mg, 0.006 mmol) in MeOH saturated with ammonia (0.5 mL) at 0 °C was added DMAP (4-dimethylaminopyridine, 0.5 mg, 0.004 mmol). The solution was stirred overnight, allowed to warm to room temperature, and concentrated. The residue was purified by flash chromatography (100% EtOAc) to give a mixture of amides 23a and 23b (2.0 mg, 86% yield). ¹H NMR (400 MHz, CDCl₃; a mixture of two rotamers in 1:4 ratio) δ 0.98 (rotamer A; s, 3H), 1.02 (rotamer B; s, 3H), 1.07 (rotamer B; s, 3H), 1.10 (rotamer A; s, 3H), 1.40 (rotamer A and B; d, J = 6.8 Hz, 3H), 2.18-2.08 (rotamer A and B; m, 1H), 2.32 (rotamer B; dd, J = 12.4, 5.6 Hz, 1H), 2.40 (rotamer A; dd, J = 12.4, 5.6 Hz, 1H), 2.81 (rotamer A and B; s br, 1H), 3.50 (rotamer A; dd, J = 10.4, 5.6 Hz, 1H), 3.54 (rotamer B; dd, J = 10.4, 5.6 Hz, 1H), 4.44 (rotamer A and B; q, J = 7.2Hz, 1H), 5.06 (rotamer A and B; d, J = 17.2 Hz, 1H), 5.14 (rotamer A and B; d, J = 10.8 Hz, 1H), 5.29 (rotamer A and B; s br, 1H), 5.41 (rotamer A; s, 1H), 5.65 (rotamer B; s, 1H), 5.90 (rotamer A; dd, J = 17.2, 6.8 Hz, 1H), 5.93 (rotamer B; dd, J = 17.2, 6.8 Hz, 1H), 6.11 (rotamer A and B; s br, 1H), 7.16-7.08 (rotamer A and B; m, 2H), 7.27-7.22 (rotamer A and B; m, 1H), 7.35 (rotamer B; d, J = 8.0 Hz, 1H), 8.04 (rotamer A; d, J = 7.6 Hz, 1H), 8.58 (rotamer A; s, 1H), 8.93 (rotamer B; s, 1H).

Diketopiperazines 26a and 26b. To a solution of **22a** and **22b** (20 mg, 0.052 mmol) in a 10:1 mixture of MeOH/H₂O (5 mL) was added LiOH (11 mg, 0.268 mmol). After stirring for 4 h, the reaction was diluted with CH₂Cl₂ (10 mL), washed with brine (10 mL), dried over Na₂SO₄, concentrated and purified by flash chromatography (17% MeOH/CH₂Cl₂) to give a viscous residue.

To a solution of the above residue in CH₂Cl₂ (5 mL) were added Et₃N (14 μ L, 0.10 mmol) and ClCOO'Bu (14 μ L, 0.10 mmol) at 0 °C. After stirring at 0 °C for 2 h, the mixture was then quenched by saturated NaHCO₃ (10 mL) and CH₂Cl₂ (10 mL). The organic layer was separated, dried over Na₂SO₄, and concentrated in vacuum. The residue was purified by column chromatography (50% EtOAc/petroleum) to yield diketopiperazines **26a** (7.0 mg, 36% yield) and **26b** (6.0 mg, 35% yield).

26a: $[\alpha]^{20}{}_{\rm D} = -162^{\circ}$ (*c* 0.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.98 (s, 3H), 1.12 (s, 3H), 1.43 (d, *J* = 6.8 Hz, 3H), 2.43 (t, *J* =12.0 Hz, 1H), 2.65 (dd, *J* = 12.8, 6.0 Hz, 1H), 3.93 (q, *J* = 5.6 Hz, 1H), 4.09-4.03 (m, 1H), 5.11 (d, *J* = 17.2 Hz, 1H), 5.16 (d, *J* = 10.8 Hz, 1H), 5.87 (dd, *J* = 17.2, 10.8 Hz, 1H), 6.13 (s, 1H), 6.26 (s br, 1H), 7.16 (t, *J* =7.6 Hz, 1H), 7.33 (q, *J* =7.2 Hz, 2H), 8.06 (d, *J* = 8.0 Hz, 1H), 9.04 (s, 1H); ¹³C NMR (50 MHz, CDCl₃) δ 21.3, 22.2, 23.0, 37.4, 41.0, 53.3, 57.7, 60.1, 77.6, 115.5, 117.0, 124.9, 129.5, 132.0, 141.4, 142.4, 161.7, 166.3, 167.6; HRMS-ESI calcd for C₂₀H₂₃N₃O₃Na (M + Na)⁺ 376.1637, found 376.1629; IR (KBr) 3307, 1744, 1674, 1458, 756 cm⁻¹.

26b: $[\alpha]^{20}_{\rm D} = -199^{\circ}$ (*c* 0.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.99 (s, 3H), 1.13 (s, 3H), 1.48 (d, *J* = 6.8 Hz, 3H), 2.47 (t, *J* =11.2 Hz, 1H), 2.63(dd, *J* =12.8, 6.0 Hz, 1H), 4.00–3.95 (m, 1H), 4.12 (q, *J* = 7.2 Hz, 1H), 5.10 (d, *J* = 17.2 Hz, 1H), 5.16 (d, *J* = 10.4 Hz, 1H), 5.85 (dd, *J* = 17.2, 10.8 Hz, 1H), 6.07 (s, 1H), 6.20 (s br, 1H), 7.16 (t, *J* =7.6 Hz, 1H), 7.35–7.30 (m, 2H), 8.03 (d, *J* = 7.6 Hz, 1H), 9.04 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 16.4, 22.2, 23.0, 36.5, 40.9, 51.2, 59.0, 60.4, 77.3, 115.3, 117.0, 124.9, 129.4, 132.2, 141.3, 142.4, 161.6, 166.1, 168.8; HRMS-ESI calcd for C₂₀H₂₃N₃O₃Na (M + Na)⁺ 376.1637, found 376.1632; IR (KBr) 3307, 1682, 1463, 1375, 1160, 757 cm⁻¹.

Compounds 27 and 28 from 26a. To a solution of diketopiperazine **26a** (10.0 mg, 0.028 mmol) in THF (3 mL) at -78 °C under N₂ was added "BuLi (2.5 M in THF, 22 μ L, 0.055 mmol). After stirring for 20 min at -78 °C, a solution of *o*-azidobenzoic anhydride (17 mg, 0.056 mmol) in THF (0.1 mL) was added via a syringe. After 10 min, the mixture was poured into a biphasic mixture of saturated NaHCO₃ (5 mL) and EtOAc (5 mL). The aqueous layer was separated and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with brine, dried

over Na_2SO_4 , and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to give **27** (6.3 mg, 44% yield) and **28** (5.7 mg, 42 % yield).

27: $[\alpha]^{20}_{\rm D} = -72^{\circ}$ (*c* 0.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.99 (s, 3H), 1.14 (s, 3H), 1.52 (d, *J* = 6.8 Hz, 3H), 2.67–2.53 (m, 2H), 4.07 (dd, *J* = 11.2, 6.4 Hz, 1H), 5.09 (q, *J* = 7.2 Hz, 1H), 5.15 (d, *J* = 17.2 Hz, 1H), 5.21 (d, *J* = 10.4 Hz, 1H), 5.87 (dd, *J* = 17.4, 3.2 Hz, 1H), 6.17 (s, 1H), 7.20–7.16 (m, 2H), 7.31–7.23 (m, 2H), 7.37 (td, *J* = 7.4, 1.2 Hz, 1H), 7.42 (dd, *J* = 7.8, 1.6 Hz, 1H), 7.52 (td, *J* = 7.8, 1.2 Hz, 1H), 8.09 (d, *J* = 7.6 Hz, 1H), 9.09 (s, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 18.4, 22.2, 22.7, 37.3, 41.1, 55.5, 59.0, 60.5, 77.3, 115.7, 117.2, 118.2, 124.9, 125.0, 125.4, 128.3, 129.5, 129.7, 131.8, 132.1, 136.3, 141.3, 142.2, 161.6, 166.3, 167.8, 168.4; HRMS-ESI calcd for C₂₇H₂₆N₆O₄Na (M + Na)⁺ 521.1908, found 521.1913; IR (KBr) 3342, 2575, 1702, 1430, 1103cm⁻¹.

28:^{6a} $[\alpha]^{20}_{\rm D} = -152^{\circ}$ (*c* 0.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.00 (s, 3H), 1.13 (s, 3H), 1.51 (d, *J* = 7.6 Hz, 3H), 2.52 (d, *J* = 4.8 Hz, 1H), 2.54 (d, *J* = 2.0 Hz, 1H), 4.05 (dd, *J* = 10.0, 7.2 Hz, 1H), 5.06 (q, *J* = 7.2 Hz, 1H), 5.08 (s, 1H), 5.11 (dd, *J* = 16.4, 1.2 Hz, 1H), 5.16 (dd, *J* = 10.8, 0.8 Hz, 1H), 5.62 (s, 1H), 5.98 (dd, *J* = 17.2, 10.8 Hz, 1H), 6.64 (d, *J* = 7.6 Hz, 1H), 6.78 (td, *J* = 7.4, 7.4, 0.8 Hz, 1H), 7.17–7.11 (m, 3H), 7.23 (td, *J* = 7.6, 7.2, 1.2 Hz, 1H), 7.39 (dd, *J* = 7.8, 1.4 Hz, 1H), 7.50 (ddd, *J* = 8.2, 7.6, 1.6 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 17.7, 22.4, 22.9, 36.9, 41.0, 55.5, 59.1, 61.7, 77.5, 109.3, 114.9, 118.2, 119.0, 125.0, 125.2, 128.5, 128.6, 129.2, 131.8, 136.4, 143.3, 149.8, 166.7, 167.9, 169.1.

Compounds 27 and 28 from 26b. To a solution of diketopiperazine **26b** (10.0 mg, 0.028 mmol) in THF (3 mL) at -78 °C under N₂ was added "BuLi (2.5 M in THF, 22 μ L, 0.055 mmol). After 20 min of stirring at -78 °C, a solution of *o*-azidobenzoic anhydride (17.0 mg, 0.056 mmol) in THF (0.1 mL) was added via a syringe. After 10 min, the mixture was poured into a biphasic mixture of saturated NaHCO₃ (5 mL) and EtOAc (5 mL). The aqueous layer was separated and extracted with EtOAc (5 mL × 3). The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to give **27** (2.1 mg, 17% yield) and **28** (1.9 mg, 16 % yield).

Synthesis of Ardeemins. (-)-**Ardeemin.**^{1,6a} Under N₂, tri(*n*butyl)phosphine (10 μ L, 0.04 mmol) was added to a solution of **28** (10 mg, 0.021 mmol) in dry benzene (2 mL). The resulting solution was stirred for overnight under N₂, and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to yield (-)-ardeemin (8.0 mg, 93% yield). [α]²⁰_D = -122° (*c* 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.04 (s, 3H), 1.18 (s, 3H), 1.49 (d, *J* = 7.2 Hz, 3H), 2.75 (dd, *J* = 12.8, 10.4 Hz, 1H), 2.95 (dd, *J* = 12.8, 6.0 Hz, 1H), 4.52 (dd, *J* = 10.4, 6.0 Hz, 1H), 5.10 (d, *J* = 8.4 Hz, 1H), 5.14 (d, *J* = 1.6 Hz, 1H), 5.46 (q, *J* = 7.2 Hz, 1H), 5.60 (s, 1H), 6.03 (dd, *J* = 17.2, 11.2 Hz, 1H), 6.66 (d, *J* = 7.6 Hz, 1H), 6.80 (t, *J* = 7.2 Hz, 1H), 7.14 (t, *J* = 7.2 Hz, 1H), 7.26 (d, *J* = 7.5 Hz, 1H), 7.50 (t, *J* = 8.0, 1H), 7.68 (d, *J* = 7.6Hz, 1H), 7.77 (ddd, J = 7.0, 6.6, 1.2 Hz, 1H), 8.27 (dd, J = 8.0, 1.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 16.9, 22.6, 22.9, 38.2, 41.0, 53.2, 58.1, 61.8, 77.8, 109.3, 114.6, 118.9, 120.7, 125.1, 127.0, 127.3, 129.1, 134.7, 143.5, 147.2, 149.8, 150.9, 160.1, 166.6.

(-)-N-Formalardeemin. Under N₂, tri(n-butyl)phosphine (10 μ L, 0.04 mmol) was added to a solution of 27 (10.0 mg, 0.02 mmol) in dry benzene (2 mL). The reaction solution was stirred for overnight under N2 and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to yield (-)-N-formalardeemin (7.8 mg, 86% yield), $[\alpha]^{20}_{D} = -52^{\circ} (c \ 0.1, \text{CHCl}_3); {}^{1}\text{H}$ NMR (400 MHz, CDCl₃) δ 1.02 (s, 3H), 1.18 (s, 3H), 1.50 (d, J =6.8 Hz, 3H), 2.73 (t, J = 12.0 Hz, 1H), 3.03 (dd, J = 13.2, 6.0 Hz, 1H), 4.57-4.53 (m, 1H), 5.17-5.11 (m, 2H), 5.46 (q, J = 7.2 Hz, 1H), 5.89 (dd, J = 17.2, 10.8 Hz, 1H), 6.14 (s, 1H), 7.20 (t, J =7.2 Hz, 1H), 7.37 (t, J = 8.0 Hz, 1H), 7.42 (d, J = 8.0 Hz, 1H), 7.51 (t, J = 7.6 Hz, 1H), 7.66 (d, J = 8.4 Hz, 1H), 7.78 (t, J = 7.6 Hz, 1H), 8.09 (d, J = 8.0 Hz, 1H), 8.28 (d, J = 7.6 Hz, 1H), 9.10 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 17.5, 22.3, 23.0, 38.7, 41.1, 53.2, 58.1, 60.6, 77.3, 115.5, 117.3, 120.5, 124.9, 125.0, 127.0, 127.2, 127.4, 129.6, 132.3, 134.8, 141.1, 142.4, 147.0, 150.1, 159.8, 161.8, 166.2; HRMS-ESI calcd for $C_{27}H_{26}N_4O_3Na (M + Na)^+$ 477.1897, found 477.1893; IR (KBr) 3310, 2895, 1715, 1643, 1410, 1367, 755 cm⁻¹.

(-)-N-Acetylardeemin.^{1,6a} To a solution of (-)-ardeemin (4 mg, 0.01 mmol) in acetic anhydride (1 mL) was added DIPEA (N, *N*-diisoproylethylamine, 5 μ L, 0.03 mmol). The reaction mixture was heated to 60 °C for 36 h and then evaporated under reduced pressure. The residue was dissolved in CH₂Cl₂ (10 mL) and washed with saturated NaHCO₃ (10 mL) and brine (10 mL). The organic layer was dried over Na₂SO₄ and concentrated. The residue was purified by chromatography (50% EtOAc/petroleum) to give (-)-*N*-acetylardeemin (3.1 mg, 72% yield). $[\alpha]^{20}_{D} = -49^{\circ}$ (c 0.1, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ1.02 (s, 3H), 1.21 (s, 3H), 1.42 (d, J = 7.6 Hz, 3H), 2.67 (s br, 3H), 2.69–2.68 (m, 1H), 3.02 (dd, J = 12.8, 5.7 Hz, 1H), 4.45-4.43 (m, 1H), 5.16-5.12 (m, 1H)2H), 5.37 (q, J = 7.6 Hz, 1H), 5.81 (dd, J = 16.8, 10.8 Hz, 1H), 6.08 (s br, 1H), 7.24-7.22 (m, 1H), 7.45-7.42 (m, 2H), 7.55-7.52 (m, 1H), 7.73 (d, J = 8.8, 1H), 7.80–7.78 (m, 1H), 8.08 (s br, 1H), 8.29 (d, J = 8.0, 1H).

Acknowledgment. This research was supported by NSFC (Nos. 20632030, 20772083, and 20825207). We thank the Analytic and Testing Center of Sichuan University for recording spectroscopic data.

Supporting Information Available: NMR spectra of compounds 7, 4, 13–23, 26–28, (–)-ardeemin, (–)-*N*-formalardeemin, and (–)-*N*-acetylardeemin and NOEDS spectra of diketopiperazine 26a and 26b. This material is available free of charge via the Internet at http://pubs.acs.org.

JO802216Z