EFFECT OF SUBSTITUENTS ON METHYL AFFINITY OF NAPHTHALENE DERIVATIVES

Sir:

An investigation of the effect of substituents on the rate of addition of methyl radicals to substituted benzenes was carried out in these laboratories a few years ago.¹ The results were in substantial agreement with the findings of Hey and his co-workers, ^{2,3,4} who studied phenylation of substituted benzenes. Benzene and its derivatives are comparatively unreactive compounds, and this caused some experimental difficulties and introduced some uncertainties in the results. To avoid them a series of more reactive substrates, namely, naphthalene derivatives, was investigated.

The rates of addition of methyl radicals were determined by the method described previously.^{5,6} The results are presented in the form k_2/k_1 (Table I) where the subscripts refer to the two reactions

$$\text{CH}_{3^{+}} + \text{solvent} \xrightarrow{k_{1}} \text{CH}_{4} + \text{solvent radical} \ \ (1)$$
 and

$$CH_3$$
· + substrate $\xrightarrow{k_2}$ CH_3 ·substrate (2)

Isoöctane was used as a solvent and the reaction was carried out at 65° . Whenever the abstraction of hydrogen atoms by methyl radicals did take place the results were recorded and listed in Table I in the form of k'_3/k_1 , k'_3 being the rate constant of abstraction per active hydrogen.

TABLE I					
Compound	k_2/k_1	k'_3/k_1	Compound	k_2/k_1	
Naphthalene	9.4				
1-Methyl-	8.1	0.83	1-Iodo-	18.4	
2-Methyl-	13.0	2,05(?)	2-Iodo-	30.1	
2,3-Dimethyl-	10.5	1.06	1,4-Dichloro-	20.6	
2,6-Dimethyl-	13.9	1.08	1-Methoxy-	8.8	
1,5-Dimethyl-	5.8	1.09	2-Methoxy-	7.5	
Acenaphthene	4.7	5.15	1-Dimethylamino-	3.5	
1-Ethyl-	8.0	4.26	1-Acetonaphthone	20.0	
2-Ethyl-	8.9	4.27	2-Acetonaphthone	44.4	
1-Fluoro-	10.5		1-Naphthonitrile	30.8	
2-Fluoro-	13.2		2-Naphthonitrile	45.9	
1-Chloro-	13.2		1-Methyl naphthoate	19.4	
2-Chloro-	21.6		1-Ethyl naphthoate	22.2	
1-Bromo-	8.0(?)		2-Methyl naphthoate	40.3	
2-Bromo-	21.2		1-Naphthylisocyanate	17.1	

Several conclusions emerge from the data listed in Table I. Substituents in the α position enhance the addition less than those placed in the β position, 1-methoxynaphthalene being the only exception. This might partially result from the "blocking" of the reactive α position, an effect responsible for the low reactivity of 1,5-dimethylnaphthalene and acenaphthene. The reactivity increases along the series F, Cl, Br, I, 1-bromonaphthalene apparently being an exception. The reaction of these compounds with methyl radicals does not involve abstraction of a halogen atom.

- (1) W. J. Heilman, A. Rembaum and M. Szwarc, J. Chem. Soc., 1127 (1957).
- (2) D. R. Augood, D. H. Hey and G. H. Williams, *ibid.*, 2094 (1952); 44 (1953).
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 (4) J. I. G. Cadogan, D. H. Hey and G. H. Williams, J. Chem. Soc.,
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- (6) R. P. Buckley and M. Szware, Proc. Roy. Soc., A240, 396 (1957).

The only substituents decreasing the reactivity are the electron donating ones, namely, methoxy, dimethylamino and ethyl. Methyl group seems to be a border case.

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RADICAL CATALYZED ADDITIONS TO DIBUTYL ETHYLENEBORONATE¹

Sir

Dibutyl ethyleneboronate (I) reacts with free radicals (X') to yield a type of intermediate radical (II) that is markedly stabilized by carbon-boron π -bonding, and which reacts further to yield boronic esters (III) containing other functional groups, such as α -halogen, previously inaccessible.

The first transfer constant² in the reaction of dibutyl ethyleneboronate (I) with carbon tetrachloride is approximately 1×10^{-3} . This low value, which would not be due to polar factors² and which is similar to that for styrene² (0.6×10^{-3}) , indicates that the energies involved for adjacent carbon p-orbital overlap with phenyl and with dialkoxyboron groups are similar in magnitude. Stabilization of the radical II is consistent with simple molecular orbital calculations, provided one chooses parameters for the electronegativity and p-orbital overlap of boron that are not too low.

Dibutyl ethyleneboronate³ (I) was prepared by addition of vinylmagnesium chloride⁴ in tetrahydrofuran to methyl borate in ether under nitrogen at -60° , acidification with a 2M phosphoric-1M hydrochloric acid solution, addition of 0.3 g. of phenothiazine to prevent polymerization on contact with air, extraction with 1-butanol, removal of excess water by freezing at -70° , distillation of butanol-water azeotrope and solvents at 20–40 mm., and distillation of the product through a Vigreux column onto 0.02 g. of phenothiazine; yield 134 g. (72%), b.p. 35– 40° (0.08 mm.), $n^{24.5}$ D 1.4167. $Anal.^{5}$ Calcd. for $C_{10}H_{21}BO_{2}$: C, 65.2; H, 11.5; B, 5.9. Found: C, 65.06; H, 11.66; B, 6.47.

Preparation of the carbon tetrachloride adduct IIIa was best accomplished by slow (40 hr.) addi-

- (1) Presented in part at the Northwest Regional Meeting of the
- American Chemical Society, June 18, 1959.

 (2) C. Walling, "Free Radicals in Solution," John Wiley and Sons. Inc. New York, 1957, pp. 157-158, 245-246, 257.
- (2) C. Waining, Tree Radician in State 11 and 12 and 13 and 14 and 14 and 15 and 16 an
- (4) H. E. Ramsden, J. R. Leebrick, S. D. Rosenberg, E. H. Miller, J. J. Walburn, A. E. Balint and R. Cserr, J. Org. Chem., 22, 1602 (1957)
 - (5) Galbraith Laboratories, Knoxville, Tenn,

tion of a dilute solution of dibutyl ethyleneboronate (I) and azobisisobutyronitrile (10 g. per mole of I) in carbon tetrachloride to refluxing carbon tetrachloride followed by distillation of the product (IIIa), yield 45%, b.p. $90-95^{\circ}$ (0.07 mm.), n^{29} D 1.4490. Anal. Calcd. for $C_{11}H_{21}BCl_4O_2$: C, 39.10; H, 6.27; Cl, 41.97; B, 3.20. Found: C, 39.42; H, 6.44; Cl, 41.67; B, 3.46. The structure IIIa was proved by treatment with 2,4-dinitrophenylhydrazine and hydrogen peroxide, which yielded 73% of the 2.4-dinitrophenylhydrazone of 3,3,3-trichloropropionaldehyde, m.p. 109–112°, loses HCl (identified by tests for H⁺ and Cl⁻), solidifies, m.p. 161–163°.6 Anal.⁷ Calcd. for C₉- $H_7Cl_3N_4O_4$: C, 31.65; H, 2.06; Cl, 31.14; N, 16.41. Found: C, 31.61; H, 2.16; Cl, 31.00; N, 16.25. Reaction of a fourfold excess of bromotrichloromethane with dibutyl ethyleneboronate (I) and a trace of initiator at 80° leads to an exothermic reaction and formation of the adduct IIIb in 94% yield, b.p. $95-100^{\circ}$ (0.07 mm.), n^{25} D 1.4720, structure proof the same as for IIIa but with a low yield, caused by decomposition in the presence of bromide ion. Anal.⁵ Calcd. for $C_{11}H_{21}BBrCl_3O_2$: C, 34.55; H, 5.53; B, 2.83; g. silver halide/g. compound, 1.615. Found: C, 34.82; H, 5.67; B, 3.13; silver halide, 1.614. The adduct IIIc was prepared by irradiating 1.1 equivalent of n-hexyl mercaptan with the ester I at -70° under carbon dioxide with a mercury vapor lamp until the product solidified, then distilling, yield 93%, b.p. 115-118° (0.07 mm.), $n^{29.5}$ D 1.4501. Anal.⁵ Calcd. for C₁₆H₃₅BO₂S: C, 63.56; H, 11.67; B, 3.58; S, 10.61. Found: C, 63.26; H, 11.70; B, 3.89; S, 10.81. The structure was proved by degradation to ethylene (confirmed by infrared) with solid potassium hydroxide at 110–127°.

(6) The 2,4-dinitrophenylhydrazone of 3,3-dichloropropenal has been reported, m.p. 164-165°, by M. S. Kharasch, O. Reinmuth and W. H. Urry, This JOUNNAL, **69**, 1105 (1947). The report of the 2,4-dinitrophenylhydrazone of 3,3,3-trichloropropionaldehyde, m.p. 173°, J. Harmon, U. S. Patent 2,396,261, Mar. 12, 1946 [Chem. Abstracts, **40**, 3466 (1946)] is presumably inaccurate.

(7) Weiler and Strauss, Oxford, England.

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STIMULATION OF THE EXCHANGE OF FORMATE INTO PYRUVATE BY A DIALYZABLE COFACTOR¹

At pH 6.5, extracts of *Micrococcus lactilyticus* oxidatively decarboxylate pyruvate² to acetate, carbon dioxide and hydrogen. At pH 8.5, however, the products of pyruvate breakdown³ are acetate and formate. Formate is not decomposed to carbon dioxide and hydrogen at either pH or under any conditions of testing⁴ and at an alkaline pH formate is exchanged into the carboxyl of pyruvate.³,⁵ The

enzymes participating in this exchange may be separated from those catalyzing oxidative decarboxylation by fractionation with ammonium sulfate. This results not only in a separation of the two types of enzymatic activities, but also yields an increase in specific activity with regard to the exchange of formate.

TABLE I

EFFECT OF BES AND OTHER PREPARATIONS ON FORMATE EXCHANGE

For measurement of formate exchange, the reaction vessels contained: $1000~\mu\mathrm{M}.$ of phosphate buffer at $p\mathrm{H}$ 8.0, $20~\mu\mathrm{M}.$ of MgCl₂, $20~\mu\mathrm{M}.$ of mercaptoethanol, $80~\mu\mathrm{g}.$ of ferrous ammonium sulfate, $50~\mu\mathrm{M}.$ of pyruvate, $0.5~\mu\mathrm{M}.$ of Cl⁴-formate (10^5 cpm.), $0.5~\mathrm{ml}.$ of BES (obtained from a crude extract containing 30 mg. protein/ml.) or $0.5~\mathrm{ml}.$ of charcoal eluate (equivalent to $2~\mathrm{ml}.$ of original crude extract), and enzyme source equivalent to $10~\mathrm{mg}.$ of protein in a total volume of $2.8~\mathrm{ml}.$ The reaction mixture was incubated $9.0~\mathrm{minutes}$ at 30° in a hydrogen atmosphere.

Preparation	Formate excha Expt. 1	ange activity ^a Expt 2.
Crude extract	125	110
Treated crude extract ^b	3	1
+ BES	119	110
+ Charcoal eluate	c	90
+ Eluate from chromatogram	71	
(NH ₄) ₂ SO ₄ ppt., 0-45% satn.	1	1
+ BES	11 0	108
BES (alone)	1	1

 a Cpm./ μ M. residual pyruvate/mg. protein, assayed by determining the radioactivity of the 2,4-dinitrophenylhydrazone of the residual pyruvate. b Prepared by treating with charcoal. c Not tested.

When crude extracts or fractions obtained by precipitation with ammonium sulfate are dialyzed for a short time, the ability to exchange formate into pyruvate is lost. Full activity may be restored by the addition of the supernatant fraction obtained by centrifugation of a heat-inactivated crude extract ("boiled extract supernatant," or "BES"). Treatment of crude extracts with charcoal (Norit A) also results in a complete loss of activity. Again, full activity may be restored either by addition of BES or of material eluted from the charcoal with 50% aqueous acetone containing 0.1 N NH₄OH (Table I). As seen in Table I, the active factor(s) can be recovered after chromatography on paper with butanol, pyridine, and water (1:1:1) as solvent.

BES cannot be replaced by yeast extract, mixtures of amino acids, or boiled extracts prepared from *Escherichia coli*, *Micrococcus aerogenes*, and *Clostridium kluyveri*. These compounds, too, when tested singly, also did not replace BES: nucleotide tri-, di-, and monophosphates, di-, and triphosphopyridine nucleotide, coenzyme A, lipoic acid, tetrahydrofolate, dihydrofolate, thiamine pyrophosphate, tocopherol derivatives, flavin adenine dinucleotide, biotin, or pyridoxal phosphate.

Further investigations are in progress into the nature of the component(s) of BES responsible for stimulating the exchange of formate into pyruvate

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