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Structural Revision of Baulamycin A and Structure-Activity Relationships of Baulamycin A Derivatives

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spectral properties of the synthetic compound differ from those reported for the natural product. On the basis of comprehensive NMR study, we proposed two other possible structures for natural baulamycin A. Total syntheses of these two substances were performed, which enabled assignment of the correct structure of baulamycin A. Key features of the convergent and fully stereo-controlled route include Evans Aldol and Brown allylation reactions to construct the left fragment, a prolinol amide-derived alkylation/desymmetrization to install the methyl substituted centers in the right fragment, and finally a Carreira alkynylation to join both fragments. In

addition, we have determined the inhibitory activities of novel baulamycin A derivatives against the enzyme SbnE. This SAR study provides useful insight into the design of novel SbnE inhibitors that overcome the drug resistance of pathogens, which cause life-threatening infections.

Introduction

Bacteria secrete low molecular weight and high-affinity iron chelators called siderophores to acquire iron needed for their virulence.¹ The biosynthetic pathways employed to generate these siderophores, such as staphyloferrin B of *Staphylococcus aureus*² and petrobactin of *Bacillus anthracis*,³ involve NRPS-independent siderophore (NIS) synthetases, which includes SbnE in staphyloferrin B and AsbA in petrobactin. Sherman *et al*⁴ recently isolated baulamycin A (BmcA) and baulamycin B (BmcB) from marine microbial-derived extracts (NPEs) from *Streptomyces tempisquensis* and showed that these substances serve as novel inhibitors of SbnE and AsbA. BmcA is an invaluable substance as it provides a promising starting-point for designing substances that are effective against drug-resistant pathogens such as MRSA and *Bacillus anthracis*.

As a part of an ongoing program devoted to total synthesis of biologically active natural products,⁵ we performed a total synthesis of Sherman's proposed⁴ stereostructure of BmcA (1). A comparison of the spectroscopic properties of the synthetic and natural materials demonstrated that the earlier proposed stereostructure of the natural product is incorrect. In the effort described below, we established the correct stereostructure of this natural product by utilizing comprehensive NMR studies and three total syntheses. It should be noted that, during the preparation of this manuscript, Goswami *et.al*^{6a} and Chandrasekhar *et.al*^{6b} reported a synthesis of

Sherman's originally proposed stereostructure of BmcA by employing a strategy that differs from ours. Aggarwal *et.al*^{6c} reported the correct structure of BmcA while our manuscript was under review. The reported stereostructure of BmcA has the following relative configurations at its seven stereogenic centers: $4R^*$, $6S^*$, $8R^*$, $11R^*$, $13R^*$, $14S^*$, and $1'R^*$ while the absolute configurations of these centers were arbitrarily represented by stereostructure **1** (Figure 1).⁴



Figure 1. Structures of BmcA and BmcB.

Results and discussion

As depicted retrosynthetically in Scheme 1, our strategy for the synthesis of 1 involved joining fragments 1 and 2 by using the Carreira alkynylation process. In the plan, fragment 1 (12 or 12a) would be derived from a protected form (4 or 4a) of 3,5-dihydroxybenzaldehyde and fragment 2 (20) would be generated from (*S*)-Roche ester.





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The synthetic pathway began with an Evans *syn* aldol reaction⁷ between 3,5-dimethoxy benzaldehyde **4** and auxiliary **5**, which produced hydroxyamide **6** (86%, dr = 96:4 by NMR) and protection as its TBS ether **7** (88%) followed by reductive cleavage promoted by treatment with DIBAL-H generate aldehyde **8** (60%, 2 steps).⁸ Brown allylation⁹ of aldehyde **8** produced the homoallylic alcohol **9** (74%, dr = 92:8 by NMR).¹⁰ Removal of the TBS group in **9** formed 1,3-diol **10** which upon reaction with 2,2-DMP, PTSA formed acetonide **11** (89%, 2 steps). The olefin group in **11** was cleaved by dihydroxylation followed by oxidative cleavage to afford aldehyde **12** (fragment **1**) (68%, 2 steps) (Scheme 2).

Scheme 2. Synthesis of fragment 1 (syn configuration)



The synthesis of fragment **2** was depicted in Scheme 3. Primary alcohol group in **13**¹¹ was converted to the corresponding iodide **14** by using TPP/Iodine. The lithium enolate, produced from L-prolinol *N*-propionamide by using *n*-BuLi, was coupled with iodide **14** to form 1,3-*anti* dimethyl¹² substituted amide **15** (85%, dr = 98:2 by NMR). Acid hydrolysis of **15**, followed by reduction formed primary alcohol **16** (69%, 2 steps), which was transformed to the iodide **17**. The lithium enolate derived from D-prolinol *N*-propionamide was coupled with iodide **17** to form 1,3,5-*anti*-trimethyl substituted amide **18** (86%, dr = 97:3 by NMR). Acid hydrolysis

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of **18**, followed by reduction generated primary alcohol **19** (65%, 2 steps). Swern oxidation¹³ of **19** followed by Bestmann-Ohira¹⁴ reaction then produced the required alkyne **20** (68%, 2 steps).



Scheme 3. Synthesis of fragment 2 (anti configuration)

The crucial coupling reaction joining **12** and **20** was conducted by using Carreira alkynylation conditions (Scheme 4).¹⁵ This process formed propargylic alcohol **21** (72%, dr = 95:5 by NMR), whose absolute configuration at the newly formed hydroxyl substituted center was confirmed as being '*S*' by utilizing the modified Mosher's ester method¹⁶ (Figure 2). The hydroxyl group in **21** was protected to form TBS ether **22** (87%), which was subjected to complete reduction by using Pd/C promoted hydrogenation to produce saturated alcohol **23** (75%). Primary alcohol **23** was then oxidized to form a crude aldehyde which was subsequently reacted with EtMgBr to generate **24** (87%, 2 steps) which was a diastereomeric mixture (1:1 by NMR). Oxidation of the secondary alcohol group in **24** afforded ketone **25** (92%), which upon removal of both the acetonide and TBS groups using PTSA in methanol generated the BmcA precursor **26** (90%). Removal of methoxy groups in **26** under several conditions¹⁷ proved to be fruitless except when aluminium iodide was employed, in which case BmcA (22%) was obtained but only after difficult purification.¹⁸



Figure 2. Modified Mosher's ester analysis.

Owing to the difficulty encountered with removal of the methoxy groups, the protection strategy was altered. Accordingly, the synthetic route was repeated using MOM protection of the phenolic hydroxy groups to produce aldehyde **12a**. Carreira alkynylation¹⁵ between **12a** and **20** was conducted to deliver propargylic alcohol **21a** (73%, dr = 96:4 by NMR) which was protected as its TBS-ether **22a** (89%). The MOM protected intermediate **22a** was then subjected to hydrogenation by using Pd/C to produce an inseparable mixture of the saturated alcohol **23a** and TBS-ether elimination product **23'** in a 8:2 ratio (confirmed by ¹H NMR analysis). After several unsuccessful attempts to separate **23a**, the mixture was used in the next step. Contrary to expectations, purification was not successful until the final step in the sequence involving global removal of the MOM, acetonide and TBS groups in **25a** by using PTSA in methanol to form **1** (90%).

Scheme 4. Synthesis of the proposed stereostructure of BmcA



The absolute configurations at the stereogenic centers in **1**, prepared by using the route described above, were firmly established by using the modified Mosher's method and HSQC NMR analysis of the corresponding acetonide derivative (acetonide **A** and **B**, Figure 3). The NMR data for synthetic **1** clearly differed from those reported previously.⁴ For instance, the ¹H and ¹³C NMR chemical shifts for C/H-1' (δ_C 76.4; δ_H 4.84), C/H-14 (δ_C 49.1; δ_H 1.70) and C/H-6 (δ_C 28.8; δ_H 1.52) were observed for the synthetic material whereas C/H-1' (δ_C 76.5; δ_H 4.47), C/H-14 (δ_C 48.5; δ_H 1.88), and C/H-6 (δ_C 29.1; δ_H 1.42) were reported for natural BmcA.⁴ These differences demonstrate that the proposed⁴ stereostructure of the natural product is incorrect. The H-14 and C-14 chemical shift discrepancies and the results of a ROESY NMR study (Figure S137) with the acetonide derivative of **1** (Figure 3-a) suggest that the configurations at the three consecutive 1,3-hydroxy substituted centers and *iso*-butyl substituted carbon are 11*R**, 13*R**, 14*R** and 1'*R** rather than the originally proposed 11*R**, 13*R**, 14*S** and 1'*R**.



Figure 3. (a) Acetonide derivatization of **1**. ¹H and ¹³C NMR chemical shifts indicating a *syn* relationship of the both acetonides. (b) The key ROESY correlations of the acetonide A of **1** showing the relative configuration of the *iso*-butyl group.

In order to address the C-6 and H-6 chemical shift discrepancies, NMR spectroscopic analysis was carried out on the trimethylheptanol derivatives **19**, **38** and **42** (Figure 4) to gain insight into the correct configurations of C-4, C-6 and C-8 in the natural product. The results show that the H-5 and H-7 ¹H chemical shifts of the protons in the two enantiomeric *syn-syn*-trimethylheptanol fragments **42** and **38**, and the *anti-anti*-trimethylheptanol fragment **19** are remarkably different (Figure 4).



Figure 4. ¹H and ¹³C chemical shifts of trimethylheptanol fragments.

The H-5 and H-7 methylene protons in the two enantiomeric *syn-syn*-trimethylhepatanol fragments, **38** and **42**, display ¹H NMR patterns that are consistent with those of the corresponding methylene protons in natural BmcA (eg., H-5a ($\delta_{\rm H}$ 1.39)/H-5b ($\delta_{\rm H}$ 0.89) and H-7a ($\delta_{\rm H}$ 1.31)/H₂-7b ($\delta_{\rm H}$ 0.88)). It was reported⁴ that H-5a ($\delta_{\rm H}$ 1.73) and H-7a ($\delta_{\rm H}$ 1.22) in the

spectrum of natural BmcA are remarkably more deshielded than are H-5b ($\delta_{\rm H}$ 0.98) and H-7b ($\delta_{\rm H}$ 0.95), respectively. In contrast, in the spectrum of the *anti-anti*-trimethylheptanol fragment **19**, the chemical shifts of H-5a ($\delta_{\rm H}$ 1.24)/H-5b ($\delta_{\rm H}$ 1.07) and H-7a ($\delta_{\rm H}$ 1.18)/H-7b ($\delta_{\rm H}$ 1.01) are not that much different. The detailed ¹H NMR analysis of fragments **19**, **38** and **42**, led us to the conclusion that *syn-syn* relative configuration of the three methyl groups is more plausible for natural BmcA than the proposed *anti-anti* relative stereochemistry. A combination of the findings outlined above led us to propose the two plausible alternative stereostructures, **1a** and **1b**, (Figure 5) for natural BmcA.



Figure 5. Proposed and revised structure of BmcA.

To resolve this issue and, consequently, assign the correct stereostructure to natural BmcA, we conducted total syntheses of both 1a and 1b. As depicted retrosynthetically in Scheme 5, the plan for these syntheses involved joining 39 and 43 with aldehyde 34 using the alkynylation protocol.¹⁵ Fragment Carreira was prepared 3,5from bis(methoxymethoxy)benzaldehyde (4a) by employing a route similar to the one used to generate 12. The redesigned alkyne fragments, 39 and 43 were synthesized starting with the common chiral precursor 36 which in turn was stereoselectively prepared from commercially available cis-4,6-dimethylcyclohexan-1,3-dione by employing an approach that uses enzymatic desymmetrization, Wittig olefination and Evans asymmetric alkylation.

Scheme 5. Retrosynthetic analysis of two possible structures (1a and 1b) of BmcA



Synthesis of **34** commenced with preparation of MOM-protected aldehyde **4a** from **2** (78%, 3 steps). Anti aldol reaction¹⁹ between amide **5** and MOM-protected aldehyde **4a** in presence of magnesium chloride produced hydroxyamide **27** (72%, dr = 97:3 by NMR). The *anti* aldol product **27** was then used in a reaction sequence that mimics the one used to prepare **12** to provide revised aldehyde fragment **34**. Accordingly, *anti* aldol product **27** was protected as its TBS ether **28** (88%). Amide group in **28** was reduced by DIBAL-H to get alcohol **29** which was subsequently oxidized to aldehyde **30** using IBX (76%, 2 steps). Brown allylation⁹ of aldehyde **30** afforded homoallylic alcohol **31** (74%, dr = 90:10 by NMR).¹⁰ It is noteworthy to mention that the diastereomeric mixture was separated in the next stage of the reaction. Deprotection of TBS group in **31** afforded 1,3-diol **32** followed by acetonide protection to provide **33** (89%, 2 steps). Dihydroxylation followed by oxidative cleavage of olefin **33** provided revised aldehyde fragment **34** (70%, 2 steps) (Scheme 6).

Scheme 6. Synthesis of revised aldehyde fragment 34 (anti configuration)

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Both the revised alkyne fragments (**39** and **43**) derived from a known precursor **36** which was synthesized from **35** by following a reported protocol.²⁰ Fragment **35**, synthesized by using a known four step protocol starting with *cis*-4,6-dimethylcyclohexan-1,3-dione,²¹ was transformed to alcohol **36**. Benzyl protection of hydroxyl group in **36** was carried out to afford **37** (82%). TBS group of **37** was deprotected to get primary alcohol **38** (85%) which was then subsequently oxidized under Swern condition¹³ followed by Bestmann-Ohira reaction¹⁴ to generate alkyne fragment **39** with all *syn* configuration (69%, 2 steps). On the other hand, acetyl protection of **36**, followed by deprotection of TBS ether delivered primary alcohol **40** (78%, 2 steps). Benzyl protection of hydroxyl group in **40** was done to afford **41** (72%), which was hydrolyzed under basic condition to afford primary alcohol **42** (95%). Primary alcohol **42** underwent Swern oxidation¹³ followed by Bestmann-Ohira reaction¹⁴ to generate alkyne fragment **43** with all *syn* configuration (66%, 2 steps) (Scheme 7).

Scheme 7. Synthesis of revised alkyne 39 and 43



Carreira alkynylation¹⁵ of **34** and **43** occurred to form propargylic alcohol **44** (76%, dr = 95:5 by NMR). Protection of the hydroxyl group in **44** formed TBS ether **45** (82%), which was then subjected to hydrogenation to produce a mixture (76%) of saturated alcohol **46** and its TBS ether elimination product (8:2, confirmed by ¹H NMR analysis) which was inseparable. The mixture of **46** and its TBS ether elimination product was oxidized to generate a crude aldehyde, which was reacted with EtMgBr to produce a secondary alcohol which upon oxidation formed a mixture (68%, 3 steps) of ketone **47** and its TBS ether elimination product which was inseparable. Finally, global deprotection of the mixture was achieved by using PTSA to produce **1a** and 11-deoxy derivative **48** (91%, combined yield) which was separable. Similarly, the BmcA isomer **1b** was synthesized by condensing aldehyde **34** with alkyne **39**.¹⁵ The formed propargylic alcohol **49** (75%, dr = 96:4 by NMR) was converted to its TBS ether **50** (83%), which when subjected to a similar reaction sequence utilized to form **1a** gave BmcA isomer **1b** along with 11-deoxy derivative **51** (41%, 5 steps) (Scheme 8). Comprehensive NMR analysis data (COSY, HSQC, HMBC, ¹H-¹H decoupling, and HETLOC) for **1a** are available in supporting information.

Scheme 8. Synthesis of 1a and 1b



The absolute configurations of the chiral centers on the left part of **1a** were confirmed by using NMR analysis of its acetonide derivatives (acetonide A and B, Figure 6). The ¹³C chemical shifts of the *gem*-dimethyl groups in acetonide A are 20.1 and 30.6 ppm, indicating the existence of a *syn*-relationship between 13-OH and 1'-OH groups. The *gem*-dimethyl groups in acetonide B have ¹³C chemical shifts of 20.3 and 30.7, indicating that 11-OH and 13-OH also have a *syn*-relationship (Figure 6).



Figure 6. Acetonide derivatization of 1a. Chemical shifts indicate a *syn* relationship of the both acetonides.

In addition, analysis of ¹H and 2D NMR spectra of acetonide A, which contains an isobutyl group in the 1,3-dioxane ring, shows that the H-14/H-1' coupling constant is 10.5 Hz. This large coupling shows that an axial-axial and thus *anti*-relationship exists between H-14 and H-1' (Figure 7). As depicted in Figure 7, H-1' ($\delta_{\rm H}$ 4.35) and H-13 ($\delta_{\rm H}$ 3.79) exhibit ROESY

correlations commonly associated with an axially oriented acetonide methyl group ($\delta_{\rm H}$ 1.53). Moreover, H-1'/H₂-15 ($\delta_{\rm H}$ 1.20 and 0.99) and H-13/H₂-15 ROESY correlations establish that the isobutyl group exists in an equatorial position, thus showing that a *syn*-relationship exists between H-1' and H-13. ROESY coupling between and H-16 ($\delta_{\rm H}$ 0.95) and H-1' also support the assignment of the relative configuration in the 1,3-dioxane, thus confirming the 11*R**, 13*R**, 14*R**, and 1'*R** configuration of **1a**. Based on the absolute configuration of C-11 in this acetonide, established by using the modified Mosher's method, the absolute stereochemistry was unequivocally confirmed to be 11*R*, 13*R*, 14*R*, and 1'*R*.



Figure 7. The key ROESY correlations of the acetonide A of **1a** showing the relative configuration of the *iso*-butyl group.

Importantly, the ¹H and ¹³C NMR spectra of **1a**, having 4*R*, 6*R*, 8*R*, 11*R*, 13*R*, 14*R*, and 1'*R* configurations, were virtually identical to those of natural BmcA even though the reported⁴ NMR spectra of natural BmcA include unidentified impurities (Figure S163). In contrast, the NMR data of **1b**, possessing 4*S*, 6*S*, 8*S*, 11*R*, 13*R*, 14*R*, and 1'*R* configurations, do not match those for natural BmcA. These observations strongly suggest that the relative configurations of BmcA need to be revised to be 4*R**, 6*R**, 8*R**, 11*R**, 13*R**, 14*R**, and 1'*R**. The optical rotation of synthetic **1a** is +10.0 (c 0.40, MeOH) whereas that of isolated natural BmcA [α]²⁰_D is -12.0 (c 0.20, MeOH). The magnitude of the two optical rotation values (**1a** and natural BmcA) was very similar but the sign was opposite, which suggests that synthetic **1a** would be enantiomer of the

natural product. However, because the ¹H NMR spectrum (Figure S163)⁴ of natural BmcA shows that it contains many unidentified impurities the reported optical rotation is questionable. As previously mentioned, the absolute configuration of natural BmcA was arbitrarily assigned in the original report. Owing to the impurities in natural BmcA and the uncertainty of the reported optical rotation value, the absolute stereochemistry of BmcA was unclear to us. Aggarwal *et. al.*^{6c} reported that synthetic **1a** is enantiomer of the natural product while our manuscript was under review. In the effort described above, we achieved the first total synthesis of correct enantiomer (**1a**) of BmcA. By using ¹H, ¹H-¹H decoupling, and HETLOC NMR experiments performed with a 900 MHz spectrometer, we have performed *J*-based configuration analysis (Figure 8) for **1a** and clarified the misinterpreted *J*-based configuration analysis in the previous report, which was possibly a result of overlapped ¹H signals in a lower field region of the ¹H NMR caused by using a 700 MHz instrument.

In order to revise the reported⁴ configurations at stereocenters of natural baulamycin A, we focused to clarify what caused misinterpretation in *J*-based configuration analysis in the previous⁴ report. The ¹H, ¹H-¹H decoupling, and HETLOC NMR experiments of **1a** performed in 900 MHz NMR spectrometer enabled us to obtain accurate ${}^{3}J_{HH}$, ${}^{3}J_{CH}$, and ${}^{2}J_{CH}$ values and compare them with those⁴ in literature (Figure 8). Based on our careful measurement of ${}^{3}J_{HH}$, ${}^{3}J_{CH}$, and ${}^{2}J_{CH}$ values and *J*-based configuration analysis, we identified critical errors in the previous⁴ report. Besides minor errors like alleged typos, wrong interpretation of *J* based configuration analysis of the stereogenic centers around C-6 and C-14 resulted in wrong assignments for the configurations of C-6 and C-14. First, we corrected ${}^{3}J_{H14,H13}$ value from 7.7 Hz to 4.0 Hz based on the ¹H-¹H decoupling experiment at H-15, which was not performed in the previous⁴ report. Furthermore, ${}^{2}J_{C13H14}$ value was revised from 7.0 Hz to 2.0 Hz, establishing the

rotamer in Figure 8-b. Additional decoupling experiments were performed at H-10, H-11, H-12, and H-13 to correct the ${}^{3}J_{H13 H12a}$ (7.2 Hz to 3.5 Hz), ${}^{3}J_{H13 H12b}$ (3.1 Hz to 9.5 Hz), ${}^{3}J_{H11 H12a}$ (8.0 Hz to 4.5 Hz), and ${}^{3}J_{H11 H12b}$ (3.2 Hz to 8.5 Hz) values. Careful analysis of HETLOC data enabled to identify ${}^{2}J_{C13,H12a}$, ${}^{2}J_{C13,H12b}$, and ${}^{2}J_{C11,H12a}$ as 1.0, 4.5, and 1.5 Hz, which were misassigned as 6.2, 1.8, and 6.8 Hz respectively in the previous⁴ report. The anti-relationship between H-13 and H-12b and large value of ${}^{2}J_{C13 H12b}$ constructed the revised rotamer depicted in Figure 8-c. The H-11/H-13, H-12a/H-13, H-12a/H-14, and H-12b/H-14 ROESY correlations supported this revised rotamer. The small coupling constant (4.5 Hz) of H-11/H-12a and the large coupling constant (8.5 Hz) of H-11/H-12b, which were previously reported in the opposite way [large (8.0 Hz) for ${}^{3}J_{H11,H12a}$ and small (3.2 Hz) for ${}^{3}J_{H11,H12b}$] were key evidence for the construction of the revised rotamer in Figure 8-d. Furthermore, several critical errors of coupling constants in the previous⁴ report were found in C-7-C-6 (Figure 8-i) and C-5-C-4 (Figure 8-k) rotamers. For the extraction of accurate coupling constants, a suite of ¹H-¹H decoupling NMR experiments were performed for H₂-7, H-6, H₂-5, and H-4. Based on the newly measured ${}^{3}J_{\rm HH}$ values, ${}^{3}J_{\text{H7a H6}}$ (3.2 Hz) and ${}^{3}J_{\text{H7b H6}}$ (3.0 Hz) were revised to 8.5 and 6.6 Hz, respectively. ${}^{3}J_{\text{C5H7b}}$ and ${}^{3}J_{C20H7b}$ values (6.0 Hz and 3.5 Hz), which were not reported in the previous⁴ report, were also clearly measured in our HETLOC NMR data. Thus, anti-relationships between H-7a/H-6 and C-5/H-7b were established in Figure 8-i. The gauche relationships of H-7b/H-6, C-8/H-6, C-5/H-7a, C-20/H-7a, and C-20/H-7b were also identified by the coupling constants, completing the rotamer depicted in Figure 8-i. Additional proton decoupling experiments for H-4, H-5a, H-5b, H₃-19 revealed that reported⁴ ${}^{3}J_{H5a H4}$ (3.2 Hz) and ${}^{3}J_{H5b H4}$ (8.0 Hz) values had to be revised to 8.5 and 5.0 Hz, respectively. Careful analysis of HETLOC data resulted in the correction of ${}^{3}J_{C21,H5a}$ (11.0 Hz) to 4.0 Hz. Thus, the *anti*-relationship between H-5a and H-4 was established

in Figure 8-k. The relationships between H-5b/H-4, C-6/H-4, C21/H-5a, and C-21/H-5b were determined as *gauche*, completing the rotamer depicted in Figure 8-k. Consequently, our comprehensive *J*-based configuration analysis utilizing ¹H, ¹H-¹H decoupling and HETLOC NMR experiments at high fields (800/900 MHz) allowed for the revision of J_{HH} and J_{CH} , thus confirming the configurations of synthetic baulamycin A (1a). In the previous⁴ report of natural baulamycin A, the coupling constants were measured at lower magnetic field without performing ¹H-¹H decoupling experiments. This may have caused the mismeasurement of the coupling constants and thus resulted in the confusion for establishing the relative configurations because of highly overlapped ¹H signals in baulamycin A. Errors in coupling constant measurement in the previous report⁴ are comprehensively noted in Figure S164.



Figure 8. J-based configuration analysis of 1a. Arrows represents ROESY correlations.

As part of these studies, twelve BmcA derivatives, including enantiomer of BmcA (1a), were subjected to SAR analysis to assess inhibitory activities against SbnE.²² Synthesis of baulamycin A derivatives 55 and 57 is depicted in Scheme 9. Carreira alkynylation¹⁵ between aldehyde fragment 12a and benzyl-protected alkyne fragment 52 derived from 7-octyne-1-ol was achieved to deliver propargylic alcohol 53 (74%, dr = 96:4 by NMR). MOM and acetonide group in 53 was deprotected by using PTSA to afford polyhydroxy compound 55 (84%).

Propargylic hydroxyl group in **53** was protected as its TBS ether **54** (81%). Hydrogenation of **54** using Pd/C was carried out to provide saturated primary alcohol **56** (81%).²³ A similar sequence of reactions like baulamycin A preparation was adopted to deliver **57**, a derivative of baulamycin A (61%, 4 steps). Baulamycin A derivatives **55** and **57** were submitted for SAR study (Table 1).

Scheme 9. Synthesis of baulamycin A derivatives 55 and 57



Synthesis of other baulamycin A derivatives **58**, **59**, **60** and **61** are outlined in Scheme 10. All of protecting groups (MOM, acetonide, and TBS) in **21a**, **23a** and **23'** were deprotected to afford compound **58**, **59** and **60** respectively and similar sequence of reactions (oxidation, Grignard reaction, oxidation, deprotection) like baulamycin A synthesis were applied on **23'** to provide **61** as a baulamycin A derivative (60%, 4 steps) (Scheme 10).

Scheme 10. Synthesis of baulamycin A derivatives 58, 59, 60 and 61



SbnE inhibition assay results, obtained by employing a malachite green-based method²⁴ are outlined in Table 1. In contrast to Sherman and co-workers report that natural BmcA has an IC_{50} value of 4.8 μ M,⁴ **1** possessing the originally proposed stereostructure of this substance has virtually no activity ($IC_{50} > 50 \mu$ M). In contrast, **1a**, whose stereostructure is now known to be that of enantiomer of natural BmcA, has an IC_{50} value of 14.4 μ M.

Compound	$IC_{50}(uM)$	Compound	IC ₅₀ (uM)
	>50		20.98
	>50		25.73
	14.40		>50
	47.27		27.77
	>50		7.16

Table 1: IC₅₀ values of BmcA derivatives against SbnE.



The results of the SAR study show that the existence of branched methyl groups on the hydrocarbon tail (C-1 through C-10) has little to no effect on inhibitory activity. Based on a comparison of the IC₅₀ values of **1**, **26**, **57** and **59** with those of **1a** and **1b**, the (R)-configuration at C-14 seems to be required for inhibitory activity for BmcA derivatives having three hydroxyl groups at C-11, C-13 and C-1' positions and saturated hydrocarbon tails (C-1 through C-10). However, BmcA derivatives **48**, **51**, **60** and **61** that do not contain the C-11 hydroxyl group possess inhibitory activities regardless of the stereochemistry at C-14. Interestingly, the acetylene derivatives **55** and **58** exhibit appreciable potencies despite having the (S)-configuration at C-14 and three hydroxyl groups at C-11, C-13 and C-1'. These compounds contain a chain-rigidifying modification caused by incorporation of a triple bond between C-9 and C-10, which may contribute to control of the hydrocarbon tail orientation for optimal interaction in the active site of SbnE. It should be noted that **55** has the highest potency among all of the tested derivatives. A more comprehensive SAR study probing the consequences of specific stereochemical and structural changes is underway and will be discussed in due course.

Conclusions

In summary, we have accomplished a synthesis of the earlier reported stereostructure of BmcA and found that discrepancies exist in the spectroscopic properties between the synthetic material and the natural BmcA. Two possibly correct stereostructures for BmcA, identified on the basis of comprehensive NMR analysis, were prepared by total synthesis. The synthetic pathways employ stereoselective Evans Aldol, Brown allylation and Carreira alkynylation reactions, and a prolinol

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amide derived alkylation/desymmetrization protocol for installing three chiral centers at C-4, C-6 and C-8. Comprehensive NMR studies with these two synthetic materials (1 and 1a) enabled the assignment of the correct enantiomer (1a) of BmcA. Moreover, inhibitory activities of novel BmcA derivatives against SbnE were determined. The SAR data provide insight about how the configuration of C-14, the existence of OH-11 and chain-rigidifying group affect inhibitory activities against SbnE. It is worth noting that the propargylic alcohol 55, having 14*S* configuration, exhibits the highest SbnE inhibitory activity even though it lacks three methyl groups at C-4, C-6 and C-8 and a ketone group, suggesting that the presence of a chainrigidifying triple bond contributes to the inhibitory activity of members of this class of substances. This structural revision with efficient synthetic strategy and SAR study lay the groundwork for discovering new BmcA derivatives that overcome drug resistant pathogens that cause life-threatening infections.

Experimental Section

General Experimental Methods: All reactions were carried out under an inert atmosphere of argon or nitrogen using standard syringe, septa, and cannula techniques unless otherwise mentioned. Reactions were monitored by using TLC with 0.25 mm E. Merck precoated silica gel plates (60 F_{254}). Reaction progress was monitored by using TLC with a UV lamp, ninhydrin, or *p*-anisaldehyde stain for detection purposes. Commercially available reagents were used without further purification. All solvents were purified by using standard techniques. Purification of products was carried out by using silica gel column chromatography using Kieselgel 60 Art. 9385 (230-400 mesh). The purity of all compounds was determined to be over 95% by using a Waters LCMS system (Waters 2998 Photodiode Array Detector, Waters 3100 Mass Detector,

Waters SFO System Fluidics Organizer, Water 2545 Binary Gradient Module, Waters Reagent Manager, Waters 2767 Sample Manager) using SunFireTM C18 column (4.6 x 50 mm, 5 µm particle size): solvent gradient = 60% (or 95%) A at 0 min, 1% A at 5 min. Solvent A = 0.035%TFA in H₂O: Solvent B = 0.035% TFA in MeOH: flow rate: 3.0 (or 2.5) mL/min. ¹H and ¹³C NMR spectra were obtained using a Bruker 400 MHz FT-NMR (400 MHz for ¹H and 100 MHz for ¹³C) spectrometer. ¹H, ¹³C, and 2D nuclear magnetic resonance (NMR) spectra were also recorded on Bruker Avance 600-MHz spectrometers at the National Center for Inter-university Research Facilities at Seoul National University (NCIRF) and on a Bruker Avance II 900-MHz NMR spectrometer at the Korea Basic Science Institute (KBSI) at Ochang. Standard abbreviations are used for denoting the signal multiplicities. Infrared spectra were measured on FT-IR Nicolet iS10 spectrometer. Samples were recorded under neat or as KBr optics. Highresolution mass spectra (HRMS) were recorded on a QTOF mass spectrometer. Optical rotations were measured by a JASCO P-200 polarimeter with a 1 cm cell. UV spectra were acquired with a Perkin Elmer Lambda 35 UV/VIS spectrometer. Electrospray ionization (ESI) low-resolution LC/MS data were acquired on an Agilent Technologies 6130 quadrupole mass spectrometer coupled with an Agilent Technologies 1200-series HPLC. Semi-preparative HPLC separations were performed with an HPLC system composed of a Gilson 305 pump and a Gilson UV/VIS-155 detector.

Methyl 3,5-Bis(methoxymethoxy)benzoate (3a). To a solution of methyl 3,5dihydroxybenzoate 2 (10 g, 59.4 mmol) in anhydrous CH_2Cl_2 (200 mL) was added diisopropylethylamine (41.8 mL, 240 mmol), followed by MOMCl (10.6 mL, 120 mmol) dropwise at 0 °C. The mixture was warmed to room temperature and stirred for 12 h. The mixture was then quenched with water (30 mL) and diluted with CH_2Cl_2 (50 mL). The organic

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layer was separated and washed with saturated NaHCO₃ solution (30 mL) and extracted with CH₂Cl₂ (3 x 100 mL). The combined organic layer was washed with brine, dried over MgSO₄ and concentrated under reduced pressure. The resulting residue was subjected to silica gel column chromatography to afford compound **3a** (13.9 g, 92%) as colorless oil. [Known literature method]²⁵; $R_f = 0.8$ (30% EtOAc/hexane).^{25 1}H NMR (400 MHz, CDCl₃): δ 7.35 (d, J = 2.3 Hz, 2H), 6.90 (t, J = 2.3 Hz, 1H), 5.18 (s, 4H), 3.88 (s, 3H), 3.47 (s, 6H); LCMS (ESI): 257 (M + H)⁺.

3,5-Bis(methoxymethoxy)benzaldehyde (4a). To a stirred solution of methyl 3,5bis(methoxymethoxy)benzoate **3a** (12 g, 46.4 mmol) in anhydrous THF (120 mL) was added dropwise a solution of lithium aluminium hydride (56 mL of a 1.0 M solution in THF, 55.7 mmol). The mixture was stirred at ambient temperature for 4 h and treated dropwise sequentially with water (1 mL), 15% aqueous NaOH (1 mL) and water (3 mL). The mixture was stirred for additional 1 h and filtered and the resulting solid was washed with THF. The filtrate was concentrated to provide 3,5-bis(methoxymethoxy)benzyl alcohol as a colorless liquid which was directly used in the next step without further purification.

To a solution of PCC (13.1 g, 61.2 mmol) and sodium acetate (17.3 g, 21.4 mmol) in CH₂Cl₂ (100 mL) was added a solution of 3,5-bis(methoxymethoxy)benzyl alcohol (7 g, 30.6 mmol) in CH₂Cl₂ (8 mL). The mixture was stirred under nitrogen for 2 h and treated with ether (300 mL). The brown mixture was filtered through filter paper over Celite. The filtrate was concentrated to provide brown oil which was subjected to silica gel column chromatography to afford compound **4a** (8.9 g, 85% over two steps) as a colorless oil. [known literature method]²⁵; $R_f = 0.4$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.89 (s, 1H), 7.19 (d, J = 2.3 Hz, 2H), 6.96 (t, J

= 2.3 Hz, 1H), 5.19 (s, 4H), 3.47 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 191.5, 158.7, 138.4, 111.1, 110.3, 94.4, 77.3, 77.0, 76.6, 56.1; LCMS (ESI): 227 (M + H)⁺.

(R)-4-Benzyl-3-(4-methylpentanoyl)oxazolidin-2-one (5). To a stirred solution of 4methylvaleric acid (4 g, 34.4 mmol) in anhydrous THF (100 mL) at -20 °C was added Et₃N (12 mL, 86.1 mmol) followed by pivaloyl chloride (4.2 mL, 34.4 mmol). The mixture was stirred for 1 h at -20 °C and treated sequentially with LiCl (2.2 g, 51.6 mmol) and (R)-4-benzyloxazolidin-2-one (6.7 g, 37.8 mmol) at same temperature. The mixture was stirred for 1 h at -20 °C and for 2 h at 0 °C and guenched with saturated NH₄Cl solution (30 mL). The mixture was diluted with EtOAc (50 mL) and extracted with EtOAc (3 x 50 mL). The combined organic layer was washed with brine, dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford 5 (8.5 g, 90%) as a yellow liquid. $R_f =$ 0.3 (30% EtOAc/hexane); $[\alpha]_D^{24} = +45.4$ (c 0.55, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.31 (m. 2H), 7.29-7.25 (m. 1H), 7.22-7.19 (m. 2H), 4.69-4.64 (m. 1H), 4.21-4.09 (m. 2H), 3.28 (dd, J = 13.4, 3.3 Hz, 1H), 3.02-2.86 (m, 2H), 2.76 (dd, J = 13.4, 9.6 Hz, 1H), 1.70-1.52 (m, 3H),0.93 (d, J = 6.4 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃); δ 173.6, 153.4, 135.3, 129.4, 128.9, 127.3, 66.1, 55.1, 37.9, 33.6, 33.1, 27.6, 22.3(2); HRMS (ESI): calcd. for $C_{16}H_{21}O_3NNa$ [M + Na]⁺ 298.1419; found 298.1415.

(R)-4-Benzyl-3-((R)-2-((R)-(3,5-dimethoxyphenyl)(hydroxy)methyl)-4-

methylpentanoyl)oxazolidin-2-one (6). To a 0.2-0.5 M solution of **5** (9.0 g, 32.7 mmol) in anhydrous CH_2Cl_2 (100 mL) under argon at 0 °C was added dibutylboron triflate (39.2 mL of a 1.0 M solution in CH_2Cl_2 , 39.2 mmol), followed by diisopropylethylamine (7.4 mL, 42.5 mmol) dropwise. After stirred for 30 min at 0 °C, the mixture was cooled to -78 °C and treated with benzaldehyde **4** (6 g, 36 mmol). The mixture was then stirred for 30 min at -78 °C and for 1.5 h

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at room temperature. The boron complex was quenched with phosphate buffer (pH~7, 50 mL) and oxidized with a mixture of 30% H₂O₂ and methanol (100 mL of a 2:1 solution) for 1 h at 0 ^oC. The mixture was stirred for 1 h at room temperature and the resulting mixture was concentrated to slurry. The residue was extracted with ether (3 x 100 mL) and the combined organic extracts were washed with 5% aqueous NaHCO₃ solution (30 mL), brine (30 mL), and then dried over MgSO₄, filtered and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford 6 (13.7 g, 86%) as a colorless liquid. R_f = 0.2 (20% EtOAc/hexane); $[\alpha]_{D}^{24} = -206.6$ (c 1.08, CHCl₃); ¹H NMR (400 MHz, CDCl₃); δ 7.34-7.27 (m, 3H), 7.19-7.17 (m, 2H), 6.56 (d, J = 2.1 Hz, 2H), 6.35 (t, J = 2.3 Hz, 1H), 4.76 (q, J = 3.1 Hz, 1H), 4.49-4.45 (m, 1H), 4.43-4.36 (m, 1H), 4.01 (dd, J = 9.0, 2.2 Hz, 1H), 3.77 (s, 6H), 3.22 (dd, J = 13.4, 3.4 Hz, 1H), 2.68 (dd, J = 13.4, 9.8 Hz, 1H), 2.35 (d, J = 3.3 Hz, 1H), 2.04-1.94 (m, 1H), 1.58-1.49 (m, 3H), 0.91 (d, J = 2.6 Hz, 3H), 0.89 (d, J = 2.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ174.6, 160.4, 152.9, 144.1, 135.0, 129.2, 128.7, 127.1, 103.7, 100.1, 75.7, 65.7, 55.5, 55.1, 48.4, 37.7, 36.7, 26.5, 23.5, 21.7; HRMS (ESI): calcd. for C₂₅H₃₁O₆NNa $[M + Na]^+$ 464.2049; found 464.2044.

(R)-4-Benzyl-3-((R)-2-((R)-((tert-butyldimethylsilyl)oxy)(3,5-dimethoxyphenyl)methyl)-4-

methylpentanoyl)oxazolidin-2-one (7). To a solution of **6** (8 g, 18.1 mmol) in anhydrous CH₂Cl₂ (50 mL) was added 2,6-lutidine (8.4 mL, 72.4 mmol), followed by TBSOTf (8.3 mL, 36.2 mmol) slowly at 0 °C. The mixture was stirred for 1 h at 0 °C and quenched with water (15 mL). The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3 x 50 mL). The combined organic layer was washed with saturated NaHCO₃ solution (30 mL) and dried over MgSO₄, concentrated under reduced pressure to give crude residue which was subjected to silica gel column chromatography to afford 7 (8.8 g, 88%) as a yellow liquid. $R_f = 0.3$ (30%)

EtOAc/hexane); $[\alpha]_D^{24} = +218.5$ (*c* 1.09, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.32-7.23 (m, 3H), 7.19-7.14 (m, 2H), 6.49 (d, J = 2.6 Hz, 2H), 6.30 (t, J = 2.2 Hz, 1H), 4.54 (d, J = 7.8 Hz, 1H), 4.36-4.31 (m, 1H), 4.17-4.12 (m, 1H), 3.88 (dd, J = 8.9, 1.6 Hz, 1H), 3.74 (s, 6H), 3.48 (t, J = 8.1 Hz, 1H), 3.11 (dd, J = 13.3, 3.2 Hz, 1H), 2.67 (dd, J = 13.3, 9.5 Hz, 1H), 1.99-1.91 (m, 1H), 1.68-1.59 (m, 1H), 0.93 (d, J = 6.4 Hz, 6H), 0.91-0.89 (m, 1H), 0.88 (s, 9H), 0.03 (s, 3H), -0.18 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 174.3, 160.3, 152.8, 145.1, 135.4, 129.4, 128.8, 127.2, 104.2, 100.2, 77.8, 65.7, 55.9, 55.4, 50.2, 38.1, 37.9, 26.8, 25.7, 23.7, 21.9, 18.1, -4.6, -5.2; HRMS (ESI): calcd. for C₃₁H₄₅O₆NSiNa [M + Na]⁺ 578.2914; found 578.2918.

(R)-2-((R)-((tert-Butyldimethylsilyl)oxy)(3,5-dimethoxyphenyl)methyl)-4-methylpentanal

(8). To a solution of 7 (8 g, 14.4 mmol) in anhydrous CH_2Cl_2 (100 mL) was added DIBAL-H (25.6 mL, 36 mmol, 20% solution in toluene) slowly for 15 min at -78 °C. The mixture was stirred for 30 min at -78 °C before being quenched with methanol (10 mL) and aqueous saturated sodium potassium tartrate solution (50 mL). The mixture was passed through a small pad of Celite and then extracted with CH_2Cl_2 (3 x 50 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure to get crude residue which was used for next step without further purification.

In the next step, to a solution of crude residue in MeOH: H₂O (4:1, 40 mL) was added solid K₂CO₃ (8 g, 58 mmol) at 0 °C and the mixture was stirred for 1 h at the same temperature. The mixture was filtered through a pad of Celite and the filter cake was washed with CH₂Cl₂ (20 mL). The filtrate was washed with water (10 mL) and brine (10 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **8** (3.3 g, 60% over two steps) as a colorless liquid. $R_f = 0.8$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +51.8$ (*c* 0.89, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 9.67 (d, *J*

= 2.8 Hz, 1H), 6.43 (d, J = 2.1 Hz, 2H), 6.34 (t, J = 2.2 Hz, 1H), 4.93 (d, J = 4.7 Hz, 1H), 3.77 (s, 6H), 2.59-2.52 (m, 1H), 1.74-1.66 (m, 1H), 1.49-1.38 (m, 1H), 1.33-1.25 (m, 1H), 0.90 (s, 9H), 0.83 (d, J = 6.5 Hz, 3H), 0.74 (d, J = 6.5 Hz, 3H), 0.03 (s, 3H), -0.13 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 205.0, 160.6, 144.7, 104.3, 99.3, 74.8, 58.2, 55.3, 33.0, 25.7, 23.3, 21.7, 18.1, -4.6, -5.3; HRMS (ESI): calcd. for C₂₁H₃₆O₄SiNa [M + Na]⁺ 403.2281; found 403.2272.

(4R,5S)-5-((R)-((tert-Butyldimethylsilyl)oxy)(3,5-dimethoxyphenyl)methyl)-7-methyloct-1-

en-4-ol (9). To a stirred solution of (+)-ipc₂BOMe (3.3 g, 10.4 mmol) in anhydrous Et₂O (20 mL) was added allylmagnesium bromide (7.9 mL of 1.0 M solution in ether, 7.89 mmol,) at 0 °C. The mixture was stirred at room temperature for 1 h before being cooled to -78 °C. The mixture was treated dropwise with aldehyde 8 (2.0 g, 5.26 mmol) at -78 °C and stirred for 1 h at same temperature and allowed to warm to room temperature. The mixture was treated with an aqueous solution of NaOH (2 M in H₂O, 10 mL) and 30% H₂O₂ solution (5 mL) at 0 °C. The biphasic solution was separated and the aqueous layer was extracted with Et₂O (3 x 30 mL). The combined organic layer was dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 9 (3.2 g, 74%) as a colorless liquid. $R_f = 0.2$ (10% EtOAc/hexane); $[\alpha]_D^{24} = +29.1$ (c 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.48 (d, J = 2.1 Hz, 2H), 6.33 (t, J = 2.2 Hz,1H), 5.84-5.73 (m, 1H), 5.12-5.06 (m, 2H), 4.78 (d, J = 4.4 Hz, 1H), 3.88 (t, J = 7.5 Hz, 1H), 3.78 (s, 6H), 2.31-2.22 (m, 1H), 2.18-2.11 (m, 1H), 1.67-1.63 (m, 1H), 1.56 (brs, 1H), 1.46-1.38 (m, 1H), 1.29-1.19 (m, 2H), 0.92 (s, 9H), 0.78 (d, J = 6.2 Hz, 3H), 0.72 (d, J = 6.2 Hz, 3H), 0.07 (s, 3H), -0.17 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 160.5, 146.2, 135.6, 117.2, 104.3, 99.1, 77.8, 72.4, 55.2, 48.0, 39.6, 32.3, 27.0, 25.8, 23.2, 22.4, 18.0, -4.5, -5.1; HRMS (ESI): calcd. for $C_{24}H_{42}O_4SiNa [M + Na]^+$ 445.2750; found 445.2745.

(1*R*,2*S*,3*R*)-1-(3,5-Dimethoxyphenyl)-2-isobutylhex-5-ene-1,3-diol (10). To a stirred solution of silyl compound **9** (3 g, 7.10 mmol) in anhydrous THF (20 mL) was added TBAF (12.8 mL of 1 M solution in THF, 12.79 mmol) at 0 °C. The mixture was stirred for 4 h at room temperature, diluted with saturated NaHCO₃ solution (5 mL). Organic layer was separated and washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **10** (2.0 g, 93%) as a colorless liquid. $R_f = 0.15$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +1.9$ (*c* 0.69, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.51 (d, *J* = 2.2 Hz, 2H), 6.33 (t, *J* = 2.2 Hz, 1H), 5.87-5.77 (m, 1H), 5.18-5.14 (m, 2H), 4.97 (brs, 1H), 4.04-4.01 (m, 1H), 3.78 (s, 6H), 3.25 (d, *J* = 1.7 Hz, 1H), 2.56 (d, *J* = 2.2 Hz, 1H), 2.32-2.21 (m, 2H), 1.74-1.70 (m, 1H), 1.44-1.35 (m, 1H), 1.28-1.21 (m, 1H), 1.01-0.93 (m, 1H), 0.72 (d, *J* = 6.6 Hz, 3H), 0.52 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 160.6, 146.0, 135.0, 118.2, 103.7, 99.1, 78.1, 75.5, 55.3, 47.0, 39.9, 30.2, 27.3, 22.6, 22.5; HRMS (ESI): calcd. for C₁₈H₂₈O₄Na [M + Na]⁺ 331.1885; found 331.1885.

(4*R*,5*S*,6*R*)-4-Allyl-6-(3,5-dimethoxyphenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxane (11). To a stirred solution of 10 (2.0 g, 6.49 mmol) in CH₂Cl₂ (20 mL) was added 2,2-dimethoxy propane (4 mL, 30 mmol) followed by PTSA (cat.) at 0 °C. The mixture was allowed to warm to room temperature and stirred for 30 min before being quenched with water (10 mL). The organic layer was extracted with CH₂Cl₂ (3 x 10 mL) and the combined organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford 11 (2.2 g, 96%) as a colorless liquid. $R_f = 0.8$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +49.4$ (*c* 0.75, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.50 (dd, *J* = 2.4, 0.6 Hz, 2H), 6.33 (t, *J* = 2.2 Hz, 1H), 5.91-5.80 (m, 1H), 5.15-5.05 (m, 2H), 4.99 (d, *J* = 1.8 Hz, 1H), 4.10 (dt, *J* = 7.8, 2.1 Hz, 1H), 3.79 (s, 6H), 2.40-2.29 (m, 1H), 2.22-2.14 (m, 1H),

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1.50-1.49 (m, 1H), 1.50 (s, 3H), 1.49 (s, 3H), 1.25-1.14 (m, 2H), 0.88-0.78 (m, 1H), 0.69 (d, J = 6.5 Hz, 3H), 0.45 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 160.6, 143.7, 135.1, 116.7, 103.9, 99.3, 98.8, 75.2, 74.2, 55.3, 40.9, 37.5, 30.2, 29.9, 27.2, 22.8, 22.5, 19.5; HRMS (ESI): calcd. for C₂₁H₃₂O₄Na [M + Na]⁺ 371.2198; found 371.2181.

2-((4R,5S,6R)-6-(3,5-Dimethoxyphenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-

yl)acetaldehyde (12). To a stirred solution of compound 11 (2 g, 5.75 mmol) in THF:H₂O (3:1, 20 mL) was added NMO (1.2 g, 10.35 mmol) followed by osmium tetroxide (600 μ L, 0.06 mmol) at 25 °C. The mixture was stirred at the same temperature for 2 h and quenched with aqueous Na₂S₂O₄ solution (5 mL). The mixture was extracted with EtOAc (3 x 10 mL) and the combined organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure to get crude diol which was directly used for next step without further purification.

To a stirred solution of crude diol in THF:H₂O (2:1, 20 mL) was added NaIO₄ (1.9 g, 9.2 mmol) portion wise at 0 °C. The mixture was stirred for 2 h at room temperature and then diluted with saturated NaHCO₃ solution (5 mL). The layers were separated and the organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **12** (1.37 g, 68% over two steps) as a colorless liquid. $R_f = 0.4$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +172.1$ (*c* 3.47, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 9.79 (t, *J* = 1.7 Hz, 1H), 6.49 (d, *J* = 1.7 Hz, 2H), 6.34 (t, *J* = 2.2 Hz, 1H), 5.07 (d, *J* = 2.0 Hz, 1H), 4.71-4.67 (m, 1H), 3.78 (s, 6H), 2.71 (ddd, *J* = 16.8, 8.8, 1.8 Hz, 1H), 2.45 (ddd, *J* = 16.9, 4.5, 1.5 Hz, 1H), 1.59-1.55 (m, 1H), 1.52 (s, 3H), 1.47 (s, 3H), 1.22-1.17 (m, 2H), 0.83-0.71 (m, 1H), 0.66 (d, *J* = 6.4 Hz, 3H), 0.45 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 200.9, 160.6, 143.1, 103.8, 99.6, 98.9, 74.8, 69.0, 55.3, 47.1, 40.8, 30.4,

29.7, 27.4, 22.7, 22.3, 19.4; HRMS (ESI): calcd. for $C_{20}H_{30}O_5Na [M + Na]^+$ 373.1991; found 373.1981.

2-((4R,5S,6R)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4yl)acetaldehyde (12a). ¹H NMR (400 MHz, CDCl₃): δ9.79 (t, *J* = 1.7 Hz, 1H), 6.67 (dd, *J* = 2.2, 0.7 Hz, 2H), 6.60 (t, *J* = 2.2 Hz, 1H), 5.15 (s, 4H), 5.07 (d, *J* = 1.9 Hz, 1H), 4.68 (ddd, *J* = 8.9, 1.4 Hz, 1.4 H

6.2, 1.9 Hz, 1H), 3.45 (s, 6H), 2.70 (ddd, J = 16.8, 8.9, 1.9 Hz, 1H), 2.44 (ddd, J = 16.8, 5.7, 1.5 Hz, 1H), 1.59-1.54 (m, 1H), 1.51 (s, 3H), 1.47 (s, 3H), 1.18 (dd, J = 6.9, 4.4 Hz, 2H), 0.84-0.74 (m, 1H), 0.68 (d, J = 6.4 Hz, 3H), 0.46 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ
200.9, 158.0, 143.2, 107.2, 103.2, 99.6, 94.4, 74.6, 69.0, 55.9, 47.1, 40.7, 30.3, 29.7, 27.4, 22.7, 22.1, 19.5; HRMS (ESI): calcd. for C₂₂H₃₄O₇Na [M + Na]⁺ 433.2202; found 433.2187.

(2R,4R)-5-(Benzyloxy)-1-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2,4-dimethylpentan-1-one

(15). To a solution of alcohol 13 (10 g, 55.5 mmol) in CH₂Cl₂ (150 mL) was added successively triphenylphosphine (17.5 g, 66.6 mmol) and imidazole (5.6 g, 83.2 mmol) followed by iodine (25 g, 99.9 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C and quenched with saturated Na₂S₂O₃ solution (50 mL). The organic layer was extracted with Et₂O (3 x 100 mL) and the combined extract was washed with brine (20 mL), dried over MgSO₄, filtered and concentrated under reduced pressure to afford the crude iodo compound 14 as a colorless liquid which was used for next step without further purification; ¹H NMR (400 MHz, CDCl₃): δ 7.39-7.26 (m, 5H), 4.53 (s, 2H), 3.40 (dd, *J* = 9.3, 5.1 Hz, 1H), 3.38-3.28 (m, 3H), 1.85-1.74 (m, 1H), 1.00 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.3, 128.3, 127.6, 74.1, 73.2, 35.1, 17.6, 13.9; LCMS (ESI): 291 (M + H)⁺.

To a stirred solution of LDA (83.3 mmol) in anhydrous THF (0.5 M) was added dropwise (S)prolinol propionamide (4.8 g, 30.6 mmol) at 0 $^{\circ}$ C. The mixture was stirred for 30 min at room

temperature, followed by the addition of HMPA (13.4 mL, 83 mmol). The mixture was cooled to -78 °C and treated dropwise with iodide **14** (11.5, 39.6 mmol) in THF (60 mL) at a rate to maintain the temperature. The mixture was stirred for 1 h at -78 °C and for 3 h at room temperature and then quenched by dropwise addition of saturated NH₄Cl solution (30 mL). The layers were separated and the organic layer was washed with brine (20 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **15** (10.7 g, 85%) as yellow liquid. $R_f = 0.3$ (30% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 5.24 (dd, J = 7.6, 2.1 Hz, 1H), 4.46 (dd, J = 18.7, 12.0 Hz, 2H), 4.25-4.15 (m, 1H), 3.67-3.58 (m, 1H), 3.57-3.47 (m, 1H), 3.47-3.42 (m, 1H), 3.40-3.32 (m, 1H), 3.30-3.22 (m, 2H), 2.75-2.63 (m, 1H), 2.05-1.67 (m, 5H), 1.60-1.39 (m, 2H), 1.11 (d, J = 6.7 Hz, 3H), 0.93 (d, J = 6.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.1, 138.4, 128.2, 127.5, 127.4, 76.1, 73.0, 67.3, 60.8, 47.4, 37.9, 35.6, 31.3, 28.0 28.0, 24.2, 17.7, 17.4; HRMS (ESI): calcd. for C₁₉H₂₉NO₃Na [M + Na]⁺ 342.2045; found 342.2052.

(2*R*,4*R*)-5-(Benzyloxy)-2,4-dimethylpentan-1-ol (16). A solution of 15 (10 g, 31.3 mmol) in 1 N HCl (50 mL) was heated at reflux for 6 h, then cooled to 0 °C and treated with 15% NaOH solution for 10 min. The mixture was acidified to $pH \sim 3$ and extracted with ether (3 x 100 mL). The combined ether extracts were dried over MgSO₄ and concentrated under reduced pressure to give crude carboxylic acid which was directly used for next step without further purification.

To a stirred solution of the crude acid in anhydrous ether (100 mL) was added dropwise lithium aluminium hydride (56 mL of 1.0 M solution in THF, 55.7 mmol) at 0 °C. The mixture was stirred for 4 h at ambient temperature. To the reaction mixture, water (2 mL), 15% aqueous NaOH (2 mL) and water (5 mL) were added sequentially. After the final addition, stirring was

continued for 1 h, and the mixture was filtered. The solid was washed with THF, and the filtrate was evaporated to provide crude alcohol. The residue was subjected to silica gel column chromatography to give compound **16** (4.79 g, 69% over two steps) as a colorless liquid. $R_f = 0.3$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +41.0$ (*c* 2.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.27 (m, 5H), 4.51 (s, 2H), 3.44 (dd, J = 9.0, 6.4 Hz, 2H), 3.29 (dd, J = 6.4, 3.6 Hz, 2H), 1.96-1.85 (m, 1H), 1.79-1.70 (m, 1H), 1.66 (brs, 1H), 1.29-1.15 (m, 2H), 0.93 (d, J = 6.7 Hz, 3H), 0.89 (d, J = 6.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.4, 128.2, 127.4, 127.4, 76.5, 72.9, 68.6, 37.1, 32.9, 30.5, 16.8, 16.2; HRMS (ESI): calcd. for C₁₄H₂₂O₂Na [M + Na]⁺ 245.1517; found 245.1517.

(2S,4R,6R)-7-(Benzyloxy)-1-((R)-2-(hydroxymethyl)pyrrolidin-1-yl)-2,4,6-trimethylheptan-

1-one (18). To a solution of alcohol **16** (4.5 g, 24.7 mmol) in CH₂Cl₂ (100 mL) was added successively triphenylphosphine (7.8 g, 29.7 mmol) and imidazole (2.5 g, 37 mmol) followed by iodine (11.3 g, 44.6 mmol) at 0 °C. The reaction mixture was stirred for 1 h at 0 °C and quenched with saturated Na₂S₂O₃ solution (50 mL). The organic layer was extracted with ether (3 x 100 mL) and the combined extracts were washed with brine (20 mL). The organic extracts were dried over MgSO₄, filtered and concentrated under reduced pressure to afford the crude iodo compound **17** as a colorless liquid which was used for next step without further purification; ¹H NMR (400 MHz, CDCl₃): δ 7.39-7.26 (m, 5H), 4.52 (s, 2H), 3.38-3.12 (m, 4H), 1.91-1.81 (m, 1H), 1.68-1.55 (m, 1H), 1.36-1.18 (m, 2H), 0.98 (d, *J* = 6.5 Hz, 3H), 0.94 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.6, 128.3, 127.4, 127.4, 76.1, 73.0, 40.7, 32.2, 31.0, 20.2, 18.3, 17.0; LCMS (ESI): 333 (M + H)⁺.

To a stirred solution of LDA (23.3 mmol) in anhydrous THF (0.5 M) was added (R)-prolinol propionamide (1.5 g, 9.3 mmol) at 0 °C slowly dropwise. The resulting solution was stirred at

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room temperature for 30 min, followed by the addition of of HMPA (4 mL, 22.3 mmol). The mixture was cooled to -78 °C and iodide **17** (3.7 g, 11.1 mmol) in THF (30 mL) added dropwise. The reaction mixture was stirred for 1 h at -78 °C and 3 h at room temperature and then quenched by dropwise addition of saturated NH₄Cl solution (10 mL). The layers were separated and the organic layer was washed with brine (20 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **18** (3.44 g, 86%) as a yellow liquid. $R_f = 0.35$ (30% EtOAc/hexane); $[\alpha]_D^{24} = +132.0$ (*c* 2.55, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 4.48 (s, 2H), 4.25-4.19 (m, 1H), 3.67-3.44 (m, 4H), 3.30-3.20 (m, 2H), 2.68-2.59 (m, 2H), 2.03-1.77 (m, 4H), 1.60-1.44 (m, 3H), 1.37-1.29 (m, 1H), 1.25-1.16 (m, 1H), 1.11 (d, *J* = 6.6 Hz, 3H), 1.08-1.01 (m, 1H), 0.88 (d, *J* = 6.6 Hz, 3H), 0.85 (d, *J* =6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.5, 138.7, 128.2, 127.4, 127.4, 76.4, 72.9, 67.8, 61.1, 47.7, 41.4, 41.2, 35.6, 30.7, 28.2, 27.7, 24.4, 19.2, 17.4, 16.8; HRMS (ESI): caled. for C₂₂H₃₅O₃NNa [M + Na]⁺ 384.2514; found 384.2576.

(2*S*,4*S*,6*R*)-7-(Benzyloxy)-2,4,6-trimethylheptan-1-ol (19). A solution of 18 (3.44 g, 9.5 mmol) in 1 N HCl (30 mL) was heated at reflux for 6 h and cooled to 0 °C and then treated with 15% NaOH solution for 10 min. The reaction mixture was acidified to pH ~ 3 and extracted with ether (3 x 50 mL). The combined ether extracts were dried over MgSO₄ and concentrated under reduced pressure to give crude carboxylic acid which was directly used for next step without further purification.

To a stirred solution of crude acid in anhydrous ether (60 mL) at 0 °C was added dropwise lithium aluminium hydride (17 mL of 1.0 M solution in THF, 17.1 mmol). The mixture was stirred for 4 h at ambient temperature and treated dropwise sequentially water (1 mL), 15% aqueous NaOH (1 mL) and water (3 mL). The mixture was stirred for 1 h and filtered and the

resulting solid was washed with THF. The filtrate was concentrated to provide crude alcohol. The residue was subjected to silica gel column chromatography to give compound **19** (1.63 g, 65% over two steps) as a colorless liquid. $R_f = 0.2$ (20% EtOAc/hexane); $[\alpha]_D^{24} = -32.6$ (*c* 0.85, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.26 (m, 5H), 4.50 (s, 2H), 3.47 (dd, J = 10.4, 5.6 Hz, 1H), 3.38 (dd, J = 10.4, 6.6 Hz, 1H), 3.31 (dd, J = 9.0, 5.7, Hz, 1H), 3.22 (dd, J = 9.0, 6.9 Hz, 1H), 1.94-1.82 (m, 1H), 1.78-1.67 (m, 1H), 1.66-1.55 (m, 1H), 1.28-1.13 (m, 2H), 1.09-0.99 (m, 2H), 0.91 (d, J = 6.6 Hz, 3H), 0.87 (d, J = 6.6 Hz, 3H), 0.82 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.7, 128.2, 127.4, 127.3, 76.4, 72.8, 68.8, 41.9, 41.3, 33.0, 30.7, 27.0, 19.0, 17.0, 16.3; HRMS (ESI): calcd. for C₁₇H₂₈O₂Na [M + Na]⁺ 287.1987; found 287.1990.

((((2R,4S,6S)-2,4,6-Trimethyloct-7-yn-1-yl)oxy)methyl)benzene (20). To a solution of oxalyl chloride (0.8 mL, 9.25 mmol) in CH₂Cl₂ (10 mL) was added DMSO (0.78 mL, 11.1 mmol) at -78 °C. The mixture was stirred for 10 min at -78 °C and treated drop-wise with a solution of alcohol 20 (1.63 g, 6.17 mmol) in CH₂Cl₂ (20 mL). The mixture was stirred for 30 min at the same temperature and then treated dropwise with Et₃N (5.1 mL, 37 mmol). The mixture was stirred for 45 min at same temperature and treated with saturated NH₄Cl solution (15 mL). The organic layer was separated and washed with saturated NaHCO₃ solution (10 mL), dried over MgSO₄ and concentrated under reduce pressure to give the crude aldehyde, which was used in the next step without further purification.

To a stirred solution of crude aldehyde and Bestmnn-Ohira reagent (1.7 g, 9.2 mmol) in methanol (10 mL) K_2CO_3 (2.48 g, 18 mmol) was added at 0 °C. The mixture was stirred for 1 h at room temperature and passed through a small pad of Celite and then extracted with CH_2Cl_2 (3 x 10 mL). The combined organic layer was washed with brine (10 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column

chromatography to give compound **20** (1.0 g, 68% over two steps) as a colorless liquid. $R_f = 0.8$ (10% EtOAc/hexane); $[\alpha]_D^{24} = +96.6$ (*c* 0.88, CHCl₃); IR (KBr): 2964, 2927, 2871, 2848, 1453, 1378, 1098, 735, 676, 630 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.27 (m, 5H), 4.51 (s, 2H), 3.33-3.23 (m, 2H), 2.56-2.47 (m, 1H), 2.02 (d, *J* = 2.4 Hz, 1H), 1.95-1.82 (m, 1H), 1.82-1.71 (m, 1H), 1.47-1.36 (m, 1H), 1.34-1.23 (m, 1H), 1.18-1.13 (m, 2H), 1.16 (d, *J* = 6.9 Hz, 3H), 0.91 (d, *J* = 6.6 Hz, 3H), 0.87 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.7, 128.3, 127.5, 127.4, 89.3, 72.9, 68.0, 45.1, 40.1, 30.8, 27.8, 23.2, 21.1, 19.7, 16.6; HRMS (ESI): calcd. for C₁₈H₂₆ONa [M + Na]⁺ 281.1881; found 281.1876.

(2S,5S,7S,9R)-10-(Benzyloxy)-1-((4R,5S,6R)-6-(3,5-dimethoxyphenyl)-5-isobutyl-2,2-

dimethyl-1,3-dioxan-4-yl)-5,7,9-trimethyldec-3-yn-2-ol (21). To a suspension of Zn(OTf)₂ (1 g, 2.70 mmol), which was dried under high vacuum at 60 to 80 °C for 20 min prior to use, and (-)-*N*-methylephedrine (516 mg, 2.80 mmol) in toluene (1 mL) was added triethylamine (400 μ L, 2.80 mmol). The mixture was stirred for 3 h at room temperature and treated dropwise with a solution of alkyne **20** (485 mg, 1.88 mmol) in toluene (1 mL) via cannula. The mixture was stirred for 45 min and treated dropwise with a solution of aldehyde **12** (300 mg, 1.11 mmol) in toluene (1 mL + 500 μ L × 2 rinse) via cannula. The mixture was stirred for 12 h at room temperature and then was quenched with saturated NH₄Cl solution (5 mL). The mixture was extracted with diethyl ether (3 x 10 mL) and the organic layer was washed with brine (5 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford compound **21** (485 mg, 72%) as a light yellow liquid. R_f = 0.4 (20% EtOAc/hexane); $[\alpha]_D^{24} = +62.7$ (*c* 0.51, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 6.48 (d, *J* = 2.3 Hz, 2H), 6.34 (t, *J* = 2.3 Hz, 1H), 5.02 (brs, 1H), 4.62-4.56 (m, 1H), 4.50 (d, *J* = 1.1 Hz, 2H), 4.36 (d, *J* = 10.3 Hz, 1H), 3.78 (s, 6H), 3.33-3.21 (m, 2H), 2.71 (brs, 1H), 2.61-2.52 (m, 1H), 2.12-2.04 (m, 1H), 1.93-1.85 (m, 1H), 1.81-1.72 (m, 1H), 1.70-1.62 (m, 1H), 1.50 (s, 3H), 1.47 (s, 3H), 1.32-1.21 (m, 3H), 1.18-1.15 (m, 4H), 1.15 (d, J = 6.7 Hz, 3H), 0.94 (d, J = 6.6 Hz, 3H), 0.89-0.80 (m, 1H), 0.88 (d, J = 6.6 Hz, 3H), 0.70 (d, J = 6.5 Hz, 3H), 0.45 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 160.5, 143.2, 138.6, 128.2, 127.4, 127.3, 103.7, 99.3, 98.8, 89.9, 80.8, 76.5, 75.0, 73.1, 72.8, 61.5, 55.2, 45.2, 41.4, 40.0, 30.8, 30.4, 29.8, 27.9, 27.3, 23.4, 22.7, 22.2, 21.2, 19.7, 19.4, 16.6; HRMS (ESI): calcd. for C₃₈H₅₆O₆Na [M + Na]⁺ 631.3975; found 631.3979.

The procedure for preparation of **21a** was same as that for the preparation of **21**. **21a** was isolated (73%) as a light yellow liquid. $R_f = 0.3$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.30 (m, 4H), 7.29-7.24 (m, 1H), 6.67 (d, J = 2.2 Hz, 2H), 6.59 (t, J = 2.2 Hz, 1H), 5.14 (s, 4H), 5.01 (brs, 1H), 4.60-4.57 (m, 1H), 4.50-4.49 (m, 2H), 4.34-4.30 (m, 1H), 3.44 (s, 6H), 3.35-3.22 (m, 2H), 2.76 (brs, 1H), 2.60-2.52 (m, 1H), 2.10-2.03 (m, 1H), 1.93-1.85 (m, 1H), 1.80-1.72 (m, 1H), 1.67-1.60 (m, 1H), 1.49-1.47 (m, 1 H), 1.49 (s, 3H), 1.46 (s, 3H), 1.46-1.36 (m, 1H), 1.32-1.27 (m, 1H), 1.18-1.08 (m, 3H), 1.15 (d, J = 6.6 Hz, 3H), 0.96-0.91 (m, 1H), 0.93 (d, J = 6.6 Hz, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.89-0.83 (m, 1H), 0.67 (d, J = 6.6 Hz, 3H), 0.44 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 157.9, 143.3, 138.7, 128.2, 127.5, 127.4(2), 107.1, 103.2, 99.4, 94.3, 90.0, 80.7, 74.9, 73.2, 72.9, 61.6, 55.8, 45.3, 41.4(2), 41.3, 40.1, 30.8, 30.4, 29.8, 27.9, 27.3, 23.4, 22.7, 22.1, 21.2, 19.7, 19.5, 16.7; LCMS (ESI): 669 (M + H)⁺.

To a solution of **21** (12 mg) in CH₂Cl₂ (1 mL) were added sequentially (*S*)-(-)- α -methoxy- α -(trifluoromethyl)phenylacetic acid (7 mg), *N*,*N'*-dicyclohexylcarbodiimide (5 mg) and 4-(dimethylamino)pyridine (0.01 mg). The mixture was stirred for 2 h at room temperature and filtered through a pad of Celite. The filtrate was washed with water (2 mL), brine (2 mL), dried

over MgSO₄, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give (S)-21b-MTPA ester (9.8 mg, 60% yield) as yellow oil. $R_f = 0.4$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.63-7.50 (m, 2H), 7.46-7.36 (m, 3H), 7.36-7.26 (m, 5H), 6.48 (d, J = 2.2 Hz, 2H), 6.34 (t, J = 2.2 Hz, 1H), 5.75-5.70 (m, 1H), 4.97 (s, 1H), 4.49 (s, 2H), 4.28 (d, J = 8.3 Hz, 1H), 3.78 (s, 6H), 3.60 (s, 3H), 3.34-3.18 (m, 2H), 2.64-2.52 (m, 1H), 2.06-1.97 (m, 1H), 1.92-1.82 (m, 1H), 1.78-1.68 (m, 3H), 1.47 (s, 3H), 1.44 (s, 3H), 1.41-1.25 (m, 3H), 1.22-1.14 (m, 4H), 1.16 (d, J = 6.8 Hz, 3H), 0.93 (d, J = 6.6 Hz, 3H), 0.88 (d, J = 6.6 Hz, 3H), 0.64 (d, J = 6.5 Hz, 3H), 0.45 (d, J = 6.5 Hz, 3H); LCMS (ESI): 825 (M + H)⁺. Similarly, the (R)-MTPA ester 21c (9 mg, 58%) was obtained using (R)-(+)- α -methoxy- α -(trifluoromethyl) phenylacetic acid (MTPA). $R_{f} = 0.4$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.60-7.52 (m, 2H), 7.42-7.36 (m, 3H), 7.36-7.27 (m, 5H), 6.48 (d, J = 2.2 Hz, 2H), 6.34(t, J = 2.2 Hz, 1H), 5.74-5.68 (m, 1H), 5.00 (brs, 1H), 4.49 (s, 2H), 4.36 (d, J = 9.9 Hz, 1H),3.78 (s. 6H), 3.57 (s. 3H), 3.32-3.18 (m. 2H), 2.61-2.48 (m. 1H), 2.13-1.99 (m. 2H), 1.92-1.65 (m, 3H), 1.48 (s, 6H), 1.22-1.14 (m, 5H), 1.13 (d, J = 6.8 Hz, 3H), 0.92 (d, J = 6.6 Hz, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.87-0.76 (m, 2H), 0.67 (d, J = 6.3 Hz, 3H), 0.46 (d, J = 6.3 Hz, 3H); LCMS(ESI): 825 $(M + H)^+$.

(2*R*,4*S*,6*R*,9*R*)-9-((*tert*-Butyldimethylsilyl)oxy)-10-((4*R*,5*S*,6*R*)-6-(3,5-dimethoxyphenyl)-5isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-2,4,6-trimethyldecan-1-ol (23). To a solution of 21 (450 mg, 0.74 mmol) in anhydrous CH_2Cl_2 (5 mL) was added slowly 2,6-lutidine (0.2 mL, 1.85 mmol) followed by TBSOTf (0.3 mL, 1.33 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C and quenched with water (3 mL) and washed with saturated NaHCO₃ solution (5 mL). Organic layer was extracted with CH_2Cl_2 (3 x 10 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel

column chromatography to give compound **22** (464 mg, 87%) as a colorless liquid. $R_f = 0.5$ (10% EtOAc/hexane); $[\alpha]_D^{24} = +72.6$ (*c* 0.52, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 6.49 (d, J = 2.2 Hz, 2H), 6.33 (t, J = 2.2 Hz, 1H), 4.99 (brs, 1H), 4.55-4.50 (m, 1H), 4.50 (s, 2H), 4.34 (d, J = 7.2 Hz, 1H), 3.78 (s, 6H), 3.34-3.20 (m, 2H), 2.61-2.47 (m, 1H), 1.98-1.74 (m, 3H), 1.70-1.62 (m, 2H), 1.47 (s, 6H), 1.21-1.13 (m, 4H), 1.15 (d, J = 6.7 Hz, 3H), 0.94 (d, J = 6.6 Hz, 3H), 0.92 (s, 9H), 0.92-0.90 (m, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.67 (d, J = 6.5 Hz, 3H), 0.15 (d, J = 6.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 160.6, 143.7, 138.7, 128.3, 127.5, 127.4, 103.8, 99.2, 98.8, 89.7, 81.4, 75.2, 72.9, 71.1, 60.9, 55.3, 45.6, 42.4, 41.2, 40.1, 30.9, 30.6, 29.8, 28.1, 27.3, 25.9, 25.6, 23.5, 22.7, 22.4, 21.3, 19.8, 19.4, 18.2, 16.7, -4.3, -4.8; LCMS (ESI): 723 (M + H)⁺.

The procedure for the preparation of **22a** was same as that for the preparation of **22**. **22a** was isolated (89%) as a colorless liquid. $R_f = 0.4$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.34-7.27 (m, 5H), 6.68 (d, J = 1.9 Hz, 2H), 6.59 (t, J = 2.3 Hz, 1H), 5.14 (s, 4H), 5.00 (brs, 1H), 4.55-4.52 (m, 1H), 4.50 (s, 2H), 4.31 (d, J = 7.7 Hz, 1H), 3.45 (s, 6H), 3.33-3.22 (m, 2H), 2.60-2.49 (m, 1H), 1.97-1.87 (m, 2H), 1.83-1.73 (m, 1 H), 1.67-1.61 (m, 1H), 1.49-1.47 (m, 1H), 1.47 (s, 3H), 1.46 (s, 3 H), 1.43-1.37 (m, 1H), 1.31-1.25 (m, 2H), 1.19 (d, J = 6.5 Hz, 3H), 1.61 (d, J = 6.8 Hz, 3H), 0.94 (d, J = 6.6 Hz, 3H), 0.92 (s, 9H), 0.91-0.89 (m, 1H), 0.90 (d, J = 6.6 Hz, 3H), 0.67 (d, J = 6.5 Hz, 3H), 0.47 (d, J = 6.5 Hz, 3H), 0.15 (d, J = 5.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 143.8, 138.8, 128.3, 127.5, 107.2, 103.1, 99.2, 94.4, 89.7, 81.4, 75.1, 72.9, 71.1, 60.9, 55.9, 53.4, 45.6, 42.4, 41.1, 40.2, 30.9, 30.5, 29.8, 28.0, 27.3, 25.9, 25.6, 23.5, 22.7, 22.3, 21.3, 19.7, 19.4, 18.2, 16.7, -4.3, -4.8; LCMS (ESI): 783 (M + H)⁺.

To a solution of **22** (450 mg, 0.62 mmol) in EtOAc (10 mL) under H_2 was added Pd/C (40 mg, 10 mol%). The mixture was stirred for 2 h at room temperature, filtered through a pad of Celite,

washed with EtOAc, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford compound **23** (290 mg, 75%) as a colorless liquid. $R_f = 0.3$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +70.7$ (*c* 0.67, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.49 (d, *J* = 1.8 Hz, 2H), 6.33 (t, *J* = 1.8 Hz, 1H), 4.98 (brs, 1H), 4.32-4.26 (m, 1H), 3.78 (m, 1H), 3.78 (s, 6H), 3.56-3.36 (m, 2H), 1.78-1.65 (m, 2H), 1.64-1.54 (m, 4H), 1.48-1.37 (m, 4H), 1.48 (s, 6H), 1.37-1.25 (m, 3H), 1.20-1.11 (m, 3H), 1.11-1.00 (m, 4H), 0.93 (s, 9H), 0.89 (d, *J* = 6.4 Hz, 3H), 0.83 (d, *J* = 6.6 Hz, 3H), 0.68 (d, *J* = 6.2 Hz, 3H), 0.45 (d, *J* = 6.2 Hz, 3H), 0.08 (d, *J* = 9.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 160.5, 143.8, 103.8, 99.1, 98.8, 75.3, 70.7, 69.3, 69.0, 55.3, 45.7, 41.5, 41.2, 41.0, 40.2, 34.7, 33.2, 32.8, 30.4, 30.2, 29.9, 27.4, 27.2, 25.9, 22.8, 22.4, 19.5, 19.2, 18.1, 16.4, -4.2, -4.4; HRMS (ESI): calcd. for C₃₇H₆₈O₆SiNa [M + Na]⁺ 659.4683; found 659.4678.

(2*R*,4*S*,6*S*)-10-((4*R*,5*S*,6*R*)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3dioxan-4-yl)-2,4,6-trimethyldec-7-yn-1-ol (23'). Compound 23' was isolated as a colorless liquid. $R_f = 0.3$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 6.68 (d, J = 2.2 Hz, 2H), 6.58 (t, J = 2.2 Hz, 1H), 5.14 (s, 4H), 4.98 (brs, 1H), 4.00-3.98 (m, 1H), 3.52-3.46 (m, 1H), 3.45 (s, 6H), 3.43-3.37 (m, 1H), 1.80-1.67 (m, 1H), 1.65-1.52 (m, 3H), 1.50-1.48 (m, 1H), 1.48 (s, 3H), 1.47 (s, 3H), 1.43-1.35 (m, 2H), 1.35-1.25 (m, 3H), 1.22-1.17 (m, 1H), 1.16-0.97 (m, 5H), 0.89 (d, J = 6.6 Hz, 3H), 0.86-0.79 (m, 6H), 0.68 (d, J = 6.4 Hz, 3H), 0.45 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 157.9, 144.0, 107.2, 103.1, 99.2, 94.3, 75.1, 74.3, 69.0, 55.9, 45.6, 41.4, 41.1, 37.6, 37.0, 33.1, 30.3, 29.9, 29.8, 27.2, 27.2, 26.9, 26.2, 22.8, 22.3, 20.0, 19.5, 19.2, 16.4; LCMS (ESI): 563 (M + H)⁺.

(4R,6S,8R,11R)-11-((*tert*-Butyldimethylsilyl)oxy)-12-((4R,5S,6R)-6-(3,5-dimethoxyphenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-4,6,8-trimethyldodecan-3-ol (24). To a solution of **23** (250 mg, 0.4 mmol) in CH₂Cl₂ (5 mL) were added NaHCO₃ (84 mg, 1 mmol) and Dess-Martin periodinane (320 mg, 0.72 mmol) at 0 °C. The mixture was stirred for 1 h at room temperature, diluted with CH₂Cl₂ (10 mL), and washed with saturated NaHCO₃ solution (5 mL). The organic layer was separated, dried over MgSO₄ and concentrated under reduced pressure. The crude aldehyde was directly used for the next step without further purification.

To a stirred solution of crude aldehyde in ether (10 mL) was added dropwise C₂H₅MgBr (1.6 mL of 1 M solution, 1.6 mmol) at 0 °C. The mixture was stirred for 1 h at the same temperature and quenched with saturated NH₄Cl solution (3 mL). Organic layer was extracted with EtOAc (3 x 10 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **24** (230 mg, 87% over two steps) as a colorless liquid. R_f = 0.4 (20% EtOAc/hexane); $[\alpha]_D^{24}$ = +65.3 (*c* 0.85, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.48 (d, *J* = 2.1 Hz, 2H), 6.33 (t, *J* = 2.2 Hz, 1H), 4.98 (brs, 1H), 4.26 (brm, 1H), 3.82-3.78 (m, 1H), 3.78 (s, 6H), 3.42-3.28 (m, 1H), 1.79-1.48 (m, 11H), 1.48 (d, *J* = 4.0 Hz, 6H), 1.22-1.02 (m, 7H), 0.94 (d, *J* = 6.8 Hz, 3H), 0.93 (s, 9H), 0.89-0.84 (m, 4H), 0.85 (d, *J* = 6.6 Hz, 3H), 0.81 (d, *J* = 6.5 Hz, 3H), 0.68 (d, *J* = 6.3 Hz, 3H), 0.45 (d, *J* = 6.3 Hz, 3H), 0.09 (d, *J* = 9.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 160.5, 143.8, 103.8, 99.1, 98.7, 78.1, 77.2, 75.3, 70.7, 69.2, 55.3, 45.7 (2), 41.7, 41.0, 35.0, 32.9, 30.4, 30.1, 29.9, 27.4, 27.2, 25.9, 22.8, 22.4, 19.6, 19.5, 19.5, 19.1, 18.1, 13.4, 10.5, 10.5, -4.2, -4.4; HRMS (ESI): calcd. for C₃₉H₇₂O₆SiNa [M + Na]⁺ 687.4996; found 687.4928.

(4R,6S,8R,11R)-11-((tert-Butyldimethylsilyl)oxy)-12-((4R,5S,6R)-6-(3,5-dimethoxyphenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-4,6,8-trimethyldodecan-3-one (25). To a solution of 24 (200 mg, 0.3 mmol) in CH₂Cl₂ (5 mL) were added NaHCO₃ (50 mg, 0.6 mmol) and Dess-Martin periodinane (230 mg, 0.54 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C and for

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1 h at room temperature and then diluted with CH₂Cl₂ (10 mL), and washed with saturated NaHCO₃ solution (5 mL). The organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **25** (180 mg, 92%) as a colorless liquid. $R_f = 0.3$ (10% EtOAc/hexane); $[\alpha]_D^{24} = +22.1$ (*c* 0.67, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.48 (brs, 2H), 6.33 (brs, 1H), 4.98 (brs, 1H), 4.29-4.24 (m, 1H), 3.78 (m, 1H), 3.78 (s, 6H), 2.67-2.55 (m, 1H), 2.45 (q, *J* = 7.2 Hz, 2H), 1.81-1.66 (m, 1H), 1.64-1.47 (m, 5H), 1.47 (d, *J* = 3.6 Hz, 6H), 1.36-1.12 (m, 7H), 1.11-0.99 (m, 5H), 1.06 (d, *J* = 6.6 Hz, 3H), 0.93 (s, 9H), 0.89-0.84 (m, 1H), 0.84 (d, *J* = 6.4 Hz, 3H), 0.82 (d, *J* = 6.4 Hz, 3H), 0.68 (d, *J* = 6.1 Hz, 3H), 0.44 (d, *J* = 6.3 Hz, 3H), 0.08 (d, *J* = 10.1 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 215.5, 160.6, 143.8, 103.8, 99.1, 98.8, 77.2, 75.3, 70.7, 69.2, 55.3, 45.1, 43.7, 41.1, 41.0, 40.2, 34.4, 34.1, 32.9, 30.3, 30.1, 29.9, 28.0, 27.3, 25.9, 22.8, 22.4, 19.5, 19.3, 19.3, 18.1, 16.5, 7.9, -0.02, -4.2, -4.4; HRMS (ESI): calcd. for C₃₉H₇₀O₆SiNa [M + Na]⁺ 685.4839; found 685.4833.

(4*R*,6*S*,8*R*,11*R*,13*R*,14*S*)-14-((*R*)-(3,5-dimethoxyphenyl)(hydroxy)methyl)-11,13-dihydroxy-4,6,8,16-tetramethylheptadecan-3-one (26). To a stirred solution of 25 (180 mg, 0.27 mmol) in MeOH (5 mL) was added PTSA (5 mg, 0.03 mmol) at room temperature. The mixture was stirred for 4 h at the same temperature and quenched with saturated NaHCO₃ solution (5 mL). The organic layer was extracted with CH₂Cl₂ (3 x 10 mL) and combined organic layer was washed with brine (2 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 26 (120 mg, 90%) as a colorless liquid. $R_f = 0.3$ (30% EtOAc/hexane); $[\alpha]_D^{24} = +23.6$ (*c* 1.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.51 (d, J = 2.3 Hz, 2H), 6.34 (t, J = 2.3 Hz, 1H), 5.04 (brs, 1H), 4.29 (d, J =10.4 Hz, 1H), 3.92-3.78 (m, 1H), 3.78 (s, 6H), 3.15-2.92 (brs, 3H), 2.66-2.55 (m, 1H), 2.46 (q, J = 7.3 Hz, 2H), 1.73-1.61 (m, 2H), 1.56-1.34 (m, 6H), 1.32-1.18 (m, 6H), 1.09-0.98 (m, 8H), 0.82 (d, J = 6.6 Hz, 3H), 0.81 (d, J = 6.6 Hz, 3H), 0.72 (d, J = 6.5 Hz, 3H), 0.53 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 215.7, 160.7, 146.0, 103.6, 99.1, 78.1, 77.8, 77.2, 73.8, 55.4, 48.2, 45.0, 43.7, 41.0, 35.7, 34.0, 33.2, 30.4, 30.0, 27.8, 27.3, 22.7, 22.4, 20.0, 19.3, 19.3, 16.4, 7.8; HRMS (ESI): calcd. for C₃₀H₅₂O₆Na [M + Na]⁺ 531.3662; found 531.3666.

(4*R*,6*S*,8*R*,11*R*,13*R*,14*S*)-14-((*R*)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-11,13-dihydroxy-

4,6,8,16-tetramethylheptadecan-3-one (1). To a stirred solution of **26** (100 mg, 0.19 mmol) in CH₂Cl₂ (5 mL) AlI₃ (380 mg, 0.95 mmol) was added at one time under nitrogen atmosphere at 0 ^oC. The mixture was sealed with a stopper and stirred for 2 days at room temperature. After the completion of the reaction monitored by LCMS, the mixture was quenched with 1 N HCl (few drops) at 0 °C and stirred for additional 2 h. Organic layer was extracted with CH₂Cl₂ (3 x 10 mL). The combined organic layer was washed with brine (5 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 1 (20 mg, 22%) as a white amorphous solid. $R_f = 0.2$ (5%) MeOH/CH₂Cl₂); $[\alpha]_{D}^{24} = +20.6$ (*c* 0.68, MeOH); $[lif[\alpha]_{D}^{20} = -12.0$ (*c* 0.20, MeOH)]; IR (KBr): 3346, 2925, 1699, 1605, 1458, 1378, 1339, 1102, 1004, 844, 668, 657, 613 cm⁻¹; ¹H NMR (600 MHz, CD₃OD): δ 6.33 (d, J = 2.2 Hz, 2H), 6.15 (t, J = 2.2 Hz, 1H), 4.84 (d, J = 4.0 Hz, 1H), 3.96 (m, 1H), 3.69 (m, 1H), 2.71 (m, 1H), 2.57-2.52 (m, 2H), 1.70 (m, 1H), 1.68-1.62 (m, 2H), 1.52-1.49 (m, 3H), 1.45-1.40 (m, 3H), 1.30-1.24 (m, 5H), 1.13-1.11 (m, 2H), 1.04 (d, J = 6.5 Hz,3H), 1.01 (t, J = 7.5 Hz, 3H), 0.86 (d, J = 6.5 Hz, 3H), 0.85 (d, J = 6.5 Hz, 3H), 0.78 (d, J = 6.5Hz, 3H), 0.64 (d, J = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CD₃OD); δ 218.1, 158.9, 148.2, 105.3, 101.6, 76.4, 74.6, 71.8, 49.1, 45.9, 44.5, 42.0, 41.9, 35.5, 34.8, 34.1, 33.1, 31.0, 28.8, 28.0, 23.1,

22.6, 19.5, 19.5, 16.7 7.8; HRMS (ESI): calcd. for $C_{28}H_{48}O_6Na [M + Na]^+$ 503.3349; found 503.3336.

(R)-4-Benzyl-3-((S)-2-((R)-(3,5-bis(methoxymethoxy)phenyl)(hydroxy)methyl)-4-

methylpentanoyl)oxazolidin-2-one (27). To a stirred solution of **5** (9.5 g, 34.5 mmol) in EtOAc (50 mL) was added MgCl₂ (500 mg, 5.76 mmol), Et₃N (8 mL, 57.6 mmol) and di-MOMprotected aldehyde **4** (6.5 g, 28.8 mmol) at 23 °C. The mixture was stirred for 24 h, filtered through a pad of Celite, washed with EtOAc and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford compound **27** (10.4 g, 72%) as a yellow liquid. $R_f = 0.3$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +13.0$ (*c* 2.77, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ7.31-7.22 (m, 3H), 7.11 (d, *J* = 6.7 Hz, 2H), 6.78 (d, *J* = 2.2 Hz, 2H), 6.66 (t, *J* = 2.2 Hz, 1H), 5.13 (dd, *J* = 15.4, 6.7 Hz, 4H), 4.75 (d, *J* = 6.7 Hz, 1H), 4.65-4.56 (m, 2H), 4.14-4.06 (m, 2H), 3.42 (s, 6H), 3.29 (brs, 1H), 3.10 (dd, *J* = 13.6, 3.2 Hz, 1H), 2.45 (dd, *J* = 13.6, 9.6 Hz, 1H), 1.79-1.72 (m, 1H), 1.62-1.51 (m, 1H), 1.41-1.34 (m, 1H), 0.88 (t, *J* = 6.1 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ176.5, 158.4, 153.4, 145.1, 135.4, 129.3, 128.9, 127.2, 107.8, 104.1, 94.5, 75.9, 65.7, 56.0, 55.4, 46.6, 38.7, 37.3, 26.1, 22.8, 22.4; HRMS (ESI): calcd. for C₂₇H₃₅NO₈Na [M + Na]⁺ 524.2260; found 524.2312.

(R)-4-Benzyl-3-((S)-2-((R)-(3,5-bis(methoxymethoxy)phenyl)((tert-

butyldimethylsilyl)oxy)methyl)-4-methylpentanoyl)oxazolidin-2-one (28). To a solution of **27** (10 g, 19.9 mmol) in anhydrous CH_2Cl_2 (60 mL) was added 2,6-lutidine (5.8 mL, 49.9 mmol), followed by TBSOTf (7.3 mL, 31.8 mmol) slowly at 0 °C. The mixture was stirred for 1 h at 0 °C and quenched with water (15 mL) and then washed with saturated NaHCO₃ solution (20 mL). Organic layer was extracted with CH_2Cl_2 (3 x 50 mL). The combined organic layer was dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel

column chromatography to afford **28** (10.7 g, 88%) as a yellow liquid. $R_f = 0.3$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +23.2$ (*c* 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.33 (m, 2H), 7.28-7.26 (m, 3H), 6.76 (d, J = 2.2 Hz, 2H), 6.64 (t, J = 2.2 Hz, 1H), 5.15 (s, 4H), 4.75 (d, J = 8.5 Hz, 1H), 4.66-4.59 (m, 1H), 4.50 (t, J = 8.2 Hz, 1H), 4.13-4.09 (m, 2H), 3.56 (dd, J = 13.2, 3.0 Hz, 1H), 3.45 (s, 6H), 2.58 (dd, J = 13.1, 11.1 Hz, 1H), 1.71-1.53 (m, 2H), 1.34-1.25 (m, 1H), 0.83 (s, 9H), 0.77 (d, J = 6.6 Hz, 3H), 0.73 (d, J = 6.6 Hz, 3H), -0.03 (s, 3H), -0.26 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 175.7, 157.8, 153.0, 145.1, 136.0, 129.3, 128.9, 127.2, 109.2, 104.2, 94.5, 77.8, 65.8, 56.2, 55.9, 49.0, 38.4, 34.6, 31.5, 26.2, 25.8, 23.6, 22.6, 21.8, 20.6, 18.0, 14.1, -4.6, -5.0; HRMS (ESI): calcd. for C₃₃H₄₉NO₈SiNa [M + Na]⁺ 638.3125; found 638.3130.

(R)-2-((R)-(3,5-bis(Methoxymethoxy)phenyl)((tert-butyldimethylsilyl)oxy)methyl)-4-

methylpentan-1-ol (29). To a stirred solution of 28 (10 g, 16.2 mmol) in anhydrous CH₂Cl₂ (100 mL) was added slowly DIBAL-H (28.9 mL, 40 mmol, 20% solution in toluene) for 15 min at -78 °C. The mixture was stirred for 30 min at -78 °C and for 0 °C for 1 h and then quenched with methanol (10 mL) and aqueous saturated sodium potassium tartarate solution (50 mL). The mixture was passed through a small pad of Celite and extracted with CH₂Cl₂ (3 x 60 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford 29 (6.4 g, 89%) as a colorless liquid. $R_f = 0.2$ (20% EtOAc/hexane); [α]_D²⁴ = +30.0 (*c* 0.36, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.60 (d, *J* = 2.1 Hz, 2H), 6.55 (t, *J* = 2.1 Hz, 1H), 5.07 (dd, *J* = 10.3, 6.7 Hz, 4H), 4.61 (d, *J* = 4.6 Hz, 1H), 3.68 (dd, *J* = 11.0, 2.1 Hz, 1H), 3.45-3.38 (m, 1H), 3.38 (s, 6H), 2.96 (brs, 1H), 1.68-1.63 (m, 1H), 1.63-1.53 (m, 1H), 1.32-1.25 (m, 1H), 1.14-1.07 (m, 1H), 0.85 (s, 9H), 0.83 (d, *J* = 6.6 Hz, 3H), 0.75 (d, *J* = 6.5 Hz, 3H), 0.01 (s, 3H), -0.21 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 157.8, 146.0, 107.8, 103.4, 94.2, 78.9, 62.8, 55.6, 45.1, 37.3, 25.6, 25.1,

23.1, 22.0, 17.8, -4.8, -5.4; HRMS (ESI): calcd. for $C_{23}H_{42}O_6SiNa [M + Na]^+ 465.2648$; found 465.2646.

(1*R*,2*R*,3*R*)-1-(3,5-Bis(methoxymethoxy)phenyl)-2-isobutylhex-5-ene-1,3-diol (32). To a stirred solution of IBX (7.3 g, 26.1 mmol) in DMSO (20 mL) was added dropwise a solution of alcohol **29** (6.4 g, 14.5 mmol) in CH₂Cl₂ (60 mL) at 25 °C. The mixture was stirred at 25 °C for 6 h and then filtered. The resulting solid was washed with diethyl ether. The filtrate was washed with saturated NaHCO₃ solution (15 mL), water (10 mL) and brine (15 mL), dried over MgSO₄, concentrated under reduced pressure to furnish crude aldehyde **30** (5.5 g, 86%) as a colorless liquid. $R_f = 0.7$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 9.69 (d, J = 3.9 Hz, 1H), 6.64 (d, J = 2.1 Hz, 2H), 6.62 (t, J = 2.1 Hz, 1H), 5.13 (s, 4H), 4.71 (d, J = 6.8 Hz, 1H), 3.45 (s, 6H), 2.65-2.58 (m, 1H), 1.59-1.50 (m, 1H), 1.50-1.37 (m, 1H), 1.09-1.00 (m, 1H), 0.84 (s, 9H), 0.82 (d, J = 6.6 Hz, 3H), 0.76 (d, J = 6.6 Hz, 3H), 0.01 (s, 3H), -0.21 (s, 3H); LCMS (ESI): 441 (M + H)⁺.

To a stirred solution of (+)-ipc₂BOMe (6.5 g, 20.4 mmol) in anhydrous Et₂O (30 mL) was added allylmagnesium bromide (17 mL, 17.1 mmol, 1.0 M in ether) at 0 °C. The mixture was stirred for 1 h at room temperature before being cooled to -78 °C. The mixture was treated dropwise with aldehyde **30** (5 g, 11.4 mmol) at -78 °C, stirred for 1 h at -78 °C, and then warmed slowly to room temperature. An aqueous solution of NaOH (2 M in H₂O) (20 mL) was added, followed by slow addition of 30% H₂O₂ solution (10 mL) at 0 °C. The biphasic solution was separated and the aqueous layer was extracted with Et₂O (3 x 50 mL). The combined organic layer was dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **31** along with diastereomeric mixture (*dr* = 90:10)

(4.1 g, 74%) as a colorless liquid. $R_f = 0.5$ (30% EtOAc/hexane) which could not be separated at this stage and was purified in the next step; LCMS (ESI): 483 (M + H)⁺, 505 (M + Na)⁺.

To a stirred solution of **31** (4 g, 8.3 mmol) in dry THF (30 mL) was added TBAF (12.4 mL of 1 M solution in THF, 12.4 mmol,) at 0 °C. The mixture was stirred for 5 h at room temperature, diluted with saturated NaHCO₃ solution (10 mL). Organic layer was separated, washed with brine (10 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford pure **32** (2.81 g, 92%) as a colorless liquid. $R_f = 0.2$ (30% EtOAc/hexane); $[\alpha]_D^{24} = +2.8$ (*c* 13.6, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.69-6.58 (m, 3H), 5.82-5.71 (m, 1H), 5.13-4.99 (m, 6H), 4.57 (d, *J* = 7.1 Hz, 1H), 3.71-3.67 (m, 1H), 3.42 (s, 6H), 2.42-2.29 (m, 1H), 2.23-2.13 (m, 1H), 1.82 (t, *J* = 6.1 Hz, 1H), 1.29-1.22 (m, 1H), 1.14-1.00 (m, 2H), 0.93-0.83 (m, 2H), 0.77 (d, *J* = 6.6 Hz, 3H), 0.65 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 157.9, 146.5, 135.4, 117.8, 108.2, 103.7, 94.2, 77.5, 74.4, 55.8, 46.4, 40.3, 38.2, 25.8, 22.5, 22.4; HRMS (ESI): calcd. for C₂₀H₃₂O₆Na [M + Na]⁺ 391.2097; found 391.2088.

(4R,5R,6R)-4-Allyl-6-(3,5-bis(methoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxane

(33). To a stirred solution of 32 (2.8 g, 7.61 mmol) in CH₂Cl₂ (20 mL) was added 2,2-dimethoxy propane (4 mL, 30 mmol) followed by PTSA (cat.) at 0 °C. The mixture was warmed to room temperature and stirred for 30 min and then quenched with water (10 mL). The organic layer was extracted with CH₂Cl₂ (3 x 10 mL). The combined organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford 33 (3.01 g, 97%) as a colorless liquid. R_f = 0.7 (20% EtOAc/hexane); $[\alpha]_D^{24}$ = +34.4 (*c* 0.77, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 6.74 (d, *J* = 2.3 Hz, 2H), 6.63 (t, *J* = 6.63 Hz, 1H), 5.99-5.89 (m, 1H), 5.15 (dd, *J* = 10.8, 6.7 Hz, 4H), 5.12-5.04

(m, 2H), 4.37 (d, J = 10.3 Hz, 1H), 3.68-3.62 (m, 1H), 3.45 (s, 6H), 2.46-2.40 (m, 1H), 2.22-2.15 (m, 1H), 1.51 (s, 3H), 1.51-1.47 (m, 1H), 1.46 (s, 3H), 0.97-0.86 (m, 3H), 0.68 (d, J = 6.1 Hz, 3H), 0.48 (d, J = 6.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 143.0, 135.3, 116.2, 109.4, 104.5, 98.4, 94.3, 78.4, 74.1, 55.8, 43.1, 37.4, 36.7, 30.1, 26.7, 22.7, 22.5, 19.6; HRMS (ESI): calcd. for C₂₃H₃₆O₆Na [M + Na]⁺ 431.2410; found 431.2428.

2-((4R,5R,6R)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-

yl)acetaldehyde (34). To a stirred solution of compound 33 (3 g, 7.35 mmol) in THF:H₂O (3:1, 20 mL) was added NMO (1.7 g, 14.7 mmol) followed by osmium tetroxide (0.2 mL, 0.24 mmol) at 25 °C. The mixture was stirred at the same temperature for 1 h and quenched with aqueous $Na_2S_2O_4$ solution (10 mL). The organic layer was extracted with EtOAc (3 x 10 mL) and the combined organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The crude residue was used for next step without further purification.

To a stirred solution of crude diol in THF:H₂O (2:1, 20 mL) was added portion wise NaIO₄ (3.14 g, 14.7 mmol) at 0 °C. The mixture was stirred for 3 h at room temperature and then diluted with saturated NaHCO₃ solution (5 mL). The organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **34** (2.11 g, 70% over two steps) as a colorless liquid. $R_f = 0.5$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +39.8$ (*c* 0.77, CHCl₃); IR (KBr): 2992, 2955, 2868, 2827, 1728, 1599, 1467, 1400, 1381, 1203, 1144, 1034, 925, 852 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.80 (m, 1H), 6.73 (d, *J* = 2.2 Hz, 2H), 6.64 (t, *J* = 2.2 Hz, 1H), 5.13 (dd, *J* = 11.8, 6.7 Hz, 4H), 4.42 (d, *J* = 10.1 Hz, 1H), 4.21-4.15 (m, 1H), 3.45 (s, 6H), 2.64-2.48 (m, 2H), 1.58-1.54 (m, 1H), 1.54 (s, 3H), 1.42 (s, 3H), 1.05-0.98 (m, 1H), 0.88-0.77 (m, 2H), 0.66 (d, *J* = 6.1 Hz, 3H), 0.48 (d, *J* = 6.1 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃); δ 201.5, 158.1, 142.4, 109.2, 104.6, 98.7, 94.3, 78.2,

70.4, 55.8, 47.0, 43.2, 36.4, 29.8, 26.7, 22.5, 22.4, 19.5; HRMS (ESI): calcd. for C₂₂H₃₄O₇Na [M + Na]⁺ 433.2202; found 433.2187.

(((2R,4R,6S)-7-(Benzyloxy)-2,4,6-trimethylheptyl)oxy)(tert-butyl)dimethylsilane (37). To a stirred solution of NaH (208 mg, 5.21 mmol) in anhydrous THF (10 mL) was added alcohol 36 (1 g, 3.47 mmol) in THF (10 mL) at 0 °C. The mixture was stirred for 30 min at room temperature and cooled to 0 °C. The mixture was treated with TBAI (110 mg, 0.3 mmol), followed by BnBr (0.4 mL, 3.82 mmol). The mixture was stirred for 5 h at room temperature and quenched with water (10 mL). The organic layer was extracted with EtOAc (3 x 10 mL) and the combined organic layer was washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford **37** (1.07 g, 82%) as a colorless oil. $R_f = 0.6$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.40-7.27 (m, 5H), 4.54 (dd, J = 15.0, 12.1 Hz, 2H), 3.50 (dd, J = 9.6, 5.0 Hz, 1H), 3.42-3.36 (m, 2H), 3.25 (dd, J = 8.9, 7.1 Hz, 1H), 1.99-1.87 (m, 1H), 1.79-1.69 (m, 1H), 1.69-1.58 (m, 1H), 1.69-1.51.43-1.32 (m, 2H), 0.99 (d, J = 6.7 Hz, 3H), 0.95 (s, 9H), 0.94 (d, J = 6.6 Hz, 3H), 0.93 (d, J =6.6 Hz, 3H), 0.93-0.84 (m, 2H), 0.09 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 138.8, 128.2, 127.4, 127.3, 75.9, 72.9, 68.0, 41.7, 41.1, 33.1, 30.9, 27.7, 25.9, 21.0, 18.3, 18.3, 17.9, -5.4; HRMS (ESI): calcd. for $C_{23}H_{42}O_2SiNa [M + Na]^+ 401.2852$; found 401.2879.

(2*R*,4*S*,6*S*)-7-(Benzyloxy)-2,4,6-trimethylheptan-1-ol (38). To a stirred solution of 37 (1 g, 2.64 mmol) in anhydrous THF (20 mL) was added TBAF (4.7 mL of 1 M solution in THF, 4.76 mmol,) at 0 °C. The mixture was stirred for 6 h at room temperature and quenched with saturated NaHCO₃ solution (5 mL). Organic layer was separated, extracted with EtOAc (3 x 10 mL), washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **38** (590 mg, 85%)

as a colorless liquid. $R_f = 0.2$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +7.2$ (*c* 0.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.26 (m, 5H), 4.49 (dd, J = 15.7, 12.1 Hz, 2H), 3.49 (dd, J = 10.5, 5.0 Hz, 1H), 3.35 (dd, J = 10.5, 5.1 Hz, 2H), 3.22 (dd, J = 9.0, 6.8 Hz, 1H), 1.91-1.82 (m, 1H), 1.75-1.67 (m, 1H), 1.65-1.53 (m, 2H), 1.39-1.24 (m, 3H), 0.96 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.7, 128.2, 127.4, 127.3, 75.7, 72.9, 68.0, 41.5, 41.1, 33.0, 30.9, 27.6, 20.9, 18.3, 17.5; LCMS (ESI): 265 (M + H)⁺.

((((2*S*,4*S*,6*R*)-2,4,6-Trimethyloct-7-yn-1-yl)oxy)methyl)benzene (39). To a solution of oxalyl chloride (0.2 mL, 2.84 mmol) in CH₂Cl₂ (3 mL) was added DMSO (0.24 mL, 3.40 mmol) at -78 °C. The mixture was stirred for 10 min at -78 °C and treated dropwise with a solution of alcohol **38** (500 mg, 1.89 mmol) in CH₂Cl₂ (6 mL). The mixture was stirred for 30 min at -78 °C and then treated dropwise with Et₃N (1.6 mL, 11.3 mmol). The mixture was stirred for 45 min at -78 °C and then treated with the saturated NH₄Cl solution (5 mL). The organic layer was separated and washed with saturated NaHCO₃ solution (5 mL), dried over MgSO₄ and concentrated under reduce pressure to afford crude aldehyde, which was used in the next step without further purification.

To a stirred solution of crude aldehyde and Bestmnn-Ohira reagent (544 mg, 2.83 mmol) in MeOH (10 mL) was added K₂CO₃ (780 mg, 5.67 mmol) at 0 °C. The mixture was stirred for 1 h at room temperature and passed through a small pad of Celite. The filtrate was extracted with CH₂Cl₂ (3 x 10 mL) and the combined organic layer was washed with brine (10 mL) dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **39** (336 mg, 69% over two steps) as a colorless liquid. $R_f = 0.8 (10\% \text{ EtOAc/hexane}); [\alpha]_D^{24} = -12.2 (c 1.1, CHCl_3); IR (KBr): 2965, 2927, 2871, 1453, 1378, 1099, 735, 676, 630 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): <math>\delta$ 7.38-7.26 (m, 5H), 4.53 (dd, J = 14.4.

12.1 Hz, 2H), 3.43-3.35 (m, 1H), 3.32-3.23 (m, 1H), 2.59-2.49 (m, 1H), 2.04 (d, J = 2.4 Hz, 1H), 1.99-1.79 (m, 2H), 1.58-1.52 (m, 1H), 1.44-1.33 (m, 1H), 1.22 (d, J = 6.8 Hz, 3H), 1.09-1.01 (m, 2H), 0.99 (d, J = 6.7 Hz, 3H), 0.94 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.8, 128.2, 127.4, 127.3, 88.8, 76.1, 72.9, 68.2, 43.7, 41.8, 30.7, 28.1, 23.5, 21.7, 20.0, 17.6; HRMS (ESI): calcd. for C₁₈H₂₆ONa [M + Na]⁺ 281.1881; found 281.1882.

(2*S*,4*S*,6*R*)-7-Hydroxy-2,4,6-trimethylheptyl acetate (40). To a stirred solution of 36 (1 g, 3.47 mmol) in CH₂Cl₂ (10 mL), Et₃N (0.9 mL, 6.94 mmol) was added followed by Ac₂O (0.5 mL, 5.20 mmol) and DMAP (cat.) at 0 °C and stirred for 30 min. The reaction mixture was quenched with water (5 mL) and extracted with CH₂Cl₂ (3 x 10 mL). The combined organic layer was washed with brine (5 mL) dried over MgSO₄, concentrated under reduced pressure to get crude reaction mixture and used for next step without further purification.

To a stirred solution of crude reaction mixture in dry THF (15 mL) was added TBAF (5.5 mL of 1 M solution in THF, 5.55 mmol,) at 0 °C. The mixture was stirred for 2 h at room temperature, diluted with saturated NaHCO₃ solution (5 mL). Organic layer was separated, washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **40** (584 mg, 78% over two steps) as a colorless liquid. $R_f = 0.2$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 3.88 (dd, J = 10.7, 5.0 Hz, 1H), 3.77 (dd, J = 10.7, 6.8 Hz, 1H), 3.43 (dd, J = 10.5, 5.1 Hz, 1H), 3.28 (dd, J = 10.4, 6.7 Hz, 1H), 2.19 (brs, 1H), 1.98 (s, 3H), 1.88-1.74 (m, 1H), 1.68-1.57 (m, 1H), 1.57-1.45 (m, 1H), 1.31-1.15 (m, 2H), 0.89-0.85 (m, 2H), 0.86 (d, J = 6.7 Hz, 3H), 0.84 (d, J = 6.6 Hz, 3H), 0.84 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 69.0, 67.7, 41.0, 40.9, 32.8, 29.7, 27.4, 20.7, 20.6, 17.9, 17.3; LCMS (ESI): 217 (M + H)⁺.

(2*S*,4*S*,6*R*)-7-(Benzyloxy)-2,4,6-trimethylheptyl acetate (41). To a stirred solution of 40 (580 mg, 2.69 mmol) and Cl₃C(=NH)OBn (1.2 g, 4.84 mmol) in cyclohexane:CH₂Cl₂ (2:1, 20 mL) was added dropwise TfOH (20 μ L, 0.27 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C and for 3 h at room temperature and quenched with saturated NaHCO₃ solution (10 mL). Organic layer was separated, extracted with EtOAc (3 x 10 mL), washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 41 (592 mg, 72%) as a colorless liquid. R_{*f*} = 0.6 (10% EtOAc/hexane); [α]_D²⁴ = -1.4 (*c* 1.38, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.34-7.26 (m, 5H), 4.49 (dd, *J* = 15.0, 12.1 Hz, 2H), 3.96 (dd, *J* = 10.7, 5.1 Hz, 1H), 3.81 (dd, *J* = 10.7, 7.1 Hz, 1H), 3.33 (dd, *J* = 8.9, 5.1 Hz, 1H), 3.21 (dd, *J* = 9.0, 6.9 Hz, 1H), 2.04 (s, 3H), 1.94-1.82 (m, 2H), 1.63-1.53 (m, 1H), 1.37-1.24 (m, 4H), 0.93 (d, *J* = 6.6 Hz, 3H), 0.92 (d, *J* = 6.7 Hz, 3H), 0.88 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 171.2, 138.7, 128.3, 127.4, 127.4, 75.8, 73.0, 69.2, 41.5, 41.1, 30.8, 29.8, 27.5, 20.9, 20.7, 18.1, 17.9; LCMS (ESI): 307 (M + H)⁺.

(2S,4R,6R)-7-(Benzyloxy)-2,4,6-trimethylheptan-1-ol (42). To a stirred solution of 41 (590 mg,

1.93 mmol) in methanol (6 mL) was added portion wise solid NaOMe (415 mg, 7.72 mmol) at 0 ^oC. The mixture was stirred for 2 h at room temperature, diluted with CH₂Cl₂ (10 mL), and quenched with saturated NH₄Cl solution (5 mL). Organic layer was separated, extracted with CH₂Cl₂ (3 x 10 mL), washed with brine (5 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **42** (592 mg, 95%) as a colorless liquid. $R_f = 0.2$ (20% EtOAc/hexane); $[\alpha]_D^{24} = -6.1$ (*c* 1.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.24 (m, 5H), 4.50 (dd, *J* = 15.7, 12.1 Hz, 2H), 3.49 (dd, *J* = 10.4, 5.0 Hz, 1H), 3.37-3.29 (m, 2H), 3.21 (dd, *J* = 9.0, 6.9 Hz, 1H), 1.94-1.80 (m, 1H), 1.77-1.63 (m, 1H), 1.63-1.47 (m, 2H), 1.39-1.23 (m, 2H), 0.95-0.90 (m, 2H), 0.95 (d, *J* =

6.7 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H), 0.90 (d, J = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 138.7, 128.2, 127.4, 127.3, 75.7, 72.9, 68.0, 41.5, 41.1, 33.0, 30.9, 27.7, 20.9, 18.3, 17.5; HRMS (ESI): calcd. for C₁₇H₂₈O₂Na [M + Na]⁺ 287.1987; found 287.1991.

((((2*R*,4*R*,6*S*)-2,4,6-Trimethyloct-7-yn-1-yl)oxy)methyl)benzene (43). To a solution of oxalyl chloride (0.27 mL, 3.12 mmol) in CH₂Cl₂ (4 mL) was added DMSO (0.26 mL, 3.74 mmol) at -78 °C. The mixture was stirred for 10 min at -78 °C and treated dropwise with a solution of alcohol 42 (550 mg, 2.08 mmol) in CH₂Cl₂ (6 mL). The mixture was stirred for 30 min at -78 °C and then treated dropwise with Et₃N (1.7 mL, 12.48 mmol). The mixture was stirred for 45 min -78 °C and treated with saturated NH₄Cl solution (5 mL). The organic layer was separated, washed with saturated NaHCO₃ solution (5 mL), dried over MgSO₄ and concentrated under reduce pressure, which was used in the next step without further purification.

To a stirred solution of crude aldehyde and Bestmnn-Ohira reagent (600 mg, 3.12 mmol) in MeOH (10 mL) was added K₂CO₃ (860 mg, 6.24 mmol) at 0 °C. The mixture was stirred for 1 h at room temperature, passed through a small pad of Celite, and extracted with CH₂Cl₂ (3 x 10 mL). The combined organic layer was washed with brine (10 mL), dried over MgSO₄ and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound **43** (354 mg, 66% over two steps) as a colorless liquid. R_f = 0.8 (10% EtOAc/hexane); $[\alpha]_D^{24}$ = +2.8 (*c* 13.6, CHCl₃); IR (KBr): 2964, 2927, 2872, 2849, 1453, 1378, 1362, 1098, 1028, 735, 676, 630 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.37-7.27 (m, 5H), 4.53 (d, *J* = 1.6 Hz, 2H), 3.42-3.33 (m, 1H), 3.30-3.22 (m, 1H), 2.58-2.52 (m, 1H), 2.05-2.02 (m, 1H), 1.97-1.81 (m, 2H), 1.58-1.51 (m, 1H), 1.43-1.30 (m, 1H), 1.20 (dd, *J* = 6.8, 1.3 Hz, 3H), 1.09-1.00 (m, 2H), 0.98 (dd, *J* = 6.7, 1.6 Hz, 3H), 0.93 (dd, *J* = 6.6, 1.3 Hz, 3H); ¹³C NMR

(100 MHz, CDCl₃): δ 138.8, 128.2, 127.4, 127.3, 88.8, 76.1, 72.9, 68.2, 43.7, 41.8, 30.7, 28.1, 23.5, 21.7, 20.0, 17.6; HRMS (ESI): calcd. for C₁₈H₂₆ONa [M + Na]⁺ 281.1881; found 281.1882.

(2S,5S,7R,9R)-10-(Benzyloxy)-1-((4R,5R,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-

isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-5,7,9-trimethyldec-3-yn-2-ol (44). To a suspension of Zn(OTf)₂ (816 mg, 2.25 mmol), which was dried under high vacuum at 60-80 °C for 20 min prior to use, and (-)-N-methylephedrine (420 mg, 2.34 mmol) in toluene (1 mL) was added triethylamine (326 μ L, 2.34 mmol). The mixture was stirred for 2.5 h at room temperature, and treated dropwise with a solution of alkyne 43 (350 mg, 1.35 mmol) in toluene (1 mL) via cannula. The mixture was stirred for 45 min, and treated dropwise with a solution of aldehyde **34** (370 mg, 0.9 mmol) in toluene (1 mL + 500 μ L x 2 rinse) via cannula. The mixture was stirred for 12 h at room temperature, quenched with saturated aqueous NH_4Cl solution (5 mL), and extracted with diethyl ether (3 x 10 mL). The organic layer was washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 44 (452 mg, 76%) as a colorless liquid. $R_f = 0.4$ (30%) EtOAc/hexane); $[\alpha]_D^{24} = +6.6$ (c 0.45, CHCl₃); IR (KBr): 2956, 2927, 1589, 1455, 1379, 1263, 1143, 1030, 925, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.34-7.26 (m, 5H), 6.74 (d, J = 2.1Hz, 2H), 6.65 (t, J = 2.1 Hz, 1H), 5.14 (dd, J = 10.9, 6.8 Hz, 4H), 4.61-4.59 (m, 1H), 4.50 (s, 2H), 4.37 (d, J = 10.3 Hz, 1H), 3.85 (t, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 2.89 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 2.89 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 2.89 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 3.49 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 3.49 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 3.49 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 3.49 (d, J = 10.3 Hz, 1H), 3.44 (s, 6H), 3.38-3.19 (m, 2H), 3.49 (s, 6H), 3.44 (s, 2.5 Hz, 1H), 2.61-2.51 (m, 1H), 2.01-1.77 (m, 4H), 1.53 (s, 3H), 1.44 (s, 3H), 1.40-1.27 (m, 3H), 1.18 (d, J = 6.7 Hz, 3H), 1.06-0.96 (m, 2H), 0.96 (d, J = 6.7 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H), 0.89 (d, J = 6.3 Hz, 3H), 0.70 (d, J = 5.7 Hz, 3H), 0.47 (d, J = 5.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): *δ* 158.0, 142.5, 138.7, 128.2, 127.4, 127.3, 109.3, 104.6, 98.5, 94.3, 89.5, 80.9, 78.3, 76.2, 73.8, 72.9, 61.6, 55.8, 43.5, 41.7, 41.4, 36.3, 31.5, 30.7, 30.0, 28.2, 26.7, 23.7, 22.6, 22.6,

22.4, 21.8, 20.3, 19.5, 17.5, 14.0; HRMS (ESI): calcd. for $C_{40}H_{60}O_8Na [M + Na]^+$ 691.4186; found 691.4185.

(((2S,5S,7R,9R)-10-(Benzyloxy)-1-((4R,5R,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-

isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-5,7,9-trimethyldec-3-yn-2-yl)oxy)(tert-

butyl)dimethylsilane (45). To a solution of 44 (450 mg, 0.67 mmol) in anhydrous CH₂Cl₂ (5 mL) was added 2,6-lutidine (0.2 mL, 1.68 mmol), followed by TBSOTf (0.23 mL, 1.00 mmol) dropwise at 0 °C. The mixture was stirred for 30 min at 0 °C, quenched with water (3 mL), and washed with saturated NaHCO₃ solution (5 mL). Organic layer was extracted with CH_2Cl_2 (3 x 10 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to give compound 45 (430 mg, 82%) as a colorless liquid. $R_f = 0.7$ (10% EtOAc/hexane); $[\alpha]_D^{24} = +6.0$ (c 0.45, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.26 (m, 5H), 6.76 (d, J = 2.2 Hz, 2H), 6.64 (t, J = 2.2 Hz, 1H), 5.14 (dd, J = 11.8, 6.7 Hz, 4H), 4.63 (ddd, J = 10.4, 4.3, 1.6 Hz, 1H), 4.51 (d, J = 10.4, 4.5 Hz, 1H), 4.5 Hz, 1H), 4.51 (d, J = 10.4, 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H), 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H), 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H), 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H), 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H, 4.5 Hz, 1H, 4.5 Hz, 1H, 4.5 Hz, 1H), 4.5 Hz, 1H, 4.5 Hz, 1H 2.7 Hz, 2H), 4.39 (d, J = 10.3 Hz, 1H), 3.81 (td, J = 10.3, 1.5 Hz, 1H), 3.45 (s, 6H), 3.41-3.31 (m, 1H), 3.27-3.20 (m, 1H), 2.64-2.49 (m, 1H), 1.99-1.80 (m, 3H), 1.77-1.69 (m, 1H), 1.51 (s, 3H), 1.44 (s, 3H), 1.38-1.27 (m, 2H), 1.18 (d, J = 6.8 Hz, 3H), 1.08-0.99 (m, 3H), 0.98 (d, J = 6.7 Hz, 3H), 0.96-0.89 (m, 15H), 0.69 (d, J = 5.7 Hz, 3H), 0.48 (d, J = 5.7 Hz, 3H), 0.14 (d, J = 4.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 142.9, 138.7, 128.2, 127.4, 127.3, 109.4, 104.5, 98.3, 94.3, 89.2, 81.5, 78.5, 76.3, 75.9, 72.9, 71.7, 60.9, 55.8, 53.3, 43.7, 43.3, 42.5, 41.8, 36.2, 30.9, 30.8, 30.0, 28.1, 26.7, 25.8, 23.6, 22.7, 22.4, 21.9, 20.1, 19.5, 18.2, 17.6, -4.5, -4.9; HRMS (ESI): calcd. for $C_{46}H_{74}O_8SiNa [M + Na]^+ 805.5051$; found 805.5067.

(2*R*,4*R*,6*R*,9*R*)-10-((4*R*,5*R*,6*R*)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-9-((*tert*-butyldimethylsilyl)oxy)-2,4,6-trimethyldecan-1-ol (46). To a

solution of **45** (430 mg, 0.55 mmol) in EtOAc (8 mL) was added Pd/C (25 mg, 10 mol%) under H₂. The mixture was stirred for 2 h at room temperature. The mixture was filtered through a pad of Celite, washed with EtOAc, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford a mixture of compound **46** and TBS-ether eliminated product (290 mg, 76% combined yield) as a colorless liquid. $R_f = 0.3$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 6.74 (d, J = 2.2 Hz, 2H), 6.62 (t, J = 2.2 Hz, 1H), 5.17-5.12 (m, 4H), 4.35 (d, J = 10.3 Hz, 1H), 3.89-3.86 (m, 1H), 3.63 (t, J = 8.4 Hz, 1H), 3.56-3.49 (m, 1H), 3.44 (s, 6H), 3.41-3.32 (m, 1H), 1.83-1.67 (m, 3H), 1.64-1.51 (m, 2H), 1.51-1.43 (m, 4H), 1.49 (s, 3H), 1.42 (s, 3H), 1.35-1.05 (m, 3H), 0.99 (d, J = 6.7 Hz, 3H), 0.93 (d, J = 6.6 Hz, 3H), 0.92-0.87 (m, 15H), 0.87 (d, J = 6.7 Hz, 3H), 0.68 (d, J = 5.9 Hz, 3H), 0.46 (d, J = 5.9 Hz, 3H), 0.06 (d, J = 3.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 160.2, 143.0, 109.4, 104.5, 98.2, 94.3, 78.8, 71.2, 69.5, 68.2, 55.8, 45.1, 43.8, 41.2, 40.6, 36.5, 33.0, 32.9, 31.6, 30.1, 27.9, 27.5, 26.8, 25.9, 25.9, 22.5, 20.9, 20.4, 19.5, 18.1, 17.6, 17.5, -4.3, -4.6; HRMS (ESI): calcd. for C₃₉H₇₂O₈SiNa [M + Na]⁺ 719.4894; found 719.4897.

(4R,6R,8R,11R)-12-((4R,5R,6R)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-

dimethyl-1,3-dioxan-4-yl)-11-((tert-butyldimethylsilyl)oxy)-4,6,8-trimethyldodecan-3-one

(47). To a solution of 46 (290 mg, 0.42 mmol) in CH_2Cl_2 (5 mL) were added NaHCO₃ (70 mg, 0.83 mmol) and Dess-Martin periodinane (320 mg, 0.75 mmol) at 0 °C. The mixture was stirred for 1 h at room temperature, diluted with CH_2Cl_2 (10 mL), and washed with saturated NaHCO₃ solution (5 mL). The organic layer was dried over MgSO₄ and concentrated under reduced pressure. The crude aldehyde was directly used for the next step.

To a stirred solution of crude aldehyde in Et_2O (10 mL) was added dropwise C_2H_5MgBr (1.7 mL of 1 M solution, 1.68 mmol) at 0 °C. The mixture was stirred for 1 h at 0 °C and quenched with

saturated NH₄Cl solution (5 mL). Organic layer was extracted with EtOAc (3 x 10 mL). The combined organic layer was dried over MgSO₄, concentrated under reduced pressure to give crude alcohol which was directly used for the next step without further purification.

To a stirred solution of crude alcohol in CH₂Cl₂ (5 mL) were added NaHCO₃ (70 mg, 0.84 mmol) and Dess-Martin periodinane (284 mg, 0.67 mmol) at 0 °C. The mixture was stirred for 1 h at room temperature and then diluted with CH₂Cl₂ (10 mL), and washed with saturated NaHCO₃ solution (5 mL). The organic layer was washed with brine (5 mL), dried over MgSO₄, and concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford a mixture (200 mg, 68% over three steps) of compound **47** and TBS-ether eliminated by-product as a colorless liquid. $R_f = 0.3$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 6.75 (d, J = 2.1 Hz, 2H), 6.62 (t, J = 2.1 Hz, 1H), 5.14 (dd, J = 11.1, 6.7 Hz, 4H), 4.64 (td, J = 16.5, 7.9, 3.3 Hz, 1H), 4.34 (d, J = 10.3 Hz, 1H), 3.56-3.46 (m, 1H), 3.45 (s, 6H), 2.69-2.59 (m, 1H), 2.56-2.36 (m, 2H), 1.86-1.58 (m, 3H), 1.55-1.40 (m, 8H), 1.50 (s, 3H), 1.44 (s, 3H), 1.08-1.00 (m, 8H), 0.95 (d, J = 6.6 Hz, 3H), 0.89-0.88 (m, 12H), 0.86 (d, J = 6.6 Hz, 3H), 0.67 (d, J = 6.1 Hz, 3H), 0.47 (d, J = 6.1 HZ, 3H), 0.09 (s, 3H), 0.07 (s, 3H); LCMS (ESI): 723 (M + H)⁺.

(4*R*,6*R*,8*R*,11*R*,13*R*,14*R*)-14-((*R*)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-11,13-dihydroxy-4,6,8,16-tetramethylheptadecan-3-one (1a). To a stirred solution of 47 (100 mg, 0.14 mmol) in MeOH (5 mL) was added PTSA (5 mg, 0.04 mmol) at room temperature. The mixture was stirred for 4 h at the same temperature and quenched with saturated NaHCO₃ solution (4 mL). The organic layer was extracted with CH_2Cl_2 (3 x 10 mL). The combined organic layer was washed with brine (5 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to separate compound **1a** (52 mg, 91%

combined yield) and 48 (8 mg) as a white solid. Spectroscopic data for compound 48 has given later. $R_f = 0.4 (5\% \text{ MeOH/CH}_2\text{Cl}_2); [\alpha]_D^{24} = +10.0 (c \ 0.40, \text{MeOH}); \text{IR (KBr)}: 3346, 2925, 1699,$ 1605, 1458, 1378, 1339, 1102, 844, 668, 657, 613 cm⁻¹; ¹H NMR (900 MHz, CD₃OD): δ 6.33 (d, J = 2.2 Hz, 2H), 6.15 (t, J = 2.2 Hz, 1H), 4.47 (d, J = 4.0 Hz, 1H), 4.01 (ddd, J = 9.5, 4.0, 3.5 Hz, 1H), 3.69 (dddd, J = 11.0, 8.5, 4.5, 4.0 Hz, 1H), 2.76 (ddq, J = 8.5, 6.5, 5.0 Hz, 1H), 2.57-2.49 11.0, 8.5, 5.0 Hz, 1H), 1.55 (ddd, J = 11.0, 9.5, 8.5 Hz, 1H), 1.53 (ddddq, J = 8.5, 8.5, 6.6, 6.5, 3.0 Hz, 1H) 1.42 (dddqd, J = 8.5, 8.5, 6.6, 6.5, 5.0 Hz, 1H) 1.41 (dddd, J = 11.5, 11.0, 8.5, 5.0 Hz) Hz, 1H), 1.39 (dddd, J = 11.5, 5.0, 4.0, 3.0 Hz, 1H), 1.38 (m, 1H), 1.33 (dddd, J = 11.0, 5.0, 3.0, 3.0 Hz, 1H), 1.22 (ddd, J = 11.0, 8.5, 8.5 Hz, 1H), 1.21 (m, 2H), 1.19 (dddd, J = 11.0, 8.5, 5.03.0 Hz, 1H), 1.06 (d, J = 6.5 Hz, 3H), 1.02 (t, J = 7.5 Hz, 3H), 0.99 (ddd, J = 11.0, 8.5, 5.0 Hz, 1H), 0.95 (ddd, J = 11.0, 6.6, 6.6 Hz, 1H), 0.88 (d, J = 6.5 Hz, 3H), 0.86 (d, J = 6.5 Hz, 3H), 0.83 (d, J = 6.5 Hz, 3H), 0.77 (d, J = 6.5 Hz, 3H); ¹³C NMR (225 MHz, CD₃OD): δ 218.3, 159.1, 148.3, 105.9, 101.9, 76.5, 73.4, 72.5, 48.5, 46.3, 44.6, 41.7, 40.7, 37.3, 35.4, 35.1, 33.1, 30.9, 29.2, 26.6, 23.3, 22.5, 20.5, 20.3, 17.9, 7.8; HRMS (ESI): calcd. for $C_{28}H_{48}O_6Na [M + Na]^+$ 503.3349; found 503.3336.

(4R,6R,8R,13R,14R)-14-((R)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-13-hydroxy-4,6,8,16tetramethylheptadecan-3-one (48). R_f = 0.5 (5% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ 6.39 (brs, 2H), 6.24 (brs, 1H), 4.52-4.46 (m, 1H), 3.67 (d, J = 3.8 Hz, 1H), 2.77-2.57 (m, 1H), 2.55-2.37 (m, 2H), 1.86-1.64 (m, 4H), 1.48-1.28 (m, 9H), 1.11-0.97 (m, 10H), 0.84 (d, J = 6.5 Hz, 3H), 0.83 (d, J = 6.5 Hz, 3H), 0.82 (d, J = 6.8 Hz, 3H), 0.78 (d, J = 6.6 Hz, 3H), 0.67 (t, J = 6.5 Hz, 3H); LCMS (ESI): 487 (M + Na)⁺. (2S,5R,7S,9S)-10-(Benzyloxy)-1-((4R,5R,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-

isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-5,7,9-trimethyldec-3-yn-2-ol (49). The procedure for the preparation of 49 was same as that for the preparation of 44. Alkyne 39 (300 mg, 1.16 mmol) and aldehyde 34 (317 mg, 0.77 mmol) were used to afford compound 49 (380 mg, 75%) as a colorless liquid. $R_f = 0.4$ (30% EtOAc/hexane); IR (KBr): 2956, 2927, 1589, 1455, 1379, 1263, 1143, 1030, 925, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.33-7.23 (m, 5H), 6.74 (d, J = 2.1 Hz, 2H), 6.64 (t, J = 2.1 Hz, 1H), 5.13 (dd, J = 10.1, 6.6 Hz, 4H), 4.61 (t, J = 6.1 Hz, 1H), 4.49 (s, 2H), 4.36 (d, J = 10.3 Hz, 1H), 3.85 (t, J = 10.1 Hz, 1H), 3.44 (s, 6H), 3.37-3.22 (m, 2H), 2.78 (brs, 1H), 2.63-2.48 (m, 1H), 1.99-1.76 (m, 4H), 1.52 (s, 3H), 1.44 (s, 3H), 1.44-1.28 (m, 3H), 1.16 (d, J = 6.6 Hz, 3H), 1.07-0.94 (m, 2H), 0.96 (d, J = 6.7 Hz, 3H), 0.95 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.6 Hz, 3H), 0.68 (d, J = 5.7 Hz, 3H), 0.48 (d, J = 5.7 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 142.6, 138.7, 128.2, 127.3, 127.3, 109.4, 104.6, 98.5, 94.3, 89.5, 81.0, 78.3, 76.2, 73.6, 72.9, 61.4, 55.7, 43.7, 43.5, 41.7, 41.5, 36.3, 30.8, 30.0, 28.3, 28.2, 26.7, 23.7, 22.6, 22.4, 21.8, 20.2, 19.5, 17.5; HRMS (ESI): calcd. for C₄₀H₆₀O₈Na [M + Na]⁺ 691.4186; found 691.4185.

(((2S,5R,7S,9S)-10-(Benzyloxy)-1-((4R,5R,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-

isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-5,7,9-trimethyldec-3-yn-2-yl)oxy)(tert-

butyl)dimethylsilane (50). The procedure for the preparation of **50** was same as that for the preparation of **45**. Alcohol **49** (350 mg, 0.52 mmol) was used to deliver compound **50** (340 mg, 83%) as a colorless liquid. $R_f = 0.7$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 6.76 (d, J = 2.2 Hz, 2H), 6.63 (t, J = 2.2 Hz, 1H), 5.15 (dd, J = 12.2, 6.7 Hz, 4H), 4.67-4.61 (m, 1H), 4.50 (d, J = 3.3 Hz, 2H), 4.38 (dd, J = 10.3, 3.6 Hz, 1H), 3.82 (t, J = 10.3 Hz, 1H), 3.45 (s, 6H), 3.42-3.30 (m, 1H), 3.28-3.20 (m, 1H), 2.63-2.47 (m, 1H), 1.98-1.78 (m, 3H),

1.78-1.67 (m, 1H), 1.51 (s, 3H), 1.43 (s, 3H), 1.40-1.29 (m, 2H), 1.17 (d, J = 6.8 Hz, 3H), 1.09-0.98 (m, 3H), 0.98 (d, J = 6.6 Hz, 3H), 0.94-0.89 (m, 15H), 0.69 (d, J = 5.7 Hz, 3H), 0.47 (d, J = 5.7 Hz, 3H), 0.13 (d, J = 4.9 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 142.9, 138.8, 128.3, 127.4, 127.3, 109.4, 104.6, 98.4, 94.3, 89.2, 81.5, 78.5, 76.3, 72.9, 71.7, 60.9, 55.8, 43.5, 43.3, 42.5, 41.8, 41.5, 36.3, 30.8, 30.0, 28.2, 26.7, 25.9, 23.6, 22.7, 22.4, 22.0, 21.1, 20.7, 20.1, 19.5, 18.3, 17.6, -4.5, -4.8; HRMS (ESI): calcd. for C₄₆H₇₄O₈SiNa [M + Na]⁺ 805.5051; found 805.5067.

(4*S*,6*S*,8*S*,11*R*,13*R*,14*R*)-14-((*R*)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-11,13-dihydroxy-4,6,8,16-tetramethylheptadecan-3-one (1b). The procedure for the preparation of 1b was same as that for the preparation of 1a from 45. Compound 50 (300 mg, 0.38 mmol) was used to synthesize compound **1b** (55 mg, 41% combined yield over five steps). Compound **51** was isolated (10 mg) as a white solid. Spectroscopic data for compound 51 has given later. $R_f = 0.4$ $(5\% \text{ MeOH/CH}_2\text{Cl}_2); [\alpha]_D^{24} = +27.2 (c \ 0.4, \text{ MeOH}); \text{ IR (KBr): } 3345, 2925, 1699, 1605, 1458,$ 1378, 1102, 844, 657, 613 cm⁻¹; ¹H NMR (600 MHz, CD₃OD): δ 6.33 (d, J = 2.2 Hz, 2H), 6.15 (t, J = 2.2 Hz, 1H), 4.47 (d, J = 4.0 Hz, 1H), 4.00 (m, 1H), 3.69 (m, 1H), 2.75 (m, 1H), 2.57-2.49 (m, 2H), 2H), 1.87 (m, 1H), 1.79 (m, 1H), 1.72 (ddd, J = 11.0, 8.5, 5.0 Hz, 1H), 1.52-1.50 (m, 2H), 1.47-1.46 (m, 2H), 1.41-1.33 (m, 3H), 1.23-1.21 (m, 3H), 1.05 (d, J = 6.5 Hz, 3H), 1.04 (m, 1H), 1.01 (t, J = 7.5 Hz, 3H), 0.98 (m, 1H), 0.93 (m, 1H), 0.87 (d, J = 6.5 Hz, 3H), 0.85 (d, J = 6.5 Hz, 3H)3H), 0.82 (d, J = 6.5 Hz, 3H), 0.76 (d, J = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CD₃OD): δ 218.4, 159.2, 148.2, 106.1, 102.0, 76.6, 73.5, 72.9, 48.7, 46.3, 44.6, 41.6, 40.8, 37.4, 35.5, 35.3, 33.3, 31.2, 29.2, 26.7, 23.4, 22.6, 20.6, 20.0, 17.8, 7.8; HRMS (ESI): calcd. for $C_{28}H_{48}O_6Na[M + Na]^+$ 503.3349; found 503.3336.

(4*S*,6*S*,8*S*,13*R*,14*R*)-14-((*R*)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-13-hydroxy-4,6,8,16tetramethylheptadecan-3-one (51). $R_f = 0.4$ (5% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, MeOD): $\delta 6.35$ (d, J = 2.1 Hz, 2H), 6.17 (t, J = 2.1 Hz, 1H), 4.46 (d, J = 7.3 Hz, 1H), 3.78-3.76 (m, 1H), 2.82-2.65 (m, 1H), 2.63-2.44 (m, 2H), 1.89-1.82 (m, 1H), 1.78-1.68 (m, 1H), 1.68-1.47 (m, 3H), 1.47-1.19 (m, 10 H), 1.15 (t, J = 6.3 Hz, 3H), 1.11-1.00 (m, 8H), 0.89 (t, J = 6.3 Hz, 3H), 0.88 (t, J = 6.7 Hz, 3H), 0.84 (t, J = 6.5 Hz, 3H), 0.73 (d, J = 6.5 Hz, 3H); HRMS (ESI): calcd. for C₂₈H₄₈O₅Na [M + Na]⁺ 487.3399; found 487.3399.

((Oct-7-yn-1-yloxy)methyl)benzene (52). To a stirred solution of NaH (336 mg, 8.4 mmol) in THF:DMF (3:1, 20 mL) was added oct-7-yn-1-ol (890 mg, 7.0 mmol) in THF (20 mL) at 0 °C. The mixture was stirred for 1 h at room temperature and cooled to 0 °C. The mixture was treated dropwise with BnBr (0.99 mL, 8.4 mmol), stirred for 3 h at room temperature, and quenched with saturated NH₄Cl solution (10 mL). The organic layer was extracted with EtOAc (3 x 10 mL). The combined organic layer was washed with brine (5 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford compound **52** (1.4 g, 92%) as a colorless liquid. $R_f = 0.8$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 4.51 (s, 2H), 3.47 (t, *J* = 6.6 Hz, 2H), 2.19 (td, *J* = 7.0, 2.6 Hz, 2H), 1.94 (t, *J* = 2.6 Hz, 1H), 1.67-1.59 (m, 2H), 1.58-1.46 (m, 2H), 1.46-1.34 (m, 4H); LCMS (ESI): 217 (M + H)⁺.

(S)-10-(Benzyloxy)-1-((4R,5S,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-

dimethyl-1,3-dioxan-4-yl)dec-3-yn-2-ol (53). The procedure for the preparation of 53 was same as that for the preparation of 44 and 21. Alkyne 52 (210 mg, 0.97 mmol) and aldehyde 12a (200 mg, 0.49 mmol) were used to afford compound 53 (226 mg, 74%) as a colorless liquid. $R_f = 0.4$ (20% EtOAc/hexane); $[\alpha]_D^{24} = +24.7$ (*c* 0.49, CHCl₃); IR (KBr): 3446, 2934, 2864, 1653, 1598, 1457, 1381, 1264, 1143, 1084, 1030, 927, 697 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.31 (m, 4H), 7.31-7.26 (m, 1H), 6.66 (d, J = 2.2 Hz, 2H), 6.59 (t, J = 2.2 Hz, 1H), 5.14 (s, 4H), 5.03-5.02 (m, 1H), 4.63-4.57 (m, 1H), 4.49 (s, 2H), 4.36-4.29 (m, 1H), 3.46 (t, J = 6.4 Hz, 2H), 3.45 (s, 6H), 2.76-2.75 (m, 1H), 2.23 (dt, J = 7.0, 1.9 Hz, 2H), 2.11-2.04 (m, 1H), 1.68-1.59 (m, 4H), 1.57-1.49 (m, 3H), 1.49 (s, 3H), 1.46 (s, 3H), 1.41-1.37 (m, 3H), 1.17-1.06 (m, 1H), 0.92-0.82 (m, 1H), 0.70 (d, J = 6.4 Hz, 3H), 0.45 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 158.0, 143.4, 138.6, 128.3, 127.6, 127.4, 107.1, 103.2, 99.4, 94.3, 85.5, 80.7, 74.9, 73.3, 72.8, 70.3, 61.8, 60.4, 55.9, 41.4, 41.3, 30.4, 29.8, 29.6, 28.7, 28.6, 27.4, 25.7, 22.8, 22.1, 19.5, 18.7, 14.2; HRMS (ESI): calcd. for C₃₇H₅₄O₈Na [M + Na]⁺ 649.3716; found 649.3734.

(((S)-10-(Benzyloxy)-1-((4R,5S,6R)-6-(3,5-bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-

dimethyl-1,3-dioxan-4-yl)dec-3-yn-2-yl)oxy)(*tert*-butyl)dimethylsilane (54). The procedure for the preparation of 54 was same as that for the preparation of 45 and 22. Alcohol 53 (150 mg, 0.24 mmol) was used to afford compound 54 (140 mg, 81%) as a colorless liquid. $R_f = 0.8$ (10% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 7.36-7.26 (m, 5H), 6.68 (d, J = 2.1 Hz, 2H), 6.59 (t, J = 2.1 Hz, 1H), 5.15 (s, 4H), 5.01 (brs, 1H), 4.54-4.49 (m, 1H), 4.49 (s, 2H), 4.29 (d, J =8.1 Hz, 1H), 3.48-3.45 (m, 2H), 3.45 (s, 6H), 2.23 (td, J = 6.9, 1.7 Hz, 2H), 1.96-1.89 (m, 1H), 1.67-1.57 (m, 2H), 1.56-1.47 (m, 4H), 1.46 (d, J = 1.9 Hz, 6H), 1.46-1.34 (m, 4H), 1.31-1.06 (m, 2H), 0.92 (s, 9H), 0.89-0.75 (m, 1H), 0.67 (d, J = 6.5 Hz, 3H), 0.45 (d, J = 6.5 Hz, 3H), 0.14 (d, J =5.7 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 157.9, 143.8, 138.6, 128.3, 127.5, 127.4, 107.2, 103.1, 99.2, 94.3, 85.1, 81.3, 75.0, 72.8, 71.0, 70.3, 60.9, 55.8, 42.3, 41.1, 30.5, 29.8, 29.7, 28.7, 28.6, 27.3, 25.8, 25.7, 25.6, 22.7, 22.2, 19.4, 18.6, 18.2, -4.4, -4.9; LCMS (ESI): 741 (M + H)⁺. (*R*)-10-((4*R*,5*S*,6*R*)-6-(3,5-Bis(methoxymethoxy)phenyl)-5-isobutyl-2,2-dimethyl-1,3-dioxan-4-yl)-9-((*tert*-butyldimethylsilyl)oxy)decan-1-ol (56). The procedure for the preparation of 56 was same as that for the preparation of **23**. Compound **54** (86 mg, 0.16 mmol) was used to afford compound **56** (140 mg, 81%) as a colorless liquid. $R_f = 0.4$ (20% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃): δ 6.66 (d, J = 1.9 Hz, 2H), 6.58 (t, J = 1.9 Hz, 1H), 5.14 (s, 4H), 4.97 (brs, 1H), 4.22 (t, J = 6.1 Hz, 1H), 3.79 (t, J = 5.7 Hz, 1H), 3.63 (t, J = 6.7 Hz, 2H), 3.45 (s, 6H), 1.77-1.62 (m, 1H), 1.62-1.46 (m, 5H), 1.46 (s, 6H), 1.34-1.21 (m, 12H), 1.18-1.11 (m, 2H), 0.92 (s, 9H), 0.83-0.72 (m, 1H), 0.67 (d, J = 6.3 Hz, 3H), 0.45 (d, J = 6.3 Hz, 3H), 0.10 (s, 3H), 0.07 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 157.9, 143.9, 107.3, 103.1, 99.1, 94.4, 75.1, 70.7, 69.0, 63.0, 55.9, 40.8, 40.3, 37.2, 37.8, 30.3, 29.9, 29.7, 29.5, 29.3, 27.3, 25.9, 25.7, 24.9, 22.8, 22.3, 19.5, 18.1, -4.2, -4.5; LCMS (ESI): 655 (M + H)⁺.

(11R,13R,14S)-14-((R)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-11,13-dihydroxy-16-

methylheptadecan-3-one (57). The procedure for the preparation of 57 was same as that for the preparation of 1a from 46. Compound 56 (120 mg, 0.18 mmol) was used to prepare compound 57 (48 mg, 61% over 4 steps). $R_f = 0.3$ (5% MeOH/CH₂Cl₂); $[\alpha]_D^{24} = +5.8$ (*c* 0.26, CHCl₃); IR (KBr): 3346, 2924, 2852, 1701, 1602, 1461, 1261, 1153, 1031, 668 cm⁻¹; ¹H NMR (400 MHz, CD₃OD): δ 6.25 (d, *J* = 2.2 Hz, 2H), 6.05 (t, *J* = 2.2 Hz, 1H), 4.75 (d, *J* = 3.5 Hz, 1H), 3.92-3.88 (m, 1H), 3.68-3.61 (m, 1H), 2.41-2.35 (m, 4H), 1.63-1.52 (m, 3H), 1.52-1.36 (m, 3H), 1.39-1.27 (m, 5H), 1.27-1.11 (m, 9H), 0.93 (t, *J* = 7.3 Hz, 3H), 0.68 (d, *J* = 6.1 Hz, 3H), 0.56 (d, *J* = 6.1 Hz, 3H); ¹³C NMR (100 MHz, CD₃OD): δ 214.8, 159.3, 148.6, 105.7, 102.0, 76.8, 75.1, 72.0, 49.3, 43.1, 42.3, 38.6, 36.6, 33.4, 30.7, 30.5, 30.3, 28.3, 26.4, 25.0, 23.4, 23.0, 8.1; HRMS (ESI): calcd. for C₂₅H₄₂O₆Na [M + Na]⁺ 461.2879; found 461.2866.

(1R,2S,3R,5S)-13-(Benzyloxy)-1-(3,5-dihydroxyphenyl)-2-isobutyltridec-6-yne-1,3,5-triol

(55). To a stirred solution of 53 (50 mg, 0.08 mmol) in MeOH (2 mL) was added PTSA (cat.) at room temperature. The mixture was stirred for 4 h at room temperature and then quenched with

saturated NaHCO₃ solution (2 mL). The organic layer was extracted with CH₂Cl₂ (3 x 5 mL). The combined organic layer was washed with brine (2 mL), dried over MgSO₄, concentrated under reduced pressure. The residue was subjected to silica gel column chromatography to afford **55** (32 mg, 84%) as a colorless liquid. $R_f = 0.6$ (5% MeOH/CH₂Cl₂); [α]_D²⁴ = +18.2 (*c* 1.5, CHCl₃); ¹H NMR (400 MHz, MeOD): δ 7.34-7.29 (m, 4H), 7.30-7.24 (m, 1H), 6.33 (d, *J* = 2.0 Hz, 2H), 6.14 (t, *J* = 2.0 Hz, 1H), 4.83 (d, *J* = 3.6 Hz, 1H), 4.49-4.45 (m, 3H), 4.01-3.98 (m, 1H), 3.48 (t, *J* = 6.4 Hz, 2H), 2.19 (td, *J* = 6.6, 1.7 Hz, 2H), 2.00-1.93 (m, 1H), 1.78-1.69 (m, 2H), 1.64-1.57 (m, 2H), 1.51-1.27 (m, 9H), 0.80 (d, *J* = 6.1 Hz, 3H), 0.67 (d, *J* = 6.1 Hz, 3H); ¹³C NMR (100 MHz, MeOD): δ 157.8, 147.4, 138.4, 128.0, 127.5, 127.3, 104.3, 100.5, 84.6, 80.9, 74.7, 72.5, 71.6, 70.0, 60.5, 48.5, 42.8, 32.3, 29.3, 28.4, 28.3, 26.6, 25.4, 22.2, 21.4, 17.9; HRMS (ESI): calcd. for C₃₀H₄2O₆Na [M + Na]⁺ 521.2879; found 521.2876.

(1R,2S,3R,5S,8S,10S,12R)-13-(Benzyloxy)-1-(3,5-dihydroxyphenyl)-2-isobutyl-8,10,12-

trimethyltridec-6-yne-1,3,5-triol (58). The procedure for the preparation of 58 was same as that for the preparation of 55. MOM-protected 21a (20 mg, 0.03 mmol) was used to afford compound 58 (12 mg, 85%) as a colorless liquid. $R_f = 0.5$ (5% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, MeOD): δ 7.33-7.32 (m, 4H); 7.28-7.22 (m, 1H), 6.34-6.32 (m, 2H), 6.17-6.14 (m, 1H), 4.90-4.87 (m, 1H), 4.51-4.44 (m, 3H), 4.04-4.01 (m, 1H), 3.35-3.21 (m, 2H), 2.60-2.46 (m, 1H), 2.06-1.93 (m, 1H), 1.92-1.72 (m, 4H), 1.45-1.22 (m, 5H), 1.17-1.14 (m, 2H), 1.11 (d, *J* = 6.8 Hz, 3H), 0.94 (d, *J* = 6.8 Hz, 3H), 0.90 (d, *J* = 6.4 Hz, 3H), 0.80 (d, *J* = 5.7 Hz, 3H), 0.65 (d, *J* = 5.7 Hz, 3H); ¹³C NMR (100 MHz, MeOD): δ 159.2, 148.7, 139.8, 129.3, 128.8, 128.5, 105.6, 101.9, 90.5, 82.5, 77.6, 75.9, 73.9, 73.1, 61.8, 49.7, 49.6, 48.3, 46.6, 44.1, 41.4, 33.5, 32.0, 29.1, 27.9, 24.6, 23.6, 22.7, 21.7, 20.3, 17.3; HRMS (ESI): calcd. for C₃₃H₄₈O₆Na [M + Na]⁺ 563.3349; found 563.3363.

(1*R*,2*S*,3*R*,5*R*,8*R*,10*S*,12*R*)-1-(3,5-Dihydroxyphenyl)-2-isobutyl-8,10,12-trimethyltridecane-1,3,5,13-tetraol (59). The procedure for the preparation of 59 was same as that for the preparation of 55. MOM-protected 23a (20 mg, 0.03 mmol) was used to afford compound 59 (10.5 mg, 82%) as a white solid. $R_f = 0.4$ (10% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, MeOD): δ 6.25 (d, J = 1.9 Hz, 2H), 6.04 (t, J = 1.9 Hz, 1H), 4.74 (d, J = 2.7 Hz, 1H), 3.90-3.88 (m, 1H), 3.63-3.59 (m, 1H), 3.38-3.22 (m, 2H), 1.67-1.48 (m, 6H), 1.48-1.24 (m, 6H), 1.24-0.88 (m, 7H), 0.79 (d, J = 6.7 Hz, 3H), 0.78 (d, J = 6.7 Hz, 3H), 0.76 (d, J = 6.6 Hz, 3H), 0.70 (d, J = 5.8 Hz, 3H), 0.56 (d, J = 5.8 Hz, 3H); ¹³C NMR (100 MHz, MeOD): δ 159.8, 149.1, 106.2, 102.4, 77.3, 75.5, 72.8, 69.6, 49.7, 47.5, 43.2, 42.8, 42.3, 36.5, 34.8, 33.9, 31.8, 29.0, 28.8, 23.9, 23.4, 20.4, 20.2, 17.5; HRMS (ESI): calcd. for C₂₆H₄₆O₆Na [M + Na]⁺ 477.3192; found 477.3187.

(1R,2S,3R,8R,10S,12R)-1-(3,5-Dihydroxyphenyl)-2-isobutyl-8,10,12-trimethyltridecane-

1,3,13-triol (60). The procedure for the preparation of **60** was same as that for the preparation of **55**. MOM-protected TBS-ether eliminated **23'** (20 mg, 0.03 mmol) was used to afford compound **60** (10.6 mg, 81%) as a viscous liquid. $R_f = 0.4$ (10% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, MeOD): δ 6.33 (d, J = 2.2 Hz, 2H), 6.14 (t, J = 2.2 Hz, 1H), 4.78 (d, J = 3.7 Hz, 1H), 3.74-3.71 (m, 1H), 3.42-3.35 (m, 1H), 3.31-3.27 (m, 1H), 1.75-1.58 (m, 3H), 1.58-1.46 (m, 4H), 1.46-1.31 (m, 5H), 1.21-0.94 (m, 6H), 0.94-0.83 (m, 12H), 0.77 (d, J = 6.1 Hz, 3H), 0.64 (d, J = 6.1 Hz, 3H); ¹³C NMR (100 MHz, CD₃OD): δ 159.2, 148.4, 105.7, 101.9, 77.7, 76.1, 69.0, 48.8, 42.7, 38.8, 36.0, 34.2, 33.3, 32.7, 31.1, 31.0, 28.5, 27.6, 23.6, 23.2, 23.0, 19.7, 17.0, 14.4; HRMS (ESI): calcd. for C₂₆H₄₆O₅Na [M + Na]⁺ 461.3243; found 461.3230.

(4*R*,6*S*,8*R*,13*R*,14*S*)-14-((*R*)-(3,5-Dihydroxyphenyl)(hydroxy)methyl)-13-hydroxy-4,6,8,16tetramethylheptadecan-3-one (61). The procedure for the preparation of 61 was same as that for the preparation of 1a from 46. MOM-protected TBS-ether eliminated 23' (20 mg, 0.04 mmol)

was used to afford compound **61** (11.3 mg, 61% over four steps) as a viscous liquid. $R_f = 0.5$ (5% MeOH/CH₂Cl₂); ¹H NMR (400 MHz, CD₃OD): δ 6.25 (d, J = 2.2 Hz, 2H), 6.06 (t, J = 2.2 Hz, 1H), 4.70 (d, J = 2.9 Hz, 1H), 3.67 (m, 1H), 2.73-2.55 (m, 1H), 2.52-2.35 (m, 2H), 1.62-1.42 (m, 1H), 1.42-1.28 (m, 8H), 1.27-1.09 (m, 8H), 1.03-0.97 (m, 3H), 0.97 (d, J = 6.8 Hz, 3H), 0.95 (d, J = 6.1 Hz, 3H), 0.78 (d, J = 6.6 Hz, 3H), 0.76 (t, J = 6.5 Hz, 3H), 0.70 (d, J = 5.8 Hz, 3H), 0.56 (d, J = 5.8 Hz, 3H); ¹³C NMR (100 MHz, CD₃OD): δ 218.3, 159.2, 148.4, 105.7, 101.9, 77.7, 76.0, 48.8, 46.1, 44.9, 42.3, 38.9, 36.0, 35.1, 33.4, 31.1, 29.2, 28.5, 27.6, 23.3, 23.1, 19.8, 17.0, 8.1; HRMS (ESI): calcd. for C₂₈H₄₈O₅Na [M + Na]⁺ 487.3399; found 487.3397.

Isolation and purification of synthetic baulamycin A products (1, 1a, and 1b): The crude product including **1** was injected to a semi-preparative reversed-phase HPLC column (Kromasil C18: 250×10 mm, 5 µm) with a gradient solvent system (35% acetonitrile/water to 75% acetonitrile/water over 40 min, UV 280 nm detection, flow rate: 2 mL/min). The major peak at the retention time of 36.0 min was obtained as pure synthetic baulamycin A (proposed structure, **1**). Similarly, the crude product containing **1a** was subjected to a semi-preparative reversed-phase HPLC column (Kromasil C18: 250×10 mm, 5 µm) with a gradient solvent system (35% acetonitrile/water to 75% acetonitrile/water over 40 min, UV 280 nm detection, flow rate: 2 mL/min). An HPLC peak bearing **1a** eluted at the retention time of 36.5 min. To remove impurities, this compound was further purified under isocratic solvent conditions (35% acetonitrile/water, UV 280 nm detection, flow rate: 2 mL/min) using a chiral column (YMC cellulose-SC: 250×4.6 mm, 5 µm). **1a** eluted as pure compound at the retention time of 20.5 min. The purification of **1b** was performed in the same manner using the combination of C18 and chiral HPLC columns (retention time through the final chiral HPLC: 20.3 min).

Experimental procedure for assessing inhibitory-activities on SbnE.

For evaluation of the inhibitory-activities of baulamycin derivatives on SbnE, a modified malachite green assay was employed. To an Eppendorf tube containing 25 mM HEPES, 5 mM MgCl₂, 100 µM ATP, 100 µM sodium citrate, 100 µM L-2,3-diaminopropionic acid, and 0.001 U/µL inorganic pyrophosphatase (IPP) was added a synthesized baulamycin derivative dissolved in DMSO to a final concentration of 46 nM, 137 nM, 412 nM, 1.23 µM, 3.70 µM, 11.1 µM, 33.3 μ M, or 100 μ M (for negative control, a DMSO vehicle was used). Each reaction was then initiated by addition of SbnE (25 nM) at 37 °C. The total reaction volume was 100 μ L. The reaction mixture was incubated for 1 h before addition of 25 µL quenching solution composed of 50 parts of malachite green solution, 12.5 parts of 7.5% ammonium molybdate, and 1 part of 11% Tween-20 solution. After additional incubation for 15 min at 37 °C, 100 µL aliquot of each mixture was loaded in a 96-well clear bottom plate and the absorbance at 630 nm was measured using a Hidex Sense microplate reader (Hidex, Finland). For positive control, a reaction containing all the same components except for SbnE was conducted. All data were collected in duplicates. The observed absorbance values were converted to the % inhibition, and then the resulting % inhibition vs log[compound] plots were fitted using GraphPad Prism version 7 to calculate the IC_{50} values (Figure S165).

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Supporting Information. Spectra for all compounds. This material is available free of charge via the Internet at ACS Publications website.

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