Spectral Assignments and Reference Data

A ¹⁵N NMR investigation of a series of benzotriazinones and related antitumour heterocycles

Keith Vaughan,¹* Derry E. V. Wilman,² Richard T. Wheelhouse² and Malcolm F. G. Stevens³

 ¹ Saint Mary's University, Halifax, Nova Scotia, Canada B3H 3C3
² Institute of Cancer Research, CRC Laboratory, 15 Cotswold Road, Belmont, Sutton, Surrey, SM2 5NG, UK

³ Cancer Research Laboratories, Department of Pharmaceutical Sciences, University of Nottingham, Nottingham NG7 2RD, UK

Received 12 July 2001; revised 6 November 2001; accepted 6 November 2001

A series of 3-substituted 1,2,3-benzotriazin-4-ones, 1 and 2, were synthesized by standard methods and the $^{15}\mathrm{N}$ NMR spectra were recorded. All spectra were obtained using the natural abundance of the nitrogen-15 isotope. The chemical shifts appear in the normal range for N-1, N-2 and N-3 of the triazine ring, and also correlate with the chemical shifts in the spectra of the imidazolotriazinone, 4, and the imidazolotetrazinone, 5. Significantly, the spectra of 1a, 2 and 4, recorded with full NOE, show inversion of the singlet assigned to N-3, demonstrating that these compounds exist in the tautomeric form shown. The structure of the 4-iminobenzotriazinone (3) was confirmed by this ^{15}N NMR analysis. The spectrum shows a signal for the NH-bearing imino-nitrogen atom, which is an inverted singlet in the NOE spectrum, whereas the signal from the N-3 atom of 3 is not inverted in the NOE spectrum. Copyright © 2002 John Wiley & Sons, Ltd.

KEYWORDS: NMR; ¹⁵N NMR; benzotriazinones; antitumour drugs

INTRODUCTION

The antitumour drug temozolomide (5) (Scheme 1) is of considerable current interest and is now used widely for the treatment of malignant glioma.¹ A previous study² reported a multinuclear NMR study of temozolomide and the related drug mitozolomide. The

*Correspondence to: K. Vaughan, Saint Mary's University, Halifax, Nova Scotia, Canada B3H 3C3. E-mail: keith.vaughan@stmarys.ca

Contract/grant sponsor: Natural Sciences and Engineering Research Council of Canada (NSERC).

Contract/grant sponsor: Saint Mary's University Senate Research Fund.



synthesis of ¹⁵N isotopically labelled temozolomide allowed the assignment of all the ¹⁵N resonances of the compound. In the present work, we recorded the ¹⁵N NMR spectra of a series of 1,2,3-benzotriazinones (**1a**–**g** and **2**), the imino-1,2,3-benzotriazine (**3**) and 2-azahypoxanthine (**4**), and assigned the chemical shifts to the respective nitrogen atoms by comparison with the chemical shifts observed for temozolomide (**5**).

EXPERIMENTAL

Sample preparation

1,2,3,-Benzotriazin-4(3H)-one (1a)

This was prepared by diazotization of anthranilamide (6 X = R = H) according to the method of Heller³ (Scheme 2).

6-Chloro-1,2,3-benzotriazin-4(3H)-one (2)

6-Chloroanthranilamide (6, X = Cl, R = H) (Aldrich Chemical) was diazotized in 2 M hydrochloric acid with sodium nitrite and neutralized to afford 6-chloro-1,2,3-benzotriazin-4(3*H*)-one (2).

3-Methyl-1,2,3-benzotriazin-4-one (1b) and related compounds

Methyl anthranilate (9) was diazotized and the resulting diazonium salt coupled with methylamine according to the method of LeBlanc and Vaughan,⁴ to afford the 1-aryl-3-methyltriazene (10 R = Me) (Scheme 3), which was cyclized by heating in ethanolic solution to afford the triazinone (1b).

The same method was used to prepare 3-(2-chloroethyl-)1,2,3benzotriazin-4-one (**1d**), 3-benzyl-1,2,3-benzotriazin-4-one (**1c**)⁴ and 3-carbamoylmethyl-1,2,3-benzotriazin-4-one (**1e**). Full details of the characterization of **1e** are reported elsewhere.⁵



Scheme 2







Spectral Assignments and Reference Data

3-Hydroxy-1,2,3-benzotriazin-4-one (1f) and the 3-benzoyloxy	
compound (1g)	

Methyl anthranilate was treated with hydroxylamine in the presence of sodium hydroxide to afford the o-aminobenzhydroxamic acid (6 X = H, R = OH), which was diazotized in hydrochloric acid with sodium nitrite according to the method of Ahern et al.6 to afford 3-hydroxy-1,2,3-benzotriazin-4-one (1f).

Reaction of 1f with benzoyl chloride in pyridine afforded 3benzoyloxy-1,2,3-benzotriazin-4-one (1g).6

3-(Carbamoylmethyl)-4-imino-1,2,3-benzotriazine (3) (Scheme 2)

Anthranilonitrile (7) was diazotized in hydrochloric acid with sodium nitrite to afford the diazonium salt, which was coupled with glycinamide. The resulting intermediate triazene cyclized spontaneously to give the 4-iminotriazine (3). Full details of the characterization of 3 are described elsewhere.

2-Azahypoxanthine (4) (Scheme 2)

5-Aminoimidazole-4-carboxamide (8) (0.50 g) was dissolved in 1 M hydrochloric acid (6.0 ml), cooled to 0 °C and then added slowly to a well-stirred ice-cold solution of sodium nitrite (2.0 g) in water (15 ml). The mixture was stirred for 10 min and then treated with concentrated ammonia to pH 10-11. The resulting clear brown solution was stirred for a few minutes until precipitation was complete. The product was filtered and dried to a fford 2-azahypoxanthine (4) (0.31 g, 57%), m.p.>250 °C.

Temozolomide (5)

This was obtained from Aston Molecules (Birmingham, UK).

Spectroscopy techniques

¹⁵N NMR spectra were recorded in DMSO solution on a Bruker AC250 spectrometer (at the Institute of Cancer Research, Sutton) at 25.36 MHz using a multinuclear 10 mm probe. Solutions were prepared by dissolving 1 mmol of each compound in dimethyl sulphoxide (2.4 ml) containing 30% (w/w) DMSO-d₆ with 30 mg of chromium acetylacetonate. A concentric capillary tube containing nitromethane was used as external reference. Spectra were recorded at 305 K. All spectra were obtained with the natural abundance of the nitrogen-15 isotope; no isotope enrichment was necessary to allow unequivocal assignment of all signals.

The microprogram INVGATE.AU (standard Bruker software) was used to acquire NOE-suppressed proton-decoupled ¹⁵N spectra by inverse-gated heteronuclear decoupling. Confirmation of proton attachment was obtained by heteronuclear gated decoupling with the GATEDEC.AU microprogram which provided ¹H coupled spectra with full NOE. The parameters used were pulse width $14 \,\mu s \, (45^\circ)$, decoupler power 18 H and 90° pulse for composite pulse decoupling 163 µs. The spectral width was 20 kHz and with 16 K data points gave an acquisition time of 0.41 s and a digital resolution of 2.44 Hz per point. Typically, a total of 32 000 scans were acquired for unlabelled compounds; the FID was exponentially multiplied (line broadening 5 Hz) before Fourier transformation.

Compound	N-1 ^a	N-2	N-3	Other
1a	-16.5	29.3	-152.5 ^b	_
1b	-16.0	32.8	-153.8	_
1c	-13.5	32.4	-143.8	_
1d	-14.0	31.8	-150.3	
1e	-14.7	33.1	-151.3	-274.6 ^c
1f	-23.8	24.7	-113.4	
1g	-17.1	25.2	-104.1	
2	-18.8	30.0	-152.5 ^d	
4	-35.3	22.4	-142.0	e

^aSee Scheme 1 for atom notation.

^b-152.6 ppm, inverted singlet when ¹H coupled with full NOE.

^c-274.7 ppm, inverted triplet when ¹H coupled with full NOE.

^d-152.5 ppm, inverted doublet when ¹H coupled with full NOE.

^eN atoms of imidazole ring not observed.

DISCUSSION

Table 1 gives the ¹⁵N chemical shifts of the benzotriazinones (1a-g and **2**) and of azahypoxanthine (**4**). The chemical shift of N-1 is in the range -13 to -19 ppm for all of the triazinones, except for **1**f at -23.8, compared with the chemical shift of ca -35 ppm in the imidazolo-fused compounds (4 and 5). The chemical shift of N-2 is in the range 20–33 ppm for all compounds and N-3 is in the range -142to -153 ppm for most compounds of the series, with the exception of temozolomide, with an upfield shift to -180 ppm, and the N-oxy compounds (1f and 1g), which show a downfield shift to -104 to -113 ppm. Compounds 1e, 3 and 5 exhibit a signal a ca -275 ppm arising from the amide N-atom; this signal has been unambiguously assigned to the amide nitrogen by full ¹H NOE, which shows the signal as an inverted triplet. Compound 5 shows the expected signals for the imidazole nitrogens at -105.6 and -199.8 ppm. Surprisingly, the N-atoms of the imidazole ring in azahypoxanthine (4) are not observed, possibly owing to a combination of signal broadening and the effect of exchange of the hydrogen atom between both nitrogen sites of the imidazole ring.

When the spectra of **1a**, **2** and **4** are recorded with ¹H coupled NOE, the signal from N-3 is inverted in each case, showing that a hydrogen atom is bonded to N-3. This observation confirms that these molecules do indeed exist in the tautomeric form shown (e.g. 1a), rather than the alternative tautomer with the hydrogen atom located at N-1 as in structure 11 (Scheme 3). The spectra of 2, with and without NOE, are shown in Fig. 1 to illustrate these observations. The appearance of a doublet for the N-3 nitrogen in the spectrum of 2 with NOE [Fig. 1(b)] allows the measurement of the coupling constant ${}^{1}J(N,H)$, which is found to be 99 ± 5 Hz in this case. This value is very close to previously reported ¹*J*(N,H) coupling constants.2

The value of these ¹⁵N chemical shift assignments is evident in the analysis of the ¹⁵N NMR spectrum of 3-(carbamoylmethyl)-4imino-1,2,3-benzotriazine (3). In an independent study,⁵ a series of 1-aryl-3-carbamoylmethyltriazenes, ArN=N—NH—CH₂CONH₂, were prepared and characterized. As part of this study, the diazotization of anthranilonitrile (7) was carried out and the resulting diazonium salt was coupled with glycinamide, NH₂CH₂CONH₂, to afford an unstable triazene, which undergoes spontaneous cyclization. The cyclization product was assigned structure **3** on the basis of IR and ¹H and ¹³C NMR spectroscopic evidence, but the structure was not unequivocal. The ¹⁵N NMR spectrum of **3** removed the ambiguity from the structural assignment; the ¹⁵N chemical shifts of 3 are shown in Table 2. Chemical shifts for N-1, N-2 and N-3 are all consistent with the previous observations, and the amide nitrogen at -275.05 ppm shows the appropriate inversion with NOE. The crucial evidence is the signal at -177 ppm, which





Spectral Assignments and Reference Data

Figure 1. 15 N NMR spectra of 6-chloro-1,2,3-benzotriazin-4(3*H*)-one (2) in DMSO, (a) without NOE and (b) with NOE.

Table 2.	¹⁵ N chemical shift data (δ , ppm) for			
3-carbamoylmethyl-4-imino- 1,2,3-				
benzotriazinone (3), with NOE				

Atom	δ
N-1	-29.0
N-2	31.0
N-3	-168.4
Amide NH ₂	-275.2
	(inverted triplet)
Imino NH	-177.8
	(inverted broad doublet)

is assigned to the imino-nitrogen atom at position 4 in the triazine ring. This nitrogen atom is shown to carry a hydrogen atom by the observed inversion with NOE.

The ¹⁵N nucleus is relatively sensitive to changes in the atom or group immediately attached to the nitrogen, which can cause significant effects.⁷ The ¹⁵N chemical shift becomes more positive (i.e. shifts to lower field) with increasing electron withdrawal by attached groups. The chemical shift data reported in this paper are consistent with this general principle. For example, compare the chemical shift of N-3 in **1b** and **1f**; the electron-withdrawing effect of the oxygen atom in **1f**, compared with the methyl carbon in **1b**, causes a large downfield shift of ca 40 ppm. On the other hand, changing the substituent from a methyl group in **1b** to a chloroethyl group in **1d** causes only a marginal downfield shift of 3.5 ppm because the halogen atom is too far removed from the nitrogen to have a

Table 3.	¹⁵ N chemical shift data		
(δ , ppm) for 8-carbamoyl-3-			
methylimidazolo [5,1-d]-1,2,3,5-			
tetrazine-4-one (5)			

Atom ^a	δ
N-1	-34.0
N-2	20.0
N-3	-180.4
N-5	-199.8
N-7	-105.6
Amide N	-274.9 ^b

^aSee Scheme 1 for atom notation.

^bInverted triplet when ¹H coupled with full NOE.

significant effect. The change from *N*-methyl in **1b** to *N*-benzyl in **1c** is more significant, causing a 10 ppm downfield shift. Introducing the chlorine substituent in the benzene ring as in **2** causes almost no change in any of the ¹⁵N resonances when compared with those of **1a**; only N-1, the closest nitrogen to the halogen, is shifted marginally by 2.4 ppm.

The ¹⁵N resonances assigned to N-1, N-2 and N-3 in the benzotriazines have chemical shifts with a similar order to the chemical shifts found in the open-chain analogues, the 1-aryl-3,3-dialkyltriazenes (12) (Scheme 3), for which typical values are N-1, -13 to -35 ppm, N-2, 68–75 ppm and N-3, -170 to -220 ppm.⁸ The most important deviation in comparing the cyclic and open-chain variants is in the chemical shift of N-2, which differs by ca 40 ppm. The most likely explanation for this difference is in the differing geometry of the N-1=N-2 double bond; the cyclic benzotriazines are locked into the *cis* geometry, whereas the 1-aryl-3,3-dialkyltriazenes are known to prefer the *trans* geometry in the solid state.⁹

Acknowledgements

This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC) and by the Saint Mary's University Senate Research Fund. We are grateful to the Institute of Cancer Research, Sutton, UK for access to the Bruker AC250 NMR spectrometer, to Aston Molecules for providing a sample of temozolomide and to the Cancer Research Laboratories at Nottingham University, UK, for laboratory resources.

REFERENCES

- 1. Friedman HS, Kerby T, Calvert H. Clin. Cancer Res. 2000; 6: 2585.
- 2. Wheelhouse RT, Wilman DEV, Thompson W, Stevens MFG. J. Chem. Soc. 1995; 249.
- 3. Heller G. J. Prakt. Chem. 1925; 111: 1.
- 4. LeBlanc RJ, Vaughan K. Can. J. Chem. 1972; 50: 2544.
- 5. Hooper DL, Jollimore JV, Vaughan K. J. Org. Chem. 1996; 61: 210.
- 6. Ahern TP, Navratil T, Vaughan K. Can. J. Chem. 1977; 55: 630.
- Levy GC, Lichter RL. Nitrogen-15 Nuclear Magnetic Resonance Spectroscopy. John Wiley & Sons: New York, 1979; 28–107.
- 8. Wilman DEV. Magn. Reson. Chem. 1990; 28: 729.
- 9. Neidle S, Wilman DEV. Acta Crystallogr., Sect. B 1992; 48: 213.