

Bioorganic & Medicinal Chemistry Letters 12 (2002) 461-464

Discovery of 5-Hydroxyalkyl-4-phenylpyridines as a New Class of Glucagon Receptor Antagonists

Gaetan H. Ladouceur,^{a,*} James H. Cook,^a Elizabeth M. Doherty,^a William R. Schoen,^a Margit L. MacDougall^b and James N. Livingston^b

^aDepartment of Chemistry Research, Bayer Research Center, 400 Morgan Lane, West Haven, CT 06516, USA ^bDepartment of Diabetes and Obesity Research, Bayer Research Center, 400 Morgan Lane, West Haven, CT 06516, USA

Received 27 September 2001; accepted 16 November 2001

Abstract—5-Hydroxyalkyl-4-phenylpyridines have been identified as a novel class of glucagon antagonists with potential utility for the treatment of diabetes. A lead structure with moderate activity was discovered through a high throughput screening assay. Structure–activity relationships led to the discovery of a potent antagonist, $IC_{50} = 0.11 \mu M$, more than 60-fold improvement over the lead structure. © 2002 Elsevier Science Ltd. All rights reserved.

Glucagon is a 29 amino acid peptidic hormone secreted by α -cells in pancreatic islets. The glucagon receptor is a member of the G-protein coupled receptor family which signals through the adenylate cyclase pathway by increasing cAMP levels in hepatocytes.^{1,2} Elevated intracellular cAMP levels stimulate liver glycogenolysis and gluconeogenesis, resulting in an increase of hepatic glucose production.³ A sustained high concentration of glucagon in the plasma is believed to be an important cause of excessive hepatic glucose production in diabetic subjects.^{4,5} This alteration contributes to the elevation in blood glucose, the underlying cause of morbidity and mortality in diabetes.

Numerous modifications in the amino acid sequence of glucagon led to the identification of several potent peptide antagonists and is the subject of recent reviews.^{6,7} Due to the poor oral bioavailability associated with peptides, their potential medical use has been limited. However, it was shown that some of the most potent peptide antagonists significantly decrease blood glucose levels in diabetic animal models when administered intravenously.^{8,9} Thus, this implied that a selective glucagon receptor antagonist could be useful for the treatment of diabetics, and consequently, prompted a strong interest from several research groups.^{10–19} Reviews covering the discovery of non-peptidic small molecule antagonists have been published.^{20,21} Our research efforts were thereby aimed at the identification of a lead structure capable of competitively altering glucagon binding to the human glucagon receptor (hGR). A high throughput screening assay resulted in the discovery of compound 1 (Fig. 1), which exhibited modest activities (binding affinity, $IC_{50}=7$ μ M and inhibition of cAMP production, $IC_{50}=2 \mu$ M). In this article, we describe the synthesis and the SAR of this new class of glucagon receptor antagonists. Compounds were tested in a binding assay using the human glucagon receptor²² and selected compounds were tested in a functional assay.²³

Most analogues were synthesized by following the general procedure described in Scheme 1, exemplified by the synthesis of compound 6.^{11,24} Several other analogues were subsequently made using compounds similar to 6 as a precursor. The synthesis of 6 started with the Hantzsch pyridine synthesis.²⁵ Condensation of enamine 3, obtained from ethyl isobutyrylacetate 2, with 4-fluorobenzaldehyde afforded pyridine 4 after oxidative treatment with DDQ (14%, three steps).



Figure 1. Lead structure found in a high throughput screening assay.

^{*}Corresponding author. Tel.: +1-203-812-2055; fax: +1-203-812-2650; e-mail: gaetan.ladouceur.b@bayer.com

⁰⁹⁶⁰⁻⁸⁹⁴X/02/\$ - see front matter \odot 2002 Elsevier Science Ltd. All rights reserved. P11: S0960-894X(01)00766-1

Selective reduction of one of the ester groups of pyridine **4** with Red-Al, followed by treatment with PCC, afforded aldehyde **5** (66%, two steps). A Wittig reaction of aldehyde **5** with the ylide of propyltriphenylphosphonium bromide provided the corresponding alkene ester, which was then successively reduced with LAH and catalytic hydrogenation, to afford alcohol **6**. Various ylides in the Wittig reaction could be used, leading to wide variation of alkyl groups at position 5 of the pyridine moiety (Tables 1 and 2).

A series of analogues with variation at position 5 of the pyridine moiety is listed in Table 1. We discovered that straight alkyl chains provided potent inhibitors with IC_{50} values in the sub-micromolar range. The optimum potency was obtained with alkyl chains from 3 to 7 carbon atoms in length. Addition of steric bulk by branching the alkyl group resulted in similar or decreased activity (e.g., 1 and 14). Likewise, less lipophilic groups attached to an alkyl chain, such as hydroxy (16) or ether (17), gave either no improvement or substantially decreased the activity.

We then investigated the SAR at positions 2 and 6 of the pyridine moiety, as seen in Table 2. To avoid the formation of regioisomers and to keep the synthesis relatively simple, we limited our study to symmetrical analogues. Smaller substituents were not favorable as compounds **19** to **22**, where \mathbb{R}^1 is methyl or ethyl, exhibited only weak activity. The diisopropyl-pyridines **23** and **24** were significantly more active with IC₅₀ values just above 1 μ M.

Steric bulk at this region of the molecule clearly increased the activity. It may also be important for the two isopropyl groups to shield the nitrogen atom of the pyridine moiety from a lipophilic area of the receptor. We were also interested in evaluating if the fluoro substituent found in the original lead structure (1) and our initial SAR (7 to 13) had any effect on binding affinity. This was addressed by comparison of 8 and 6, which are the corresponding *para*-fluorophenyl analogues of compounds 23 and 24. The compounds, 8 and 6, exhibited approximately a two-fold increase in activity over their des-fluoro counterparts.



Scheme 1. Synthesis of phenylpyridine 6. Reagents and conditions: (a) AcO–NH₄⁺, cyclohexane, $-H_2O$, 92%; (b) ethyl isobutyrylacetate, 4-fluorobenzaldehyde, 20%; (c) DDQ, CH₂Cl₂, 78%; (d) Red-Al, THF, 67%; (e) PCC, CH₂Cl₂, 99%; (f) *n*BuPPh₃Br, KO*t*Bu, THF, 90%; (g) LiAlH₄, THF, 80%; (h) H₂, Pd/C, EtOH, 98%.

SAR at position 3 of the pyridine moiety was then explored using compound 6 as an intermediate to generate new analogues (Scheme 2). The lack of significant activity found with the methyl ether 28, the acetate 27, the aldehyde 25, and the fluoromethyl derivative 30 suggested that a hydrogen donor group is required at that position. Attempts to replace the hydroxy group by other hydrogen donors also failed. Indeed, the thiol 29 and the carboxylic acid 27 exhibited no significant activities up to a concentration of 20 μ M.

The hydroxymethyl group at position 3 of the pyridine moiety was therefore the key pharmacophoric group for activity and became the focus of further investigation (Scheme 3). A breakthrough result was obtained with the discovery of compound **31**, which exhibited an IC₅₀ of 0.3 μ M, or a 2-fold increase of activity over its cor-

Table 1. SAR for position 5 of the pyridine moiety



Table 2. SAR for positions 2,6 of the pyridine moiety



				IC ₅₀ (µM)
19	Me	Et	Н	> 20
20	Me	Pent	Н	> 20
21	Et	Et	Н	> 20
22	Et	Pent	Н	20
23	iPr	Et	Н	1.8
24	iPr	Pent	Н	1.4
8	iPr	Et	F	1.0
6	iPr	Pent	F	0.7



Scheme 2. Synthesis of phenylpyridine analogues. Reagents and conditions: (a) PCC, CH_2Cl_2 , 74%; (b) Jones reagent, acetone, 58%; (c) AcCl, Et_3N , CH_2Cl_2 , 41%; (d) NaH, MeI, THF, 75%; (e) Lawesson's reagent, toluene, 26%; (f) DAST, CH_2Cl_2 , 76%.



Scheme 3. Synthesis of hydroxyalkylpyridine analogues. Reagents and conditions: (a) MeLi, THF, 70%; (b) EtMgBr, THF, 70%; (c) $CH_2=CHMgBr$, THF, 70%; (d) $CH_3OCH_2PPh_3Br$, KOtBu, THF, 60%; (e) HCl, THF, 50%; (f) NaBH₄, EtOH, 42%; (g) TMSCF₃, TBAF, THF, 70%.



Figure 2. Absolute configuration of 36a.

responding primary alcohol **6**. This analogue was synthesized by direct alkylation of aldehyde **25** with methyllithium to produce a racemic mixture of secondary alcohols. Larger secondary alcohols were thereafter explored (e.g., **32** and **33**), but resulted in a loss of activity, suggesting that the corresponding binding site of the receptor is relatively compact in that area. A transposition of the hydroxy group to the adjacent carbon atom (**34**) also resulted in a loss of activity. In an attempt to modify the acidity of the hydroxy group, the trifluoro isostere **35** was prepared.²⁶ It, however, exhibited a much weaker activity (IC₅₀ = 13.0 μ M) than its methyl isostere.

Since the racemic secondary alcohol **31** was the most active analogue at that point, it was resolved by chiral chromatography²⁷ to evaluate the activity of each enantiomer. We discovered that the faster eluting enantiomer was significantly more active (**31a**, $IC_{50}=0.15 \mu M$) than the slower enantiomer (**31b**, $IC_{50}=3.0 \mu M$).

 Table 3.
 Racemates versus most active enantiomers of secondary alcohols



Compd (racemate)	hGR IC ₅₀ (µM)	R	Compd (More potent enantiomer)	hGR IC ₅₀ (µM)
36	0.22	Pr	36a	0.11
37	0.25	Bu	-	-
31	0.30	Pent	31a	0.15
38	0.24	Hex	38a	0.12

This interesting result prompted us to evaluate a series of hydroxyethyl analogues bearing different alkyl chains at position 5 of the pyridine moiety (Table 3). In the listed examples, a similar enantiomeric preference was observed in which compound **36a** exhibited the best activity ($IC_{50}=0.11 \mu M$).

Finally, the absolute configuration of the secondary alcohol group was assigned by X-ray analysis of the Mosher ester derivative.²⁸ Thus, the secondary alcohol of compound **36a** was assigned the *R* configuration (Fig. 2) and by analogy, **31a** and **38a** were assumed to possess the same configuration. Compound **36a** was also evaluated for functional activity. In that assay, **36a** exhibited approximately the same activity (IC₅₀=0.065 μ M) as measured in the receptor binding assay.

In conclusion, SAR optimization of a lead structure found in a high throughput screening led to the discovery of new small molecule glucagon receptor antagonists with an approximately 70-fold increase of potency. Further optimization of this novel series will be reported in due course.

Acknowledgements

We are grateful to A. Paiva and L. Huang for efficient LC/MS analyses, D. Eustice for contributions to the initial screening work and A. Goehrt for X-ray analysis.

References and Notes

1. Dohlman, H. G.; Thorner, J.; Caron, M. G.; Lefkowitz,

- R. J. Annu. Rev. Biochem. 1991, 60, 653.
- 2. Christophe, J. Biochim. Biophys. Acta 1995, 1241, 45.
- 3. Unger, R. H.; Orci, L. New Engl. J. Med. 1981, 1518.
- 4. Gerich, J. E.; Lorenzi, M.; Bier, D.; Schneider, V.; Tsalikian,
- E.; Karam, J.; Forsham, P. H. New Engl. J. Med. 1975, 292, 985.
- 5. Unger, R. H. Metabolism 1978, 27, 1691.
- 6. Johnson, D. G.; Goebel, C. U.; Hruby, V. J.; Bergman,
- M. D.; Trivedi, D. Science 1982, 215, 1115.

- 7. Unson, C. G.; Andreau, D.; Gurzenda, E. M.; Merrifield,
- R. B. Proc. Natl. Acad. Sci. U.S.A. 1987, 84, 4083.
- 8. Unson, C. G.; Gurzenda, E. M.; Merrifield, R. B. Peptides 1989, 10, 1171.
- 9. Salzig, T. M.; Unson, C.; Merrifield, B. Diabetes 1996, 45, 220.
- 10. Collins, J. L.; Dambek, P. J.; Goldstein, S. W.; Faraci, W. S. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 915.
- 11. Schoen, W. R.; Ladouceur, G. H.; Cook, J. H.; Lease, T. G.; Wolanin, D. J.; Kramss, R. H.; Hertzog, D. L.; Osterhout, M. H. US Patent 6,218,431, 2001. *Chem. Abstr.*
- **2001**, *134*, 311111. 12. De Laszlo, S. E.; Chang, L. L.; Kim, D.; Mantlo, N. B.
- PCT Int. Appl. WO 97/16442, 1997. Chem. Abstr. 1997, 127, 50543.
- 13. Cascieri, M. A.; Koch, G. E.; Ber, E.; Sadowski, S. J.; Louizides, D.; de Laszlo, S. E.; Hacker, C.; Hagmann, W. K.; MacCoss, M.; Chicchi, G. G.; Vicario, P. P. J. Biol. Chem. **1999**, 274, 8694.
- 14. Azizeh, B. Y.; van Tine, B. A.; Sturm, N. S.; Hutzler, A. M.; David, C.; Trivedi, D.; Hruby, V. J. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1849.
- 15. Azizeh, B. Y.; Shenderovich, M. D.; Trivedi, D.; Li, G.; Sturm, N. S.; Hruby, V. J. J. Med. Chem. **1996**, *39*, 2449.
- 16. Ling, A.; Kuki, A.; Shi, S.; Plewe, M. B.; Feng, J.; Trues-
- dale, L. K.; May, J.; Kiel, D.; Madsen, P.; Sams, C.; Lau, J. PCT Int. Appl. WO 00/39088, 2000. *Chem. Abstr.* 2000, 133, 73951.
- 17. Madsen, P.; Knudsen, L. B.; Wiberg, F. C.; Carr, R. D. J. Med. Chem. 1998, 41, 5150.
- 18. Gonzales, J.; Sams, C.; Teng, M.; Ling, A.; Gregor, V.; Hong, Y.; Kiel, D.; Kuki, A; Shi, S.; Naerum, L.; Madsen, P.; Lau, J.; Plewe, M. B.; Feng, J.; Jonhson, M. D.; Teston, K. A.; Siedelmann, U. G.; Knudsen, L. B. PCT Int. Appl. WO 99/ 01423, 1999. *Chem. Abstr.* **1999**, *130*, 110061.
- 19. Ling, A.; Yufeng, H.; Gonzalez, J.; Gregor, V.; Polinsky, A.; Kuki, A.; Shi, S.; Teston, K.; Murphy, D.; Porter, J.; Kiel, D.; Lakis, J.; Anderes, K.; May, J. J. Med. Chem. 2001, 44, 3141.
- 20. Livingston, J. N.; Schoen, W. R. Annu. Rep. Med. Chem. 1999, 34, 189.
- 21. Zhang, B. B.; Moller, D. E. Curr. Opin. Chem. Biol. 2000, 4, 461.
- 22. Stable lines of hGR-CHO (Chinese hamster ovary) cells were prepared as previously described (Heurich, R.; Buggy, J.; Vandenberg, M.; Rossomando, A. *Biochem. Biophys. Res. Comm.* **1996**, 220, 905). Cells were grown in roller bottles then pelleted. The cell pellet was subjected to Polytron homo-

genization in a 4°C hypotonic lysis buffer that contained 10 mM Tris, pH 7.4, 2 mM EDTA, 5 mM MgCl₂, and 1 mM PMSF. Nuclei were removed by centrifugation at 800g for 15 min at 4°C. Membranes were collected by centrifugation of the supernate at 15,000g for 15 min. The membranes were washed, repelleted and suspended in 0.25 M sucrose, 10 mM Tris, pH 7.4, 5 mM EDTA and stored at -80 °C. The glucagon binding assay was carried out in 96-well filtration plates (1.2, glass fiber type C, Millipore) pretreated for 1 h with 0.3% polyethyleneimine. Each well contained 5 µg of membrane protein in 150 µL of binding buffer (20 mM Tris, pH 7.4, 1 mM EDTA, 1 mg/mL BSA, 1 mg/mL bacitracin). Five L of compound were added in duplicate at concentrations of 20, 2, 0.2, and 0.02 M final, followed by the addition of 9 fmol ¹²⁵Iglucagon (New England Nuclear) contained in 50 µL binding buffer. Control wells contained membranes, 0.2% DMSO, radiolabeled glucagon without and with excess native glucagon (1 µM) to establish nonspecific binding. The plates were incubated for 60 min at 21 °C, and filtered on the Millipore vacuum apparatus. Plates were washed three times with cold PBS/0.1% BSA, pH 7.4. The filters were punched into 12×75 tubes, and the membrane-associated radioactivity determined. 23. hGR-CHO cells were grown to confluency in 24-well plates. The cells were washed twice with PBS and preincubated for 10 min at 37 °C in assay buffer as described (Feth, F.; Rascher, W.; Michel, M. C. Arch. Pharmacol. 1991, 344, 1). The cells were washed once and incubated for an additional 10 min in 250 L of assay buffer supplemented with 100 M isobutylmethylxanthine. Control DMSO (0.1%) or test compound (10 nM to 10 µM final concentration) was added to the wells followed by 50 L of glucagon to give a 10 pM final concentration. The incubation was continued for 15 min, then stopped by removal of the assay buffer and the immediate addition of 250 L ethanol (65%, v/v). After 5 min the supernatant was transferred to a microfuge tube and the ethanol was evaporated by Speed Vac. cAMP content was measured using the Amersham SPA cAMP assay kit (RPA 556) as described by the manufacturer. Results were reported as pmol per assay.

24. All data reported herein reflect purified and characterized (¹H NMR, MS) samples.

25. Hantzsch, A. Justus Liebigs Ann. Chem. 1882, 215, 1.

26. Sura Prakash, G. K.; Krishnamurty, R.; Olah, G. J. Am. Chem. Soc. **1989**, 111, 393.

27. Chiralpak AD column; isocratic elution 1.5 mL/min (99% hexane/methyl *t*-butyl ether), retention times (**33a**, 7.3 min; **33b**, 9.9 min).

28. Dale, J. A.; Mosher, H. S. J. Am. Chem. Soc. 1973, 95, 512.