



Identification of pyridazin-3-one derivatives as potent, selective histamine H₃ receptor inverse agonists with robust wake activity

Robert L. Hudkins*, Lisa D. Aimone, Thomas R. Bailey, Robert J. Bendesky, Reddeppa reddy Dandu, Derek Dunn, John A. Gruner, Kurt A. Josef, Yin-Guo Lin, Jacquelyn Lyons, Val R. Marcy, Joanne R. Mathiasen, Babu G. Sundar, Ming Tao, Allison L. Zulli, Rita Raddatz, Edward R. Bacon

Discovery Research, Cephalon, Inc., 145 Brandywine Parkway, West Chester, PA 19380, USA

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ABSTRACT

H₃R structure–activity relationships on a novel class of pyridazin-3-one H₃R antagonists/inverse agonists are disclosed. Modifications of the pyridazinone core, central phenyl ring and linker led to the identification of molecules with excellent target potency, selectivity and pharmacokinetic properties. Compounds **13** and **21** displayed potent functional H₃R antagonism in vivo in the rat dipsogenia model and demonstrated robust wake activity in the rat EEG/EMG model.

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Histamine elicits physiological responses mediated by four G-protein coupled receptors (H₁R–H₄R) and exerts a variety of functions in the central nervous system (CNS).¹ H₁ and H₂ receptors in the periphery are involved in the allergic response and gastric acid secretion, respectively, and have been some of the more successful drug target classes over the past 50 years. The H₄ receptor is expressed mainly in mast cells, eosinophils, and tissues involved in the immune response, and may play a role in inflammation and pain.² The H₃ receptor (H₃R) in the brain is primarily localized pre-synaptically, where it functions both as an autoreceptor to modulate histamine release and as an inhibitory heteroreceptor regulating the release of multiple neurotransmitters, including acetylcholine (ACh), dopamine (DA), norepinephrine (NE) and serotonin (5-HT).¹ While activation of the H₃R results in the inhibition of neurotransmitter release, blockade of H₃R by selective antagonists or inverse agonists can reverse the histamine-mediated inhibition of neurotransmitter release, leading to enhanced release. H₃Rs by virtue of their localization and function regulate a variety of neurotransmitters that are thought to be involved in attention, sleep and cognition.^{1c–i} The discovery of H₃R antagonists are of current interest, with potential utility in addressing a variety of CNS disorders associated with attention and cognitive deficits, including deficits in wakefulness, attention-deficit hyperactivity disorder (ADHD), Alzheimer's disease (AD), mild cognitive

impairment, and schizophrenia. Several clinical candidates (1a–1e) have advanced into trials (Fig. 1).^{1g–m}

We identified a novel class of pyridazin-3-one H₃R antagonists/inverse agonists with exceptional drug properties, safety and in vivo profiles.³ The preliminary structure–activity relationships (SAR) and characterization of **1** (CEP-26401, irdabisant) and N-methyl **2** were recently disclosed.⁴ In this paper we report SAR around the pyridazinone core, central phenyl ring and the linker leading to the identification of molecules with excellent target

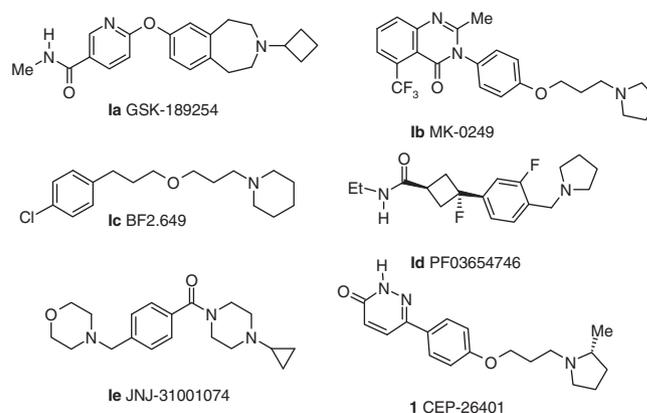


Figure 1. Structures of clinical H₃R antagonists.

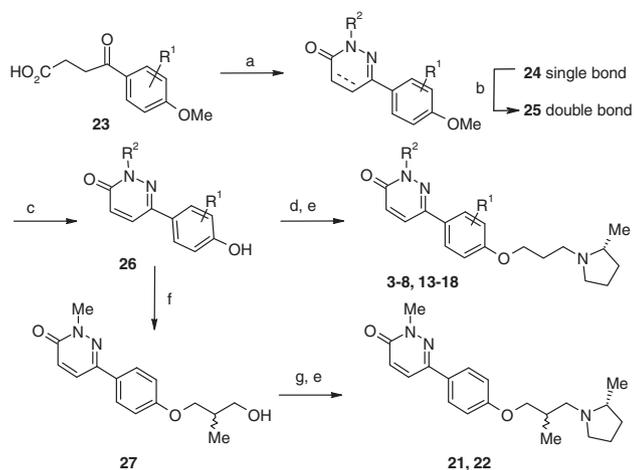
* Corresponding author. Tel.: +1 610 738 6283.

E-mail address: rhudkins@cephalon.com (R.L. Hudkins).

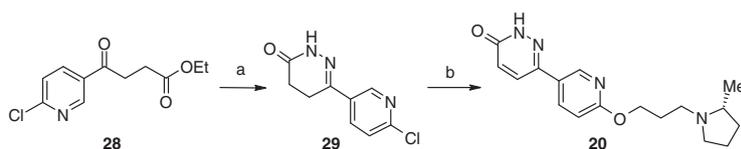
potency and selectivity, pharmacokinetic properties and potent in vivo wake-promoting activity in the rat.

The N^2 -substituted targets (e.g. **3–8**) were synthesized using our previously described method outlined in Scheme 1.^{3,4} Cyclocondensation of 4-(4-methoxyphenyl)-4-oxobutyric acid **23** (or ester) with (N -substituted)hydrazine,^{5a} oxidation of the resulting 4,5-dihydropyridazinone **24** (MnO_2 , $CuCl_2$ or $SeO_2/HOAc$) to **25**,^{5b} followed by O -demethylation produced phenol intermediates **26** in very good yield and purity. Alkylation of **26** with 3-bromo-1-chloropropane provided chloroalkyl intermediates that were reacted with R -2-methylpyrrolidine to produce targets **3–8**. In an analogous manner, ring substituted analogues (e.g. **13–18**) were synthesized starting with a phenyl substituted keto-acid or ester **23**. An alternative approach was employed for 3-cyano **19** using a previously described Suzuki coupling. 5-Boranyl-2-[3-((R)-2-methyl-pyrrolidin-1-yl)-propoxy]-benzonitrile with 3,6-dichloropyridazine and hydrolysis of the intermediate chloropyridazine gave **19**.⁴ The S - and R -methyl linker isomers **21** and **22** were synthesized by alkylation of **26** ($R^2 = Me$, $R^1 = H$) with R - and S -3-bromo-2-methyl-propanol to provide **27**, conversion to the mesylate then (R)-2-methylpyrrolidine alkylation (Scheme 1). Synthesis of the central ring 3-pyridyl **20** was produced by ethyl 4-(6-chloropyridin-3-yl)-4-oxo-butylate **28** and hydrazine hydrate cyclization to the 4,5-dihydro-2H-pyridazinone **29**, followed by alkylation with 3-((R)-2-methylpyrrolidin-1-yl)propan-1-ol using DMSO and base to directly produce aromatized target **20** (Scheme 2). The 4-methyl analogue **9** was synthesized by the procedure outlined in Scheme 1, starting with 4-(4-methoxyphenyl)-3-methyl-4-oxobutyric acid.

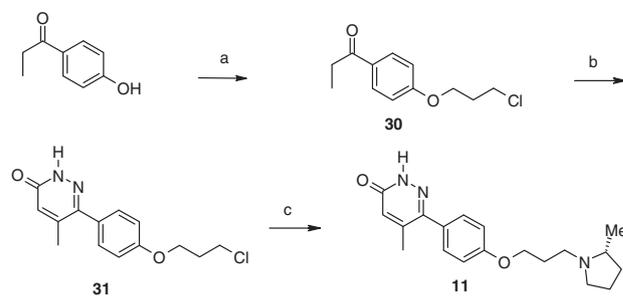
Additional approaches to selectively modify either the 4- or 5-position are shown in Schemes 3–5. The 5-methyl analogue **11** was produced using a modification of our previously described



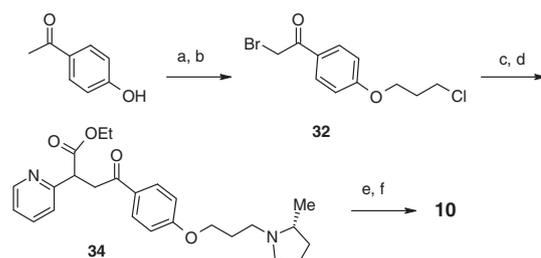
Scheme 1. Reagents and conditions: (a) R^2NHNH_2 , 2-propanol, reflux, 80–95%; (b) MnO_2 , xylenes, 155 °C, $SeO_2/HOAc$ or $Cu(II)Cl_2$, CH_3CN , reflux, 50–85%; (c) BBr_3 , DCM, 5 °C → rt, 70–95%; (d) K_2CO_3 , $Br(CH_2)_3Cl$, acetone, 65 °C; (e) (R)-2-methylpyrrolidine, NaI, K_2CO_3 , CH_3CN 80 °C, 35–60% two steps; (f) R or S 3-bromo-2-methylpropanol, K_2CO_3 , CH_3CN 80 °C, 50–75%; (g) (i.) MsCl, TEA, DCM; (ii.) (R)-2-methylpyrrolidine, NaI, K_2CO_3 , CH_3CN 80 °C, 40–60% two steps.



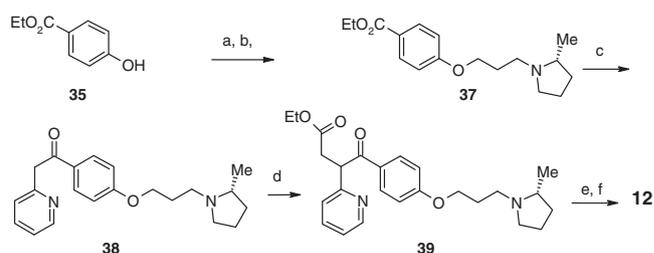
Scheme 2. Reagents and conditions: (a) $H_2NNH_2 \cdot H_2O$, ethanol, 80 °C, 87%; (b) 1 M $KtOBu$, $t-BuOH$, 3-((R)-2-methylpyrrolidin-1-yl)propan-1-ol, DMSO, 100 °C, 8%.



Scheme 3. Reagents and conditions: (a) $Br(CH_2)_3Cl$, K_2CO_3 , acetone, 80 °C, 65%; (b) (i.) $HC(O)COOH \cdot H_2O$, 85 °C; (ii.) $NH_2NH_2 \cdot H_2O$, ethanol, 85 °C, two steps 35%; (c) R -2-methylpyrrolidine, K_2CO_3 , CH_3CN ; 67%.



Scheme 4. Reagents and conditions: (a) K_2CO_3 , $Br(CH_2)_3Cl$, acetone, 80 °C, 70%; (b) $CuBr$, $EtOAc$, $CHCl_3$, reflux, 70%; (c) ethyl 2-pyridylacetate, NaH, DMF, 72% → 33; (d) (R)-2-methylpyrrolidine, NaI, K_2CO_3 , CH_3CN 80 °C; (e) $NH_2NH_2 \cdot H_2O$; ethanol, HOAc, 46%; (f) DMSO, K_2CO_3 , 80 °C, 75%.



Scheme 5. Reagents and conditions: (a) K_2CO_3 , $Br(CH_2)_3Cl$, acetone, 80 °C, 89% → 36; (b) (R)-2-methylpyrrolidine, NaI, K_2CO_3 , CH_3CN 80 °C, 96%; (c) LiHMDS, THF, 2-methylpyridine, 0 → 45 °C, 70–80%; (d) NaH, $BrCH_2CO_2Et$, DMSO, toluene, 52%; (e) $NH_2NH_2 \cdot H_2O$; ethanol, 25%; (f) NaOH, water, $EtOH$, Na 3-nitrobenzenesulfonic acid, 35%.

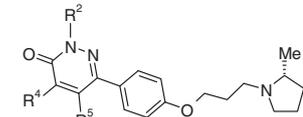
Aldol/hydrazine cyclocondensation sequence to construct the pyridazinone ring^{4,6} (Scheme 3).

Chloro **30** was condensed with glyoxalic acid monohydrate in acetic acid at 135 °C, followed by hydrazine cyclocondensation of the resulting Aldol adduct to give the 4,5-dihydro-2H-pyridazin-3-one **31** in low yield. Incorporation of (R)-2-methylpyrrolidine gave **11**. The critical step toward the synthesis of the 4-pyridyl **10** (Scheme 4) was alkylation of 2-bromo-1-[4-(3-chloropropoxy)phenyl]ethanone **32** with ethyl 2-pyridinyl acetate to produce

33 (72% yield). Installation of (*R*)-2-methylpyrrolidine (**34**), hydrazine cyclization, followed by DMSO oxidation produced **10** in 30% yield. The 5-(2-pyridyl) **12** synthesis commenced with ethyl 4-hydroxybenzoate **35** (Scheme 5), installation of the amine side chain (**36** → **37**), and 2-methylpyridine addition to ketone **38**, followed by subsequent alkylation with ethyl bromoacetate to produce the keto-ester intermediate **39**. Hydrazine condensation and oxidation of the 4,5-dihydropyridazinone using sodium 3-nitrobenzenesulfonate produced **12**.

The pyridazinone analogues were tested using in vitro binding assays by displacement of [³H]NAMH in membranes isolated from CHO cells transfected with cloned human H₃ or rat H₃ receptors as shown in Table 1 and Table 2.^{3,4} The (*R*)-2-methylpyrrolidine amine was previously established to be optimum⁴ and was fixed for comparison while exploring further SAR. Also previously disclosed was substitution at the N² position could accommodate significant steric bulk without affecting H₃R binding affinity, although increasing the size of the R² group greater than methyl (**2**) resulted in compounds with log *P* values greater than **3**. This negatively affected molecular weight and decreased the ligand efficiency (LE) and the ligand lipophilic efficiency (LLE).⁷ Amphiphilic, high log *P* compounds have been shown to enhance hERG activity and drive high tissue distribution inducing phospholipidosis.⁸ Motivated to synthesize compounds with lower *c log P*, the fluorinated ethyl (**3** and **4**) and polar N-ethanol **5** were designed. Similar to other simple alkyl derivatives (e.g. **2**)⁴ compounds **3**, **4** and **5** also had high affinity for both human and rat H₃R. Further discovery flow profiling with these analogues however showed poor oral bioavailability in the rat (*F* < 10%). Lowering the *c log P* also was the motivation for replacement of the lipophilic R² phenyl **6** with more polar 2-pyridyl **7** and 2-pyrimidinyl **8** heteroaryls. The 2-Pym **8** showed balanced and slightly higher affinity compared with **7** for both human and rat H₃Rs (hH₃R K_i = 1.4 nM, rH₃R K_i = 7.9 nM) with lower lipophilicity (calculated *c log P* = 1.8 by the Tripos method). Compound **8** had acceptable in vitro metabolic stability across species in liver microsomes (*t*_{1/2} > 40 min in mouse, rat, dog, monkey and human) and IC₅₀ values > 30 μM for inhibition of cytochrome P450 enzymes CYP1A2, 2C9, 2C19, 2D6 and 3A4, indicating minimal potential for drug–drug interactions. In a hERG patch–express assay **8** had an IC₅₀ value > 10 μM. Compound **8** also showed excellent selectivity (>1000-fold) for hH₁, hH₂, and hH₄ receptor subtypes and against a panel of 70 GPCRs, ion channels and enzymes. In a rat pharmacokinetic experiment **8** showed accept-

Table 1
Pyridazin-3-one H₃R binding data^{a,b}

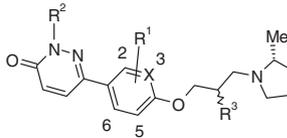


Entry	R ²	R ⁴	R ⁵	hH ₃ (K _i , nM)	rH ₃ (K _i , nM)	<i>c log P</i>
1	H	H	H	2.0 ± 0.4	7.2 ± 0.4	2.3
2	Me	H	H	1.4 ± 0.1	6.3 ± 1.1	2.8
3	CH ₂ CH ₂ F	H	H	3.4 ± 0.5	11 ± 0.5	2.6
4	CH ₂ CF ₃	H	H	5.7 ± 0.8	11 ± 0.5	2.7
5	CH ₂ CH ₂ OH	H	H	3.0 ± 0.1	9.5 ± 0.7	1.5
6	Ph	H	H	2.5 ± 0.9	7.9 ± 1.6	4.3
7	2-Pyr	H	H	10 ± 2	22 ± 3	2.8
8	2-Pym	H	H	1.4 ± 0.1	7.9 ± 0.8	1.9
9	H	Me	H	3.4 ± 0.1	8.7 ± 1.0	2.8
10	H	2-Pyr	H	3.1 ± 0.03	10 ± 1.0	3.1
11	H	H	Me	2.9 ± 0.1	12 ± 0.1	2.5
12	H	H	2-Pyr	12 ± 1	70 ± 1	2.3

^a K_i values nM ± SEM.

^b Pyr = 2-pyridine; 2-Pym = 2-pyrimidine.

Table 2
Pyridazin-3-one central ring and linker H₃R binding data^a



Entry	R ¹	R ²	X	R ³	hH ₃ (K _i , nM)	rH ₃ (K _i , nM)
13	3-F	Me	C	H	5.8 ± 1.2	27 ± 7
14	2-F	H	CH	H	16 ± 2	69 ± 24
15	3-Cl	Me	C	H	4.4 ± 0.5	14 ± 2
16	3,5-F	Me	C	H	11 ± 3	41 ± 5
17	3,5-F	H	C	H	7.1 ± 1.4	40 ± 4
18	2-OMe	H	CH	H	152 ± 19	229 ± 65
19	3-CN	H	C	H	38 ± 12	111 ± 17
20	H	H	N	H	35 ± 14	70 ± 15
21	H	Me	CH	S-Me	2.4 ± 0.4	4.6 ± 1
22	H	Me	CH	R-Me	15 ± 2	52 ± 5

^a K_i values nM ± SEM.

able i.v. intrinsic pharmacokinetic properties (*t*_{1/2} = 1.5 h, CL = 29 mL/min/kg, V_d = 3.9 L/kg) and oral bioavailability (*F* = 56%). However, the brain exposure was low (brain/plasma ratio (B/P) = 0.5) for potential CNS indications, and this compound was not further advanced. Subsequently, the 4- and 5-positions were probed with methyl and 2-Pyr substitution. Methyl substitutions (**9** and **11**) were equally tolerated at both positions, while the 5-Pyr **12** had 4-fold and 7-fold lower affinity for hH₃R and rH₃R, respectively, compared to the 4-Pyr **10**. Compounds **10** and **12** showed acceptable in vitro metabolic stability, but did not show improvement in oral bioavailability (*F* < 10%) and/or brain exposure (B/P << 1) compared to **1** or **2**.

Data in Table 2 reveals the effect of varying the substitution on the central phenyl ring. Substitution in the 3-position with fluorine (**13**), chlorine (**15**) or with 3,5-difluoro (**16**, and **17**) were tolerated for H₃R binding affinity, while 2-substitution showed significantly reduced H₃R affinity (e.g. **14** and **18**). Replacing the central phenyl ring with 3-pyridyl **20** led to a ≥ 10-fold decrease in human and rat H₃R affinity. The effect of incorporating a chiral methyl group on the propyl linker was also investigated with *S*-21 and *R*-22. The *S*-isomer **21** retained hH₃R (K_i = 2.4 nM) and rH₃R (K_i = 4.6 nM) affinity equivalent to **2**, while *R*-22 had 6- and 11-fold weaker affinity for hH₃R and rH₃R.

Based on target affinity and physical property data (pH₂ and pH_{7.4} solubility >0.25 mg/mL, *c log P* = 3.3 and 3.0) *S*-methyl **21** and 3-fluoro **13** were further profiled for selectivity against hH₁, hH₂, and hH₄ receptor subtypes and against a panel of 70 GPCRs, ion channels and enzymes (MDS Pharma Services, LeadProfiler). Compound **13** showed <30% inhibition against hH₁, hH₂, and hH₄ subtypes at 10 μM concentration, and inhibited only the norepinephrine transporter (NET) by greater than 70% at 10 μM concentrations in the broader screening panel. Compound **21** also showed excellent selectivity (> 1000-fold) for hH₁, hH₂, and hH₄ subtypes and displayed <30% inhibition at 10 μM concentration in the MDS panel of 70 targets, including NET (19% inh.). Functionally, **13** and **21** showed potent antagonist activity and displayed full inverse agonist activity in the [³⁵S]GTPγS hH₃R binding assay.^{3,4} 3-Fluoro **13** and *S*-Methyl **21** decreased basal activity with EC₅₀ values of 1.3 ± 0.1 nM and 2.1 ± 0.1 nM, respectively.

Based on the target H₃R affinity, selectivity, in vitro metabolic stability in liver microsomes (*t*_{1/2} > 40 min across species) and CYP inhibition selectivity (IC₅₀ > 30 μM), **21** and **13** were further evaluated for pharmacokinetic properties in the rat (Table 3) in comparison with **2**.⁴ Compound **13** showed an i.v. *t*_{1/2} of 1.2 h, clearance of 38 mL/min/kg, and oral bioavailability of 42% based on 6 h

AUC data with good brain exposure (B/P = 3.4). *S*-Me 21 had similar oral bioavailability (%*F* = 37 based on 6 h AUC) with good brain exposure in the rat (B/P = 2.5) (Table 3).

The rat dipsogenia model was used in the project as an *in vivo* surrogate measure of H₃R functional inhibition in the brain following peripheral administration. Histamine and the H₃-selective agonist, *R*- α -methylhistamine (RAMH), induce water drinking in the rat when administered either peripherally or centrally, an effect that is blocked by H₃R antagonists.^{1h,9} In this model both **13** and **21** potently and dose-dependently inhibited RAMH-induced dipsogenia with ED₅₀ values of 0.13 (0.02–0.91) mg/kg *i.p.* and 0.02 (0.005–0.07) mg/kg *i.p.*, respectively. Following the demonstration of potent *in vivo* H₃R functional activity in the brain, **13** and **21** were further evaluated for wake promoting activity in the rat.¹⁰ Histamine-producing neurons are an important part of the monoaminergic arousal system and H₃R antagonists have been documented to increase wakefulness in a number of species, although at doses and cortical H₃R occupancy levels much higher than those producing activity in the dipsogenia model or efficacy in cognition models.¹¹ Wake promoting activity was measured as previously described using male Sprague Dawley rats surgically implanted for chronic recording of EEG (electroencephalographic) and EMG (electromyographic) signals.¹² The cumulative wake time at 4 h after dosing was evaluated during the normal quiet period of the rat. Compound **21** increased waking at 3 (166 ± 6 min) and 10 mg/kg *i.p.* (231 ± 6 min) by 4 h AUC values (*P* < 0.001 ANOVA). At 10 mg/kg, waking was enhanced out to 5.5 h post dosing, and

Table 3
Pharmacokinetic properties in rat^a

		2 ^b	13 ^b	21 ^c
<i>i.v.</i>	<i>t</i> _{1/2} (h)	1.6 ± 0.3	1.2 ± 0.3	0.8 ± 0.2
	<i>V</i> _d (L/kg)	6.5 ± 2.9	4.5 ± 1.4	2.5 ± 1.0
	CL (mL/min/kg)	45 ± 11	38 ± 7	37 ± 8
<i>p.o.</i>	AUC (ng h/mL)	538 ± 75	936 ± 166	892 ± 198
	<i>C</i> _{max} (ng/mL)	123 ± 6	284 ± 34	146 ± 9
	<i>t</i> _{1/2} (h)	2.3	1.6	4.3
	<i>F</i> (%)	28 ± 4	42 ± 7	38 ± 8
	B/P ^d	3.5 ± 0.4	3.4 ± 0.3	2.5 ± 0.1

^a Administration at 1 mg/mg *i.v.* and 5 mg/kg *p.o.*; data calculated from 6 h AUC values.

^b *i.v.* formulation (3% DMSO, 30% solutol, 67% phosphate buffered saline) oral formulation (saline).

^c *i.v.* formulation (3% DMSO, 30% solutol, 67% phosphate buffered saline) oral formulation (50% Tween 80, 40% propylene carbonate and 10% propylene glycol).

^d B/P = brain to plasma ratio.

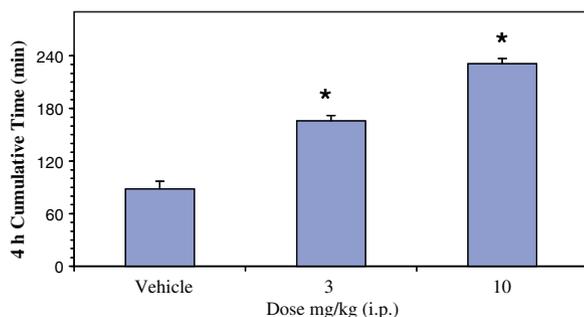


Figure 2. Compound **21**-induced wake promotion; administered *i.p.* to male rats chronically implanted with electrodes for recording EEG and EMG activity. Cumulative wake 4 h AUC values shown for each dose (mean ± SEM, *n* = 7–8/group). **p* < 0.05 Dunnett's post hoc versus vehicle.

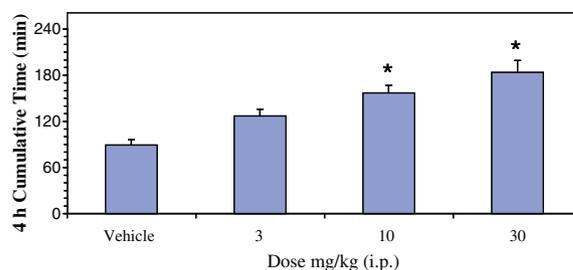


Figure 3. Compound **13**-induced wake promotion; administered *i.p.* to male rats chronically implanted with electrodes for recording EEG and EMG activity. Cumulative wake 4 h AUC values shown for each dose (mean ± SEM, *n* = 7–8/group). **p* < 0.05, Dunnett's post hoc versus vehicle.

maximal cumulative wake surplus was 196 min reached at 7 h. EEG activity, behavior and body temperature were all normal at the 3 or 10 mg/kg doses. At 10 mg/kg, **21** demonstrated robust wake promotion, with the treated animals being awake 96% of the time up to 4 h post dose and averaging a 62% increase in wake time over the vehicle treated animals (Fig. 2). Compound **13** increased wake activity in a dose-related manner at 10 (157 ± 9 min) and 30 mg/kg (184 ± 15 min) by 4 h AUC values (*P* < 0.001 ANOVA) (Fig. 3). Compound **13** showed less robust wake activity compared to **21**, consistent with weaker potency in the dipsogenia model.

In summary, H₃R structure-activity relationships were disclosed on the pyridazin-3-one phenoxypropyl amine core leading to the identification of new molecules displaying excellent H₃R target potency, selectivity and rat pharmacokinetic properties. Compounds **13** and **21** were profiled in greater detail and advanced into *in vivo* evaluations. Both compounds displayed potent H₃R antagonist activity in the brain using the rat dipsogenia model as a functional H₃R readout and demonstrated potent wake-promoting activity in the rat EEG/EMG model.

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