

PREPARATION OF FLUORINE-MODIFIED 25-HYDROXYVITAMIN D₂ 28,28,28-TRIFLUORO-, 26,26,26, 27,27,27-HEXAFLUORO- AND 28-NOR-26,26,26,27,27,27-HEXAFLUORO-25-HYDROXYVITAMIN D₂

Takeo Taguchi,^a Ryoichi Namba,^a Masakazu Nakazawa,^a Masaharu Nakajima,^a Yumiko Nakama,^a
 Yoshiro Kobayashi,^{a,*} Noriyuki Hara,^b and Nobuo Ikekawa^b

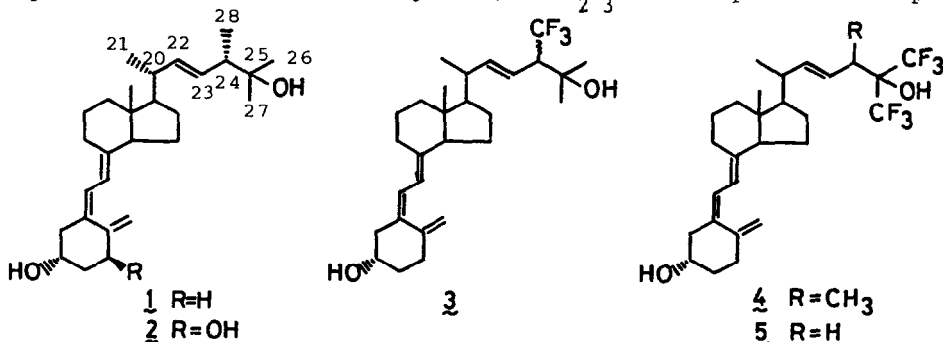
a) Tokyo College of Pharmacy, 1432-1 Horinouchi, Hachioji, Tokyo 192-03, Japan,

b) Department of Chemistry, Tokyo Institute of Technology, 2-12-1 Ookayama, Meguro-ku,
 Tokyo 152, Japan

Summary Preparations of the titled fluorinated vitamin D₂ analogs (3, 4 and 5) were efficiently achieved through reactions of the C₂₂-aldehyde (6) with phenylsulfone derivatives (7, 8 and 9).

As in the case of vitamin D₃,¹ it was demonstrated that vitamin D₂ must be metabolically hydroxylated at C-25 followed by C-1 to yield the active form, 1,25-dihydroxyvitamin D₂ (2, 1,25-(OH)₂D₂).² Hydroxylation at C-24 and C-26 of 25-OH-D₂ (1) is an alternative metabolic pathway.^{3,4} Recently, DeLuca and Ikekawa reported a unique methyl migration on the side chain of 24-epi-25-OH-D₂ to form biologically potent 22-dehydro-1,25-dihydroxy-26-homovitamin D₃ (1,25(OH)₂-Δ²²-26-homo-D₃), although the mechanism of this biotransformation is the present subject.⁵ Biological potencies of vitamin D₂ and/or its metabolites are equal to those of the corresponding vitamin D₃ forms in rat and human, but about one-tenth as active in bird.^{2,6} It is suggested that the differential biological activity of vitamin D₂ in bird is due to the presence of 24-methyl rather than an unsaturated side chain,⁷ the functional importance of 24-methyl of vitamin D₂ has still remained to be clarified.

On the basis of metabolism and mode of function of vitamin D₃, and the characteristic properties of fluorinated molecules with respect to biological response, fluorinated vitamin D₃ analogs were designed, synthesized and their biological response was investigated to clarify the physiological significance of metabolic hydroxylation.^{1d,8} The hexafluoro- and 24,24-difluoro analogs of 1,25(OH)₂D₃ showed higher activities than those of 1,25(OH)₂D₃, presumably due to the metabolic stability of these fluoro analogs.^{9,10} Moreover, these fluorinated analogs¹¹ as well as 24- or 26-homologs of 1,25(OH)₂D₃¹² were reported to show preferential

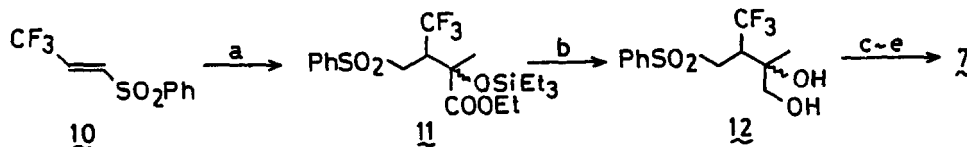


activity in inducing differentiation of human leukemia cells HL-60. These results prompted us to carry out the preparation of the fluorinated vitamin D₂ analogs replaced the hydrogens at C-28 or C-26,27 by fluorines to clarify the biological significance of methyl group at C-24 of vitamin D₂ and to investigate the importance of the modification of the side chain in such biological responses. In this paper we report the synthesis of the titled fluorinated vitamin D₂ analogs (3, 4 and 5).

Synthetic strategy for these fluorinated analogs is as follows ¹³ 1) Addition of the sulfone derivatives (7-9) to the C₂₂-aldehyde (6), and 2) reductive desulfonylation leading to trans-olefin. Since the presence of fluorine substituents in 7 or 8 caused to decrease in reactivity in the reaction with the aldehyde (6) and to facilitate retro-reaction of the β -hydroxysulfone intermediates (13, 16, 19) at the desulfonylation step, several modifications were required for these reactions.

Preparation of 28,28,28-F₃-25-OH-D₂ (3)

The phenylsulfone (7) was prepared using the trifluoropropene derivative (10)¹⁴ as follows Reaction of lithium enolate derived from the lactate with 10 gave the adduct (11), which was, in turn, reduced to the diol (12), followed by conversion to the tertiary alcohol via the epoxide, then protected the hydroxyl group as THP ether (7).



a, CH₃CH(OSiEt₃)COOEt, LDA/THF (92%); b, 1) DIBAL/Et₂O 11) 10% HCl, MeOH (98%); c, 1) MsCl, Et₃N 11) DBU/CH₂Cl₂ (82%); d, LiAlH₄/Et₂O (98%); e, DHP, p-TsOH/1,4-dioxane (74%)

Reaction of the lithio derivative of 7 with the C₂₂-aldehyde (6)^{13a,15} gave the desired adduct (13) in low yield (7%), probably due to steric effect.¹⁶ Addition of ethereal solution of MgBr₂·Et₂O prepared by reaction of 1,2-dibromoethane with Mg in ether was found quite effective. Thus, lithiation of 7 (LDA, THF, 0°C, 30 min) followed by the addition of 2.2 equiv. of ethereal solution of MgBr₂ (-78°C, 5 min), and then the aldehyde (0.7 equiv. of 6, -78°C, 1.5 h) gave 13 as an unseparable mixture of diastereomers in 89% yield. Desulfonylation of 13 with freshly prepared 3% Na-Hg¹⁷ (THF-MeOH, Na₂HPO₄, 0°C) gave the THP ether (14) in 67% yield, which was deprotected (MeOH, p-TsOH, rt) to give the trifluor-ergosterol (15, 98%).¹⁸ Photoirradiation of 15 (200 W medium-pressure Hg lamp, Vycor filter, EtOH-benzene, 5 min) followed by thermal isomerization (reflux, 1h) afforded 3 in 32% yield after chromatographic purification [3 λ_{max} (EtOH) 265, 228 nm, m/e· 466, 433, 271, 253, 136, 118, 59].

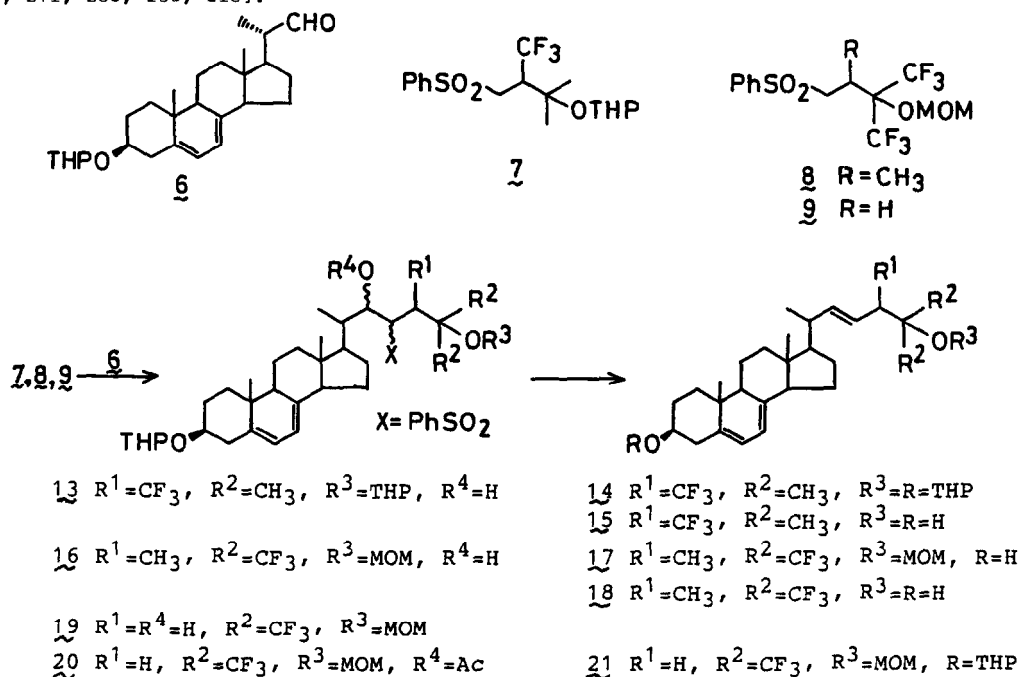
Preparation of Hexafluoro Analogs (4, 5)

For the preparation of the hexafluoro analogs (4, 5) the sulfone intermediates (8, 9) were synthesized through the reaction of lithium enolate of ethyl propionate or Reformatsky reagent from ethyl bromoacetate with hexafluoroacetone. Protection of the hydroxyl group as methoxymethyl(MOM) ether was the only one choice for these hexafluorocarinols due to the

difficulty of introducing other protecting groups such as THP or silyl group. Without protection no adduct was obtained by the reaction of the dianion with the aldehyde (6).

Reaction of the anion of **8** (LDA, -78°C , 30 min, then $\text{MgBr}_2 \cdot \text{Et}_2\text{O}$, 5 min) with **6** (THF- Et_2O , -78°C , 1h) gave the adduct (**16**, 98%) as chromatographically separable four diastereomers (**16a-d**) in a ratio, 23:28:27:21 (**16a**:**16b**:**16c**:**16d**) according to the order of elution on flash column chromatography (hexane-AcOEt). Desulfonylation of each isomer by freshly prepared Na-Hg afforded the trans-olefinic compound in each case (35-71% yield). Removal of THP group (MeOH-p-TsOH, rt, **17**, 93-96%) and then MOM group (MeOH- CH_2Cl_2 , p-TsOH, reflux) gave 25-hydroxy-ergosterol derivatives (**18**). On comparing the nmr spectrum and chromatographic behavior of each isomer corresponding to **17** and **18**, two (**16a**, **16d**) of the four isomers of **16** were converted to one of **18** (**18a**) and the other two (**16b**, **16c**) to the other isomer (**18b**).¹⁹ As with the conversion of **15** to **3**, each isomer of **18** was converted to the vitamin D_2 form (**4a** from **18a** and **4b** from **18b**, respectively).²⁰

Preparation of the 28-nor-hexafluoro analog (**5**) was also carried out.²¹ The absence of a branched methyl group in **9** facilitated the reaction with **6**. Thus, reaction of the lithio derivative of **9** with **6** (THF, -78°C , 1.5h) gave the adduct (**19**) in 82% yield. Acetylation of **19** proceeded smoothly to give **20** (Ac_2O , Py, DMAP, 90%) and subsequent desulfonylation with Na-Hg gave the trans-olefinic compound [**21**, (CDCl_3) δ : 5.50(dd, $J=15.2$ and 8.7 Hz, 22-H)] in 73% yield. Deprotection of the THP and MOM groups and subsequent conversion to the vitamin D_2 form (**5**) were carried out by the similar method as above [**5** λ_{max} (EtOH) 264, 228 nm; m/e 506, 473, 271, 253, 136, 118].



References and Notes

1. a) H. F. DeLuca and H. K. Schnoes, *Annu. Rev. Biochem.*, **45**, 631(1976), b) A. W. Norman, J. Roth, and L. Orci, *Endocrine Rev.*, **3**, 331(1982), c) R. Brommage and H. F. DeLuca,

- Endocrine Rev., 6, 491(1985), d) N. Ikekawa, Medicinal Res. Rev., 7, 333(1987).
2. G. Jones, H. K. Schnoes, and H. F. DeLuca, Biochemistry, 14, 1250(1975).
 3. T. Suda, H. F. DeLuca, H. K. Schnoes and J. W. Blunt, Biochemistry, 8, 3515(1969),
 4. A. Hay and G. Jones, Clin. Chem., 25, 473(1979).
 5. a) Y. Tanaka, R. R. Sicinski, H. F. DeLuca, H. Sai, and N. Ikekawa, Biochemistry, 25, 5512 (1986), b) S. Sai, S. Takatsuto, N. Ikekawa, Y. Tanaka, and H. F. DeLuca, Chem. Pharm. Bull., 34, 4508(1986).
 6. D. Drescher, H. F. DeLuca and H. H. Imrie, Arch. Biochem. Biophys., 130, 657(1969).
 7. H. F. DeLuca, M. Weller, J. W. Blunt, P. F. Neville, *ibid.*, 124, 122(1968).
 8. Y. Kobayashi and T. Taguchi, J. Synth. Org. Chem., 43, 1073(1985).
 9. S. Okamoto, Y. Tanaka, H. F. DeLuca, Y. Kobayashi, and N. Ikekawa, Am. J. Physiol., 224, E159(1983).
 10. Y. Tanaka, H. F. DeLuca, Y. Kobayashi, and N. Ikekawa, Arch. Biochem. Biophys., 229, 348 (1984).
 11. H. P. Koeffler, T. Amatruda, N. Ikekawa, Y. Kobayashi, and H. F. DeLuca, Cancer Res., 44, 5624(1985).
 12. V. K. Ostrem, Y. Tanaka, J. Prah, H. F. DeLuca, and N. Ikekawa, Proc. Natl. Acad. Sci. USA, 84, 2610(1987).
 13. a) S. Yamada, M. Shiraishi, M. Ohmori, and H. Takayama, Tetrahedron Lett., 25, 3347(1984), b) J. W. Morzycki, H. K. Schnoes, and H. F. DeLuca, J. Org. Chem., 49, 2148(1984).
 14. T. Taguchi, G. Tomizawa, M. Nakajima, and Y. Kobayashi, Chem. Pharm. Bull., 33, 4077(1985); T. Taguchi, A. Hosoda, G. Tomizawa, A. Kawara, T. Masuo, Y. Suda, M. Nakajima, and Y. Kobayashi, *ibid.*, 35, 909(1987).
 15. D. H. R. Barton, T. Shioiri, and D. A. Widdowson, J. Chem. Soc.(C), 1968(1971).
 16. Similar reaction of lithio derivative of 2 with benzaldehyde and cyclohexanecarboxaldehyde gave the adduct in 56% and 74% yield, respectively.
 17. Reagents for Organic Synthesis, ed. by L. F. Fieser and M. Fieser, John Wiley and Sons, Inc., New York, Vol 1(1967). p1030.
 18. On the basis of ^1H -nmr(400 MHz) and ^{19}F -nmr(188 MHz) spectrum, 15 may be a 1:1 mixture of stereoisomers at C-24. ^1H -nmr(CDCl_3) δ : 0.638, 0.644(3H, both s, 18- H_3), 2.720, 2.726(1H, both quintet, $J=9.7$ Hz, 24-H); ^{19}F -nmr(CDCl_3 , relative to benzotrifluoride) -0.87 ppm(d, $J=9.7$ Hz), -1.04 ppm(d, $J=9.7$ Hz).
 19. Although the ^1H -nmr spectrum of 18a was quite similar to that of 18b, the chemical shifts of two unequivalent trifluoromethyl groups in 18a clearly differed from those of 18b. 18a ^1H -nmr(CDCl_3) δ 5.59(dd, $J=15.2$, 9.0 Hz, 22-H); ^{19}F -nmr(CDCl_3) -8.78 ppm(q, $J=9$ Hz), -9.75 ppm(q, $J=9$ Hz), 18b: ^1H -nmr(CDCl_3) δ 5.60(dd, $J=15.4$, 8.7 Hz); ^{19}F -nmr(CDCl_3) -8.56 ppm(q, $J=9$ Hz), -9.82 ppm(q, $J=9$ Hz).
 20. Each isomer of 4 showed a retention time on HPLC (Zorbax Sil column, 4.6x25 cm; 5% 1-PrOH in hexane, flow rate 2 ml/min) different from that of the other; 5.1 min for one isomer(4a) and 5.4 min for another one(4b). Although both compounds showed biological activity in calcium mobilization from bone in rat, 4a was more active than 4b. These results as well as the nmr data indicate 4b to possibly be an epimer at C-24 of 4a.
 21. 1 α -Hydroxy derivative of 5 was reported by Hoffmann-La Roche group. E. Baggiolini, G. Pizzolato, M. Uskokovic, and G. Truitt, Eur. Pat. Appl. EP 182,298(1986).

(Received in Japan 17 September 1987)