Preliminary communication

Synthesis of 8-(methoxycarbonyl)octyl glycosides of O- α -L-rhamnopyranosyl-(1 \rightarrow 3)-O- α -L-rhamnopyranosyl-(1 \rightarrow 3)-2-acetamido-2,6-dideoxy-Dglucopyranose; models for the antigen of *Pseudomonas aeruginosa* Fisher immunotype 5

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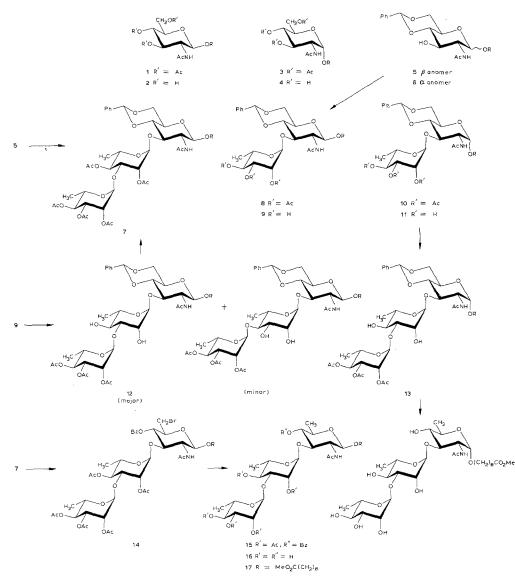
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The cell-surface lipopolysaccharide antigens of the seven Fisher immunotypes of *Pseudomonas aeruginosa* show major, qualitative differences in their polysaccharide chain-components¹; trisaccharide repeating-units in this component have been established² for Types 1, 2, and 5. Use of these whole antigens as human vaccines³ is complicated by the fact that their lipid A component is strongly immunogenic. The isolated polysaccharide chain-components have molecular weights⁴ too low⁵ for them to serve as effective antigens unless conjugated to a carrier of high molecular weight⁶. The present work was undertaken to provide a single repeat of the Type 5 sequence in the form of a long-chain glycoside in which the aglycon is presumed⁷ to be non-immunogenic. The terminal methoxycarbonyl functionality should permit attachment of the trisaccharide hapten, by way of the derived⁷ acyl azide, to such protein acceptors as human, or other mammalian, serum proteins, in order to furnish artificial antigens, and to suitable, functionalized solid supports that may be effective affinity-adsorbents for *P. aeruginosa* Type 5 antibodies.

8-(Methoxycarbonyl)octanol reacted with 2-acetamido-3,4,6-tri-O-acetyl-2-deoxy- α -D-glucopyranosyl chloride⁸ in benzene containing Hg(CN)₂ at 25° to give 90% of the acetylated β -glycoside 1, m.p. 102°, $[\alpha]_D -10°$ (CHCl₃), convertible⁹ by ZnCl₂ and an excess of 8-(methoxycarbonyl)octanol at 130° (vacuum) into 60% of syrupy α -glycoside 3, $[\alpha]_D^{25}$ +78° (MeOH). O-Deacetylation (Zemplén) of 1 gave ~100% of 9, m.p. 156°, $[\alpha]_D -21°$ (MeOH), and similarly, 3 gave 80% of its O-deacetylated analog 4, m.p. 139°, $[\alpha]_D +112°$ (MeOH). Benzylidenation of 2 with α,α -dimethoxytoluene at 25° (TsOH) gave 87% of the 4,6-benzylidene acetal⁷ 5, m.p. 222°, $[\alpha]_D -51°$ (HCONMe₂), and similar treatment of 4 gave 90% of the 4,6-benzylidene acetal 6, m.p. 138°, $[\alpha]_D +61°$ (MeOH).

Methyl α -L-rhamnopyranoside underwent selective¹⁰ glycosylation at O-3 by tri-O-acetyl- α -L-rhamnopyranosyl bromide, and the product obtained by acetylation and

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subsequent acetolysis with Ac₂O containing 1% of H₂SO₄ was treated with HBr–HOAc, to give 2,4-di-O-acetyl-3-O-(2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl)- α -L-rhamnopyranosyl bromide¹¹. The latter underwent reaction with 5 in benzene [Hg(CN)₂] at ~25°, to give 8-(methoxycarbonyl)octyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O-[2,4-di-O-acetyl-3-O-(2,3,4-tri-O-acetyl- α -L-rhamnopyranosyl)- α -L-rhamnopyranosyl]- β -D-glucopyranoside (7), isolated chromatographically homogeneous, but in rather low yield. In an alternative approach, glycoside 5 was condensed at 50° with tri-O-acetyl- α -L-rhamnopyranosyl bromide in 1:1 benzene–acetonitrile [Hg(CN)₂] to give 90% of the disaccharide derivative¹² 8, m.p. 142°, [α]_D -104° (MeOH), and the α precursor 6 likewise gave 90–100% of the syrupy disaccharide analog 10, [α]_D +7° (MeOH).

Zemplén O-deacetylation of 8 gave 80% of 8-(methoxycarbonyl)octyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O- α -L-rhamnopyranosyl- β -D-glucopyranoside (9), m.p. 192°, $[\alpha]_D$ -86° (MeOH), which underwent reaction with tri-O-acetyl- α -L-rhamnopyranosyl bromide in MeCN-MeNO₂ [Hg(CN)₂], to give 25% of the 3-O-substituted product (12), m.p. 123-124°, $[\alpha]_D$ -80° (MeOH) and 10% of the 4-O-substituted isomer, m.p. 119°, $[\alpha]_D$ -92° (MeOH). The latter reacted with periodate, and was converted by acetone and a trace of acid into an isopropylidene acetal (n.m.r.), whereas 12 did not react with periodate or acetone-H⁺, and was converted by acetylation (100%) into trisaccharide derivative 7, m.p. 119-120°, $[\alpha]_D$ -64° (MeOH), identical (by ¹Hn.m.r.) with that prepared previously.

O-Deacylation of 10 gave 96% of the corresponding triol 11, m.p. 194°, $[\alpha]_D$ +4° (MeOH), which, on treatment with tri-*O*-acetyl- α -L-rhamnopyranosyl bromide in benzene-MeCN [Hg(CN)₂] at 30°, gave 63% of the 3-*O*-glycosylated product (13) as a foam, m.p. 102°, $[\alpha]_D$ -16° (MeOH), accompanied by minor proportions of the 4-*O*-substituted product (14%) and a tetrasaccharide derivative.

The fully acetylated β -trisaccharide derivative 7 reacted with 1.1 mol of *N*bromosuccinimide in boiling, 1:9 (CHCl₂)₂--CCl₄ (tungsten light) during 5 min to open the benzylidene acetal ring and give 82% of the corresponding 6-bromide 4-benzoate 14, m.p. 163-164°, $[\alpha]_D$ -49° (MeOH), and reduction of the latter with hydrogen (50 lb. in.⁻²)-Raney nickel gave 80% of the 6-deoxy analog 15. m.p. 176--177°, $[\alpha]_D$ -55° (MeOH). Final Zemplén deprotection of 15 afforded 100% of the target β -glycoside, namely 8-(methoxycarbonyl)octyl 2-acetamido-2,6-dideoxy-3-O-[3-O-(\alpha-L-rhamnopyranosyl)- α -L-rhamnopyranosyl]- β -D-glucopyranoside (16), m.p. 98-100° (dec.), $[\alpha]_D$ -66° (MeOH). Similarly, acetylation of the α -trisaccharide derivative 13, followed by treatment of the acetate ($[\alpha]_D$ +7° in MeOH) with 1.1 mol of *N*-bromosuccinimide in boiling CCl₄ for 5 min (tungsten light) gave 88% of the syrupy 6-bromide 4-benzoate α analog ($[\alpha]_D$ ~0° in MeOH) of 14, which, on hydrogenolysis (86%; product $[\alpha]_D$ -3° in MeOH), followed by *O*-deacylation (83%), gave the α anomer (17) of 16.

The foregoing syntheses gave the target trisaccharide glycosides in acceptable net yields at the 0.5-g level, and all products were adequately identified by ¹H- and ¹³C- n.m.r. spectroscopy, and by other analytical data. The practical feasibility of the 6-deoxy-genation step¹³ at the trisaccharide level is noteworthy, and the consequent availability of the 4,6-protecting group on the amino sugar considerably simplified the synthetic strategy.

REFERENCES

- 1 D. Horton, G. Rodemeyer, and T. H. Haskell, Carbohydr. Res., 55 (1977) 35-47.
- 2 D. Horton, D. A. Riley, S. Samreth, and M. G. Schweitzer, Abstr. Pap. Int. Glycoconjugate Symp., Tokyo, Sept. 20-25, 1981, pp. 53-54.
- 3 H. J. Jennings, Adv. Carbohydr. Chem. Biochem., 41, in press.
- 4 D. Horton, D. A. Riley, and P. M. T. Hansen, Biopolymers, 19 (1980) 1801-1814.

- 5 E. C. Gotschlich, T. Y. Liu, and M. S. Artenstein, J. Exp. Med., 129 (1969) 1349-1365.
- 6 R. C. Seid and J. C. Sadoff, J. Biol. Chem., 256 (1981) 7305-7310; H. J. Jorbeck,
 S. B. Svenson, and A. A. Lindberg, Infect. Immun., 32 (1981) 497-502.
- 7 R. U. Lemieux, D. R. Bundle, and D. A. Baker, J. Am. Chem. Soc., 97 (1975) 4076-4083.
- 8 D. Horton, Org. Synth., 46 (1966) 1-5.
- 9 B. Weissman, J. Org. Chem., 35 (1970) 1690-1691.
- 10 R. R. King and C. T. Bishop, Can. J. Chem., 52 (1974) 3913-3917.
- 11 C. Laffite, A.-M. Nguyen Phuoc Du, F. Winternitz, R. Wylde, and F. Pratviel-Sosa, *Carbohydr.* Res., 67 (1978) 91-103.
- 12 D. R. Bundle and A. Josephson, Can. J. Chem., 57 (1979) 662-668.
- 13 S. Hanessian and N. R. Plessas, J. Org. Chem., 34 (1969) 1035-1045.